#### AN ABSTRACT OF THE THESIS OF

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Studies were undertaken of the adsorption of chlorinated phenols from aqueous solution on granular activated carbon (Filtrasorb-400, 30x40 mesh). Single-component equilibrium adsorption data on the eight compounds in two concentration ranges at pH 7.0 fit the Langmuir equation better than the Freundlich equation. The adsorptive capacities at pH 7.0 increase from pentachlorophenol to trichlorophenols and are fairly constant from trichlorophenols to monochlorophenols. Equilibrium studies were performed at various temperatures at pH 7.0. The adsorption process was found to be exothermic for pentachlorophenol and 2,4,6-trichlorophenol, and endothermic for 2,4-dichlorophenol and 4-chlorophenol.

Equilibrium measurements were also conducted for 2,4,5-trichlorophenol, 2,4dichlorophenol, and 4-chlorophenol over a wide pH range. A surface complexation model was proposed to describe the effect of pH on adsorption equilibria of chlorophenols on activated carbon. Activated carbon surface functional sites are divided into acidic groups and basic groups with which molecular and ionized forms of chlorophenols interact, respectively, to form two neutral surface complexes, of which the complex of basic groups is more substantial and more stable than that of acidic groups. The simulations of the model are in excellent agreement with the experimental data.

Batch kinetics studies were conducted of the adsorption of chlorinated phenols on granular activated carbon. The external film diffusion model, linear-driving-force approximation, and surface reaction kinetics model were employed to fit the adsorption kinetics data of chlorophenols. The results show that the surface reaction model best describes both the short-term and long-term kinetics, while the external film diffusion model describes the short-term kinetics data very well and the linear-driving-force approximation improved its performance for the long-term kinetics. The mass transfer coefficient was found to increase from more chlorinated compounds to less chlorinated compounds. Two-component adsorption kinetics experiments revealed that the adsorption of chlorophenols on activated carbon is to some extent an irreversible process and non-ideal competition between two components exists.

Multicomponent adsorption equilibria of chlorophenols on granular activated carbon was investigated in the micromolar equilibrium concentration range. The Langmuir competitive and Ideal Adsorbed Solution (IAS) models were tested for their performances on the three binary systems of pentachlorophenol/2,4,6-trichlorophenol, 2,4,6-trichlorophenol/2,4-dichlorophenol/4-chlorophenol, and the tertiary system of 2,4,6-trichlorophenol/2,4-dichlorophenol/4-chlorophenol, and found to fail to predict the two-component adsorption equilibria of the former two binary systems and the tertiary system. A new prediction method and a modification of the IAS model, both based on thermodynamic considerations, were proposed. The required parameters are the single-component Langmuir isotherm constants and initial concentrations for each component. The proposed new method and the modification of the IAS model were found to significantly improve the accuracy of the predictions of two-component adsorption equilibria of chlorophenols. This new method also performs much better than the Langmuir competitive and IAS models in the three-component system.

Studies were also conducted on the desorption equilibria and kinetics of chlorophenols. The results indicate that the Langmuir equation fits both adsorption and desorption equilibrium data better than the Freundlich equation and the degree of the irreversibility of adsorption increases from 4-chlorophenol to pentachlorophenol. A linear-driving-force desorption rate equation was proposed, which describes the desorption kinetic data very well. The desorption mass transfer coefficients were found to increase from pentachlorophenol to 4-chlorophenol.

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## Adsorption of Chlorophenols on Granular Activated Carbon

#### Chapter 1 INTRODUCTION

Chlorinated phenols are of environmental concern due to anthropogenic inputs from industrial wastes, degradation of chlorinated pesticides, and use of pentachlorophenol as a wood preservative. Chlorophenols in the nanomolar to micromolar concentration range also form during municipal water purification *via* the phenol-chlorine reaction. Chlorophenol removal is necessary because chlorophenols are odor-producing and some have been found to be carcinogenic. 2-Chlorophenol, 2,4-dichlorophenol, and 2,6dichlorophenol are the major odor-causing species and produce detectable odor at the 2 and 3  $\mu$ g/l levels.

Adsorption is a fundamental process for separating inorganic and organic contaminants from waters and wastewaters. Activated carbon remains the principal adsorbent in full-scale treatment. At present, the applications of activated carbon adsorption in water treatment in the United States are predominantly traditional taste and odor control. However, activated carbon adsorption is increasingly being considered for removal of synthetic organic chemicals, color-forming organics, and disinfection byproducts and their naturally occurring precursors. It has been found that the chlorophenols can be strongly adsorbed by activated carbon and will not break through conventional granular activated carbon (GAC) columns for years (Montgomery, 1985). It is expected that the application of activated carbon to control contamination of drinking water by toxic or carcinogenic compounds at low concentrations will increase.

The recent concerns with the removal of chlorinated phenols have focused attention on the biodegradation of these compounds on activated carbon (Speitel et al., 1989). The highly chlorinated phenols, i.e., tetra- and pentachlorophenol, can be degraded anaerobically and various lower chlorinated phenols, i.e., tri-, di- and monochlorophenol, will be formed as their metabolites (Nicholson et al., 1990). Biodegradation and adsorption will occur simultaneously in these systems. Therefore, it is of significance to investigate the adsorption behavior of various chlorophenols on granular activated carbon.

Both kinetics and thermodynamics or equilibrium are important to the process of activated carbon adsorption. The chlorophenols can partially or totally ionize in aqueous solution and pH will play an important role in activated carbon adsorption. The objectives of this thesis are:

(1) to investigate equilibrium characteristics of activated carbon adsorption of chlorinated phenols and to determine the isotherm parameters for single-component systems;

(2) to develop and verify a surface complexation model for describing the effect of pH on equilibrium adsorption of organic electrolytes on activated carbon;

(3) to investigate competitive adsorption equilibrium characteristics of chlorophenols in multicomponent systems;

(4) to develop and verify a non-ideal competitive model for predicting

multicomponent adsorption equilibria of chlorophenols;

(5) to modify the Ideal Adsorbed Solution (IAS) model and to verify the modified IAS model for predicting multicomponent adsorption equilibria of chlorophenols;

(6) to investigate the adsorption kinetics of chlorophenols and to determine which kinetic model can best describe the kinetic processes of chlorophenol adsorption;

(7) to investigate the desorption equilibrium and kinetics of chlorophenols on activated carbon;

(8) to investigate the influence of pH on adsorption equilibrium and kinetics of chlorophenols;

(9) to investigate the influence of temperature on adsorption equilibrium of chlorophenols.

#### Chapter 2 BACKGROUND

#### Model Isotherms for Single-Component Adsorption

In this context, single-component actually means one contaminant component in aqueous solution. The solvent (water) is simply assumed to be inert, the adsorption of the contaminant component is assumed to be unaffected by the water, and therefore, the system is treated as a single-component system.

To represent the equilibrium relation for single-component adsorption, a number of model isotherms reported in the literature will be reviewed.

Langmuir Isotherm. Langmuir adsorption isotherm describes equilibrium between surface and solution as a reversible chemical equilibrium between species (Langmuir, 1918). The adsorbent surface is considered to be made up of fixed individual sites where molecules of adsorbate may be chemically bound. Denote an unoccupied surface site as -S and the adsorbate in dilute solution as species A, with concentration C, and consider the reaction between the two to form occupied sites (-SA):

$$-S + A \neq -SA \tag{2-1}$$

Assume that this reaction has a fixed free energy of adsorption equal to  $\Delta G_a^{\ o}$  that is not dependent on the extent of adsorption and not affected by interaction among sites. Each site is assumed to be capable of binding at most one molecule of adsorbate; the Langmuir

model allows accumulation only up to a monolayer.

If Q is the maximum number of moles adsorbed per mass adsorbent when the surface sites are saturated with adsorbate (i.e., a full monolayer), and q is the number of moles of adsorbate per mass adsorbent at equilibrium. According to the law of mass action, we have

$$b = \frac{[-SA]}{[-S] [A]} = \frac{q}{(Q-q)C}$$
(2-2)

where  $b = exp(-\Delta G_a^{0}/RT)$ , an equilibrium constant and C is equilibrium concentration in solution. The rearrangement of equation 2-2 leads to

$$q = \frac{QbC}{1+bC}$$
(2-3)

Correspondence of experimental data to the Langmuir equation does not mean that the stated assumptions are valid for the particular system being studied, because departure from the assumptions can have a canceling effect. An advantage of this model is that it can approach Henry's law at low concentrations. The constants in the Langmuir equation can be determined by plotting C/q versus C and making use of equation 2-3 rewritten as

$$\frac{C}{q} = \frac{1}{Qb} + \frac{C}{Q} \tag{3-4}$$

**BET** Isotherm. The BET adsorption isotherm (Brunauer et al., 1938) extends the Langmuir model from a monolayer to several molecular layers. This isotherm has the form

$$\frac{q}{Q} = \frac{BC}{(C_s - C) \left[1 + (B - 1)\frac{C}{C_s}\right]}$$
(2-5)

Where B is a dimensionless constant related to the difference in free energy between adsorbate on the first and successive layers and  $C_s$  is the saturation concentration of the adsorbate in solution.

*Freundlich Isotherm*. The Langmuir and BET models incorporate an assumption that the energy of adsorption is the same for all surface sites and not dependent on degree of coverage. In reality, the energy of adsorption may vary because real surfaces are heterogeneous. The Freundlich adsorption isotherm (Freundlich, 1926) attempts to account for this. The Fruendlich isotherm has the form

$$\boldsymbol{q} = K C^n \tag{2-6}$$

q and C are the equilibrium solid- and solution-phase concentrations of solute; K and n are empirical constants, which can be evaluated by plotting log q versus log C. A potential disadvantage of this model is that it does not approach Henry's law at low concentrations.

Langmuir-Freundlich Isotherm. Sips (1948) modified the Langmuir isotherm by the introduction of a power law expression of Freundlich form:

$$q = \frac{QbC^n}{1 + bC^n} \tag{2-7}$$

which reduces to the Freundlich type for low concentration and exhibits saturation for high concentration. This isotherm contains three parameters.

Toth Isotherm. Toth (Toth, 1971; Jossens and Prausnitz, 1978) has considered only adsorption of gases but his ideas can be extended to adsorption of solutes from dilute aqueous solution. Toth isotherm has the form

$$q = \frac{QC}{(b + C^{M})^{1/M}}$$
(2-8)

which contains three parameters. Toth equation reduces to Henry's law at very low concentrations and exhibits saturation at high concentrations.

#### Prediction Methods for Multicomponent Adsorption Equilibria

Polluted water usually contains more than one contaminant. Multicomponent adsorption involves competition to occupy the limited adsorbent surface available and the interactions between different adsorbates. A number of methods have been developed that aim at predicting multicomponent adsorption equilibria using data from single-component adsorption isotherms. For simple systems considerable successes have been achieved but there is still no established method with universal proven applicability, and this problem remains one of the more challenging obstacles to the development of improved methods of process design (Ruthven, 1984).

*Multicomponent Langmuir Isotherm*. A common model for predicting adsorption equilibria in multicomponent systems is the Langmuir model for competitive adsorption, which was first developed by Butler and Ockrent (1930). This model is based on the same assumptions as the Langmuir model for single adsorbates. Assuming, as did Langmuir, that the rate of adsorption of a species at equilibrium is equal to its rate of desorption, the equation

$$Q_{i} = \frac{Q_{i}b_{i}C_{i}}{1 + \sum_{i=1}^{n}b_{i}C_{i}}$$
(2-9)

was developed, in which  $Q_i$  and  $b_i$  are the Langmuir constants determined from the singlesolute adsorption isotherm of species *i*. Because of its mathematical simplicity, this model is widely used (Hseih et al., 1977; Crittenden et al., 1980; Murin and Snoeyink, 1979).

Broughton (1948) observed that the extension of the Langmuir theory to adsorption from binary adsorbate systems is thermodynamically consistent only in the special case where  $Q_1 = Q_2$ . However, Young and Crowell (1962) have pointed out that thermodynamic consistency is of secondary importance if the equation provides the correct analytical description of the adsorption phenomena.

Modified Multicomponent Langmuir Isotherm. Jain and Snoeyink (1973) have noted

that if the Langmuir model for competitive adsorption satisfactorily predicts the extent of adsorption from a bisolute system when  $Q_1 \neq Q_2$ , it is probably because there is competition for all available sites. They have proposed a model which can be used to predict the extent of adsorption of each species from a bisolute solution if a portion of the adsorption occurs without competition. The model was based on the hypothesis that adsorption without competition occurs when  $Q_1 \neq Q_2$ . Further, it was assumed that the number of sites for which there was no competition was equal to the quantity  $(Q_1 - Q_2)$ , where  $Q_1 > Q_2$ . On this basis, the following equations were proposed:

$$q_{1} = \frac{(Q_{1} - Q_{2})b_{1}C_{1}}{1 + b_{1}C_{1}} + \frac{Q_{2}b_{1}C_{1}}{1 + b_{1}C_{1} + b_{2}C_{2}}$$
(2-10)

$$q_2 = \frac{Q_2 b_2 C_2}{1 + b_1 C_1 + b_2 C_2}$$
(2-11)

The first term on the right side of equation 2-10 is the Langmuir expression for the number of moles of species 1 that adsorb without competition on the surface area proportional to  $(Q_1 - Q_2)$ . The second term represents the number of moles of species 1 adsorbed on the surface area proportional to  $Q_2$  under competition with species 2 and is based on the Langmuir model for competitive adsorption. The number of moles of species 2 adsorbed on surface area proportional to  $Q_2$  under competition with species 1 can be calculated from equation 2-11. *Multicomponent Langmuir-Freundlich Isotherm*. The Sips equation (eqn 2-7) may be easily extended to binary or multicomponent systems (Ruthven 1984; Yu and Neretnieks, 1990). The resulting expression for the isotherm is

$$q_{i} = \frac{Q_{i}b_{i}C_{i}^{n_{i}}}{1 + \sum b_{i}C_{i}^{n_{i}}}$$
(2-12)

The simple formula makes this method very attractive. Although not thermodynamically consistent, this expression has been shown to provide a reasonably good empirical correlation of binary equilibrium data for a number of simple gases on molecular sieve adsorbents (Yon and Turnock, 1971) and is widely used for design purposes (Maurer, 1980). However, because of the lack of a proper theoretical foundation this approach should be treated with caution.

Ideal Adsorbed Solution (IAS) Model. The most common model for describing adsorption equilibrium in multicomponent systems is the Ideal Adsorbed Solution (IAS) model, which was developed by Radke and Prausnitz (Radke and Prausnitz, 1972). This model relies upon the assumption that the adsorbed phase forms an ideal solution and hence the name Ideal Adsorbed Solution (IAS) model has been adopted. The following part should only be a summary of the main equations and assumptions of this theory.

Equation 2-13 used within this theory relates the concentration of solute *i* in the mixture,  $C_i$ , to a corresponding concentration of this solute in an single solute system,  $C_i^{0}$ :

$$C_i(\Pi, T, Z_i) = Z_i C_i^0(\Pi, T)$$
 (2-13)

Here,  $Z_i$  represents the mole fraction of surface coverage by component *i*,  $\pi$ , the spreading pressure on the surface and *T*, the absolute temperature. The spreading pressure defines the lowering of surface tension at the adsorbate-solution interface:

$$\mathbf{\Pi} = \mathbf{\gamma}^{\mathbf{0}} - \mathbf{\gamma} \tag{2-14}$$

 $\gamma^0$  is the surface tension of the pure solvent (water) and  $\gamma$  the surface tension created by the mixture of solvent and solutes.

Equation 2-13 holds only when  $\pi$  and T in the mixture are the same as those in the respective single-solute systems.

Spreading pressure can be related to the characteristic adsorption equilibria of each single solute system according to:

$$\Pi_{i} = \frac{RT}{A} \int_{0}^{c_{i}^{0}} q_{i}^{0} \frac{dC_{i}^{0}}{C_{i}^{0}}$$
(2-15)

In equation 2-15, R is the universal gas constant, A, the surface area per unit weight,  $C_i^o$ , the liquid-phase concentration of species *i* in single-solute systems which gives the same spreading pressure as that of the mixture and  $q_i^o$ , the solid-phase loading corresponding to  $C_i^o$ .

Equivalence of the spreading pressures of all the solutes in the mixture gives

$$\int_{0}^{C_{1}^{0}} q_{1}^{0} \frac{dC_{1}^{0}}{C_{1}^{0}} = \int_{0}^{C_{2}^{0}} q_{2}^{0} \frac{dC_{2}^{0}}{C_{2}^{0}} = \int_{0}^{C_{3}^{0}} q_{3}^{0} \frac{dC_{3}^{0}}{C_{0}^{0}} = \dots$$
(2-16)

$$q_{i}^{0} = f_{i}(C_{i}^{0}) \tag{2-17}$$

Combining the IAS theory with the Gibbs equation for isothermal adsorption gives the relationship necessary for equilibrium calculations:

$$\frac{1}{q_{T}} = \sum_{i}^{n} \frac{Z_{i}}{q_{i}^{0}}$$
(2-18)

#### Other two equations required for IAS model calculations are

$$\sum_{i}^{n} Z_{i} = 1$$
 (2-19)

$$q_i = Z_i q_T \tag{2-20}$$

Equations 2-13, 2-16, 2-17, 2-18, 2-19 and 2-20 constitute a set of simultaneous equations from which the IAS model calculation can be made.

This model has received widespread use in multisolute adsorption research for a variety of reasons (Jossens and Prausnitz, 1978; Yu and Neretnieks, 1990; Yen and Singer, 1984; Crittenden et al., 1985a; Crittenden et al., 1985b; Smith and Weber, 1988; Weber and Smith, 1987; Fritz and Schlunder, 1981). Besides the fact that application of

IAS model necessitates only single-solute data, the model is flexible in that multicomponent calculations can be performed using several different single-solute isotherm relationships. In addition, this model has a solid theoretical foundation, providing a useful understanding of the thermodynamic approach to adsorption. In this regard it is similar to the Gibbs adsorption equation upon which it is based. This is in contrast to the Langmuir competitive model, which is founded on the same limiting assumptions as the single-solute Langmuir model (i.e., monolayer adsorption and a homogeneous adsorbent surface).

However, Radke and Prausnitz (1972) have pointed out that the IAS model for predicting multisolute adsorption is most reliable for those systems where solute adsorption loading is moderate. If solute adsorption loading is large, the deviations of the predictions from experimentally observed data may be significant. Similar to the Langmuir and other multisolute equilibria models, the IAS model predicts that the adsorbate more favorably adsorbed in single-solute solutions also adsorbs to a greater extent when in competition at equimolar concentration. This is true only when adsorption is reversible and competition for adsorption sites is ideal. The criterion of ideal competition implies that the adsorbent is homogeneous with respect to adsorption sites and that the sites are equally accessible. However, activated carbon cannot be considered homogeneous because of its extensive microporous structure and the occurrence of different functional groups on the activated carbon surface. An ideal competition assumption, therefore, is invalid. Many researchers (Grant and King, 1990; Yonge et al., 1985; Snoeyink et al., 1969) have also shown that the adsorption of some compounds, such as phenolic compounds, are highly irreversible. This implies that it is difficult to replace each other once one of the components is prior adsorbed on activated carbon. It is evident that adsorption kinetics will affect the multicomponent adsorption if the adsorption rates of components are not proportional to their respective adsorptive capacities. Consequently, IAS and other existing multisolute equilibria models will fail to accurately predict solid-phase loadings under system conditions that exhibit significant non-ideal, that is, unequal competition and irreversible adsorption effects (Thacker et al., 1984; Smith and Weber, 1988; Yonge and Keinath, 1986).

*Modifications of IAS Model.* A number of attempts have been made to modify the IAS model to improve its accuracy and to reduce computational efforts. Using the IAS model, DiGiano et al. (1978) derived a simplified competitive equilibrium adsorption model (SCAM). This model, which is based on the Freundlich isotherm, assumes the single-solute isotherms of all the components are equal and it utilizes average isotherm constants when this assumption is not valid. The IAS equations have been reduced to a single expression:

$$q_{i} = K^{\prime \frac{n^{\prime}-1}{n^{\prime}}} \left[K_{i}C_{i}^{n_{i}}\right]^{\frac{1}{n^{\prime}}} \left[\sum_{i=1}^{N} \left(\frac{K_{i}}{K^{\prime}}C_{i}^{n_{i}}\right)^{\frac{1}{n^{\prime}}}\right]^{(n^{\prime}-1)}$$
(2-21)

where  $q_i$  is the solid-phase equilibrium concentration of solute *i*;  $n_i$  and  $K_i$ , the empirical Freundich constants for single solute *i*;  $C_i$ , the liquid-phase equilibrium concentration of solute *i*; n', the average value of  $n_i$ ; and K', the average value of  $K_i$ .

This model significantly simplifies the computations of the IAS model, although it does not improve its accuracy (Yonge and Keinath, 1986; Wilmanski and Breemen, 1990).

One popularized approach to modify the IAS model is to incorporate an empirical coefficient,  $R_i$ , into equation 2-20 to more accurately describe experimental equilibria (Thacker et al., 1984; Smith and Weber, 1988; Yonge and Keinath, 1986; Speitel et al., 1989):

$$\boldsymbol{q}_i = \boldsymbol{R}_i \boldsymbol{Z}_i \boldsymbol{q}_T \tag{2-22}$$

The modification factors,  $R_i$ , are determined from multisolute equilibrium data with a minimization procedure. This modification provides a significantly better description of the data. This improvement, however, is the result of parameters that are determined from the multisolute data itself.

#### Surface Comlexation Modeling

In the past quarter century there has been increasing interest in the sorption properties and surface chemistry of hydrous oxides. Common hydrous oxides, such as those of iron, aluminum, manganese, and silicon, can sorb a large number of chemical species. Experimental sorption data have been described by various empirical means, including partition coefficients, isotherm equations, and conditional equilibrium sorption constants. Much effort has been also invested in the development of theoretical models of the oxide/water interface. This work has been conducted to gain understanding of the sorption mechanisms involved, describe the available data in an efficient manner, and enable prediction of sorption.

Schindler, Stumm, and coworkers (Schindler and Kamber, 1968; Schindler and Gamsjager, 1972; Stumm et al., 1970; Huang and Stumm, 1973) proposed new ways of describing surface charge development and sorption models that embody the central features of the well-known electric-double-layer (EDL) theory but emphasize chemical reactions of sorbing ions with surface functional groups: the *surface complexation approach*. In this approach, sorbing ions are considered to react chemically with specific surface hydroxyl groups after coming through the interfacial electric field at the surface. The electric field, which may be positive or negative, results from a positive or negative surface charge caused by the chemical reactions at the surface.

The reactions with surface hydroxyl groups are considered analogous to the formation of soluble complexes and are described by mass law equations. However, the corresponding equilibrium constants are not, in fact, constant. What distinguishes surface reactions from reactions among (monomeric) solutes is the variable electrostatic energy of interaction caused by the variable charge on the surface. These electrostatic effects are taken into account by applying a coulombic correction factor, derived from EDL theory, to the equilibrium constants of the surface complexation reactions.

Surface complexation models have been widely applied to hydrous oxides/inorganics or organics systems, while very few studies have been conducted to apply the surface complexation models to activated carbon/inorgaics systems. In order to gain better insight into the mechanism of cadmium removal, Huang and Ostovic (1978)

have characterized the surface properties of activated carbon in terms of its surface acidity and hydroxo group. The association between cadmium ions and activated carbon was interpreted and described by a surface complexation model. No studies were found that applied surface complexation models to activated carbon/organics systems.

#### **Kinetic Models**

To properly interpret the kinetic experimental data, it is necessary to determine for the experimental system which of the steps in the adsorption process governs the overall adsorption rate. The process of adsorption of an organic compound by a porous adsorbent can be categorized as three consecutive steps. The first step is transport of solute across the boundary layer or surface film to the exterior surface of the adsorbent particle. The second step is transport of solute within the pores of the adsorbent particle, from the exterior of the particle to the interior surfaces of the particle. Similarly, solute may be transported along surfaces of pore walls. The final step is the physical or chemical binding of solute to the interior surface of the adsorbent.

*External Film Diffusion Model*. If external film diffusion is the rate-controlling step, the rate equation (Wermeulen, 1958; Kuo et al., 1987) can be expressed by

$$\frac{dq}{dt} = \frac{K_f a}{M} \left( C - C^* \right) \tag{2-23}$$

where  $K_{,a}$  is the mass transfer coefficient; C, the adsorbate concentration in bulk liquidphase;  $C^*$ , the adsorbate concentration of the liquid that is in equilibrium with the solidphase concentration q; and M, the carbon dosage.

Assuming the adsorption isotherm can be expressed by the Langmuir equation, i.e.,  $q = QbC^*/(1 + bC^*)$ , and taking advantage of the mass balance  $q = (C_0 - C)/M$  where  $C_0$ is the initial adsorbate concentration, equation 23 can be changed to

$$-\frac{dC}{dt} = K_{f}a(C - \frac{C_{0} - C}{b[QM - (C_{0} - C)]}) \qquad (2-23')$$

Usually, external transport is the rate-limiting step in systems which have: (a) poor mixing; (b) dilute concentration of adsorbate; (c) small particle sizes of adsorbent; and (d) high affinity of adsorbate for adsorbent (Zogorski et al., 1976). Some experiments conducted at low concentrations have shown that film diffusion solely controls the adsorption kinetics of low molecular weight substances (Fritz et al., 1981; Merk et al., 1981).

Internal Surface Diffusion Model. The adsorbate can diffuse by two mechanisms within the adsorbent, pore and surface diffusions. For pore diffusion, the adsorbate is transported within the pore fluid. For surface diffusion, the adsorbate continues to move along the surface of the adsorbent to available adsorption sites as long as it has enough energy to leave its present site. Investigations (Brecher et al., 1967; Furusawa and Smith, 1973; Komiyama and Smith, 1974) have demonstrated that surface diffusion is the dominant mechanism, so the contribution of pore diffusion is neglected. Many researchers (Crittenden and Weber, 1978a; Crittenden and Weber, 1978b; Crittenden et al., 1980; van Vliet and Weber, 1981; Wu and Gschwend, 1986; Traegner and Suidan, 1989) have used

the surface diffusion model for describing the kinetic data or for design of adsorbers.

The partial differential equation for this model is written in spherical coordinates as:

$$\frac{\partial q(r,t)}{\partial t} = D_s \left( \frac{\partial^2 q(r,t)}{\partial r^2} + \frac{2}{r} \frac{\partial q(r,t)}{\partial r} \right) \qquad (2-24)$$

in which q(r,t) represents the solid-phase concentration along the inner surface of the particle; r, the radial coordinate with an origin at the center of the particle; and  $D_s$ , the surface diffusion coefficient. The magnitude of  $D_s$  is a measure of how fast the molecules diffuse inside the carbon particle and therefore sets a time scale for the adsorption process. Two boundary conditions and one initial condition have to be specified in order to obtain a unique solution to equation 2-24. Initially the particle is free of adsorbate, i.e.:

$$q(r, t=0) = 0$$
 (2-25)

The boundary condition at the center of the particle is:

$$\frac{\partial q(r=0,t)}{\partial r} = 0 \qquad (2-26)$$

i.e., no adsorbate flux across the center. Finally, the continuity of flux at the solid-liquid interface has to be satisfied:

$$\rho_p D_s \frac{\partial q(r=d_p/2, t)}{\partial r} = K_f (C_b - C_s) \qquad (2-27)$$

with  $C_b$  and  $C_s$  denoting the bulk liquid and solid-liquid interface adsorbate concentrations, respectively;  $d_p$ , the particle diameter; and  $\rho_p$ , the apparent density of the carbon particle. This boundary condition contains the second important kinetic parameter  $K_p$ , which represents the liquid film mass transfer coefficient. The parameter  $K_f$  is a measure for how fast the molecules diffuse across the stagnant liquid film layer. It is assumed that local equilibrium occurs at the exterior carbon surface.

The average carbon loading, which is only a function of time, is given by:

$$q_{avg} = \frac{3}{(d_p/2)^3} \int_0^{d_p/2} q(r,t) r^2 dr \qquad (2-28)$$

*Linear-Driving-Force Approximation*. The surface diffusion model (eqn 2-24) is usually approximated by the linear-driving-force relation (Vermeulen, 1958; Kuo et al., 1987):

$$\frac{dq}{dt} = K_{p}a(q^* - q) \qquad (2-29)$$

where  $K_p a$  (= 60  $D_s/d_p^2$ ) is the mass transfer coefficient and  $q^*$  is the solid-phase concentration in equilibrium with the instantaneous fluid-phase concentration outside the particle.

If adsorption isotherm can be expressed by the Langmuir equation, i.e.,  $q^* = QbC/(1 + bC)$  and mass balance  $q = (C_0 - C)/M$  is used, equation 2-29 becomes:

$$-\frac{dC}{dt} = K_{p}a(\frac{MQbC}{1+bC} + C - C_{0}) \qquad (2-29')$$

Surface Reaction Model. For the case in which surface reaction is the rate-controlling step, the rate of adsorption can be expressed as (Vermeulen, 1958; Kuo et al., 1987)

$$\frac{dq}{dt} = K_a [C(Q - q) - \frac{q}{b}]$$
(2-30)

where  $K_a$  is the surface reaction rate constant, and Q and b are the Langmuir adsorptive capacity and equilibrium constant, respectively. Using the mass balance  $q = (C_0 - C)/M$ , the above equation changes to

$$-\frac{dC}{dt} = K_a [C(QM - C_0 + C) - \frac{1}{b}(C_0 - C)] \qquad (3-30')$$

The adsorption process can be pictured as one in which molecules leave solution and are held on the solid surface by chemical and physical binding. If the bonds that form between the adsorbate and adsorbent are very strong, the process is almost always irreversible, and chemical adsorption or chemisorption is said to have occurred. On the other hand, if the bonds that are formed are very weak, as is characteristic of bonds formed by the dispersion interactions or hydrogen bonding, physical adsorption is said to have occurred. The molecules adsorbed by this means are easily removed, or desorbed, by a change in the solution concentration of the adsorbate, and for this reason, the process

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is said to be reversible. There is a difference in the activation energy of adsorption reaction between physisorption and chemisorption. For chemical bonding, activation energy is higher than 10 kcal/mole, and for dispersion interactions and hydrogen bonding, it ranges from 2 to 10 kcal/mole.

Kuo et al. (1987) showed that the rate of the adsorption of dissolved organics from *in situ* tar sand by-product waters could be described by the surface reaction kinetics, i.e., equation 3-30. Grant and King (Grant and King, 1990) found that the oxidative coupling of phenolic compounds on carbon surfaces was a plausible explanation for irreversible adsorption. Their data indicated that a chemical reaction occurred and that the phenolate ion was more reactive than phenol. Because chemical reaction is slow compared to physisorption, surface reaction kinetics may control the adsorption process.

# Chapter 3 SINGLE-COMPONENT ADSORPTION EQUILIBRIA OF CHLOROPHENOLS

#### **Materials and Methods**

*Chemicals*. Eight chlorinated phenols were studied, pentachlorophenol (PCP), 2,3,4,6-tetrachlorophenol (2,3,4,6-TeCP), 2,4,6-trichlorophenol (2,4,6-TCP), 2,4,5-trichlorophenol (2,4,5-TCP), 2,4-dichlorophenol (2,4-DCP), 3,4-dichlorophenol (3,4-DCP), 2-chlorophenol (2-CP) and 4-chlorophenol (4-CP). The pKa values for these compounds are given in Table 3-1. Aqueous solutions of each chlorinated phenol were prepared by dissolving it in glass-distilled, deionized water, having varying concentrations of 100 to 300 mg/l. Preliminary studies indicated that in the neutral pH range (6.5 to 7.5), a 5 mM phosphate (NaH<sub>2</sub>PO<sub>4</sub> and Na<sub>2</sub>HPO<sub>4</sub>) buffer could maintain the pH of solution with adsorbate concentration of less than 300 mg/l within 0.2 unit fluctuation during adsorption. A 5 mM phosphate buffer was included in all subsequent experiments, unless noted otherwise. Using low concentration of phosphate aims at minimizing its influence on the adsorption of chlorophenols. The pH of each solution was adjusted with minimum amounts of HCl and NaOH solutions.

Activated Carbon. The adsorbent used throughout this study was Calgon Filtrasorb-400 activated carbon, the properties of which are described elsewhere (Montgomery, 1985). The carbon received from the manufacturer was ground in an analytical mill and sieved

to yield a 30 x40 mesh size (0.59 - 0.42 mm). After sieving, the carbon was washed with glass-distilled, deionized water to remove all fines, dried overnight in an oven at 105°C, and stored in an air-tight desiccator until use.

Compound	рКа
Pentachlorophenol	5.25ª
	4.74 <sup>b</sup>
2,3,4,6-Tetrachlorophenol	5.40ª
2,4,6-Trichlorophenol	6.15ª
	5.99 <sup>b</sup>
2,4,5-Trichlorophenol	6.94ª
2,4-Dichlorophenol	7.85*
4-Chlorophenol	9.18°
2-Chlorophenol	8.49°
	8.52 <sup>b</sup>

TADIC J.I UNA VALUES IVI CHIVIVUICHU	Table 3	3.1 p	Ka V	alues	for (	Chloro	phenols
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<sup>a</sup> after Schellenberg et al. (1984)

<sup>b</sup> after Westall et al. (1985)

<sup>c</sup> after CRC Handbook of Chem. Phys. (1990-1991)

*Equilibrium Studies*. Batch studies were used to obtain the equilibrium data. 40-ml portions of aqueous solution with the desired pH and adsorbate concentration were placed in 40-ml Teflon tubes containing accurately weighed amounts of carbon. Then the tubes were continuously shaken in a shaker bath for 6 days. This equilibration time was determined from preliminary kinetic studies. The temperature was kept constant at 30°C

for all experiments, except for studies on the effect of temperature. After adsorption equilibrium was reached, the equilibrium concentration was measured and the extent of adsorption was calculated. If the desired solution pH was outside the phosphate buffering region (pH 6.5-7.5) and the acidic and basic buffering region (pH < 4 and pH > 10), occasionally it was necessary to adjust pH during the equilibration period to maintain the desired pH. The final pH was measured.

Analytical Techniques. 5-ml aliquots were used for analysis of chlorophenols. The samples were first acetylated and then extracted into hexane. Each sample was added to 45 ml of glass-distilled water in a 150-ml separatory funnel using a 5-ml volumetric pipette. 50  $\mu$ l of internal standard (either 2,4,6-tribromophenol for 2,4,5-trichlorophenol, or 2,6-dibromophenol for other chlorophenols) was added using a 50- $\mu$ l syringe. After one ml of a 0.73 g/ml solution of potassium carbonate and one ml of acetic anhydride were added, the funnel was shaken for exactly two minutes. The complete acetylation reaction would take about two hours, after which 5 ml of hexane was added using a dedicated 5-ml volumetric pipette and again the funnel was shaken for another two minutes. After 30 minutes, the water layer was drained and the hexane layer was withdrawn using a new pasteur pipette and transferred to a 2-ml amber glass vial with Teflon-lined cap, which was stored in a refrigerator until gas chromatograph analysis was conducted.

Using an autosampler, 1 µl of hexane solution was injected into a Hewlett Packard Model 5890A Gas Chromatograph equipped with a 63 Ni electron capture detector (ECD) and 30 m by 0.323 mm i.d. DB-5 fused-silica capillary column (J+W Scientific, Orangeville, CA). Helium (5 psi) was used as the carrier gas and a 95% argon/5% methane mixture was used as the ECD auxiliary gas. The detector temperature was set at 320°C and injector temperature was 250°C. The samples were run using a temperature program as follows: an initial oven temperature of 45°C was held for 2 minutes, increased by 15°C/min to 150°C, and then by 5°C/min to a final temperature of 215°C that was held for 5 minutes.

#### **Results and Discussion**

Isotherms of Chlorophenols at pH 7. Equilibrium adsorption data at pH 7.0 for all compounds fitted the Langmuir equation, q = ObC/(1 + bC). The constants in the Langmuir equation were determined by plotting C/q versus C and making use of the above equation rewritten as a linearized form, C/q = 1/(Qb) + C/Q. Representative Langmuir plots of PCP, 2,4,6-TCP, and 3,4-DCP for high and low concentration ranges are shown in Figures 3.1 and 3.2, respectively. The Freundlich equation ( $q = k C^{n}$ ) was also used to fit the data, and the empirical constants k and n were evaluated by plotting log q versus log C. The values of Q, b, k, and n are given in Table 3.2, together with the linear regression correlation coefficients. The values of R-squared indicate that except 4-CP for high concentration range, the Langmuir equation fitted the equilibrium data much better than the Freundlich equation. This suggests a monolayer coverage of the accessible sites on the surface of the carbon. Except for PCP, the Freundlich equation better describes the equilibrium data in the high concentration range than in the low concentration range. The adsorption constants are concentration-dependent, because there are significant differences in the magnitudes of the constants between the two
concentration ranges. The higher concentration range has a larger Q and smaller b, k and n. Noteworthy also is the fact that the Langmuir adsorptive capacities (Q) increase from pentachlorophenol to trichlorophenols and do not change significantly from trichlorophenols to monochlorophenols. For molecular species, the more chlorinated a chlorophenol, the less soluble in water or more hydrophobic, and then the more adsorptive the compound. A comparison of the pKa values of the compounds reveals that at pH 7 the percentage of neutral molecules in all species increases from pentachlorophenol to monochlorophenols. The dominant species is the ionized form for pentachlorophenol and the molecular form for monochlorophenols, respectively. The adsorptive capacity for the neutral molecules is larger than that for the ionized forms. It is the combination of two factors, chlorination level and dissociation extent, that results in such a change of the adsorptive capacities for chlorophenols. In addition, from the comparisons of adsorption constants between 2,4,5-TCP and 2,4,6-TCP, 2,4-DCP and 3,4-DCP, and 4-CP and 2-CP, it can be deduced that the position of chlorine on the phenyl ring has no influence on the adsorption of chlorophenols.

Isotherms of 2,4,5-TCP at Various pH Values. Isotherms for adsorption of 2,4,5-TCP at pH values of  $4.15 \pm 0.2$ ,  $5.22 \pm 0.1$ ,  $6.58 \pm 0.01$ ,  $8.17 \pm 0.2$ ,  $9.3 \pm 0.2$ ,  $10.25 \pm 0.1$  were determined. Representative isotherms are presented in Figure 3.3. The solid lines shown in Figure 3.3 represent the Langmuir plots of best fit. There are only slight differences between the isotherms for pH  $4.15 \pm 0.2$  and pH  $5.22 \pm 0.1$ , but significant differences exist for other pH values. Similar results have been observed by Snoeyink et al. (1969).



Figure 3.1 Langmuir isotherms for adsorption of PCP, 2,4,6-TCP and 3,4-DCP (higher concentration range)



Figure 3.2 Langmuir isotherms for adsorption of PCP, 2,4,6-TCP and 3,4-DCP (lower concentration range)

		Langmuir		Freundlich			
Adsorbate	Concentra- tion Range 10 <sup>-3</sup> mmol/l	Q mmol/g	b I/mmo	R <sup>2</sup>	K	n	R <sup>2</sup>
РСР	0.843-12.53	1.419	1012.3	0.998	3.397	0.210	0.996
	13.2-208.4	1.954	135.6	1.000	2.308	0.114	0.987
2,3,4,6-	0.317-5.55	1.473	2874.2	0.998	3.956	0.195	0.956
TeCP	5.55-492.9	2.211	91.69	0.998	2.360	0.098	0.983
2,4,6-	0.39-11.70	2.135	1350.5	0.997	10.27	0.328	0.769
TCP	11.70-614.4	2.920	82.35	0.999	3.090	0.092	0.928
2,4,5-	0.157-10.33	1.943	1979.9	0.999	8.999	0.304	0.911
TCP	2.60-171.2	2.628	250.3	0.999	3.176	0.116	0.997
2,4-DCP	0.712-15.52	2.014	1007.9	0.998	5.583	0.243	0.866
	15.83-960.7	2.911	43.85	0.998	2.896	0.090	0.980
3,4-DCP	1.47-16.01	2.166	299.4	0.995	12.51	0.442	0.900
	17.3-816.4	2.867	62.89	0.999	2.949	0.100	0.964
4-CP	2.06-23.42	1.810	300.8	0.997	5.038	0.297	0.992
	20.15-781.9	3.090	17.04	0.988	3.087	0.208	0.993
2-CP	1.90-21.86	1.939	504.3	0.994	3.708	0.190	0.978
	21.86-791.3	3.132	31.85	0.998	3.208	0.145	0.990

 Table 3.2
 Adsorption Constants for Chlorophenols at pH 7



Figure 3.3 Langmuir isotherms for adsorption of 2,4,5-TCP at different pH

Langmuir constants Q and b for 2,4,5-TCP at different pH values were determined by regressing the isotherm data using the linearized form of the Langmuir equation. To minimize the effect of concentration range on the magnitude of the constants, only data within close concentration range for various pH values (except pH 10.25  $\pm$  0.1) were used. A summary of Langmuir capacity, Q, and constant, b, at various pH values, plus the concentration range within which the data were obtained, are given in Table 3.3. Several observations are evident from Table 3.3. The magnitude of b is fairly constant among the various pH values with close concentration ranges. However, it decreases significantly for pH 10.25  $\pm$  0.1, which can be attributed to the much higher concentration range. Decreasing pH value below the compound's pKa (6.94) appears to have no subsequent influence on the adsorptive capacity of 2,4,5-TCP. But a linear decrease in the adsorptive capacity of the compound occurs at pH values greater than its pKa.

PH	Q(mmol/g)	b(l/mmol)	Concentration Range (mmol/l)
4.15	2.798	189.0	0.00293-0.151
5.22	2.846	216.1	0.00227-0.141
6.58	2.743	188.9	0.00200-0.128
8.17	2.334	154.9	0.00185-0.166
9.30	1.861	147.0	0.00387-0.171
10.25	1.643	50.35	0.0295-0.249

 Table 3.3 Effect of pH on Langmuir Constants for 2,4,5-TCP

Surface Complexation Model. For the purposes of simplicity, activated carbon surface functional groups can be divided into two generalized types (Jankowska et al., 1991; Muller et al., 1980): acidic groups which may undergo neutralization by bases and basic groups which may be neutralized by acids. Carboxyl, phenolic, quinonoid, and normal lactone are the principal types of acidic surface functional groups, while usually structures corresponding to chromene or pyrone-like structures, are attributed to the basic group. The surface ionization reaction are therefore expressed by

$$\equiv AH \iff \equiv A^{-} + H_{g}^{+} \qquad K_{g}^{intr} \qquad (3-1)$$
$$\equiv BH^{+} \iff \equiv B + H_{g}^{+} \qquad K_{b}^{intr} \qquad (3-2)$$

where  $\equiv$ AH and  $\equiv$ A<sup>-</sup> represent neutral and negatively charged surface acidic groups,  $\equiv$ BH<sup>+</sup> and  $\equiv$ B represent positively charged and neutral basic groups,  $K_a^{intr}$  and  $K_b^{intr}$  are intrinsic equilibrium constants, and  $H_s^+$  denotes a proton at the carbon surface. The mass law equations corresponding to reactions 3-1 and 3-2 are

$$K_{a}^{intr} = \frac{\left[=A^{-}\right] \left[H^{+}\right]_{s}}{\left[=AH\right]}$$
(3-3)

$$K_{b}^{intr} = \frac{\left[=B\right] \left[H^{+}\right]_{s}}{\left[=BH^{+}\right]} \tag{3-4}$$

**1** \

The concentration of protons at some location, i, in the electrical double layer is related to the bulk concentration by the Boltzmann distribution (Davis et al., 1978; Muller et al., 1980), e. g.,

$$[H^{+}]_{i} = [H^{+}] \exp\left(\frac{-F\Psi}{RT}\right)$$
(3-5)

where  $\Psi$  is the surface potential, F is the Faraday's constant, R is the gas constant, and T is the absolute temperature.

Hence

$$K_{a}^{intr} = \frac{[=A^{-}][H^{+}]\exp(-F\psi/RT)}{[=AH]}$$
(3-6)

and

$$K_{b}^{intr} = \frac{[=B] [H^{+}] \exp(-F\psi/RT)}{[=BH^{+}]}$$
(3-7)

In the presence of chlorophenols, the following surface complexation reactions can be proposed:

$$=AH + H^{+} + ClP^{-} \iff =AH - HClP \qquad K_{A}^{intr} \qquad (3-8)$$

$$=BH^{+} + ClP^{-} \iff =BH^{-}ClP \qquad K_{B}^{intr} \qquad (3-9)$$

The concentrations of CIP in the electrical double layer are expressed in terms of the Boltzmann distribution, e. g.,

$$[ClP^{-}]_{i} = [ClP^{-}] \exp(F\psi/RT)$$
 (3-10)

Thus, the mass law equations corresponding to reactions 8 and 9 are

$$K_{\mathbf{A}}^{intr} = \frac{[=AH-HClP]}{[=AH] [H^+] [ClP^-]}$$
(3-11)

$$K_B^{intr} = \frac{[=BH-ClP]}{[=BH^+] [ClP^-] \exp(F\psi/RT)}$$
(3-12)

The formation of the surface complexes,  $\equiv$ AH-HClP and  $\equiv$ BH-ClP readjusts the acid-base equilibrium and affects the surface charge. The formation of  $\equiv$ BH-ClP serves to directly decrease (make more negative) the surface charge and hence the surface potential. The net surface charge density, in coulombs per square meter, is given by

$$\boldsymbol{\sigma} = \frac{F}{AS} \left( \left[ \equiv BH^+ \right] - \left[ \equiv A^- \right] \right) \tag{3-13}$$

where A is the specific surface area  $(m^2/g)$  and S the solid (carbon) concentration (g/l). To specify the influence of the excess surface charge density on the formation of surface complexes, we require an estimate of the surface potential. According to the Gouy-Chapman theory (for a symmetrical electrolyte with valence Z), the surface charge density  $\sigma$  is related to the surface potential  $\Psi$  (in volts) by

$$\sigma = \sqrt{8\epsilon RTI} \sinh \frac{ZF\Psi}{2RT}$$
(3-14)

where  $\varepsilon$  is the bulk permittivity of water and I the ionic strength.

Table 3.4 summarizes the surface complexation and aqueous reactions and corresponding equilibrium constants for activated carbon/2,4,5-TCP system. The two surface complexation constants,  $K_A^{intr}$  and  $K_B^{intr}$ , and total concentrations of acidic and basic groups can be extracted from adsorption data via the nonlinear regression program FITEQL (Westall, 1982). These constants and concentrations can in turn be used to simulate the adsorption data. The two properties of the electrical double layer, specific surface area and interface capacitance are assigned, respectively, 941 m<sup>2</sup>/g (after Huang and Ostovic, 1978) and 1.2 F/m<sup>2</sup> (Westall, 1982; Davis et al., 1978).

The optimal surface complexation constants for 2,4,5-TCP, 2,4-DCP and 4-CP are given in Table 3.5, together with overall variances (SOS/DF), which are the main indicator of the goodness of fit in the output from FITEQL. Westall (1982) notes that values of variance between 0.1 and 20 indicate a reasonably good fit. The three values of variances are all in close proximity to 1, indicating that the proposed model is very successful in fitting the adsorption data (Westall, 1982; Dzombak and Morel, 1990).

Surface complexation reactions	log K <sup>intr</sup>
$\equiv A^{-} + H^{+} = \equiv AH$ $\equiv B + H^{+} = \equiv BH^{+}$	7.78ª 5.70ª
$\equiv AH + H^{+} + ClP^{-} = \equiv AH - HClF$ $\equiv BH^{+} + ClP^{-} = \equiv BH - ClP$	$\log K_{A}^{intr}$ $\log K_{B}^{intr}$
Aqueous reactions	log K
$HClP = H^{+} + ClP^{-}$ $H_{2}O = H^{+} + OH^{-}$	-6.94 -14.0

## Table 3.4Surface Complexation and Aqueous Reactions in<br/>Activated Carbon/2,4,5-Trichlorophenol System

<sup>a</sup> after Huang and Ostovic (1978)

Adsorbate	$\log K_A^{intr}$	$\log K_B^{intr}$	SOS/DF
2,4,5- <b>TC</b> P	21.48	16.65	2.28
2,4-DCP	25.19	13.05	0.38
4-CP	27.41	16.68	1.41

Table 3.5Best Estimates for Chlorophenol/Carbon Surface<br/>Complexation Constants

Equilibrium measurements were made for adsorption of 2,4,5-TCP from solutions at pH range of 4 to 10 with phosphate ( $Na_2HPO_4$  and  $NaH_2PO_4$ ) concentrations of zero

and 5 mM. The solutions with or without phosphate contained the same initial 2,4,5-TCP concentrations of 1.525 mmol/l and the same carbon dosages were used. This implies that for the two systems with and without phosphate, the equilibrium concentrations are essentially identical if the amounts adsorbed are very close to each other, and therefore, the equilibrium data are highly comparable because the influence of concentration range on adsorption is eliminated. The data shown in Figure 3.4 indicate that no phosphate effect can be observed in whole pH range studied for any of these systems. It seems to be a reasonable explanation that phosphate is a too weak adsorbate to compete with chlorophenol. Some investigators have reported similar results for systems with salts of low concentration. Zogorski et al. (1976) noted that 0.05 M phosphate buffer exhibited no discernible influence on the rate of adsorption of 2,4-dichlorophenol and 2,4dinitrophenol, whether in an undissociated or dissociated form. Snoeyink et al. (1969) studied the effect of NaCl on the adsorption of phenol and p-nitrophenol (PNP) and found no distinguishable differences in capacity for NaCl concentrations of zero and 0.01 M at pH 2.0 and pH 10.0. However, they have observed a significant increase in capacity at lower surface coverage for the PNP system with 1.0 M NaCl at pH 10.0. This has been explained by the formation of ion pairing of the cation with the PNP anion, which would have the effect of increasing adsorptive capacity because the ion pair would behave more like the acid form of the PNP, or the reduction of repulsive forces between the adsorbed anions, which allows more molecules on the surface at lower solution concentrations.

The curve in Figure 3.4 represents the optimum fit of the surface complexation model to both sets of 2,4,5-TCP adsorption data obtained from the solutions with and

without phosphate. Figure 3.5 shows 2,4-DCP and 4-CP adsorption data as a function of pH (without phosphate) and the optimum surface complexation model fits. Similar to the relationship between adsorptive capacity and equilibrium pH for 2,4,5-TCP (Table 3.3), no subsequent influence on adsorption is observed for decreasing pH value below the compounds' pKa values, while marked decrease in amounts adsorbed occurs as pH increases above the pKa values. The two figures demonstrate that the proposed surface complexation model can describe very well the adsorption data of chlorophenols on activated carbon.

Figure 3.6 gives surface speciation of activated carbon in 2,4,5-TCP solution calculated as a function of pH with the surface complexation model. Total concentrations of basic and acidic groups derived from adsorption data with the optimization procedure are respectively 0.926 and 0.443 mmol/l (equivalent to 1.853 and 0.886 mmol/g), indicating that basic sites are about twice as many as acidic sites. Site saturation is essentially attained below pH 9.21 for basic groups. Surface complex of basic groups ( $\equiv$ BH-CIP) is relatively stable as pH increases. However, surface complex of acidic groups ( $\equiv$ AH-HCIP) is very susceptible to the change of pH and site saturation occurs only at pH 4.19. These imply that the electrostatic interaction between positively charged basic groups ( $\equiv$ BH<sup>+</sup>) and negatively charged chlorophenolate ion (CIP<sup>-</sup>), leading to the interaction between neutral acidic groups ( $\equiv$ AH) and neutral molecular chlorophenol (HCIP).



Figure 3.4 Effect of phosphate on adsorption of 2,4,5-TCP as a function of pH and optimum surface complexation model fit



Figure 3.5 Amounts adsorbed of 4-CP and 2,4-DCP as a function of pH and optimum surface complexation model fits



Figure 3.6 Calculated surface speciation of activated carbon (0.5 g/l) in the presence of 1.525 mmol/l 2,4,5-TCP (  $[=BH-CIP]+[=BH^*]+[=B] = 0.926 \text{ mmol/l},$  $[=AH-HCIP]+[=AH]+[=A^-] = 0.443 \text{ mmol/l})$ 

Some investigators have given an insight into the mechanisms of adsorption of phenolic compounds on activated carbon. It is suggested that the chlorophenols are adsorbed with the phenyl ring parallel to the surface of the adsorbent and interaction is between the  $\pi$ -electron system of the phenyl ring and the aromatic surface structure (Coughlin and Ezra, 1968) of the carbon. In addition, Mattson et al.(1970) substantiate the presence of significant numbers of carbonyl and carboxyl functional groups on the surfaces of activated carbon using infrared internal reflectance spectrophotometric techniques. Chlorophenols may be adsorbed at the carbonyl oxygens on the activated carbon surface according to a donor-acceptor complexation mechanism. The carbonyl oxygen acts as the electron donor and the aromatic ring of the solute acts as the electron acceptor. The functional groups containing oxygen, such as phenolic, carbonyl, carboxyl, lactone, and quinone (Boehm, 1964) may also offer adsorptive sites for hydrogen bonding of the phenolic protons, particularly at low pH levels where the chlorophenols are predominantly in molecular form. From this study, these likely mechanisms seem applicable only to the interaction between neutral chlorophenol molecules and neutral acidic sites. The electrostatic bonding of chlorophenolate ions with positively charged basic sites may be more substantial for adsorption of chlorophenols on activated carbon.

Effect of Dissolved Organic Matter on Adsorption Equilibrium. Figure 3.7 shows the effect of background dissolved organic matter (DOM) on adsorption equilibrium of 3,4-DCP. Background solution containing DOM (acetate and its metabolites) was collected from biodegradation reactor and prefiltered through a glass fiber filter prior to use to remove suspended particles. The results indicate that the presence of 5 mg/l (as TOC) of

background DOM reduces the Langmuir adsorptive capacity about 13%. The precise mechanisms responsible for reduction by DOM of target compound adsorption are not readily apparent. DOM may alter such solution properties as solubility and hydrophobicity and, therefore, adsorption characteristics. Direct competition for adsorption sites between target compounds and the adsorbing components of the background DOM is certainly another potential mechanism for reduction of the adsorptive capacity.

Effect of Temperature on Adsorption Equilibrium. Equilibrium studies were performed at various temperatures at pH 7.0. The data obtained from these studies are plotted in Figures 3.8 -3.11. There are two patterns for the effect of temperature on the adsorption of chlorophenols on granular activated carbon. For PCP and 2,4,6-TCP, the amount adsorbed, for a given equilibrium concentration is seen to increase with decreasing temperature, and therefore, the adsorption is an exothermic process. Contrary to these results, for 2,4-DCP and 4-CP, an increase in temperature resulted in increased adsorption, and the adsorption reaction is endothermic. The parameters of the Langmuir equation for each isotherm of these four compounds have been computed and are tabulated in Table 3.6.

The change in the heat content of a system in which adsorption occurs, that is, the total amount of heat evolved in the adsorption of a definite quantity of solute on adsorbent, is termed the heat of adsorption,  $\Delta H$ . The differential heats of adsorption for the four chlorophenols have been determined using the values of Q, computed from the Langmuir equation, corresponding to the selected temperatures and the Van't Hoff-Arrhenius equation in the form (Weber and Morris, 1964):

$$\Delta H = 2.3R \frac{T_1 T_2}{T_2 - T_1} (\log Q_2 - \log Q_1)$$
(3-1)

An example computation of adsorption heat for 2,4-DCP is demonstrated here. The logarithms of Q for the data of 2,4-DCP from Table 3.4 are plotted versus the reciprocal of kelvin temperature in Figure 3.12. From this plot the slope,  $-\Delta H/(2.3R)$ , has been taken and  $\Delta H$  has been computed as follows:

$$\Delta H = -2.3 (1.986) (-269.4) = 1233 \text{ cal./mole}$$

The heats of adsorption for the four compounds are also included in Table 3.6.

Temperature effects on adsorption equilibrium generally are not significant, particularly over the range of temperature encountered in water and waste waters. This fact is shown by Figures 3.8 - 3.11 and the values of isotherm parameters in Table 3.6 (except for 2,4,6-TCP). In practical operations, variation of temperature will be small relative to that which has been imposed on the experimental systems. The observed temperature-dependence is undoubtedly the net effect of temperature on the adsorbate-adsorbent bonds, the solvent-adsorbent bonds, the solvent-adsorbent bonds, the solvent-adsorbent bonds, the adsorbate interactions, and the solvent-adsorbate interactions. The solvent-adsorbent bond is important because adsorption of a solute molecule probably involves concomitant displacement of solvent molecules from the surface. Snoeyink et al. (1969) reported that the reaction of acid with coconut-shell carbon is endothermic. Grant and King (1990) reported that the adsorption of phenol on Filtrasorb-100 activated carbon is endothermic, an observation that is consistent with the temperature-dependence of 2,4-DCP and 4-CP adsorption, and



Figure 3.7 Effect of dissolved organic matter on adsorption equilibrium



Figure 3.8 Langmuir isotherms for adsorption of PCP at various temperatures



Figure 3.9 Langmuir isotherms for adsorption of 2,4,6-TCP at various temperatures



Figure 3.10 Langmuir isotherms for adsorption of 2,4-DCP at various temperatures



Figure 3.11 Langmuir isotherms for adsorption of 4-CP at various temperatures



Figure 3.12 Effect of temperature on adsorption of 2,4-DCP

Adsorbate	Temperature °C	Q mmol/g	b l/mmol	∆H cal/mole
	25	1.483	1112.9	
PCP	30	1.438	1130.3	-1565
	40	1.310	1063.8	
	30	1.986	3585.4	
2,4,6-TCP	40	1.527	3108.6	-5426
	50	1.137	1964.4	
	22.5	2.067	962.0	
2,4-DCP	30	2.182	1327.8	1233
	40	2.299	1712.8	
	50	2.483	1358.7	
	23	1.647	946.1	
4-CP	30	1.856	800.0	2672
	40	2.110	574.4	

Table 3.6Langmuir Parameters for Adsorption of Chlorophenolsas a Function of Temperature and Heat of Adsorption

regarded this phenomenon as evidence for the occurrence of a chemical reaction. At pH 7.0, predominant species are anionic for PCP and 2,4,6-TCP, and neutral for 2,4-DCP and 4-CP. This might explain the adsorption being exothermic for the former, and endothermic for the latter. However, the total process and realistic mechanism are very

complex, and the effect of temperature on adsorption equilibria is difficult to interpret.

**Reproducibility of Measurements.** Six identical and independent analytical measurements including acetylation, extraction and gas chromatography were performed for a 2,4-DCP solution sample of 0.026 mmol/l. The ratios of area of 2,4-DCP peak to that of internal standard and standard deviations have been calculated and are given in Table 3.7.

Ratio of areas	Standard deviation	Coefficient of variation (%)
0.04804		
0.04808		
0.04798	0.00036	0.75
0.04802		
0.04800		
0.04891		

 Table 3.7
 Reproducibility for Analytical Measurements

From the small relative standard deviation it can be concluded that the reproducibility of analytical measurements is very good and the error resulting from analytical procedures is minor.

To provide an estimate of experimental error for the isotherm parameters, five isotherms for adsorption of 2,4-DCP have been independently obtained from the bottle

	Q mmol/g	b I/mmol
	2.3439	1249.3
Langmuir parameters	2.3564	1193.0
	2.3428	1132.3
	2.3421	1225.4
	2.2473	1195.7
Means	2.3265	1199.1
Standard deviations	0.0447	43.9
Coefficient of variation (%)	2.05	3.66
95% confidence	2.2710	1144.6
intervals	-2.3820	-1253.6

# Table 3.8Standard Deviations and 95% ConfidenceIntervals for Langmuir Parameters

experiments, in which the same stock solution was used and for each isotherm each bottle was dosed with a definite amount of carbon. Table 3.8 shows the standard deviations and 95% confidence intervals for the Langmuir parameters. The errors may result from

experimental procedures, such as pipetting of solution and weighing of carbon, but most of them may be attributed to the heterogeneity of activated carbon.

**Blank Test.** Blank tests have been conducted with PCP, 2,4-DCP and 4-CP of 0.5 mmol/l at 30 and 40°C, and the results are given in Table 3.9. From the table, the ratios of peak areas for chlorophenols and internal standard at 30 and 40°C are very close to those of controls, and thus the effects of biodegradation and volatilization can be neglected.

Adsorbate	Temperature °C	Ratio of Areas	
	control	0.846	
PCP	30	0.844	
	40	0.868	
	control	0.150	
2,4-DCP	30	0.153	
	40	0.149	
	control	0.0222	
4-CP	30	0.0229	
	40	0.0224	

Table 3.9 Blank Test Results

### Conclusions

Studies were undertaken of the adsorption of chlorinated phenols from aqueous solution on granular activated carbon. Single-component equilibrium adsorption data on the eight compounds in two concentration ranges at pH 7.0 fit the Langmuir equation better than the Freundlich equation, while the Freundlich equation improves its performance in high concentration range. There are pronounced differences in the magnitudes of the constants between the two concentration ranges, and thus the adsorption constants are concentration-dependent. The adsorptive capacities at pH 7.0 increase from pentachlorophenol to trichlorophenols and are fairly constant from trichlorophenols to monochlorophenols. The position of chlorine on the phenyl ring has no influence on the adsorption of chlorophenols. No effect of phosphate on the adsorption was observed in the pH range of 4 - 10. The presence of background dissolved organic matter could reduce the adsorptive capacity of chlorophenols. Equilibrium studies were performed at various temperatures at pH 7.0. The adsorption process was found to be exothermic for pentachlorophenol and 2,4,6-trichlorophenol, and endothermic for 2,4-dichlorophenol and 4-chlorophenol. Different dissociation extents of these compounds seem to be responsible for such temperature effects.

Equilibrium measurements were also conducted for 2,4,5-trichlorophenol, 2,4dichlorophenol, and 4-chlorophenol over a wide pH range. No subsequent influence on adsorption is observed for decreasing pH values below the compounds' pKa values, while marked decrease in amounts adsorbed occurs as pH increases above the pKa values. A surface complexation model has been proposed to describe the effect of pH on adsorption equilibria of chlorophenols on activated carbon. Activated carbon surface functional sites are divided into acidic groups and basic groups which may be neutralized by bases or acids. Molecular and ionized forms of chlorophenols interact, respectively, with these two kinds of surface groups to form two neutral surface complexes, of which the complex of basic groups is more substantial and more stable than that of acidic groups. The simulations of the model are in excellent agreement with the experimental data.

#### Chapter 4 ADSORPTION KINETICS OF CHLOROPHENOLS

#### **Materials and Methods**

*Kinetic studies.* Batch kinetic experiments were conducted by using a 3-liter glass reactor. The solution was stirred with a two-blade stirring rod attached to a motor. The temperature of the adsorbate solution was kept constant at 30°C during the experiment by placing the reactor in a constant temperature chamber. A 5 mM phosphate buffer was included in all experiments. At appropriate intervals, solution samples (2 to 8 ml) were withdrawn for analysis.

#### **Results and Discussion**

*Effect of Stirring Speed on Adsorption Rate.* Three stirring speeds of approximately 200, 400 and 800 rpm have been tried to see the effect of stirring speed on adsorption rate. Kinetic studies with 2,4,6-trichlorophenol indicated that the rate of adsorption was independent of stirring for a shaft speed greater than 400 rpm, as shown in Figure 4.1. Since too vigorous stirring could cause a severe attrition of carbon, all subsequent kinetic experiments were conducted at a stirring speed of 600 rpm.

*Effect of pH on Adsorption Rate.* Figure 4.2 shows the effect of solution pH on the observed rate of adsorption of 2,4,5-TCP by granular activated carbon. Because the pKa of 2,4,5-TCP is 6.94, essentially all species are present in the neutral form at pH 3.95 or



Figure 4.1 Effect of stirring speeds on adsorption rate of 2,4,6-TCP



Figure 4.2 Effect of pH on adsorption rate of 2,4,5-TCP

in the ionized form at pH 10.0, both forms being approximately equal at pH 7.0. The rate of adsorption of 2,4,5-TCP at pH 10.0 is slower than that at pH 3.95 and 7.0, and there is only a slight difference between the adsorption rates for pH 3.95 and 7.0. Similar kinetic behavior was reported for the adsorption of alkylbenzenesulfonic acids and phenols (Weber and Morris, 1963; Zogorski et al., 1976). The uncharged molecular species of 2,4,5-TCP may be adsorbed by the carbon, which bears net negative charges, more rapidly than the negatively charged ionic species. At higher pH, both the repulsive forces between the adsorbate anion and the carbon surface, and between the adsorbed species themselves, would tend to reduce the adsorption rate. Thus, the adsorption rate of chlorophenols in acidic solutions is much higher than that in basic solutions.

That the adsorption rates for pH 3.95 and 7.0 are essentially identical may be explained by the existence of the following chemical equilibria in the solution:

$$H_2 PO_4^{-} \rightleftharpoons H^+ + HPO_4^{-2}$$
(4-1)

$$H^{+} + \bigcup_{CI} \bigcup_{CI}^{O^{-}} \bigcap_{CI}^{CI} \longrightarrow_{CI}^{OH} \bigcap_{CI}^{CI} (4-2)$$

Because of the rapidity of acid-base reactions, these two equilibria can be reached nearly instantaneously. Even though the molecular species of 2,4,5-TCP at pH 7.0 accounts for only 47%, most species adsorbed by GAC can be the molecular 2,4,5-TCP, due to the

proton donation by the phosphate buffer.

Adsorption Kinetics. Figures 4.3 and 4.4 shows the typical data from short-term kinetic studies on the adsorption of 2,4-DCP and 4-CP by 30x40 mesh Filtrasorb-400 granular activated carbon as a function of time, and the curves predicted by the external film diffusion model (eqn 2-23), linear-driving-force approximation (eqn 2-29), and surface reaction kinetics (eqn 2-30). Adsorption is indicated by the decreasing concentration of solute remaining in solution. The curves are all the best fits to the data with the R-squared values of 0.999, 0.981, and 0.999 for the external film diffusion model, linear-driving-force approximation, and surface reaction kinetics for 2,4-DCP, and 0.993, 0.983, and 0.998 for these three models for 4-CP, respectively. The required Langmuir isotherm constants are given in Table 3.2. It is evident from the R-squared values that while both the surface reaction kinetics and the external film diffusion model appear to describe the kinetic data very well, surface reaction kinetics fits the data a little better than external film diffusion model.

The performances of the models are compared using statistical techniques. This comparison was achieved by calculating both the sum of the squares of the errors (SSE) and the square of the correlation coefficient (R-SQUARE). In mathematical forms these quantities are

$$SSE = \sum_{i=1}^{n} (\hat{C}_{i} - C_{i})^{2}$$
 (4-3)


Figure 4.3 Comparison of kinetic models for adsorption of 2,4-DCP



Figure 4.4 Comparison of kinetic models for adsorption of 4-CP

in which  $C_i$  = the experimentally determined concentration of kinetic data point *i*, and  $\hat{C}_i$ = the predicted concentration; and

$$R-SQUARE = \frac{\sum C_i^2 - SSE}{\sum C_i^2}$$
(4-4)

Figures 4.5 - 4.8 show the adsorption kinetic data of 2,4-DCP and 2,4,6-TCP at different adsorbate concentrations and different carbon dosages, and their respective best fit curves predicted by the surface reaction kinetics and external film diffusion models. Mass transfer coefficients and corresponding R-square values for these two models are summarized in Table 4.1. Mass transfer coefficients for the surface reaction kinetics varied slightly with initial adsorbate concentration and carbon dosage. They varied directly with initial concentration and inversely with carbon dosage. However, mass transfer coefficients for the external film diffusion model are very close to each other for all the cases if carbon dosage is not included in the rate expression. It can be seen that except for 2,4-DCP at the carbon dosage of 100 mg/l the R-square values for the surface reaction kinetics are all higher than those for the external film diffusion model for these three chlorophenols.

In Table 4.1 two observations are made. The external film diffusion model improved its performance on the 2,4,6-TCP and 2,4-DCP systems when the initial adsorbate concentration decreased to about one-fifth. The R-squared values are highest among theses three cases for both adsorbates and almost the same as those of the surface



Figure 4.5 Changes of 2,4-DCP concentration with time and predictions of surface reaction model



Figure 4.6 Changes of 2,4-DCP concentration with time and predictions of external film diffusion model



Figure 4.7 Changes of 2,4,6-TCP concentration with time and predictions of surface reaction model



Figure 4.8 Changes of 2,4,6-TCP concentration with time and predictions of external film diffusion model

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Adsorbate	Init.conc. mmol/l	Carbon dosage mg/l	Surface reaction		External diffusion	
			Ka l/mmol-	R <sup>2</sup>	K <sub>r</sub> a 1/hr	R <sup>2</sup>
	0.1136	30	2.014	1.000	0.082	0.997
2,4,6-TCP	0.1136	100	1.777	0.998	0.305	0.997
	0.0238	100	1.382	0.999	0.280	0.999
	0.1154	100	1.874	1.000	0.302	0.998
2,4-DCP	0.1154	200	1.728	0.999	0.617	0.999
	0.0287	100	1.662	0.999	0.310	0.999
4-CP	0.1257	100	1.980	0.998	0.280	0.993

 Table 4.1
 Mass Transfer Coefficients for Three Chlorophenols

reaction kinetics. This implies that the external transport resistance increases as adsorbate concentration decreases. Also, at the same carbon dosage (100 mg/l) and approximately the same initial adsorbate concentrations, the mass transfer coefficient for the surface reaction kinetics increases from 1.777 l/mmol-hr for 2,4,6-TCP to 1.874 l/mmol-hr for 2,4-DCP and 1.980 l/mmol-hr for 4-CP. This means that at pH 7.0, the order in which adsorption equilibrium is attained will be first 4-CP, then 2,4-DCP and finally 2,4,6-TCP. This implies that the more chlorinated a chlorophenol, the slower it reaches adsorption

equilibrium. Decreasing dissociation extent from 4-CP to 2,4,6-TCP may account for this observation.

A high initial adsorbate concentration study on the adsorption kinetics of 2,4-DCP and two long-term studies on the adsorption kinetics of 2,4,6-TCP and PCP have also been performed, as shown in Figures 4.9, 4.10 and 4.11, respectively. It is seen from the figures that surface reaction kinetics still best describe the kinetic data of these three experiments. Noteworthy also is the observation that the linear-driving-force approximation has improved its performance and the external film diffusion model has become the worst for the long-term kinetics.

The distinction between external diffusion or surface reaction and intraparticle diffusion controlled processes can be made by performing an interruption test. This test has been used by Zogorski et al. (1976) and Weber and Morris (1963). In an interruption test, the adsorbent is removed from the adsorbate solution for a period and then reimmersed. When concentration gradients are present, the pause gives time for the gradients to level off within the pores of the adsorbent. When intraparticle diffusion is the rate-limiting step, the rate of removal immediately after reimmersion is greater than the rate prior to interruption. The interruption period has no influence on the rate of adsorption after reimmersion when external transport or surface reaction is rate limiting. Thus, the interruption test is a simple test for determining if large concentration gradients exist within the pores of the adsorbent.

A interruption test was conducted with 2,4,6-TCP to help define the nature of the rate-limiting step in the adsorption process of chlorophenols. After 3 hours from the



Figure 4.9 Comparison of kinetic models for adsorption of 2,4-DCP (high initial concentration)



Figure 4.10 Comparison of kinetic models for adsorption of 2,4,6-TCP (long term)



Figure 4.11 Comparison of kinetic models for adsorption of PCP (long term)



Figure 4.12 Interruption test for adsorption of 2,4,6-TCP

starting of the kinetic experiment, the adsorbate solution was decanted, and the adsorbent was left in the reactor with a few milliliters of solution. The experiment started again 17 hours later. The results of this interruption test are shown in Figure 4.12, in which the concentration of 2,4,6-TCP remaining in solution is plotted against accumulated immersion time. The figure also indicates that surface reaction kinetics can fit the experimental data over the entire kinetic process. The rate of adsorption of 2,4,6-TCP appears not to be limited by internal diffusion. This observation is based on the lack of influence of the interruption period on the concentration profile after reimmersion.

Several characteristics of the adsorbent, adsorbate, and solution phase are of importance in determining the rate-limiting step. These factors include the particle size of the adsorbent, concentration of adsorbate, degree of mixing, interaction of adsorbate with adsorbent, and mass transfer coefficients in each individual adsorption step. Usually, external film transport is the rate-limiting step in systems which have dilute concentration of adsorbate and small particle sizes of adsorbent. In contrast, the intraparticle step limits the overall transfer for those systems which have large particle sizes of adsorbent and high concentration of adsorbate. Surface reaction controls the overall adsorption process for systems in which surface reaction is a chemical reaction because it is usually slow compared to physisorption. It can be concluded from the above kinetic results that both the external film diffusion model and the surface reaction kinetics model fit the kinetic data, but surface reaction kinetics is better. Several researchers (Mattson et al., 1969a; Mattson et al., 1969b; Grant and King, 1990) have indicated the formation of chargetransfer complexes between phenolic compounds and carbon surface functional groups, or an oxidative coupling reaction of phenolic compounds on the carbon surface. As shown in chapter 3, the adsorption equilibrium of chlorophenols on activated carbon can be described very well by the proposed surface complexation model, in which molecular and ionized forms of chlorophenols interact respectively with acidic and basic surface functional groups to form two neutral surface complexes, of which the complex of basic groups is more substantial and more stable than that of acidic groups. Thus, chemisorption appears to be the most reasonable explanation for the kinetic characteristics of the adsorption of chlorophenols on activated carbon.

Sensitivity Analysis. An understanding of the relative importance of each parameter in the surface reaction kinetics model can be seen from the sensitivity analysis presented in Figures 13, 14 and 15, where typical chlorophenol kinetics parameters have been used in the simulations. As shown in Figures 13 and 14, the surface reaction kinetics model is very sensitive to the parameters Ka and Q. This gives an indication of the necessity of obtaining accurate Langmuir adsorptive capacity. Figure 15 shows that the effect of Langmuir constant b on kinetic process is very small.

*Two-component Adsorption Kinetic Process*. The experiments have been performed to verify that the adsorption of chlorophenols on activated carbon is to some extent an irreversible process and non-ideal competition between two components exists. In kinetic studies with 2,4-DCP and 2,4,6-TCP, two adsorbates were introduced simultaneously or one of two was added 10 hours later. The duration of the experiments is 6 days, which is the same as the equilibration time for equilibrium studies, and the results are given in Figures 4.16 - 4.18. It can be seen that the final concentrations are always lower for



Figure 4.13 Sensitivity analysis: Ka parameter



Figure 4.14 Sensitivity analysis: Q parameter



Figure 4.15 Sensitivity analysis: b parameter



Figure 4.16 Changes of 2,4,6-TCP and 2,4-DCP concentrations with time in batch reactor (two components were introduced simultaneously)



Figure 4.17 Changes of 2,4,6-TCP and 2,4-DCP concentrations with time in batch reactor (2,4,6-TCP was introduced 10 hours earlier)

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Figure 4.18 Changes of 2,4,6-TCP and 2,4-DCP concentrations with time in batch reactor (2,4-DCP was introduced 10 hours earlier)

adsorbates added later and higher for adsorbates added earlier as compared with the respective concentrations of adsorbates in simultaneously introduced system. The difference of 10 hours contact time in a total 144 hours could not result in so significant a disparity in final concentration. It must be due to the irreversibility of adsorption, that is, that molecular or anionic species of earlier added chlorophenol preferentially occupy higher-energy active sites on the carbon surface and some of them cannot be replaced by later added species. In another sense, the preferential adsorption can be said to occur in systems of simultaneously introduced two components if the adsorption rates of components are not proportional to their respective adsorptive capacities. From Figure 4.16 this disproportion of adsorption rates with adsorptive capacities is seen to exist in 2,4-DCP/2,4,6-TCP system because the adsorption rate of 2,4-DCP is higher than that of 2,4,6-TCP. This may explain why the Langmuir, IAS and other existing multicomponent equilibrium models fail to predict solid-phase loadings in systems of chlorophenols.

## Conclusions

Batch kinetics studies were conducted of the adsorption of chlorinated phenols on granular activated carbon. pH played an important role in the adsorption kinetics of chlorophenols at pH values above the pKa values of the compounds, while little influence on adsorption rate was observed if pH was decreased below the pKa values. The external film diffusion model, the linear-driving-force approximation, and the surface reaction kinetics model have been employed to fit the adsorption kinetics data of chlorophenols. The results show that the surface reaction model best describes both short-term and longterm kinetics, while the external film diffusion model describes the short-term kinetics data very well and the linear-driving-force approximation improves its performance for the long-term kinetics. The interruption test also indicates that the rate of adsorption is not limited by internal surface diffusion. The mass transfer coefficient increases from more chlorinated compounds to less chlorinated compounds, which implies that the more chloro groups in a chlorophenol, the slower it reaches adsorption equilibrium. The twocomponent adsorption kinetics experiments have revealed that the adsorption of chlorophenols on activated carbon is to some extent irreversible and that non-ideal competition between two components exists. Chemisorption appears to be the most reasonable explanation for the kinetics characteristics of the adsorption of chlorophenols on activated carbon.

# Chapter 5 MULTICOMPONENT ADSORPTION EQUILIBRIA OF CHLOROPHENOLS

#### A New Method for Predicting Multicomponent Adsorption Equilibria

It has been pointed out that the Langmuir competitive model, IAS model, and other existing multicomponent equilibria models fail to predict solid-phase loadings under system conditions that exhibit significant irreversible adsorption effects and non-ideal competition. As we can see from the previous and following chapters, chlorophenols show a very high affinity for activated carbon, the adsorption process is highly irreversible, and thus these systems are non-ideal. It seems impossible for a model to accurately predict multicomponent adsorption equilibria in non-ideal systems if the model does not use other parameters in addition to single-component isotherm constants. From the view point of thermodynamics, all liquid- and solid-phase solutions can be treated as a non-ideal solution while an ideal solution is merely a particular case of non-ideal solutions. The derivation of the proposed prediction method will start with the fundamentals of solution thermodynamics.

Let us consider in detail adsorption on a adsorbent surface from a binary liquid mixture. We can assume that at adsorption equilibrium we have two binary solutions (two phases): the solid or surface solution and the bulk solution. At thermodynamic equilibrium:

$$\mu_1^s = \mu_1$$
 ,  $\mu_2^s = \mu_2$  (5-1)

where  $\mu_1^s$  and  $\mu_2^s$  are the chemical potentials of components 1 and 2, respectively, in the surface solution, and  $\mu_1$  and  $\mu_2$  are the corresponding chemical potentials in the bulk solution.

Adopting the thermodynamic description of adsorption equilibria used by Fu et al. (1948) we can rewrite equilibria (eqn 5-1) in the form:

$$\mu_1^{0,s} + RT \ln x_1^s f_1^s = \mu_1^0 + RT \ln x_1 f_1$$
 (5-2)

$$\mu_2^{0,s} + RT \ln x_2^s f_2^s = \mu_2^0 + RT \ln x_2 f_2$$
 (5-3)

where  $\mu_1^{0,s}$  and  $\mu_2^{0,s}$  are the standard chemical potentials of components 1 and 2 in the surface solution,  $\mu_1^{0}$  and  $\mu_2^{0}$  are those in the bulk solution,  $x_i^{s}$  and  $x_i$ , and  $f_i^{s}$  and  $f_i$  are accordingly the mole fractions and the activity coefficients of the respective components in the surface and bulk solutions. Of course, the standard chemical potentials of the components depends on the reference system used.

From equations 5-2 and 5-3 we get

$$\frac{x_1^s}{x_1} = \frac{f_1}{f_1^s} \exp \left[-\frac{1}{RT} (\mu_1^{0,s} - \mu_1^0)\right]$$
(5-4)

$$\frac{x_2^{s}}{x_2} = \frac{f_2}{f_2^{s}} \exp \left[-\frac{1}{RT} (\mu_2^{0,s} - \mu_2^{0})\right]$$
(5-5)

Dividing equation 5-4 by equation 5-5, side by side, we get:

$$\frac{x_1^s x_2}{x_1 x_2^s} = \frac{f_1 f_2^s}{f_1^s f_2} \exp\left\{-\frac{1}{RT} \left[ \left(\mu_1^{0,s} - \mu_1^0\right) - \left(\mu_2^{0,s} - \mu_2^0\right) \right] \right\}$$
(5-6)

Let us denote:

$$\alpha = \frac{x_1^{s} x_2}{x_1 x_2^{s}}$$
(5-7)

$$D = \frac{f_1 f_2^s}{f_1^s f_2}$$
(5-8)

$$K = \exp \left\{-\frac{1}{RT} \left[ \left(\mu_1^{0,s} - \mu_1^{0}\right) - \left(\mu_2^{0,s} - \mu_2^{0}\right) \right] \right\}$$
(5-9)

Using the notations 5-7, 5-8 and 5-9, we can write equation 5-6 in the form:

$$\frac{x_1^{s} x_2}{x_1 x_2^{s}} = DK = \alpha$$
 (5-10)

Since  $x_1^s = q_1 / (q_1 + q_2)$ ,  $x_2^s = q_2 / (q_1 + q_2)$ ,  $x_1 = C_1 / (C_1 + C_2)$ , and  $x_2 = C_2 / (C_1 + C_2)$ , we have:

$$\frac{q_1 C_2}{q_2 C_1} = DK = \alpha^{-1}$$
(5-11)

where  $q_1$  and  $q_2$  are the amounts adsorbed, and  $C_1$  and  $C_2$  are the equilibrium concentrations for components 1 and 2, respectively. The quantity  $\alpha$  is known as the distribution coefficient or distribution function. K is a function only of temperature. At a given temperature, K is a constant. When adsorption takes place from an ideal solution and the surface solution is also ideal, then D = 1, and:

$$\frac{q_1C_2}{q_2C_1} = K = \alpha = constant$$
(5-12)

In case of adsorption from a non-ideal solution, the value of D, and therefore the

<sup>1</sup> From the Langmuir competitive model (eqn 2-9), we have:

$$\frac{q_1C_2}{q_2C_1} = \frac{Q_1b_1}{Q_2b_2} = constant$$

Hence it is not surprising that the Langmuir competitive model holds only in ideal systems.

value of  $\alpha$ , vary with the concentrations of components 1 and 2 in the surface and bulk solutions.

Now, consider two-component competitive adsorption from another aspect. The two-component adsorption can be described as the following competitive reactions:



in which -S represents the unoccupied surface sites, and SA and SB represent the surface sites occupied by components A and B, respectively.

Denote single-component adsorptive capacities of components A and B as  $Q_1$  and  $Q_2$ , and single-component adsorption constants as  $b_1$  and  $b_2$ , respectively. Applying the law of mass action to the single-component system, for component 1 we have  $q_1 = b_1 C_1 (Q_1 - q_1)$ , which, after rearrangement, leads to the single-component Langmuir equation. For a two-component system in which component 1 is the dominantly adsorbed component, the following equation can be obtained from a mass balance:

$$q_1 = b_1 C_1 (Q_1 - q_1) - f q_2 \qquad (5-14)$$

where  $f = Q_1 / Q_2$ , a factor for converting  $q_2$  into  $q_1$ . The first term on the left side would represent the amount adsorbed for component 1, if single-component adsorption took place, and the second term represents correction to the amount adsorbed due to competition from  $q_2$ . The difference between the two terms produces the real solidloading for component 1,  $q_1$ . Combining equations 5-11 and 5-14, and solving for  $q_1$ , we get

$$q_{1} = \frac{Q_{1}b_{1}C_{1}}{1 + b_{1}C_{1} + \frac{Q_{1}C_{2}}{Q_{2}C_{1}\alpha}}$$
(5-15)

which we call a Langmuir-type equation. Equations 5-11 and 5-15 can be used to calculate  $q_1$  and  $q_2$  if A is the dominantly adsorbed component and  $\alpha$  is known from two-component adsorption data.

Similarly, if component B is the dominantly adsorbed component, we have

$$q_2 = b_2 C_2 (Q_2 - q_2) - \frac{q_1}{f}$$
 (5-16)

From equations 5-11 and 5-16, we also get

$$q_{2} = \frac{Q_{2}b_{2}C_{2}}{1 + b_{2}C_{2} + \frac{Q_{2}C_{1}\alpha}{Q_{1}C_{2}}}$$
(5-17)

which is a Langmuir-type equation, too. Equations 5-11 and 5-17 can also be used to calculate  $q_2$  and  $q_1$  if B is the dominantly adsorbed component and  $\alpha$  is known. The ratio of  $q_1$  and  $q_2$ , that is, which component is dominantly adsorbed, may be estimated

from the initial and equilibrium concentrations of components.

If  $\alpha$  is unknown, however, we can not use the above equations for calculation. From the mass balance, we have

$$q_1 = \frac{C_{1,0} - C_1}{M}$$
,  $q_2 = \frac{C_{2,0} - C_2}{M}$  (5-18)

where M is the carbon dosage, and  $C_{1,0}$  and  $C_{2,0}$  are the initial concentrations of components A and B, respectively. From equation 5-18, we have

$$\frac{q_1}{q_2} = \frac{C_{1,0} - C_1}{C_{2,0} - C_2} \tag{5-19}$$

Substitution of equation 5-19 into equation 5-11 leads to

$$\frac{(C_{1,0} - C_1)C_2}{(C_{2,0} - C_2)C_1} = \alpha$$

Hence, equations 5-15 and 5-17 change to:

$$q_{1} = \frac{Q_{1}b_{1}C_{1}}{1 + b_{1}C_{1} + \frac{Q_{1}}{Q_{2}}\frac{C_{2,0} - C_{2}}{C_{1,0} - C_{1}}}$$
(5-20)

$$q_{2} = \frac{Q_{2}b_{2}C_{2}}{1 + b_{2}C_{2} + \frac{Q_{2}}{Q_{1}}\frac{C_{1,0} - C_{1}}{C_{2,0} - C_{2}}}$$
(5-21)

In practice, we can use equations 5-19 and 5-20 or 5-19 and 5-21 for calculations of two-component adsorption equilibria. The conversion factor, f, which is incorporated in equations 5-15, 5-17, 5-20 and 5-21, can be  $Q_1b_1C_1(1 + b_2C_2)/\{Q_2b_2C_2(1 + b_1C_1)\}$ , instead of  $Q_1/Q_2$ . But this will increase the complexity of calculations, even though sometimes it may improve the prediction results. This prediction method can also be readily extended to three- or more component system.

For three-component adsorption, we have the mass balance equations 5-19 and

$$\frac{q_1}{q_3} = \frac{C_{1,0} - C_1}{C_{3,0} - C_3} \tag{5-22}$$

Assume component 1 is the dominant adsorbed component. According to the law of mass action and the mass balance, we have

$$q_1 = b_1 C_1 (Q_1 - q_1) - \frac{Q_1}{Q_2} q_2 - \frac{Q_1}{Q_3} q_3$$
 (5-23)

From equations 5-19, 5-22 and 5-23, we have

$$q_{1} = \frac{Q_{1}b_{1}C_{1}}{1 + b_{1}C_{1} + \frac{Q_{1}}{Q_{2}}\frac{C_{2,0} - C_{2}}{C_{1,0} - C_{1}} + \frac{Q_{1}}{Q_{3}}\frac{C_{3,0} - C_{3}}{C_{1,0} - C_{1}}}$$
(5-24)

When component 1 is the dominant adsorbed component, equations 5-19, 5-22 and 5-24 can be used to calculate  $q_1$ ,  $q_2$  and  $q_3$ .

If component 2 is the dominant adsorbed component, we can use equation 5-19 and the following for calculations:

$$\frac{q_2}{q_3} = \frac{C_{2,0} - C_2}{C_{3,0} - C_3}$$
(5-25)

$$q_{2} = \frac{Q_{2}b_{2}C_{2}}{1 + b_{2}C_{2} + \frac{Q_{2}}{Q_{1}}\frac{C_{1,0} - C_{1}}{C_{2,0} - C_{2}} + \frac{Q_{2}}{Q_{3}}\frac{C_{3,0} - C_{3}}{C_{2,0} - C_{2}}}$$
(5-26)

If component 3 is the dominant adsorbed component, equations 5-22, 5-25 and the following can be used:

$$q_{3} = \frac{Q_{3}b_{3}C_{3}}{1 + b_{3}C_{3} + \frac{Q_{3}}{Q_{1}}\frac{C_{1,0} - C_{1}}{C_{3,0} - C_{3}} + \frac{Q_{3}}{Q_{2}}\frac{C_{2,0} - C_{2}}{C_{3,0} - C_{3}}}$$
(5-27)

### Modification of IAS Model

Because of the influence of non-ideal competition and irreversibility of multicomponent adsorption, the spreading pressures of the components may not be equal. To take this into account, we multiply either side of the spreading pressure equivalence equation by a factor F for the two component system:

$$F \int_{0}^{C_{1}^{0}} q_{1}^{0} \frac{dC_{1}^{0}}{C_{1}^{0}} = \int_{0}^{C_{2}^{0}} q_{2}^{0} \frac{dC_{2}^{0}}{C_{2}^{0}}$$
(5-28)

where  $C_i^0$  is the liquid-phase concentration of species *i* in single-component systems which gives the same spreading pressure as that of the mixture and  $q_i^0$ , the solid-phase loading corresponding to  $C_i^0$ .

If the relations between  $q_i$  and  $C_i$  can be expressed by the Langmuir isotherm, i.e.,  $q_1 = Q_1 b_1 C_1 / (1 + b_1 C_1)$  and  $q_2 = Q_2 b_2 C_2 / (1 + b_2 C_2)$ , integration of equation 5-28 gives

$$F Q_1 \ln(1 + b_1 C_1^0) = Q_2 \ln(1 + b_2 C_2^0)$$
 (5-29)

F can be determined by testing whether equation 5-11,  $q_1C_2/(q_2C_1) = \alpha$ , if  $\alpha$  is known, or equation 5-19,  $q_1/q_2 = (C_{0,1} - C_1)/(C_{0,2} - C_2)$ , if  $\alpha$  is unknown, is satisfied.

The other equations required for calculation are

$$C_i(\Pi, T, Z_i) = Z_i C_i^0(\Pi, T)$$
 (2-13)



Figure 5.1 Flow diagram of computations for modified IAS model

$$\frac{1}{q_T} = \sum_{i}^{n} \frac{Z_i}{q_i^0} \tag{2-18}$$

 $\sum_{i}^{n} Z_{i} = 1$  (2-19)

$$\boldsymbol{q_i} = \boldsymbol{Z_i} \boldsymbol{q_T} \tag{2-20}$$

This modified IAS method can be solved by implementing a Newton-Raphson algorithm. The flow diagram of the computation is given in Figure 5-1. It can also be extended to three- or more component system, but the complexity of computations may increase considerably.

#### **Results and Discussion**

Multicomponent Adsorption Equilibria at pH 7. Two-component adsorption equilibrium data for PCP/2,4,6-TCP, 2,4,6-TCP/2,4-DCP and 2,4-DCP/4-CP systems of different initial concentration ratios, together with the prediction results of the various models, are given in Tables 5.1, 5.2 and 5.3, respectively. Typical experimental data and prediction results of various models are also shown in Figures 5.2 - 5.5. To reduce systematic errors, single-component Langmuir isotherms were measured each time twocomponent adsorption was conducted for each system, and only data within the

concentration range from which most two-component adsorption equilibrium data are obtained were used for extraction of the isotherms constants. The data in Table 5.1 and 5.2 show that the predictions of the Langmuir competitive and IAS models are close to each other, and except for 2,4,6-TCP of case 1 (with  $C_{0,TCP} = 0.822$  mmol/l and  $C_{0,DCP} =$ 0.247 mmol/l) in Table 5.2, for which both are close to the experimental values with sums of relative deviations of 73.3% and 81.1%, respectively, both deviate from the experimental values to relatively large extents. The predictions of 2,4,6-TCP adsorption from the PCP/2,4,6-TCP and 2,4,6-TCP/2,4-DCP systems are always higher than the respective observed values, and those of PCP and 2,4-DCP adsorption from these two systems are always lower than the observed values. These facts imply that the Langmuir competitive and IAS models predict that the adsorbate more favorably adsorbed in singlecomponent solutions also adsorbs to a greater extent when in competition with another compound. But this is true only when adsorption is reversible and competition for adsorption sites is ideal. The ideal competition implies that the adsorbent is homogeneous with respective to adsorption sites and that the sites are equally accessible. However, activated carbon cannot be considered homogeneous because of its extensive microporous structure and the occurrence of different functional groups on the activated carbon surface. An ideal competition assumption, therefore, is invalid. Many researchers (Grant and King, 1990; Yonge et al., 1985; Snoeyink et al., 1969) have also shown that the adsorption of some compounds, such as phenolics, are highly irreversible. Our experimental results of adsorption and desorption equilibria for the four compounds revealed that PCP and 2,4,6-TCP are adsorbed on activated carbon much more irreversibly than 2,4-DCP and 4-CP.
This may explain why the Langmuir competitive and IAS models failed to accurately predict solid-phase loadings in PCP/2,4,6-TCP and 2,4,6-TCP/2,4-DCP solutions and their predictions are significantly improved for the 2,4-DCP/4-CP system (Table 5.3).

It is noteworthy that the new method significantly improves the predictions for the PCP/2,4,6-TCP and 2,4,6-TCP/2,4-DCP systems. The relative deviations between the predicted values of the new method and the experimental values are much lower than those for the Langmuir competitive and IAS models. In all situations equilibrium adsorbate concentrations are very small compared to the initial concentrations. Since in cases 1 and 2 of Table 5.1 the initial PCP concentrations are higher than 2,4,6-TCP and a higher solid-phase loading can be expected for PCP than 2,4,6-TCP, we apply the Langmuir-type equation to PCP. For the same reason, we use the Langmuir-type equation to predict the adsorption of 2,4,6-TCP in the last case (with  $C_{0PCP} = 0.205$  mmol/l and  $C_{0,TCP}$  = 0.645 mmol/l). Similarly, for the 2,4,6-TCP/2,4-DCP system, we apply the Langmuir-type equation to 2,4,6-TCP in case 1 and to 2,4-DCP in cases 2 and 3. For most points of cases 1 and 3 in the 2,4,6-TCP/2,4-DCP system, the relative deviations are within 10%. It may be concluded from this fact that the new prediction method is the most successful in predicting the adsorption from a two-component system in which one of two components is dominantly adsorbed. It is evident that the improvements made by the new method must be attributed to the thermodynamic condition (equation 5-11) which is imposed on the two-component adsorption equilibria. Additionally, except for some situations, such as case 1 of PCP/2,4,6-TCP system, the  $\alpha$  (distribution coefficient) values are fairly constant within each case, even though they change to some extent from case

## Table 5.1 Two-Component (PCP/2,4,6-TCP) Adsorption\*

 $Q_{PCP} = 1.3607 \text{ mmol/g}$   $b_{PCP} = 581.43 \text{ l/mmol}$ 

(Concentration range: 0.000904 - 0.01324 mmo1/1)

 $Q_{TCP} = 2.4035 \text{ mmol/g}$   $b_{TCP} = 1793.4 \text{ l/mmol}$ 

(Concentration range: 0.000276 - 0.01437 mmo1/1)

Case 1.  $C_{0,PCP} = 0.580 \text{ mmol/l}$   $C_{0,TCP} = 0.138 \text{ mmol/l}$ 

Pentachlorophenol

			Langmuir		IAS Model		Equation 5-20		Modi	fied IAS	Model	
Ceg. PCP	Ceg. TCP	gobad	q -	R	q	R	α	q	R	F	q	R
µmo1/1	µmo1/1	mmo1/g	mmol/g	육	mmol/g	8		mmol/g	æ		mmol/g	÷
0.989	0.109	0.39Ź	0.442	12.8	0.417	6.38	0.465	0.458	16.8	2.302	0.509	29.8
1.058	0.135	0.469	0.450	4.05	0.420	10.4	0.539	0.478	1.92	2.540	0.533	13.6
1.297	0,208	0,529	0.482	8.88	0.435	17.8	0.675	0.543	2.65	2.866	0.598	13.0
1.638	0.229	0.715	0.548	23.4	0.492	31.2	0.588	0.621	13.1	2.589	0.655	8.39
1.877	0.459	0.804	0.510	36.6	0.416	48.3	1.027	0.667	17.0	3.437	0.714	11.2
7.168	2.067	1.020	0.639	37.4	0.366	64.1	1.215	1.070	4.90	2.926	1.019	0.00
14.47	4.185	1.127	0.677	39.9	0.298	73.6	1,222	1.199	6.39	2.680	1.103	2.13
sum of R,	¥			163		252			62.8			78.1

#### 2,4,6-Trichlorophenol

			Lang	muir	IAS M	odel	Equation	5-19	Modified IAS	Mode1
Cag, TCP	Ceq, PCP	qobad	q	R	q	R	q	R	a	R
µmo1/1	µmo1/1	mmo1/g	mmo1/g	÷	mmol/g	÷	mmo1/g	÷	mmol/a	*
0.109	0.989	0.093	0.267	187	0.294	216	0.109	17.2	0.121	30.1
0.135	1.058	0.111	0.315	184	0.349	214	0.114	2.70	0.127	14.4
0.208	1.297	0.126	0.422	235	0.479	280	0.129	2.38	0.143	13.5
0.229	1.638	0.145	0.419	189	0.487	236	0.148	2.07	0.159	9.66
0.459	1.877	0.191	0.679	255	0.800	319	0.159	16.8	0.170	11.0
2.067	7.168	0.242	1.004	315	1.421	487	0.254	4.96	0.242	0.00
4.185	14.47	0.267	1.066	299	1.678	528	0.284	6.37	0,261	2.25
sum of R,	8			1664		2280		52.5		80.9

## Table 5.1 (continued)

# Case 2. $C_{0, PCP} = 0.478 \text{ mmo1/1}$ $C_{0, TCP} = 0.276 \text{ mmo1/1}$

#### Pentachloropheno1

C C ~		Langmuir		IAS Model		Equation 5-20		1 5-20	Modi	fied IAS	Model	
C <sub>eq, PCP</sub>	Ceq, TCP	qobsd	q	R	q	R	α	q	R	F	a	R
$\mu$ mol/1	μmo1/1	mmo1/g	mmo1/g	÷	mmo1/g	8		mmol/a	8		$mmo1/\alpha$	*
2.205	1.735	0.734	0.323	56.0	0.176	76.0	1.364	0.669	8.86	3.595	0.732	0.27
3.618	3.237	0.792	0.321	59.5	0.132	83.3	1.554	0.835	5.43	3.414	0.826	4.29
3.891	3.412	0.844	0.328	61.1	0.132	84.4	1.523	0.858	1.66	3.345	0.837	0.83
9.079	8.426	0.921	0.336	63.5	0.081	91.2	1.624	1.088	18.1	2.971	0.937	1.74
11.98	8.932	0.976	0.395	43.0	0.096	90.2	1.299	1.143	17.1	2.708	0.956	2.05
16.35	12.71	1.036	0.388	59.5	0.075	92.8	1.361	1.194	15.3	2.650	0.975	5.89
sum of R	, 8			343		518			66.5			15.1

#### 2,4,6-Trichlorophenol

			Lang	muir	IAS Mode	91	Equation	5-19	Modified IAS	Mode1
Ceq, TCP	Ceq, PCP	qobad	q	R	q	R	q	R	q	R
μmö1/1	µmo1/1	mmo1/g	mmol/g	₹	mmol/g	÷	mmo1/g	¥	mmo1/g	÷
1.735	2.205	0.422	1.387	229	1.604	280	0.386	8.53	0.423	0.24
3.237	3.618	0.455	1.566	244	1.863	309	0.481	5.71	0.476	4.62
3.412	3.891	0.485	1.568	223	1.878	287	0.494	1.86	0.482	0.62
8.426	9.079	0.522	1.698	225	2.124	307	0.622	19.2	0.536	2.68
8.932	11.98	0.554	1.605	190	2.107	280	0.656	18.4	0.549	0.90
12.71	16.35	0.586	1.645	181	2.178	272	0.682	16.4	0.557	5.12
sum of R,	8			1292	1	735		70.1		14.2

#### Table 5.1 (continued)

Case 3.  $C_{0,PCP} = 0.205 \text{ mmol/l}$   $C_{0,TCP} = 0.645 \text{ mmol/l}$ 

Pentachlorophenol

<b>a</b>			Langmuir		IAS Model			Equation	n 5-19	Modi	fied IAS	Mode1
Ceq, PCP	Ceq, TCP	qobsd	q	R	q	R	α	q	R	F	' a	R
$\mu$ mol/l	μmo1/1	mmo1/g	mmo1/g	Ł	mmo1/g	Ł		mmol/g	· 8		mmo1/q	÷
0.083	0.436	0.270	0.036	86.7	0.028	89.6	1.664	0.255	5.56	6.853	0.143	47.0
0.136	0.838	0.299	0.042	86.0	0.028	90.6	1.946	0.375	25.4	6.799	0.202	32.4
0.209	1.105	0.331	0.054	83.7	0.032	90.3	1.674	0.427	29.0	5.557	0.250	24.5
0.248	2.085	0.372	0.040	89.2	0.018	95.2	2.668	0.540	45.2	6.711	0.283	23.9
0.583	4.521	0.387	0.049	87.3	0.015	96.1	2.471	0.643	66.1	4.956	0.372	3.88
0.887	7.781	0.403	0.045	88.8	0.010	97.5	2.807	0.692	71.7	4.595	0.408	1.24
1.088	9.439	0.429	0.046	89.3	0.009	97.9	2.780	0.706	64.6	4.339	0.421	1.86
sum of R,	¥			611		657			308			135

#### 2,4,6-Trichlorophenol

•			Langm	uir	IAS M	odel	Equation	5-20	Modified IAS	Mode 1
C <sub>eq, TCP</sub>	Ceq, PCP	Gopad	q	R	q	R	q	R	q	R
μmö1/1	μmo1/1	mmo1/g	mmol/g	8	mmo1/g	₽;	mmol/g	*	mmo1/g	÷
0.436	0.083	0.850	1.027	20.8	1.036	21.9	0.802	5.65	0.451	46.9
0.838	0.136	0.940	1.399	48.8	1.418	50.9	1.179	25.4	0.637	32.2
1.105	0.209	1.042	1.535	47.3	1.564	50.1	1.344	29.0	0.786	24.6
2.085	0.248	1.168	1.840	57.5	1.873	60.4	1.695	45.1	0.889	23.9
4.521	0.583	1.213	2.063	70.1	2.118	74.6	2.015	66.1	1.168	3.71
7.781	0.887	1.261	2.168	71.9	2.227	76.6	2.161	71.4	1.275	1.11
9.439	1.088	1.337	2.192	63.9	2.255	68.7	2.200	64.5	1.315	1.65
sum of R,	÷			380		403		307		134

<sup>a</sup>  $C_{eq}$  = equilibrium concentration,  $C_0$  = initial concentration,  $q_{obsd}$ = observed amount adsorbed, Langmuir = Langmuir Competitive Model (equation 2-9), R = relative deviation ( $|q_{obsd} - q_{pred}|/q_{obsd}$ ),  $\alpha$  = distribution coefficient ( $q_{PCP}C_{TCP}/(q_{TCP}C_{PCP})$ ), F = Modified factor (on the PCP side of equation 5-22)

#### Table 5.2 Two-Component (2,4,6-TCP/2,4-DCP) Adsorption<sup>\*</sup>

 $Q_{TCP} = 2.1079 \text{ mmol/g}$   $b_{TCP} = 2564.4 \text{ l/mmol}$ 

(Concentration range: 0.000143 - 0.00838 mmol/1)

 $Q_{DCP} = 1.8959 \text{ mmol/g}$   $b_{DCP} = 2313.4 \text{ l/mmol}$ 

(Concentration range: 0.000371 - 0.00873 mmol/l)

Case 1.  $C_{0, TCP} = 0.822 \text{ mmol/l}$   $C_{0, DCP} = 0.247 \text{ mmol/l}$ 

2,4,6-Trichlorophenol

			Langmuir		IAS Model		Equation 5-20		Modified IAS	Model	
Cer. TCP	Cer. DCP	gobad	q	R	q	R	α	q	R	р	R
$\mu$ mol/l	$\mu$ mol/l	mmol/g	mmol/g	ક્ર	mmol/g	*		mmol/g	ક	mmol/g	8
0.243	0.043	0.652	0.762	16.9	0.766	17.5	0.600	0.671	2.97	0.630	3.29
0.426	0.045	0.782	1.048	33.9	1.053	34.6	0.358	0.949	21.3	0.758	3.07
1.129	0.114	1.379	1.467	6.43	1.478	7.23	0.336	1.443	4.67	1.113	19.3
1.300	0.129	1.498	1.517	1.23	1.529	2.08	0.331	1.505	0.48	1.155	22.9
1.962	0.185	1.561	1.642	5.15	1.658	6.17	0.313	1.666	6.70	1.263	19.1
2.318	0.194	1.647	1.694	2.91	1.711	3.90	0.279	1.721	4.53	1.288	21.8
4.310	0.377	1.730	1.802	4.19	1.826	5.57	0.290	1.881	8.71	1.410	18.5
5.503	0.495	1.784	1.830	2.56	1.857	4.09	0.298	1.926	7.94	1.445	19.0
sum of R,	, <del>%</del>			73.3		81.1			57.3		127

2,4-Dichlorophenol

			Langmuir	IAS Model		Equation 5-19		Modified IAS		Model	
C	C	Gabed	q	R	q	R	q	R	F	q	R
		mmol/g	mmol/g	÷	mmol/q	¥	mmol/g	₹		mmol/g	8
0 043	0.243	0.196	0.112	43.0	0.106	45.7	0.202	3.08	1.844	0.190	3.25
0 045	0.426	0.235	0.092	61.1	0.086	63.5	0.286	21.3	2.644	0.228	3.14
0.045	1,129	0.415	0.121	71.0	0.110	73.6	0.435	4.63	2.282	0.335	19.3
0 129	1.300	0.451	0.123	72.8	0.111	75.5	0.453	0.46	2.247	0.348	23.0
0.125	1 962	0.470	0.126	73.3	0.111	76.4	0.502	6.78	2.146	0.380	19.1
0.103	2 318	0.497	0.115	76.8	0.100	79.8	0.518	4.47	2.236	0.388	21.9
0.127	4 310	0 522	0.128	75.5	0.106	79.7	0.568	8.77	1.948	0.426	18.4
0.495	5.503	0.539	0.134	75.2	0.109	79.9	0.582	7.95	1.851	0.437	19.0
sum of R,	ક્ર			549		574		57.5			127

## Table 5.2 (continued)

# Case 2. $C_{0,TCP} = 0.513 \text{ mmol/l}$ $C_{0,DCP} = 0.618 \text{ mmol/l}$

2,4,6-Trichlorophenol

			Langmu	ıir	IAS Model			Equation	5-19	Modified IAS	Model
C <sub>eq, TCP</sub>	Ceg, DCP	<b>q</b> obsd	q	R	q	R	α	q	R	q	R
$\mu$ mol/l	$\mu$ mol/l	mmol/g	mmol/g	÷	mmol/g	8		mmol/g	웅	mmol/g	8
0.852	0.467	0.770	1.079	40.1	1.113	44.4	0.455	0.602	21.9	0.653	15.2
3.414	1.445	0.887	1.409	58.9	1.481	66.9	0.350	1.030	16.2	0.806	9.16
3.974	1.504	0.979	1.464	49.5	1.536	56.8	0.313	1.043	6.52	0.810	17.3
6.087	2.206	1.025	1.515	47.9	1.600	56.2	0.298	1.164	13.6	0.835	18.5
9.545	3.463	1.078	1.541	43.0	1.641	52.3	0.297	1.277	18.5	0.854	20.8
sum of R,	¥			239		277			76.6		80.9

2,4-Dichlorophenol

		Langmuir		IAS Model		Equation	n 5-20	Modif	ied IAS	Model	
Ceq, DCP	Ceq, TCP	qobsd	q	R	q,	R	q	R	F	q	R
μmo1/1	$\mu mo1/1$	mmo1/g	mmo1/g	*	mmo1/g	*	mmo1/g	*6		mmo1/g	*
0.467	0.852	0.929	0.481	48.3	0.449	51.7	0.725	21.9	1.795	0.788	15.2
1.445	3.414	1.073	0.484	54.9	0.418	61.0	1.246	16.2	1.708	0.974	9.16
1.504	3.974	1.186	0.450	62.1	0.384	67.7	1.263	6.48	1.760	0.981	17.3
2.206	6.087	1.245	0.446	64.2	0.368	70.4	1.414	13.6	1.702	1.014	18.5
3.463	9.545	1.315	0.454	65.5	0.362	72.5	1.558	18.5	1.625	1.042	20.8
sum of R,	£			295		323		76.6			81.0

#### Table 5.2 (continued)

Case 3.  $C_{0, TCP} = 0.206 \text{ mmol/l}$   $C_{0, DCP} = 0.989 \text{ mmol/l}$ 

2,4,6-Trichlorophenol

			Langmu	iir	IAS Mo	del		Equation	5-19	Modified IAS	Mode1
C <sub>eq, TCP</sub>	Ceq, DCP	q <sub>obsd</sub>	q	R	q	R	α	q	R	q	R
$\mu$ mo1/1	µmo1/1	mmo1/g	mmo1/g	8	mmo1/g	8		mmo1/g	8	mmo1/q	ૠ
0.101	0.448	0.191	0.239	25.3	0.247	29.4	0.916	0.184	3.73	0.187	2.21
0.108	0.579	0.235	0.224	4.75	0.234	0.65	1.109	0.209	11.2	0.205	12.8
0.203	1.091	0.270	0.271	0.39	0.291	7.77	1.117	0.268	0.87	0.250	7.51
0.450	2.129	0.306	0.344	12.5	0.381	24.6	0.981	0.317	3.77	0.285	6.82
0.793	3.504	0.345	0.385	11.5	0.439	27.1	0.917	0.343	0.54	0.302	12.7
1.870	7.138	0.381	0.453	19.0	0.539	41.5	0.791	0.367	3.70	0.316	17.0
2.444	9.026	0.402	0.469	16.7	0.566	40.8	0.765	0.372	7.62	0.319	20.7
sum of R,	8			90.1		172			31.4		79.7

2,4-Dichlorophenol

0			Langmuir		IAS Model		Equation	n 5-20	Modi	fied IAS	Mode1
Ceq, DCP	Ceq, TCP	qobsd	q	R	q	R	q	R	F	a	R
$\mu$ mol/l	µmo1/1	mmo1/g	mmo1/g	8	mmo1/g	*	mmo1/g	÷		mmol/a	8
0.448	0.101	0.920	0.856	6.90	0.847	7.90	0.885	3.80	1.258	0.899	2.30
0.579	0.108	1.133	0.971	14.3	0.960	15.3	1.006	11.2	1.108	0.987	12.9
1.091	0.203	1.300	1.183	9.00	1.164	10.5	1.290	0.82	1.107	1.202	7.54
2.129	0.450	1.474	1.319	10.5	1.284	12.9	1.528	3.67	1.171	1.371	6.98
3.504	0.793	1.664	1.379	17.1	1.330	20.1	1.654	0.60	1.193	1.453	12.7
7.138	1.870	1.836	1.403	23.6	1.325	27.8	1.769	3.66	1.230	1.525	16.9
9.026	2.444	1.941	1.406	27.5	1.318	32.1	1.794	7.57	1.234	1.540	20.7
sum of R,	8			109		127		31.4			80.0

•  $C_{eq}$  = equilibrium concentration,  $C_0$  = initial concentration,  $q_{obsd}$ = observed amount adsorbed, Langmuir = Langmuir Competitive Model (equation 2-9), R = relative deviation ( $|q_{obsd} - q_{pred}|/q_{obsd}$ ),  $\alpha$  = distribution coefficient ( $q_{rcP}C_{pcP}/(q_{pcP}C_{TCP})$ ), F = Modified factor (on the DCP side of equation 5-22)

## Table 5.3 Two-Component (2,4-DCP/4-CP) Adsorption<sup>a</sup>

 $Q_{DCP} = 1.9640 \text{ mmol/g}$   $b_{DCP} = 1898.01 \text{ l/mmol}$ 

(Concentration range: 0.000371 - 0.0155 mmol/1)

 $Q_{CP} = 2.0677 \text{ mmol/g}$   $b_{CP} = 191.67 \text{ l/mmol}$ 

(Concentration range: 0.00206 - 0.0725 mmol/1)

Case 1.  $C_{0, DCP} = 0.740 \text{ mmol/l}$   $C_{0, CP} = 0.248 \text{ mmol/l}$ 

#### 2,4-Dichlorophenol

			Langm	Langmuir		IAS Model		Equatio	on 5-20 Modif		fied IAS Model	
C <sub>eq,DCP</sub> µmo1/1	C <sub>eq.CP</sub> µmol/1	g <sub>obsd</sub> mmol/g	q mmol/g	R १	q mmol/g	R %	α	q mmol/g	R %	F	q mmol/a	R %
0.802	3.403	1.048	0.942	10.1	0.932	11.1	12.84	1.054	0.58	1.224	1.019	2.82
1.256	5.161	1.206	1.071	11.2	1.056	12.4	12.52	1.267	5.08	1.186	1.151	4.52
3.020	11.71	1.397	1.254	10.2	1.227	12.1	12.10	1.600	14.5	1.140	1.337	4.32
6.784	23.27	1.629	1.379	15.3	1.341	17.7	11.21	1.785	9.59	1.095	1.438	11.7
13.86	41.84	1.827	1.463	19.9	1.415	22.5	10.65	1.874	2.58	1.073	1.503	17.7
sum of R,	8			66.7		75.8			32.3			41.1

#### 4-Chlorophenol

			Langmu	ıir	IAS M	odel	Equation	n 5-19	Modified IAS	Model
C <sub>eq,CP</sub> µmol/1	C <sub>eq,DCP</sub> µmol/1	q <sub>obsd</sub> mmol/g	q mmol/g	R %	q mmol/g	R St	q mmol/g	R %	g mmol/g	R %
3.403	0.802	0.346	0.425	22.6	0.435	25.7	0.348	0.60	0.337	2.65
5.161	1.256	0.396	0.468	18.2	0.483	22.0	0.416	5.12	0.379	4.33
11.71	3.020	0.447	0.517	15.5	0.545	21.8	0.512	14.5	0.428	4.26
23.27	6.784	0.498	0.503	0.91	0.543	8.98	0.546	9.66	0.440	11.6
41.84	13.86	0.518	0.469	9.36	0.520	0.35	0.531	2.59	0.427	17.5
sum of R,	8			66.6		78.8		32.5		40.3

# Table 5.3 (continued)

Case 2.  $C_{0, DCP} = 0.493 \text{ mmol/l}$   $C_{0, CP} = 0.495 \text{ mmol/l}$ 

2,4-Dichlorophenol

			Langmuir		IAS Model			Equation 5-20			Modified IAS Mod		
C <sub>eq,DCP</sub> Wmo1/1	C <sub>eq,CP</sub> Mmo1/1	g <sub>obsd</sub> mmol/g	q mmol/q	R %	q mmol/g	R *	α	q mmol/q	R %	F	g mmol/a	R *	
0.405	3.875	0.646	0.602	6.79	0.594	8.04	9.586	0.557	13.7	1.029	0.607	6.02	
0.896 1.374	8.678 12.87	$0.819 \\ 0.909$	0.766 0.844	6.50 7.23	0.748 0.819	8.68	9.806 9.555	0.919 1 129	12.2 24 2	1.046 1 034	0.781	4.68	
3.864	33.81	1.112	0.972	12.6	0.928	16.6	9.295	1.561	40.3	1.030	0.967	13.1	
7.020	61.87	1.249	1.000	19.9	0.941	24.6	9.904	1.725	38.2	1.053	1.024	18.0	
sum of R,	8			50.0		67.9			129			48.4	

4-Ch]	lorop	henol
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_	-		Langmu	ıir	IAS M	odel	Equatio	n 5-19	Modified IAS	Model
С <sub>ед, СР</sub> µmol/1 3.875	$C_{eq, DCP}$ $\mu mol/1$ 0.405	$\frac{q_{obsd}}{mmol/g}$ 0.643	q mmol/g 0.611	R % 5.02	q mmo1/g 0.619	R % 3.73	q mmol/g 0.555	R % 13.8	q mmol/g	R % 5 91
8.678 12.87 33.81 61.87	0.896 1.374 3.864 7.020	0.808 0.891 1.047 1.111	0.788 0.839 0.904 0.936	2.55 5.79 13.6 15.7	0.806 0.865 0.951 0.997	0.27 2.93 9.19 10.2	0.907 1.106 1.470 1.535	12.1 24.1 40.3 38.2	0.772 0.834 0.910 0.911	4.49 6.41 13.1 18.0
sum of R,	8			42.7		26.3		129		47.8

### Table 5.3 (continued)

#### Case 3. $C_{0,DCP} = 0.239 \text{ mmol/l}$ $C_{0,CP} = 0.750 \text{ mmol/l}$

2,4-Dichlorophenol

			Langm	uir	IAS M	odel		Equatio	n 5-19	Mod:	ified IAS	Model
Ceq, DCP	Ceq, CP	Qobsd	q	R	đ	R	α	q	R	F	a	R
$\mu$ mo1/1	$\mu$ mo1/1	mmol/g	mmol/g	8	mmol/g	8		mmol/g	8		mmol/g	8
0.251	7.855	0.318	0.315	1.01	0.307	3.49	10.03	0.352	10.7	1.059	0.330	3.71
0.672	20.50	0.459	0.404	12.0	0.386	15.9	9.967	0.503	9.59	1.054	0.421	8.28
1.209	34.34	0.518	0.457	11.8	0.430	17.0	9.439	0.570	10.0	1.033	0.457	11.7
3.020	64.94	0.570	0.587	2.98	0.544	4.56	7.410	0.642	12.6	0.961	0.495	13.2
sum of R,	ê			27.8		41.0			42.9			36.9

#### 4-Chlorophenol

C <sub>eq,CP</sub> μmo1/1 7.855 20.50 34.34	C <sub>eq,DCP</sub> µmo1/1 0.251 0.672	<b>Q</b> obed mmol/g 0.989 1.403	Lang q mmo1/g 1.043 1.309	muir R \$ 5.49 6.70	IAS Mod q mmol/g 1.052 6 1.328 5	el R % .32 .35	Equation q mmol/g 1.094 1.541	n 5-20 R % 10.7 9.84	Modified IAS q mmol/g 1.028 1.292	Model R % 3.89 7.91
64.94 sum of R,	3.020 %	1.655	1.342	18.9 42.7	1.387 1	.,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	1.864	10.1 12.6 43.2	1.377	11.6 13.2 36.6

<sup>a</sup>  $C_{eq}$  = equilibrium concentration,  $C_0$  = initial concentration,  $q_{obsd}$ = observed amount adsorbed, Langmuir = Langmuir Competitive Model (equation 2-9), R = relative deviation ( $|q_{obsd} - q_{pred}|/q_{obsd}$ ),  $\alpha$  = distribution coefficient ( $q_{DCP}C_{CP}/(q_{CP}C_{DCP})$ ), F = modified factor(on the DCP side of equation 5-20)

## Table 5.4 Three-Component (2,4,6-TCP/2,4-DCP/4-CP) Adsorption<sup>a</sup>

$Q_{\rm TCP} = 2.1079$	mmol/g b <sub>TCP</sub>	= 2564.4	l/mmol	(Concentration	range:	0.000143	- 0.00838	mmol/l)
$Q_{DCP} = 1.8959$	mmol/g b <sub>DCP</sub>	= 2313.4	1/mmol	(Concentration	range:	0.000371	- 0.00873	mmol/l)
$Q_{\rm CP} = 2.0677$ m	mmol/g b <sub>cp</sub>	= 191.67	l/mmol	(Concentration	range:	0.00206 -	0.0725 m	mo1/1)

Case 1.  $C_{0, TCP} = 0.165 \text{ mmol/l}$   $C_{0, DCP} = 0.615 \text{ mmol/l}$   $C_{0, CP} = 0.257 \text{ mmol/l}$ 

2,4,6-Trichlorophenol

				Langn	wir	IAS MO	del	Equation 5-		
C <sub>eq,TCP</sub> µmo1/1 0.147 0.304 1.043	C <sub>eq,DCP</sub> µmol/1 0.506 0.871 2.927	C <sub>eq,CP</sub> µmo1/1 1.401 2.669 12.06	$q_{obsd}$ mmol/g 0.215 0.251 0.327	q mmol/g 0.282 0.382 0.442	R % 31.1 52.0 35.1	q mmo1/g 0.294 0.404 0.493	R % 36.7 61.0 50.8	q mmol/g 0.213 0.281 0.409	R % 0.94 12.0 25.1	
2.107	5.682	24.51	0.359	0.470	30.8	0.540	50.4	0.451	25.5	
sum of R,	8				149		198.9		63.5	

#### 2,4-Dichlorophenol

				Langn	nuir	IAS Mo	od <b>el</b>	Equation	5-26
C <sub>eq,DCP</sub> µmol/1	C <sub>eq,TCP</sub> µmo1/1	C <sub>eq,CP</sub> µmol/1	g <sub>obsd</sub> mmol/g	q mmol/g	R %	q mmol/g	R %	q mmol/g	R %
0.506	0.147	1.401	0.804	0.788	1.92	0.771	4.10	0.795	1.12
2.927	1.043	12.06	1.225	1.006	17.8	0.933	23.8	1.532	25.1
5.682	2.107	24.51	1.347	1.028	23.7	0.926	31.3	1.691	25.5
sum of R	, 8				48.8		67.9		63.7

## Table 5.4 (continued)

#### 4-Chlorophenol

c	<u> </u>	<u> </u>		Langn	uir	IAS Model		Equation	5-25	
μmol/1 1.401 2.669 12.06 24.51	μmol/1 0.147 0.304 1.043 2.107	μmol/1 0.506 0.871 2.927 5.682	q <sub>obed</sub> mmol/g 0.334 0.388 0.490 0.514	q mmol/g 0.197 0.246 0.375 0.401	R % 41.1 36.8 23.6 22.1	q mmol/g 0.203 0.256 0.403 0.441	R % 39.2 34.0 17.8 14.2	q mmol/g 0.331 0.435 0.613 0.645	R % 0.90 12.0 25.1 25.5	
sum of R,	8				123.6		105.2		63.5	

 $C_{0,CP} = 0.771 \text{ mmol/l}$ 

Case 2.  $C_{0,TCP} = 0.137 \text{ mmol/l}$   $C_{0,DCP} = 0.128 \text{ mmol/l}$ 

2,4,6-Trichlorophenol

_	-	- ·		Langn	uir	IAS MO	del	Equation	5-22	
C <sub>eq,TCP</sub> µmol/1 0.0113 0.202 0.679 1.109 1.656	C <sub>eq,DCP</sub> µmo1/1 0.142 0.232 0.681 1.289 1.595	C <sub>eq,C</sub> ₽ µmol/1 5.353 9.725 31.59 42.01 65.98	Q <sub>obed</sub> mm01/g 0.181 0.209 0.268 0.293 0.325	q mmol/g 0.0256 0.278 0.354 0.403 0.415	R 85.8 33.0 32.2 37.7 27.8	q mmol/g 0.0261 0.284 0.371 0.429 0.443	R 85.6 35.9 38.4 46.4 36.3	q mmol/g 0.159 0.215 0.311 0.330 0.358	R % 12.1 2.81 16.3 12.6 10.3	
sum of R,	ક				216.5		242.6		54.1	

2,4-Dichlorophenol

c	0	•		Langn	uir	IAS MO	del	Equation	5-25
μmol/1 0.142 0.232 0.681 1.289 1.595	μmol/1 0.0113 0.202 0.679 1.109 1.656	C.,cp µmo1/1 5.353 9.725 31.59 42.01 65.98	<b>Q</b> obsd mm01/g 0.169 0.250 0.273 0.303	q mmo1/g 0.262 0.260 0.288 0.380 0.324	R 8 55.4 32.9 15.2 39.2 6.86	q mmol/g 0.253 0.242 0.254 0.329 0.272	R % 24.1 1.60 20.5 10.2	q mmol/g 0.149 0.201 0.291 0.308 0.335	R % 11.7 2.81 16.3 12.6 10.3
sum of R,	ક				149.6		106.1		53.7

#### Table 5.4 (continued)

4-Chlorophenol

C	C	C	-	Langm	uir	IAS MO	del	Equation	5-27
μmol/1 5.353 9,725 31.59 42.01 65.98	μmol/1 0.0113 0.202 0.679 1.109 1.656	μmol/1 0.142 0.232 0.681 1.289 1.595	Gobsd mm01/g 1.008 1.163 1.451 1.569 1.690	q mmol/g 0.890 0.984 1.207 1.119 1.212	R % 11.8 15.4 16.8 28.7 28.3	q mm01/g 0.899 0.996 1.227 1.148 1.240	R % 10.8 14.4 15.4 26.8 26.6	q mmol/g 0.890 1.196 1.686 1.767 1.864	R 8 2.79 16.2 12.6 10.3
sum of R,	ક				101		94		53.7

<sup>a</sup>  $C_{eq}$  = equilibrium concentration,  $C_0$  = initial concentration,  $q_{obsd}$ = observed amount adsorbed, Langmuir = Langmuir Competitive Model (equation 2-9), R = relative deviation ( $|q_{obsd} - q_{pred}|/q_{obsd}$ )















Figure 5.5 Adsorption equilibria of TCP in the presence of PCP
(Initial concentrations: PCP, 0.478 mmol/l; TCP, 0.276 mmol/l)
■ Experimental data; + Langmuir; > IAS; > New method; × Modified IAS



Figure 5.6 Langmuir isotherms of TCP in the presence of DCP

to case. For a wastewater, if the concentration ratio of two contaminants does not change remarkably and the  $\alpha$  value is prior determined, we can use the Langmuir-type equations 5-15 or 5-17 without resorting to the initial concentrations of two components. The simplicity and higher accuracy may make this method very attractive.

The modified IAS model also significantly improves the accuracy of the predictions for two-component adsorption. The relative deviations are relatively low compared to those of the Langmuir competitive and IAS models. In some cases, such as cases 2 and 3 of PCP/2,4,6-TCP system and case 2 of 2,4-DCP/4-CP system, it is better than the new method. This may be explained by that the modified IAS model uses all the single-component isotherm constants of both components while the new method only uses three isotherm constants. The *F* values are fairly constant within each case, even though they change to some extent from case to case. As mentioned above, we can use the modified IAS model to predict the two-component adsorption equilibria without knowing the initial concentrations of two components if  $\alpha$  or *F* is known. Values of *F* greater than 1 are indications of the influence of irreversibility and non-ideal competition of two-component adsorption.

Table 5.3 shows that the Langmuir competitive and IAS models predict the 2,4-DCP/4-CP two component adsorption equilibria very well. The reason for this may be that the adsorption of 2,4-DCP and 4-CP on activated carbon is fairly reversible. The new method predicts the solid loadings better than the Langmuir competitive and IAS models in case 1 and worse than these two models in case 2, while the predictions of these three models for case 3 are very close. Responsible for higher deviations of the predictions of the new method in case 2 may be much higher concentration range of 2,4-DCP singlecomponent isotherm data than that of 2,4-DCP two-component equilibrium data. The modified IAS model improves the accuracy of the predictions for all cases, and thus is the most successful among the four models tested for this system. The values of the modification factor F are very close to 1, which indicates that the competition between 2,4-DCP and 4-CP is basically ideal.

Table 5.4 shows three component adsorption equilibrium data for the 2,4,6-TCP/2,4-DCP/4-CP system. Similar to two component systems, the performances of the Langmuir competitive and IAS models are essentially identical for the three-component system. The relative deviations between the predicted values of the new method and experimental values are relatively low in comparison with those for the Langmuir competitive and IAS models. The proposed new method appears to be the best of the three models. This implies that the new method based on thermodynamic consideration of two-component adsorption equilibria can be successfully extended to three-component system.

The important thing to note is that even though the new method and modified IAS model presented in this thesis use initial concentrations in addition to single-component isotherm constants, they still require fewer parameters than the widely used IAS modification shown in equation 2-22, in which the modification factors,  $R_i$ , are determined from multicomponent equilibrium data with a statistical technique. From the mass balance expression  $q = (C_o - C)/M$ , one can see that to calculate the solid-phase loading q, carbon dosage M must be used in addition to initial concentration  $C_o$ .

For multicomponent adsorption conducted with the same two-component solution but varying carbon dosages, the equilibrium data of each component conform to singlecomponent adsorption isotherms. The equilibrium data of each component in all cases of three systems can be described by single-component Langmuir equation. From Figure 5.6 which is just such an example, it can be seen that when 2,4,6-TCP is the dominantly adsorbed component, the Langmuir capacity is almost the same as single-component adsorption capacity but the adsorption constant becomes smaller, and if 2,4-DCP is the dominant adsorbed component both capacity and constant for 2,4,6-TCP change. This can be explained by equations 5-11, 5-15 and 5-17. For the case in which 2,4,6-TCP is the dominant adsorbed component, we use the Langmuir-type equation (eqn 5-15) to predict 2,4,6-TCP adsorption equilibrium. The term  $Q_1C_2/(Q_2C_1\alpha)$ , which is equivalent to  $Q_1 q_2 / (Q_2 q_1)$  or  $Q_1 (C_{0,2} - C_2) / (Q_2 (C_{0,1} - C_1))$ , in equation 5-15, is almost constant. Hence the Langmuir-type equation can be changed to the single-component Langmuir equation, and Q remains the same but b decreases. For the case in which 2,4-DCP is the dominant adsorbed component, equation 5-11 is used to predict the 2,4,6-TCP adsorption equilibrium. Substitution of equation 5-17 into equation 5-11 and rearrangement lead to

$$q_{1} = \frac{Q_{2}b_{2}\alpha C_{1}}{1 + \frac{Q_{2}q_{1}}{Q_{1}q_{2}} + b_{2}\alpha \frac{q_{2}}{q_{1}}C_{1}}$$
(5-30)

which can also be treated as a Langmuir equation because the terms  $\alpha$ ,  $1 + Q_2 q_1/(Q_1 q_2)$ and  $b_2 \alpha q_2/q_1$  are basically constant. In this case, both Q and b change: Q decreases and b may decrease or increase.

*Multicomponent Adsorption Equilibria at Various pH Values*. Two-component adsorption was conducted for the 2,4,5-TCP/2,4-DCP system at various pH values. Figure 5.7 gives 2,4,5-TCP and 2,4-DCP equilibrium adsorption data and the surface complexation model fits, and Table 5.5 summarizes the surface complexation constants and overall variance. For convergence of optimization procedure, only two complexation constants ( $K_A^{intr}$  for 2,4-DCP and  $K_B^{intr}$  for 2,4,5-TCP) were extracted using FITEQL, whereas the other two complexation constants ( $K_A^{intr}$  for 2,4,5-TCP and 2,4-DCP adsorption data follow the same patterns as in single-component adsorption, although the presence of competing species decreases the respective solid-phase loadings. The constants provide good simulation of 2,4-DCP

Co	mplexation Constant	S	
Adsorbate	log K <sub>A</sub> <sup>intr</sup>	$\log K_{B}^{intr}$	SOS/DF
2,4,5-TCP	21.48 <sup>a</sup>	14.58	40.2
2,4-DCP	24.12	13.05ª	42.5
		·	

 Table 5.5
 Best Estimates for 2,4,5-TCP/2,4-DCP Two-Component Surface

 Complexation Constants

<sup>a</sup> from Table 3.5

Adsorbate	$\log K_A^{intr}$	$\log K_B^{intr}$	SOS/DF	
2,4,5-TCP	24.86	12.14	2.32	
2,4-DCP	20.34	15.05	0.99	

 Table 5.6
 Best Estimates for Conditional Surface Complexation Constants

 (2,4,5-TCP/2,4-DCP System)

adsorption but underestimate below pH 4.5 and overestimate within pH 7.4 - 8.2 2,4,5-TCP adsorption with about 10% deviations. Compared to the four multicomponent adsorption models tested in this chapter, the Langmuir competitive model, the IAS model, the proposed new method, and the modified IAS model, the surface complexation model performs very well in predicting two-component adsorption equilibria of chlorophenols/activated carbon system.

As shown in Figure 5.8, 2,4,5-TCP and 2,4-DCP adsorption data can separately excellently be fitted by the surface complexation model. Since the adsorption equilibria of each component were attained in the presence of the other component, the constants extracted from optimization procedure are called "conditional surface complexation constants". Table 5.6 gives the conditional surface complexation constants, which are somewhat different in magnitude from those of single-component adsorption. In effect, the conditional surface complexation constants are not supposed to be equal to the surface complexation constants obtained from single-component adsorption data.



Figure 5.7 2,4,5-TCP/2,4-DCP two-component adsorption as a function of pH and optimum surface complexation model fits



Figure 5.8 2,4,5-TCP/2,4-DCP two-component adsorption as a function of pH and separate SCM fits to each component adsorption data

## Conclusions

Two- and three- component adsorption equilibria of chlorophenols on granular activated carbon were investigated in the micromolar equilibrium concentration range. The Langmuir competitive and Ideal Adsorbed Solution (IAS) models were tested for their performances on the three binary systems of pentachlorophenol/2,4,6-trichlorophenol, 2,4,6-trichlorophenol/2,4-dichlorophenol, and 2,4-dichlorophenol/4-chlorophenol, and the tertiary system of 2,4,6-trichlorophenol/2,4-dichlorophenol/4-chlorophenol, and found to fail to predict the two-component adsorption equilibria of the former two binary systems and the tertiary system. A new prediction method and a modification of the IAS model, both based on thermodynamic considerations, have been proposed. The required parameters are the single-component Langmuir isotherm constants and initial concentrations for each component. The proposed new method and the modified IAS model were found to significantly improve the accuracy of the predictions of twocomponent adsorption equilibria of chlorophenols. The new method also performs much better than the Langmuir competitive and IAS models in the three-component system. The observation that the equilibrium data of each component in two-component systems can be described by single-component adsorption isotherms has also been explained by rearranging the equations of the new method. 2,4,5-TCP/2,4-DCP two-component adsorption equilibria at various pH values were described by the surface complexation model with as high as 10% deviation at some points, while the separate SCM fits to each component are in excellent agreement with the adsorption data.

## Chapter 6 DESORPTION EQUILIBRIUM AND KINETICS OF CHLOROPHENOLS

## Introduction

In recent years, the failures of multicomponent adsorption equilibrium models in predicting solid-phase loadings have been reported for a large number of activated carbonorganic compound systems (Thacker et al., 1984; Smith and Weber, 1988; Yonge and Keinath, 1986). This is because these models assume that both liquid- and solid-phase solutions are ideal and thus ideal competition occurs. Many systems, however, cannot be dealt with as ideal ones because of high liquid- and/or solid-phase adsorbate concentrations (Radke and Prausnitz, 1992) or strong interaction between adsorbates and activated carbon. The latter frequently bring about irreversibility of adsorption, which seems an important reason for deviations of multicomponent adsorption from ideal competition. Yonge et al. (1985) showed that the degree of irreversibility was as high as 85 - 97% for certain phenolic compounds and postulated high-energy bonding (chemisorption) to be the cause of strong irreversibility.

It has been shown in chapter 3 that chlorophenols can be favorably adsorbed by activated carbon and the adsorptive capacities are fairly high. Thus chlorophenols are compounds which have a very strong affinity with activated carbon. Chapter 5 has demonstrated that the Langmuir competitive and IAS models fail to accurately predict solid-phase loadings on activated carbon for most systems. These facts imply that the chlorophenols-activated carbon systems are not ideal and the irreversibility of adsorption must be taken into account. Investigations have been conducted on desorption equilibria and kinetics in order to gain a more complete understanding of single-component adsorption irreversibility. This chapter presents desorption equilibrium and kinetic data and proposes a desorption rate equation.

### Materials and Methods

Desorption Kinetics. Batch desorption kinetic experiments were conducted for PCP, 2,4-DCP and 4-CP. The experimental protocol consisted of equilibrating a 300 mg GAC sample at a initial liquid-phase adsorbate concentration of approximately 2 mM. A larger mass of GAC was used so as to minimize the effects of unintentional loss of GAC which could occur during the desorption procedure and to result in a detectable adsorbate concentration during the operation of desorption kinetics. Following equilibration, the liquid and solid phases were separated by using an all-glass Millipore filtration unit designed to hold 47-mm diameter disks. The GAC retained on the glass-fiber filters was rinsed back into the reactor with 3 liters of adsorbate free buffer solution agitated as previously described. 2 - 5 ml solution samples were withdrawn at appropriate intervals. Preliminary experiments were performed to determine if a significant quantity of GAC remained on the filters following this rinse procedure. This was done by drying the filters, following the rinse procedure, at 105°C to a constant weight and comparing this weight to the tare weight of the filters. Less than 3 mg of carbon was found to adhere to the filters.

Desorption Equilibrium. Single-solute desorption isotherms were developed to determine the extent of single-solute irreversible adsorption for the three compounds. The adsorption isotherm data were first generated by using the procedure described previously. Following equilibration, the solution was decanted and the GAC and tube wall were washed with deionized, double-distilled water. These processes were completed very carefully and quickly to avoid any loss of GAC and any detectable desorption. 40-ml portions of deionized, double-distilled water with 5 mM phosphate buffer and pH 7.0 were pipetted into the tubes containing equilibrated GAC. The tubes were then shaken for 6 days. From kinetic results, no considerable desorption was observed after 2 days. This indicates that a 6-day equilibration period was sufficient for the attainment of equilibrium in the desorption isotherm studies. The equilibrium solid-phase loading on the GAC was calculated after desorption using

$$q = \frac{q_0 m - CV}{m} \tag{6-1}$$

where  $q_0$  = equilibrium solid-phase loading after the initial adsorption step (moles per gram), m = mass of GAC in the tubes (grams), C = equilibrium liquid-phase adsorbate concentration after desorption (moles per liter), V = volume of solution (liters), and q = equilibrium solid-phase loading after desorption (moles per gram).

#### **Results and Discussion**

Desorption Equilibrium. The adsorption and desorption isotherms are shown in

Figures 6.1 - 6.3. The fact that adsorption isotherms all lie below the respective desorption isotherms implies the occurrence of irreversible adsorption, since the two isotherms should coincide for completely reversible adsorption. The Langmuir and Freundlich constants for these adsorption and desorption isotherms are tabulated in Table 6.1. The adsorptive capacities are seen to be lower than the desorption capacities for all these three compounds. The data listed in Table 6.2 summarize the relative degree of irreversibility for the three adsorbates. Percent irreversibility is defined as  $1 - C_d/C_a$ , where  $C_d$  and  $C_a$  represent desorption and adsorption equilibrium liquid-phase concentrations at the same solid-phase loading, respectively. The results from this definition are comparable between components at given adsorbate concentrations, although they are different from this concentration to the other. The data at 0.02 mmol/l adsorption equilibrium concentration were used for the calculation. These data indicate that the degree of irreversibility increases from 4-CP to PCP.

On the basis of the findings of Mattson et al. (1969a), which indicated the formation of charge-transfer complexes between adsorbates and activated carbon surface functional groups, chemisorption appears to be the most logical explanation for irreversible adsorption. Depending on the type of surface functional groups as well as the type of adsorbate, a sufficiently strong bond that resists desorption can be formed. The degree of irreversibility then would be directly related to the number of high-energy (chemisorption) bonds. That fraction of adsorption that was reversible would then likely be that fraction of adsorption that occurred as a result of van der Waals forces and/or weaker charge-transfer complexes that occur at other adsorption sites on the activated



Figure 6.1 Langmuir isotherms for adsorption and desorption of PCP



Figure 6.2 Langmuir isotherms for adsorption and desorption of 2,4-DCP



Figure 6.3 Langmuir isotherms for adsorption and desorption of 4-CP

			Langmuir			Freundlich			
Adsorbate		Conc. range 10 <sup>-3</sup> mmol/l	Q mmol/g	b l/mmol	R <sup>2</sup>	K	n	R <sup>2</sup>	
PCP	Ads <sup>a</sup> Des <sup>b</sup>	0.574-24.44 0.122-18.23	1.569 1.729	768.1 1678.3	0.999 0.997	4.622 4.886	0.271 0.236	0.845 0.925	
2,4-DCP	Ads Des	0.385-39.27 0.182-30.19	2.310 2.452	965.1 1664.0	0.999 0.999	4.053 3.870	0.164 0.125	0.950 0.994	
4-CP	Ads Des	2.06-50.00 3.16-37.00	1.977 2.234	226.4 298.0	0.997 0.999	4.369 4.744	0.270 0.238	0.986 0.972	

# Table 6.1Comparison of Langmuir and Freundlich Constants<br/>for Adsorption and Desorption of Three Compounds

\* Adsorption

<sup>b</sup> Desorption

# Table 6.2 Single-Solute Desorption Irreversibility

Adsorbate	Irreversibility, %				
PCP	83				
2,4-DCP	74				
4-CP	56				

carbon surface.

Grant and King (1990) found that most of the adsorption of phenol at pH 12.1 was irreversible and irreversible uptakes at pH 1.8 were much lower than those at higher pH values. They also found that irreversible adsorption of phenolic compounds on carbon surfaces was caused by oxidative coupling. From the pKa values of chlorophenols, it can be known that most species are present in ionized forms for PCP and in molecular forms for 4-CP. That phenolate ions were more reactive than phenols seems to be responsible for that the degree of irreversibility of adsorption increases from 4-CP to PCP.

The interaction of the aromatic ring with the surface of the activated carbon may give another plausible explanation for the change of the degree of adsorption irreversibility from 4-CP to PCP. It is well known that the electron density of an aromatic ring is strongly influenced by the nature of the substituent. A chloro group acts as a strong electron withdrawing group in reducing the overall electron density in the pisystem of the ring. Thus, chloro-substituted aromatic compounds act as acceptors in donor-acceptor complexes (i. e., charge-transfer complexes), and the aromatic compounds with more chloro groups form stronger complexes with a given donor than the compounds with fewer chloro groups. This may explain why the adsorption of PCP is most irreversible and that of 4-CP is least irreversible.

**Desorption Kinetics.** The desorption rate can be observed from the desorption kinetic data to decrease as desorption propagates and finally (approximately 3 - 4 days) becomes so small that equilibrium can be considered to be reached. The desorption of all three compounds followed a biphasic pattern: a fast desorption phase following by a slow

phase. The fast phase lasted only a few hours for 4-CP, about 1 day for 2,4-DCP, and 2 days for PCP. If the difference between actual solid-phase loading and solid-phase loading that is in equilibrium with the instantaneous liquid-phase concentration outside the particle is the driving force for desorption, the desorption rate equation can be expressed by

$$-\frac{dq}{dt} = k_d (q - \frac{Q_d b_d C}{1 + b_d C})$$
(6-2)

where  $k_d$  is the mass transfer coefficient;  $Q_d$  and  $b_d$ , the Langmuir constants for desorption isotherm; and C, the adsorbate concentration in the bulk liquid-phase. The term  $Q_d b_d C/(1+b_d C)$  represents the solid-phase loading in equilibrium with C.

Using the mass balance  $q = q_0 - C/M$ , where  $q_0$  is the initial solid-phase loading and M is the carbon dosage, equation (6-2) can be changed to

$$\frac{dC}{dt} = k_d \left( q_0 M - C - \frac{Q_d b_d CM}{1 + b_d C} \right)$$
(6-3)

Figures 6.4 - 6.6 show the typical desorption kinetic data and curves fitted by equation (6-2) for PCP, 2,4-DCP and 4-CP. Table 6.3 summarizes the parameters used for fitting, and the R-square values. The Langmuir isotherm constants for desorption are obtained from the desorption isotherms shown in Figures 6.1 - 6.3. It is evident that the desorption kinetic model (equation 6-2) can describe the desorption kinetic data very well. Noteworthy is the fact that 4-CP reached desorption equilibrium most rapidly while PCP slowest. The increases in desorption mass transfer coefficient  $k_d$  from PCP to 4-CP are


Figure 6.4 Changes of PCP concentration with time during desorption and predictions of desorption kinetic rate equation



Figure 6.5 Changes of 2,4-DCP concentration with time during desorption and predictions of desorption kinetic rate equation



Figure 6.6 Changes of 4-CP concentration with time during desorption and predictions of desorption kinetic rate equation

Adsorbate	Q₄ mmol/g	b <sub>d</sub> l/mmol	q₀ mmol/g	M g/l	k <sub>d</sub> 1/hr	R <sup>2</sup>
PCP	1.729	1678.3	1.781	0.1	0.087	0.999
2,4-DCP	2.452	1664.0	2.805	0.1	0.250	0.998
4-CP	2.234	298.0	2.336	0.1	0.270	1.000

 Table 6.3 Desorption Kinetic Parameters

consistent with this phenomenon. This order is identical with the one for the changes of adsorption mass transfer coefficient. It is not unexpected that the less chlorinated a chlorophenol, the faster it reaches both adsorption and desorption equilibria. The difference in the strength of the interaction between these three chlorophenols and activated carbon may be again responsible for the changes of desorption rates.

#### Conclusions

Studies were conducted on the desorption equilibria and kinetics of chlorophenols. The results indicate that the Langmuir equation fits both adsorption and desorption equilibrium data better than the Freundlich equation and the degree of the irreversibility of adsorption increases from 4-chlorophenol to pentachlorophenol. A desorption rate equation has been proposed which describes the desorption kinetics data very well. The desorption mass transfer coefficients were found to increase from pentachlorophenol to 4-chlorophenol. The more chlorinated a chlorophenol, the slower and more irreversible is its desorption process. The differences in the strength of interaction between chlorophenols and activated carbon may be responsible for the changes of adsorption irreversibility and desorption rate.

### Chapter 7 SUMMARY AND CONCLUSIONS

A number of single-component adsorption equilibrium models have been developed in the past decades, while the Langmuir and Freundlich equations can satisfactorily fit equilibrium data for most systems. Existent adsorption kinetics models, the external film diffusion model, internal surface diffusion model, linear-driving-force approximation, or surface reaction kinetics model, have been shown to successfully describe adsorption kinetics processes. Considerable achievements have also been made in prediction of multicomponent adsorption equilibria for simple systems, but it remains a very challenging task to accurately predict multicomponent adsorption equilibria under non-ideal competitive conditions. Little effort has been given to desorption mechanisms and kinetics, especially for systems like chlorophenols, which show a very high affinity for activated carbon.

Studies were undertaken of the adsorption of chlorinated phenols from aqueous solution on granular activated carbon. Single-component equilibrium adsorption data on the eight compounds in two concentration ranges at pH 7.0 fit the Langmuir equation better than the Freundlich equation, while the Freundlich equation exhibits improved performance in higher concentration ranges. There are pronounced differences in the magnitudes of the constants between the two concentration ranges, and thus the adsorption constants are concentration-dependent. The adsorptive capacities at pH 7.0 increase from

pentachlorophenol to trichlorophenols and are fairly constant from trichlorophenols to monochlorophenols. No effect of phosphate on the adsorption was observed in the pH range of 4 - 10. The presence of background dissolved organic matter could reduce the adsorptive capacity of chlorophenols. Equilibrium studies were performed at various temperatures at pH 7.0. The adsorption process was found to be exothermic for pentachlorophenol and 2,4,6-trichlorophenol, and endothermic for 2,4-dichlorophenol and 4-chlorophenol.

Equilibrium measurements were also conducted for 2,4,5-trichlorophenol, 2,4dichlorophenol, and 4-chlorophenol over a wide pH range. No subsequent influence on adsorption is observed for decreasing pH values below the compounds' pKa values, while marked decrease in amounts adsorbed occurs as pH increases above the pKa values. A surface complexation model (SCM) has been proposed to describe the effect of pH on adsorption equilibria of chlorophenols on activated carbon. Activated carbon surface functional sites are divided into acidic groups and basic groups which may be neutralized by bases or acids. Molecular and ionized forms of chlorophenols interact, respectively, with these two kinds of surface groups to form two neutral surface complexes, of which the complex of basic groups is more substantial and more stable than that of acidic groups. The simulations of the model are in excellent agreement with the experimental data.

Batch kinetics studies were conducted of the adsorption of chlorinated phenols on granular activated carbon. pH played an important role in the adsorption kinetics of chlorophenols at pH values above the pKa values of the compounds, while little influence on adsorption rate was observed if pH was decreased below the pKa values. The external film diffusion model, the linear-driving-force approximation, and the surface reaction kinetics model have been employed to fit the adsorption kinetics data of chlorophenols. The surface reaction model best describes both short-term and long-term kinetics, while the external film diffusion model describes the short-term kinetics data very well and the linear-driving-force approximation exhibits improved performance for the long-term kinetics. The interruption test also indicates that the rate of adsorption is not limited by internal surface diffusion. The mass transfer coefficient increases from more chlorinated compounds to less chlorinated compounds, which implies that the more chloro groups in a chlorophenol, the slower it reaches adsorption equilibrium. The two-component adsorption kinetic experiments have revealed that the adsorption of chlorophenols on activated carbon is to some extent irreversible and that non-ideal competition between two components exists.

Two- and three- component adsorption equilibria of chlorophenols on granular activated carbon were investigated in the micromolar equilibrium concentration range. The Langmuir competitive and Ideal Adsorbed Solution (IAS) models were tested for their performances on the three binary systems of pentachlorophenol/2,4,6-trichlorophenol, 2,4,6-trichlorophenol/2,4-dichlorophenol, and 2,4-dichlorophenol/4-chlorophenol, and the tertiary system of 2,4,6-trichlorophenol/2,4-dichlorophenol/2,4-dichlorophenol/4-chlorophenol, and found to fail to predict the two-component adsorption equilibria of the former two binary systems and the tertiary system. A new prediction method and a modification of the IAS model, both based on thermodynamic considerations, have been proposed. The required

parameters are the single-component Langmuir isotherm constants and initial concentrations for each component. The proposed new method and the modified IAS model were found to significantly improve the accuracy of the predictions of two-component adsorption equilibria of chlorophenols. The new method also performs much better than the Langmuir competitive and IAS models in the three-component system. 2,4,5-TCP/2,4-DCP two-component adsorption equilibria at various pH values were described by the surface complexation model with as high as 10% deviation at some points, and the separate SCM fits to each component are in excellent agreement with the adsorption data.

Studies were conducted on the desorption equilibria and kinetics of chlorophenols. The Langmuir equation fits both adsorption and desorption equilibrium data better than the Freundlich equation and the degree of the irreversibility of adsorption increases from 4-chlorophenol to pentachlorophenol. A linear-driving-force desorption rate equation has been proposed which describes the desorption kinetics data very well. The desorption mass transfer coefficients were found to increase from pentachlorophenol to 4chlorophenol. The more chlorinated a chlorophenol, the slower and more irreversible is its desorption process. The differences in the strength of interaction between chlorophenols and activated carbon may be responsible for the changes of adsorption irreversibility and desorption rate.

In summary, the following important conclusions can be drawn for the adsorption of chlorinated phenols on granular activated carbon:

(1) The Langmuir equation fits single-component equilibrium adsorption data

better than the Freundlich equation;

(2) The adsorptive capacities at pH 7.0 increase from PCP to TCP and are fairly constant from TCP to CP;

(3) The proposed surface complexation model describes very well the experimental data for the effect of pH on adsorption equilibria;

(4) The adsorption process is exothermic for PCP and 2,4,6-TCP, and endothermic for 2,4-DCP and 4-CP;

(5) The surface reaction model best describes kinetics adsorption data;

(6) The mass transfer coefficient increases from 2,4,6-TCP to 4-CP;

(7) The adsorption process is to some extent irreversible;

(8) Multicomponent adsorption is non-ideal, and the Langmuir competitive and

IAS models fail to predict the adsorption equilibria of PCP-TCP, TCP-DCP and PCP-TCP-CP systems;

(9) The proposed new method and the modified IAS model significantly improve the accuracy of the predictions of multicomponent adsorption equilibria;

(10) The Langmuir equation fits desorption equilibrium data better than the Freundlich equation;

(11) The degree of the irreversibility of adsorption increases from 4-CP to PCP;

(12) The proposed desorption rate equation describes desorption kinetics data very well;

(13) The desorption mass transfer coefficient increases from PCP to 4-CP.

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## APPENDICES

#### Appendix A Computer Program for IAS Model

```
REM
          THIS PROGRAM SOLVES IDEAL ADSORBED SOLUTION
   REM
         (IAS) MODEL FOR TWO COMPONENT SYSTEM.
   REM
         NEWTON-RAPHSON METHOD IS USED. LANGMUIR ISOTHERM
   REM
         PARAMETERS NEED TO BE INPUTTED.
   PRINT "HOW MANY DATA POINTS":
   INPUT N
   DIM Q(2), B(2), CO1(N), CO2(N), C1(N), C2(N), X01(N)
   DIM XO2(N), XI(N), X2(N), ZI(N), Z2(N)
   REM INPUT LANGMUIR ISOTHERM PARAMETERS FOR EACH OF
   REM COMPONENTS
   PRINT "ADSORPTIVE CAPACITY (COMPONENT 1) ";
   INPUT Q(1)
   PRINT "ADSORPTIVE CAPACITY (COMPONENT 2) ";
   INPUT Q(2)
   PRINT "CONSTANT B (COMPONENT 1)";
   INPUT B(1)
   PRINT "CONSTANT B (COMPONENT 2)";
   INPUT B(2)
   REM INPUT EXPERIMENTAL DATA POINTS (CONCENTRATIONS)
   PRINT "C1"
   FOR I = 1 TO N
       INPUT C1(I)
   NEXT I
   PRINT "C2"
   FOR I = 1 TO N
       INPUT C2(I)
   NEXT I
   REM COMPUTATION FOR IAS
   DEF FNA (X) = Q(1) * LOG(1 + B(1) * X) - Q(2)
       * LOG(1 + B(2) * X * C2(I) / (X - C1(I)))
   DEF FNB (X) = Q(1) * B(1) / (1 + B(1) * X) + Q(2) * B(2)
       * C1(I) * C2(I) / ((X + B(2) * C2(I) * X - C1(I)))
       * (X - C1(I))
   FOR I = 1 TO N
       X1 = C1(I) * 1.02
       X2 = X1 - FNA(X1) / FNB(X1)
10
       RESIDUE = ABS(X1 - X2)
       IF RESIDUE < .000001 THEN 20
       X1 = X2
       GOTO 10
20
       C01(I) = X2
       CO2(I) = CO1(I) * C2(I) / (CO1(I) - C1(I))
       Z1(I) = C1(I) / C01(I)
       Z2(I) = C2(I) / C02(I)
       X01(I) = Q(1) * B(1) * C01(I) / (1 + B(1) * C01(I))
       X02(I) = Q(2) * B(2) * C02(I) / (1 + B(2) * C02(I))
```

X1(I) = Z1(I) / (Z1(I) / X01(I) + Z2(I) / X02(I))X2(I) = Z2(I) / (Z1(I) / X01(I) + Z2(I) / X02(I))NEXT I REM OUTPUT COMPUTATIONAL RESULTS OPEN "A:DC1.IAS" FOR OUTPUT AS #1 PRINT TAB(5); "C1"; TAB(14); "C2"; TAB(22); "C01"; TAB(31); "C02"; TAB(41); "Z1"; TAB(50); "Z2"; TAB(58); "X1"; TAB(67); "X2" PRINT #1, TAB(5); "C1"; TAB(14); "C2"; TAB(22); "C01"; TAB(31); "C02"; TAB(41); "Z1"; TAB(50); "Z2"; TAB(58); "X1"; TAB(67); "X2" PRINT FOR I = 1 TO N PRINT USING "###.#### "; C1(I); C2(I); C01(I); C02(I); Z1(I); Z2(I); X1(I); X2(I) PRINT #1, USING "###.#### "; C1(I); C2(I); C01(I); CO2(I); Z1(I); Z2(I); X1(I); X2(I)NEXT I CLOSE #1 END

#### Appendix B Computer Program for Modified IAS Model

REM THIS PROGRAM SOLVES MODIFIED IDEAL ADSORBED SOLUTION (IAS) MODEL FOR TWO COMPONENT SYSTEM. REM NEWTON-RAPHSON METHOD IS USED. LANGMUIR ISOTHERM REM PARAMETERS AND INITIAL LIQUID-PHASE REM REM CONCENTRATIONS NEED TO BE INPUTTED. PRINT "HOW MANY DATA POINTS"; INPUT N DIM Q(2), B(2), CO1(N), CO2(N), C1(N), C2(N), XO1(N)DIM XO2(N), X1(N), X2(N), Z1(N), Z2(N), F(N)REM INPUT LANGMUIR ISOTHERM PARAMETERS FOR EACH OF REM OMPONENTS PRINT "ADSORPTIVE CAPACITY (COMPONENT 1)"; INPUT O(1)PRINT "ADSORPTIVE CAPACITY (COMPONENT 2) "; INPUT O(2)PRINT "CONSTANT B (COMPONENT 1)": INPUT B(1) PRINT "CONSTANT B (COMPONENT 2)"; INPUT B(2)REM INPUT EXPERIMENTAL DATA POINTS (CONCENTRATIONS) PRINT "INITIAL CONCENTRATION (COMPONENT 1)"; INPUT CI1 PRINT "INITIAL CONCENTRATION (COMPONENT 2)"; INPUT CI2 PRINT "C1" FOR I = 1 TO N INPUT C1(I) NEXT I PRINT "C2" FOR I = 1 TO N INPUT C2(I) NEXT I REM COMPUTATION FOR MODIFIED IAS DEF FNA (X) = Q(1) \* LOG(1 + B(1) \* X) - F(1) \* O(2)\*  $LOG(1 + \tilde{B}(2) * X * C2(I) / (X - C1(I)))$ DEF FNB (X) = Q(1) \* B(1) / (1 + B(1) \* X) + F(I) \* Q(2)\* B(2) \* C1(I) \* C2(I) / ((X + B(2) \* C2(I) \* X))- C1(I) + (X - C1(I))FOR I = 1 TO N F(I) = 1 $\mathbf{J} = \mathbf{0}$ W2 = 0a = (CI1 - C1(I)) \* C2(I) / ((CI2 - C2(I)) \* C1(I))X1 = C1(I) \* 1.0210 X2 = X1 - FNA(X1) / FNB(X1)RESIDUE = ABS(X1 - X2)

```
IF RESIDUE < .0000001 THEN 20
       X1 = X2
       GOTO 10
20
       C01(I) = X2
       CO2(I) = CO1(I) * C2(I) / (CO1(I) - C1(I))
       Z1(I) = C1(I) / C01(I)
       Z2(I) = C2(I) / C02(I)
       XO1(I) = Q(1) * B(1) * CO1(I) / (1 + B(1) * CO1(I))
       X02(I) = Q(2) * B(2) * C02(I) / (1 + B(2) * C02(I))
X1(I) = Z1(I) / (Z1(I) / X01(I) + Z2(I) / X02(I))
       X2(I) = Z2(I) / (Z1(I) / X01(I) + Z2(I) / X02(I))
       K = X1(I) * C2(I) / (X2(I) * C1(I))
       IF ABS(a / K - 1) < .005 THEN 30
       W = ABS(a / K - 1)
       J = J + 1
       IF J = 1 THEN W1 = W
       IF J = 2 THEN W2 = W
       IF W_2 > W_1 THEN 25
       F(I) = F(I) + .001
       PRINT F(I)
       GOTO 5
25
       F(I) = F(I) - .001
       PRINT F(I)
       GOTO 5
30 NEXT I
   REM OUTPUT COMPUTATIONAL RESULTS
   OPEN "A:DC1.MIA" FOR OUTPUT AS #1
   PRINT TAB(5); "C1"; TAB(13); "C2"; TAB(20); "C01";
         TAB(28); "C02"; TAB(37); "Z1"; TAB(45); "Z2";
         TAB(52); "X1"; TAB(60); "X2"; TAB(68); "F"
   PRINT #1, TAB(5); "C1"; TAB(13); "C2"; TAB(20); "C01";
         TAB(28); "C02"; TAB(37); "Z1"; TAB(45); "Z2";
         TAB(52); "X1"; TAB(60); "X2"; TAB(69); "F"
   PRINT
   FOR I = 1 TO N
       PRINT USING "###.#### "; C1(I); C2(I); C01(I);
             CO2(I); Z1(I); Z2(I); X1(I); X2(I); F(I)
       PRINT #1, USING "###.#### "; C1(I); C2(I); C01(I);
             CO2(I); Z1(I); Z2(I); X1(I); X2(I); F(I)
  NEXT I
   CLOSE #1
   END
```

00004 00006 00160 00011 00001 00008	1 1 1 40 3 -3.0 -3.0 -1.0 0.0 0.0 0.0	1.00 1.00 0.00 0.00 0.00 0.00	1 E-3 E00 E00 E00	HL B P: X( H- C	A SI ClP + LP-	2		:	1		
00001 00002 00003 00004 00005 00006 00007 00008 00009 00010	$\begin{array}{c} 0.0 \\ -14.0 \\ 7.78 \\ 0.0 \\ 5.7 \\ 0.0 \\ 6.94 \\ 0.0 \\ 10.0 \\ 15.0 \end{array}$	01 04 04 06 06 01 08 04	01 -1 01 01 01 01 01 01	160 160 08 01 160	01 01 01 02 -1	01 01 08 01	01 01 01 01	11 08	01 01	11	01
1.2	941.0 2 009 010 004 006 12	0.5	2			2		Ċ	)		
0.0013 0.0013 0.0013 0.0012 0.00008 0.000	011 52 50 42 44 93 37 41 004 39 78 38 58 001								-		

-3.763 -3.758 -3.738 -3.742 -3.634 -3.541	008
-3.283 -3.271 -3.189	
-3.196 -3.183	
	00

## INPUT DATA FOR VERIFICATION

#### CHEMICAL EQUILIBRIUM PROBLEM

ID	х	LOGX	T	GROUP	NAME
4	1.00E-03	-3.00	1.00E-03	I -T	HA
6	1.00E-03	-3.00	1.00E-03	I -T	в
160	1.00E-01	-1.00	0.00E-01	I —T	PSI
11	1.00E+00	0.00	0.00E-01	II -T,X	XClP
1	1.00E+00	0.00	0.00E-01	III-X	H+
8	1.00E+00	0.00	0.00E-01	III-X	ClP-

ID	LOGK	4	6	160	11	1	8
1	0.00	0.00	0.00	0.00	0.00	1.00	0.00
2	-14.00	0.00	0.00	0.00	0.00	-1.00	0.00
3	7.78	1.00	0.00	1.00	0.00	1.00	0.00
4	0.00	1.00	0.00	0.00	0.00	0.00	0.00
5	5.70	0.00	1.00	1.00	0.00	1.00	0.00
6	0.00	0.00	1.00	0.00	0.00	0.00	0.00
7	6.94	0.00	0.00	0.00	0.00	1.00	1.00
8	0.00	0.00	0.00	0.00	0.00	0.00	1.00
9	10.00	1.00	0.00	0.00	1.00	2.00	1.00
10	15.00	0.00	1.00	-1.00	1.00	1.00	1.00

SURFACE PROPERTIES: C(F/M\*\*2) = 1.200 S(M\*\*2/G) = 941.0A(G/L) = 0.5

## PARAMETERS FOR OPTIMIZATION:

LOG K	FOR SPECIES:	9	10
T FOR	COMPONENTS:	4	· 6

INPUT DATA FOR VERIFICATION

SERIAL DATA AND ESTIMATED STANDARD DEVATION

TOTAL CONCENTRATION FOR COMPONENT:

	11
1	1.352E-03
2	1.350E-03
3	1.342E-03
4	1.344E-03
5	1.293E-03
6	1.237E-03
7	1.141E-03
8	1.004E-03
9	9.890E-04
10	8.780E-04
11	8.880E-04
12	8.680E-04

LOG FREE CONCENTRATION FOR COMPONENT:

	_	_
	1	8
1	-4.19	-3.76
2	-5.86	-3.76
3	-6.11	-3.74
4	-6.35	-3.74
5	-6.85	-3.63
6	-7.57	-3.54
7	-8.04	-3.42
8	-9.19	-3.28
9	-9.21	-3.27
10	-9.98	-3.19
11	-10.00	-3.20
12	-10.28	-3.18

STANDARD DEVIATION OF FREE CONCENTRATION:

COMPONENT	1	RELATIVE 0.0230	ABSOLUTE	0.000E-01					
COMPONENT	8	RELATIVE 0.0230	ABSOLUTE	0.000E-01					
STANDARD DEVIATION OF TOTAL CONCENTRATION:									
COMPONENT	11	RELATIVE 0.0100	ABSOLUTE	1.000E-06					

VALUES OF ADJUSTABLE PARAMETERS AT EACH ITERATION: LOG K, T, LOG X

I	V(Y): SOS/DF	9	10	4	6
0 1 2 3 4 5 6	5.950E+02 1.300E+02 3.306E+02 1.547E+02 1.724E+02 6.074E+01 9.540E+01	1.000E+01 1.461E+01 1.851E+01 1.929E+01 2.013E+01 2.138E+01 2.227E+01	1.500E+01 1.572E+01 1.603E+01 1.503E+01 1.544E+01 1.572E+01 1.581E+01	1.000E-03 1.000E-04 1.000E-05 4.080E-04 6.284E-05 8.342E-05 4.494E-04	1.000E-03 1.281E-03 1.884E-03 1.292E-03 1.324E-03 1.339E-03 8.342E-04
7	4.594E+01	2.127E+01	1.634E+01	4.622E-04	8.642E-04
8	2.532E+00 2.291E+00	2.142E+01 2.146E+01	1.654E+01 1.660E+01	4.403E-04 4.413E-04	9.320E-04 9.301E-04
10 11	2.283E+00 2.282E+00	2.147E+01 2.148E+01	1.663E+01	4.425E-04 4.429E-04	9.277E-04
12	2.282E+00	2.148E+01	1.664E+01	4.430E-04	9.265E-04
13	2.282E+00	2.148E+01	1.664E+01	4.431E-04	9.264E-04
15	2.283E+00	2.148E+01	1.665E+01	4.431E-04	9.264E-04
****	OPTIMIZATION	PROCEDURE C	ONVERGED		
15	2.283E+00	2.148E+01	1.665E+01	4.431E-04	9.264E-04

DESCRIPTION OF CHEMICAL EQUILIBRIUM: ELECTROSTATICS

	T(SIGMA)	LOG X(PSI)	SIGMA(C/M**2)	PSI(V)
1	-9.055E-04	2.616	-0.1857	-0.1547
2	-8.949E-04	2.585	-0.1835	-0.1529
3	-8.887E-04	2.568	-0.1823	-0.1519
4	-8.779E-04	2.536	-0.1800	-0.1500
5	-8.520E-04	2.461	-0.1747	-0.1456
6	-7.819E-04	2.259	-0.1603	-0.1336
7	-7.345E-04	2.122	-0.1506	-0.1255
8	-6.429E-04	1.857	-0.1319	-0.1099
9	-6.442E-04	1.861	-0.1321	-0.1101
10	-7.104E-04	2.052	-0.1457	-0.1214
11	-7.117E-04	2.056	-0.1460	-0.1216
12	-7.282E-04	2.104	-0.1493	-0.1244

#### DESCRIPTION OF CHEMICAL EQUILIBRIUM: -LOG C

	1	2	3	4	5	6	7	8	9	10
1	4.19	9.81	6.48	12.69	4.99	9.11	1.01	3.76	3.35	3.04
2	5.86	8.14	4.86	9.37	5.05	7.48	2.68	3.76	3.37	3.04
3	6.11	7.89	4.66	8.89	5.11	7.27	2.91	3.74	3.38	3.04
4	6.35	7.65	4.46	8.42	5.17	7.05	3.15	3.74	3.39	3.04
5	6.85	7.15	4.18	7.57	5.42	6.74	3.54	3.63	3.42	3.04
6	7.57	6.43	3.85	6.32	5.92	6.31	4.17	3.54	3.52	3.03
7	8.04	5.96	3.72	5.58	6.32	6.10	4.52	3.42	3.60	3.03
8	9.19	4.81	3.55	4.00	6.98	5.35	5.53	3.28	4.19	3.04
9	9.21	4.79	3.56	3.99	6.99	5.34	5.54	3.27	4.20	3.04
10	9.98	4.02	3.74	3.59	6.70	4.47	6.23	3.19	5.26	3.05
11	10.00	4.00	3.75	3.59	6.69	4.44	6.26	3.20	5.30	3.05
12	10.28	3.72	3.90	3.50	6.62	4.15	6.52	3.18	5.77	3.07

## DESCRIPTION OF CHEMICAL EQUILIBRIUM: C (MOL/L)

	1	2	3	4	5	6
1	6.457E-05	1.549E-10	3.295E-07	2.051E-13	1.027E-05	7.683E-10
2	1.380E-06	7.244E-09	1.377E-05	4.301E-10	8.829E-06	3.316E-08
3	7.762E-07	1.288E-08	2.201E-05	1.274E-09	7.777E-06	5.411E-08
4	4.467E-07	2.239E-08	3.482E-05	3.765E-09	6.798E-06	8.837E-08
5	1.413E-07	7.079E-08	6.665E-05	2.707E-08	3.770E-06	1.841E-07
6	2.692E-08	3.715E-07	1.416E-04	4.812E-07	1.200E-06	4.902E-07
7	9.120E-09	1.096E-06	1.902E-04	2.614E-06	4.791E-07	7.919E-07
8	6.457E-10	1.549E-05	2.787E-04	9.949E-05	1.040E-07	4.464E-06
9	6.166E-10	1.622E-05	2.774E-04	1.028E-04	1.028E-07	4.584E-06
10	1.047E-10	9.550E-05	1.820E-04	2.557E-04	1.990E-07	3.362E-05
11	1.000E-10	1.000E-04	1.782E-04	2.599E-04	2.053E-07	3.600E-05
12	5.248E-11	1.905E-04	1.264E-04	3.150E-04	2.381E-07	7.131E-05

7 8 9 10 1 9.705E-02 1.726E-04 4.428E-04 9.161E-04 2 2.099E-03 1.746E-04 4.293E-04 9.175E-04 3 1.236E-03 1.828E-04 4.211E-04 9.185E-04 4 7.047E-04 1.811E-04 4.083E-04 9.195E-04 5 2.858E-04 2.323E-04 3.764E-04 9.225E-04 6 6.745E-05 2.877E-04 3.010E-04 9.247E-04 7 3.048E-05 3.837E-04 2.503E-04 9.251E-04 8 2.931E-06 5.212E-04 6.487E-05 9.218E-04 9 2.877E-06 5.358E-04 6.286E-05 9.217E-04 10 5.902E-07 6.471E-04 5.444E-06 8.926E-04 11 5.546E-07 6.368E-04 4.967E-06 8.902E-04 12 2.999E-07 6.561E-04 1.708E-06 8.548E-04? Appendix D Single-Component Adsorption Equilibrium Data

Effect of Phosphate at Various pH Values

Initial 2,4,5-TCP concentration: 301 mg/l Carbon dosage: 20 mg/40 ml

phosphate (5 mM)			no phosphate		
рĦ	q mmol/g	Ce mmol/l	q mmol/g	Ce mmol/l	
4.19			2.704	0.1724	
5.86	2.699	0.1747			
6.11			2.683	0.1829	
6.35	2.687	0.1812			
6.85	2.585	0.2324			
7.57			2.473	0.288	
8.04	2.281	0.3839			
9.19	2.007	0.521			
9.21			1.978	0.5353	
9.98	1.755	0.6468			
10			1.776	0.6364	
10.28			1.736	0.6562	

# Effect of pH on Adsorption of 2,4-DCP and 4-CP

2,4-DCP

4-CP

PH	q mmol/g	Ce mmol/l	q mmol/g	Ce mmol/l
3.6	2.93	0.824		
3.71	2.93	0.8459		
3.86			3.237	2.115
4.01	2.924	0.6887		
4.17			3.297	2.031
7.18	2.862	0.5432		
7.77			3.263	2.011
7.82			3.301	1.95
9.25			3.07	1.945
9.51	2.199	0.9261		
10.39	1.879	0.9958		
10.46			2.562	2.047
10.48			2.549	2.037
10.61	1.769	0.9292		
10.67	1.757	1.02		
11			2.237	2.074
11.11			2.258	2.036
12.07			1.786	2.112

#### PCP Isotherm Data at pH 7.0

Ce mg/l	q mg/g	Ce mmol/l	q mmol/g	Ce/q	LOG Ce	LOG q
0.225	201.5	0.000843	0.755726	0.001115	-3.07417	-0.12163
0.379	230.5	0.001423	0.864614	0.001645	-2.84678	-0.06317
0.811	267	0.003041	1.009001	0.003013	-2.51698	0.003891
1.67	317.3	0.00628	1.190252	0.005276	-2.20204	0.075638
3.34	354.5	0.012527	1.32986	0.009419	-1.90215	0.123805
3.53	389.6	0.013249	1.406583	0.009419	-1.87781	0.148165
6.51	415.9	0.024441	1.501535	0.016277	-1.61188	0.176535
13.55	457.7	0.050869	1.652446	0.030784	-1.29354	0.218127
21.35	488.7	0.080149	1.764366	0.045426	-1.09610	0.246588
36.12	514.4	0.135603	1.857152	0.073016	-0.86773	0.268847
55.5	524.16	0.208388	1.892389	0.110119	-0.68112	0.277010

Langmuir Constants

Regression	Output:
Constant	0.000696
Std Err of Y Est	0.000181
R Squared	0.997846
No. of Observations	5
Degrees of Freedom	3

X Coefficient(s) 0.704563 Std Err of Coef. 0.018895

> Q = 1.419318 b = 1012.303

## Regression Output:

Constant	0.003775
Std Err of Y Est	0.000830
R Squared	0.999620
No. of Observations	6
Degrees of Freedom	4

X Coefficient(s) 0.511778 Std Err of Coef. 0.004983

> Q = 1.953968 b = 135.5526

## Freundlich Constants

Regressio Constant Std Err of Y Est R Squared No. of Observation Degrees of Freedom	on Output: ns m	0.531087 0.007609 0.995633 5 3
X Coefficient(s) Std Err of Coef.	0.210244 0.008038 K = n =	3.396933 0.210244

Regression	Output:
Constant	0.363306
Std Err of Y Est	0.006653
R Squared	0.986742
No. of Observations	6
Degrees of Freedom	4

X Coefficient(s) 0.113697 Std Err of Coef. 0.006589

K	=	2.308373
n	=	0.113697

2,3,4,6-TeCP Isotherm Data at pH 7.0

Langmuir Constants

Regression Output:Constant0.000236Std Err of Y Est0.000081R Squared0.997740No. of Observations5Degrees of Freedom3

X Coefficient(s) 0.678595 Std Err of Coef. 0.018646

> Q = 1.473632 b = 2874.171

## Regression Output:

NCGTCDDTOH	oucpuc.
Constant	0.004933
Std Err of Y Est	0.004186
R Squared	0.998096
No. of Observations	7
Degrees of Freedom	5

X Coefficient(s) 0.452336 Std Err of Coef. 0.008834

> Q = 2.210741 b = 91.68653

## Freundlich Constants

Regression Constant Std Err of Y Est R Squared No. of Observations Degrees of Freedom	Output	: 0.597243 0.024490 0.956126 5 3
X Coefficient(s) 0 Std Err of Coef. 0	.195332 .024157	
	K = n =	3.955879 0.195332
Regression Constant Std Err of Y Est R Squared No. of Observations Degrees of Freedom X Coefficient(s) 0	Output:	0.372990 0.010127 0.982934 7 5
Std Err of Coef. 0	.005752 K = n =	2.360427 0.097627

2,4,6-TCP Isotherm Data at pH 7.0

Ce mg/l	q mg∕g	Ce mmol/l	q mmol/g	Ce/q	LOG Ce	LOG q
0.077	105.6	0.000390	0.534873	0.000729	-3.40892	-0.27174
0.105	173.7	0.000531	0.879805	0.000604	-3.27422	-0.05561
0.363	329.5	0.001838	1.668945	0.001101	-3.19504 -2.73550	0.074173
1.01	370.3	0.005115	1.875601	0.002727	-2.29109	0.273140
2.31	394.8	0.011700	1.999696	0.005851	-1.93180	0.300963
2.31	394.8	0.011700	1.999696	0.005851	-1.93180	0.300963
11.13	465.5	0.056374	2.357797	0.023909	-1.24891	0.372506
17.01	516.6	0.086157	2.616623	0.032926	-1.06470	0.417741
41.06	543.9	0.207972	2.754900	0.075491	-0.68199	0.440105
64.89	532.2	0.328673	2.695638	0.121927	-0.48323	0.430661
121.3	572.6	0.614394	2.900268	0.211840	-0.21155	0.462438

Langmuir Constants

Regression Output:Constant0.000346Std Err of Y Est0.000126R Squared0.997083No. of Observations6Degrees of Freedom4

X Coefficient(s) 0.468492 Std Err of Coef. 0.012667

Q	=	2.134508
b	=	1350.473

Regression	Output:
Constant	0.004158
Std Err of Y Est	0.003189
R Squared	0.998644
No. of Observations	6
Degrees of Freedom	4

X Coefficient(s) 0.342455 Std Err of Coef. 0.006309

> Q = 2.920090 b = 82.35189

#### Freundlich Constants

Regression	Output:
Constant	1.011691
Std Err of Y Est	0.119733
R Squared	0.768680
No. of Observations	6
Degrees of Freedom	4

X Coefficient(s) 0.328260 Std Err of Coef. 0.090037

K =	10.27286
n =	0.328260

Regression	Output:
Constant	0.489994
Std Err of Y Est	0.017651
R Squared	0.927649
No. of Observations	6
Degrees of Freedom	4
5	

X Coefficient(s) 0.091698 Std Err of Coef. 0.012804

Κ	=	3.090258
n	=	0.091698

## 2,4,5-TCP Isotherm Data at pH 7.0

Ce mg/l	q mg/g	Ce mmol/l	q mmol/g	Ce/q	LOG Ce	LOG q
0.031	100.6	0.000157	0.509547	0.000308	-3.80405	-0.29281
0.054	132.6	0.000273	0.671630	0.000407	-3.56301	-0.17286
0.074	170.4	0.000374	0.863090	0.000434	-3.42618	-0.06394
0.095	189.6	0.000481	0.960340	0.000501	-3.31768	-0.01757
0.202	234.3	0.001023	1.186749	0.000862	-2.99006	0.074359
0.266	273.2	0.001347	1.383781	0.000973	-2.87053	0.141067
0.514	314.6	0.002603	1.593476	0.001633	-2.58445	0.202345
2.04	366.8	0.010332	1.857873	0.005561	-1.98578	0.269016
0.514	314.6	0.002603	1.593476	0.001633	-2.58445	0.202345
2.04	366.8	0.010332	1.857873	0.005561	-1.98578	0.269016
5.37	420.3	0.027199	2.128855	0.012776	-1.56543	0.328146
33.8	508	0.171199	2.573063	0.066535	-0.76649	0.410450

## Langmuir Constants

### Regression Output:

Constant	0.000259
Std Err of Y Est	0.000043
R Squared	0.999425
No. of Observations	9
Degrees of Freedom	7

X Coefficient(s) 0.514639 Std Err of Coef. 0.004662

> Q = 1.943109 b = 1979.875

#### Regression Output:

Constant	0.001520
Std Err of Y Est	0.000899
R Squared	0.999411
No. of Observations	4
Degrees of Freedom	2

X Coefficient(s) 0.380562 Std Err of Coef. 0.006529

> Q = 2.627691 b = 250.3250
# Freundlich Constants

Regression	Output:
Constant	0.954205
Std Err of Y Est	0.059246
R Squared	0.910676
No. of Observations	9
Degrees of Freedom	7

X Coefficient(s) 0.304492 Std Err of Coef. 0.036043

K	=	8.999234	
n	=	0.304492	

Regression	Output:
Constant	0.501937
Std Err of Y Est	0.005989
R Squared	0.996941
No. of Observations	4
Degrees of Freedom	2
X Coefficient(s) 0 Std Err of Coef. 0	.115585 .004526

K	=	3.176415
n	-	0.115585

2,4-DCP Isotherm Data at pH 7.0

Ce mg/l	q mg/g	Ce mmol/l	q mmol/g	Ce/q	LOG Ce	LOG q
0.116	139.3	0.000711	0.854706	0.000832	-3.14767	-0.06818
0.169	158.2	0.001036	0.970671	0.001068	-2.98424	-0.01292
0.202	223.2	0.001239	1.369493	0.000905	-2.90678	0.136559
0.956	270	0.005865	1.656644	0.003540	-2.23167	0.219229
1.27	283.4	0.007792	1.738863	0.004481	-2.10833	0.240265
2.53	312.2	0.015523	1.915572	0.008103	-1.80901	0.282298
2.58	318.4	0.015830	1.953613	0.008103	-1.80051	0.290838
10.11	375.8	0.062032	2.305804	0.026902	-1.20738	0.362822
10.57	375.2	0.064854	2.302122	0.028171	-1.18805	0.362128
43.1	408.3	0.264449	2.505215	0.105559	-0.57765	0.398845
66.4	430.5	0.407411	2.641428	0.154239	-0.38996	0.421838
90.7	454	0.556510	2.785617	0.199779	-0.25452	0.444921
156.6	469.9	0.960854	2.883175	0.333262	-0.01734	0.459871

Langmuir Constants

Regression Ou	tput:
Constant	0.000492
Std Err of Y Est	0.000147
R Squared	0.997886
No. of Observations	6
Degrees of Freedom	4

X Coefficient(s) 0.496604 Std Err of Coef. 0.011427

> Q = 2.013674 b = 1007.900

Regression Output: Constant 0.007834

Std Err of Y Est	0.005415
R Squared	0.998229
No. of Observations	7
Degrees of Freedom	5

X Coefficient(s) 0.343566 Std Err of Coef. 0.006471

> Q = 2.910647 b = 43.85290

# Freundlich Constants

Regression	Output:
Constant	0.746829
Std Err of Y Est	0.058743
R Squared	0.866008
No. of Observations	6
Degrees of Freedom	4

X Coefficient(s) 0.242546 Std Err of Coef. 0.047702

Κ	=	5.582505
n	=	0.242546

0
4
8
7
5
: )

X Coefficient(s) 0.090318 Std Err of Coef. 0.005763

K	=	2.895615
n	=	0.090318

3,4-DCP Isotherm Data at pH 7.0

Ce mg/l	₫ mg/g	Ce mmol/l	q mmol/g	Ce/q	LOG Ce	LOG q
0.24 0.302 0.325 0.4 0.57 0.95	95 119 137.6 156.8 189.6 232.3	0.001472 0.001852 0.001994 0.002454 0.003497 0.005828	0.582893 0.730150 0.844275 0.962081 1.163332 1.425328	0.002526 0.002537 0.002361 0.002551 0.003006 0.004089	-2.83192 -2.73212 -2.70025 -2.61007 -2.45625 -2.23441	-0.23441 -0.13658 -0.07351 -0.01678 0.065704 0.153914
2.61	289.8	0.016014	1.778132	0.009006	-1.79549	0.249964
2.82 6.57 12.42 44.7	313 342.3 391.9 428.3	0.017302 0.040311 0.076205 0.274266 0.530126	1.920481 2.100257 2.404589 2.627929	0.009009 0.019193 0.031691 0.104366	-1.76188 -1.39456 -1.11801 -0.56182	0.283410 0.322272 0.381040 0.419613 0.436713
133.1	445.5	0.816664	2.843293	0.287224	-0.08795	0.453821

Langmuir Constants

Regression Output:

Constant	0.001541
Std Err of Y Est	0.000192
R Squared	0.994623
No. of Observations	7
Degrees of Freedom	5

X Coefficient(s) 0.461608 Std Err of Coef. 0.015177

Q	=	2.166337
b	=	299.3787

Regression Output:Constant0.005546Std Err of Y Est0.003128R Squared0.999378No. of Observations6Degrees of Freedom4

X Coefficient(s) 0.348825 Std Err of Coef. 0.004348

> Q = 2.866766 b = 62.88925

# Freundlich Constants

Output:
1.097391
0.058202
0.900232
7
5

X Coefficient(s) 0.442005 Std Err of Coef. 0.065805

K	=	12.51387
n	=	0.442005

Regression	Output:
Constant	0.469702
Std Err of Y Est	0.014295
R Squared	0.964227
No. of Observations	6
Degrees of Freedom	4
X Coefficient(s) 0.	.100260
Std Err of Coef. 0.	009655

K	=	2.949187
n	=	0.100260

4-DCP Isotherm Data at pH 7.0

Ce mg/l	q mg/g	Ce mmol/l	q mmol/g	Ce/q	LOG Ce	LOG q
0.264	101.4	0.002053	0.788920	0.002603	-2.68740	-0.10296
0.439	119.2	0.003415	0.927409	0.003682	-2.46653	-0.03272
0.743	140.4	0.005780	1.092351	0.005292	-2.23801	0.038362
1.04	161.6	0.008091	1.257294	0.006435	-2.09197	0.099436
1.89	185.5	0.014704	1.443242	0.010188	-1.83254	0.159339
3.01	206.8	0.023418	1.608962	0.014555	-1.63043	0.206546
2.59	175.3	0.020150	1.363883	0.014774	-1.69570	0.134777
5.95	210	0.046292	1.633859	0.028333	-1.33448	0.213214
10	236.8	0.077802	1.842371	0.042229	-1.10900	0.265377
18.8	265.9	0.146269	2.068777	0.070703	-0.83484	0.315713
29.9	294.7	0.232630	2.292849	0.101459	-0.63333	0.360375
53.1	315.9	0.413133	2.457791	0.168091	-0.38390	0.390545
73.9	350	0.574963	2.723099	0.211142	-0.24036	0.435063
100.5	390.7	0.781918	3.039757	0.257230	-0.10683	0.482838

Langmuir Constants

#### Regression Output:

oulpul:
0.001836
0.000283
0.996800
6
4

X Coefficient(s) 0.552359 Std Err of Coef. 0.015647

> Q = 1.810415 b = 300.8365

# Regression Output:Constant0.018987Std Err of Y Est0.010916R Squared0.987500No. of Observations8Degrees of Freedom6

X Coefficient(s) 0.323599. Std Err of Coef. 0.014863

> Q = 3.090240 b = 17.04283

#### Freundlich Constants

Regression Output:Constant0.702295Std Err of Y Est0.011531R Squared0.992236No. of Observations6Degrees of Freedom4

X Coefficient(s) 0.297042 Std Err of Coef. 0.013137

Κ	=	5.038427
n	=	0.297042

Regression	Output:
Constant	0.489496
Std Err of Y Est	0.010662
R Squared	0.992811
No. of Observations	8
Degrees of Freedom	6

X Coefficient(s) 0.207946 Std Err of Coef. 0.007223

Κ	Ħ	3.086714
n	=	0.207946

2-DCP Isotherm Data at pH 7.0

Ce mg/l	q mg/g	Ce mmol/l	q mmol/g	Ce/q	LOG Ce	LOG q
0.244	145.9	0.001898	1.135143	0.001672	-2.72161	0.055050
0.437	163	0.003399	1.268186	0.002680	-2.46852	0.103183
1.05	182.7	0.008169	1.421458	0.005747	-2.08781	0.152734
1.36	206.3	0.010581	1.605072	0.006592	-1.97546	0.205494
2.81	232.4	0.021862	1.808138	0.012091	-1.66029	0.257231
2.81	232.4	0.021862	1.808138	0.012091	-1.66029	0.257231
4.76	259.3	0.037034	2.017427	0.018357	-1.43139	0.304798
13.82	295.5	0.107523	2.299074	0.046768	-0.96849	0.361552
22.5	331.8	0.175056	2.581498	0.067811	-0.75682	0.411871
47.9	350.4	0.372675	2.726211	0.136700	-0.42866	0.435559
62.7	373.3	0.487823	2.904380	0.167961	-0.31173	0.463053
77.4	385.4	0.602194	2.998521	0.200830	-0.22026	0.476907
101.7	393.6	0.791254	3.062320	0.258384	-0.10168	0.486050

Langmuir Constants

Regression	Output:
Constant	0.001022
Std Err of Y Est	0.000380
R Squared	0.993514
No. of Observations	5
Degrees of Freedom	3

X Coefficient(s) 0.515604 Std Err of Coef. 0.024052

> Q = 1.939472 b = 504.3157

Regression Output:

Constant	0.010023
Std Err of Y Est	0.004655
R Squared	0.997758
No. of Observations	8
Degrees of Freedom	6

X Coefficient(s) 0.319298 Std Err of Coef. 0.006179

> Q = 3.131861 b = 31.85469

## Freundlich Constants

Regression	Output:
Constant	0.569159
Std Err of Y Est	0.013616
R Squared	0.978341
No. of Observations	5
Degrees of Freedom	3

X Coefficient(s) 0.189862 Std Err of Coef. 0.016309

Κ	=	3.708172
n	=	0.189862

Regression	Output:
Constant	0.506249
Std Err of Y Est	0.008880
R Squared	0.990465
No. of Observations	8
Degrees of Freedom	6
5	

X Coefficient(s) 0.145079 Std Err of Coef. 0.005811

Κ	=	3.208114
n	=	0.145079

2,4,5-TCP Isotherms at Various pH Values

pH = 4.15Regression Output: Constant 0.001891 Std Err of Y Est 0.000936 Ce Ce/a q R Squared 0.998107 0.578 242.3 0.002385 No. of Observations 8 302.2 0.003871 Degrees of Freedom 6 1.17 2.16 370.8 0.005825 3.39 422.5 0.008023 X Coefficient(s) 0.001810 5.47 458.4 0.011932 Std Err of Coef. 0.000032 485.6 0.017236 8.37 22.8 507.9 0.044890 Q 552.3619 mg/g = 547.2 0.054459 2.797760 mmol/g 29.8 or 0.957304 1/mg b = 189.0006 l/mmol or

pH = 5.22Regression Output: 0.001626 Constant Ce/q Std Err of Y Est 0.000671 Ce q 0.998652 R Squared 241.4 0.001855 No. of Observations 8 0.448 6 0.675 302 0.002235 Degrees of Freedom 377.5 0.003655 1.38 3.84 427.9 0.008974 X Coefficient(s) 0.001780 4.86 457.5 0.010622 Std Err of Coef. 0.000026 9.38 488 0.019221 15.3 521.4 0.029344 0 561.7905 mg/g = 2.845517 mmol/g 27.9 551 0.050635 or 1.094642 l/mg b = 216.1152 l/mmol or

pH = 6.58Regression Output: Constant 0.001930 Ce Ce/q Std Err of Y Est 0.000966 q R Squared 0.996816 0.394 241.9 0.001628 No. of Observations 8 1.05 301.6 0.003481 Degrees of Freedom 6 1.4 361.7 0.003870 5.07 412.9 0.012279 X Coefficient(s) 0.001846 440.7 0.013410 Std Err of Coef. 0.000042 5.91 11.3 467.4 0.024176 14.5 504.6 0.028735 0 541.4717 mg/g = 25.3 529.7 0.047762 2.742600 mmol/gor b = 0.956760 l/mg 188.8932 l/mmol or

	pH	= 8.	.17				Re	ear	ressi	on Output	:	
	-				Const	tan	t			<u>-</u>	0.00276	5
Ce		q		Ce/q	Std I	Err	of	Y	Est		0.00143	2
					R Squ	uare	ed				0.99740	0
0.365		242	.4	0.001505	No. d	of (	Obse	erv	ratic	ns		7
2.62		298.	. 5	0.008777	Degre	ees	of	Fr	reedc	m		5
5.4		352.	. 7	0.015310	•							
7.77		392.	.9	0.019776	X Coe	eff:	icie	ent	:(s)	0.002169		
17.4		41	LO	0.042439	Std H	Err	of	Co	ef.	0.000049		
21.8		434.	.9	0.050126								
32.7		451.	.2	0.072473				0	=	460.8999	mq/q	
								-	or	2.334498	mmol/q	
								b	=	0.784558	l/mq	
								-	or	154.8954	l/mmol	

	pH = 9.3		Regression	1 Output:	
			Constant	0	.003656
Ce	. <b>q</b>	Ce/q	Std Err of Y Est	0	.001890
	-	_	R Squared	0	.998220
0.765	198.8	0.003848	No. of Observations	5	5
4.31	265.9	0.016209	Degrees of Freedom		3
13.7	322.8	0.042441	-		
27	344.4	0.078397	X Coefficient(s) (	.002722	
33.8	359.5	0.094019	Std Err of Coef. (	.000066	
22.0	222.2	0.094019	SLU BIL OF COEL. U		

Q	=	367.3288	mg/g
	or	1.860552	mmol/g
b	=	0.744483	l/mg
	or	146.9834	l/mmol

	pН	= 10.2	25		Re	gr	ession	l Oi	utput	:	
				Constant	t				-	0.012	880
Ce		q	Ce/q	Std Err	of	Y	Est			0.001	.127
				R Square	ed					0.999	747
5.83		196.7	0.029639	No. of (	Obse	rva	ations				4
15.8		255.1	0.061936	Degrees	of	Fr	eedom				2
29.5		289.1	0.102040	-							
49.2		299.9	0.164054	X Coeff:	icie	nt	(s) 0	.00	03083		
				Std Err	of	Co	ef. 0	.00	00034		
						0	_ 2	21	3570	ma / a	

Q.	=	324.35/9	mg/g
	or	1.642900	mmol/g
b	=	0.255046	l/mg
	or	50.35374	l/mmol

# Comparison of 3,4-DCP Isotherms with and without DOM

Without DOM

Ce	P	Ce/q
0.016014	1.778132	0.009006
0.037305	1.944410	0.019185
0.070560	2.226653	0.031689
0.254018	2.433427	0.104387
0.490857	2.530985	0.193939

Regression	Output:
Constant	0.004177
Std Err of Y Est	0.001341
R Squared	0.999777
No. of Observations	5
Degrees of Freedom	3

X Coefficient(s) 0.388280 Std Err of Coef. 0.003341

> Q = 2.575460 b = 92.94793

With DOM

Ce	р	Ce/q
0.008774	1.352313	0.006488
0.040066	1.652349	0.024248
0.084672	1.926003	0.043963
0.167505	2.175727	0.076988
0.457111	2.195361	0.208216

Regression Output:Constant0.004507Std Err of Y Est0.002223R Squared0.999428No. of Observations5Degrees of Freedom3X Coefficient(s)0.445000Std Err of Coef.0.006146

Q = 2.247188 b = 98.73054

# Effect of Temperature on PCP Adsorption

25 C Regression Output: Ce q Ce/g Constant 0.000606 mmol/lmmol/q Std Err of Y Est 0.000093 0.000714 0.676936 0.001056 R Squared 0.996778 0.000812 0.756724 0.001073 No. of Observations 7 0.001368 0.852759 0.001604 Degrees of Freedom 5 0.001444 0.933269 0.001547 0.003249 1.134364 0.002864 X Coefficient(s) 0.674468 0.004693 1.208376 0.003884 Std Err of Coef. 0.017148 0.006498 1.331849 0.004879 Q = 1.482648 b = 1112.944 30 C Regression Output: Ce q Ce/q Constant 0.000615 mmol/l mmol/qStd Err of Y Est 0.000070 R Squared 0.998879 0.001238 0.798243 0.001551 No. of Observations 4 0.001530 0.924604 0.001655 Degrees of Freedom 2 0.001938 1.019195 0.001902 0.006498 1.263614 0.005142 X Coefficient(s) 0.695277 Std Err of Coef. 0.016467 Q = 1.4382731130.338 b = 40 C Regression Output: Ce q Ce/q Constant 0.000717 mmol/lmmol/gStd Err of Y Est 0.000043 0.001426 0.765750 0.001862 R Squared 0.999304 0.001631 0.843372 0.001934 No. of Observations 5 0.002148 0.926771 0.002317 Degrees of Freedom 3 0.003310 1.017390 0.003254 0.005957 1.130393 0.005269 X Coefficient(s) 0.763491 Std Err of Coef. 0.011630 0 = 1.309772 b = 1063.809 T(K) LOG Q 0 Regression Output: 1.48265 0.171038 Constant 298.2 -0.97337303.2 1.43827 0.157840 Std Err of Y Est 0.004292 313.2 1.30977 0.117195 R Squared 0.988301 No. of Observations 3 Degrees of Freedom 1 X Coefficient(s) 341.9399 Std Err of Coef. 37.20210 Reaction Heat = -1564.73 cal/mol

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Effect of Temperature on 2,4,6-TCP Adsorption

30 C Regression Output: Constant 0.000140 Ce Ce/q Std Err of Y Est đ 0.000020 mmol/lmmol/qR Squared 0.998735 0.000253 0.843168 0.000300 No. of Observations 6 0.000319 1.082577 0.000294 Degrees of Freedom 4 0.000400 1.205152 0.000332 0.000678 1.475966 0.000459 X Coefficient(s) 0.503472 0.001200 1.605632 0.000747 Std Err of Coef. 0.008955 0.002947 1.810937 0.001627 Q= 1.986203 b= 3585.369

	40 C		Regression	Output:
Ce	q	Ce/q	Constant	0.000210
mmol/l	mmol/g	-	Std Err of Y Est	0.00028
0.000384	0.896857	0.000429	R Squared	0.997362
0.000724	1.028212	0.000704	No. of Observations	5
0.001068	1.137956	0.000939	Degrees of Freedom	3
0.001316	1.228114	0.001072	<b>J</b>	
0.002324	1.351027	0.001720	X Coefficient(s) 0.	655060
			Std Err of Coef. 0.	019449

Q= 1.526576 b= 3108.558

	50 C		Regress	sion Output	:
Ce	q	Ce/q	Constant	-	0.000447
mmol/l	mmol/g	. –	Std Err of Y Est		0.000092
0.00076	0.67	0.001134	R Squared		0.991941
0.001043	0.723969	0.001441	No. of Observati	lons	4
0.001190	0.856337	0.001389	Degrees of Freed	lom	2
0.002871	0.962028	0.002985	0		_
			X Coefficient(s)	0.879645	
			Std Err of Coef.	0.056064	
			Q=	1.136821	
			b=	1964.430	
T (K)	Q	Log Q	Regress	ion Output	:
323.2	1.1368	0.055684	Constant	-	-3.61101
313.2	1.5266	0.183725	Std Err of Y Est	•	0.008768
303.2	1.9862	0.298022	R Squared		0.997384
			No. of Observati	ons	3
			Degrees of Freed	om	1

X Coefficient(s) 1186.271 Std Err of Coef. 60.74584

Reaction Heat = -5425.71 cal/mol

Effect of Temperature on 2,4-DCP Adsorption

22.5 C Regression Output: Constant 0.000502 Ce Ce/q Std Err of Y Est 0.000084 a 0.998524 mmol/lmmol/g R Squared 0.001374 1.225374 0.001121 No. of Observations 6 0.001740 1.372867 0.001267 Degrees of Freedom 4 0.003227 1.536290 0.002100 0.005286 1.669034 0.003167 X Coefficient(s) 0.483681 0.007315 1.788799 0.004089 Std Err of Coef. 0.009296 0.012035 1.924492 0.006253 Q= 2.067478 b= 962.0086

30 C		Regressi	on Output:	:	
			Constant	-	0.000345
Ce	q	Ce/q	Std Err of Y Est		0.000047
mmol/l	mmol/g		R Squared		0.999533
0.001002	1.359298	0.000737	No. of Observatio	ns	6
0.001823	1.523900	0.001196	Degrees of Freedo	m	4
0.003109	1.710922	0.001817			
0.004790	1.857825	0.002578	X Coefficient(s)	0.458226	
0.006725	1.977000	0.003401	Std Err of Coef.	0.004947	
0.012802	2.064316	0.006201			
			Q=	2.182328	
			b=	1327.798	

	40 C		Regressio	n Output:	:
			Constant	-	0.000253
Ce	q	Ce/g	Std Err of Y Est		0.000100
mmol/l	mmol/g	-	R Squared		0.996369
0.000743	1.529800	0.000485	No. of Observation	S	6
0.001634	1.701482	0.000960	Degrees of Freedom	L .	4
0.002247	1.815347	0.001238	-		
0.004831	1.957531	0.002468	X Coefficient(s)	0.435020	
0.006666	2.057236	0.003240	Std Err of Coef.	0.013130	
0.009675	2.222429	0.004353			
				Q=	2.298744

b= 1712.834

	50 C		Regressi	on Output	:
			Constant	-	0.000296
Ce	q	Ce/q	Std Err of Y Est		0.000051
mmol/l	mmol/g	_	R Squared		0.999267
0.001327	1.684963	0.000787	No. of Observatio	ns	6
0.001805	1.797058	0.001004	Degrees of Freedo	m	4
0.003327	1.929802	0.001724	-		
0.003640	2.087325	0.001743	X Coefficient(s)	0.402773	
0.007551	2.255467	0.003348	Std Err of Coef.	0.005452	
0.012389	2.351043	0.005269			
			O=	2.482785	
			b=	1358.731	

T (K)	Q	Log Q	Regression Output	::
295.7	2.06748	0.315441	Constant	1.226138
303.2	2.18233	0.338920	Std Err of Y Est	0.003705
313.2	2.29874	0.361489	R Squared	0.992016
323.2	2.48279	0.394939	No. of Observations	4
			Degrees of Freedom	2
			X Coefficient(s) -269.437	,

Std Err of Coef. 17.09201

Reaction Heat= 1232.962

**Regression Output:** 23 C Constant 0.000641 Ce Ce/q Std Err of Y Est 0.000155 q mmol/lmmol/a R Squared 0.999235 0.000957 0.898473 0.001065 No. of Observations 5 0.002147 1.114675 0.001926 Degrees of Freedom 3 0.006007 1.339106 0.004486 0.010623 1.485735 0.007150 X Coefficient(s) 0.607245 0.020797 1.576255 0.013194 Std Err of Coef. 0.009698 Q = 1.646780 b =946.1319

30 C Regression Output: Ce Ce/q Constant q 0.000673 mmol/lStd Err of Y Est mmol/q 0.000381 0.000815 0.990490 0.000823 R Squared 0.993305 0.001578 1.121408 0.001407 No. of Observations 7 0.002371 1.218661 0.001945 Degrees of Freedom 5 0.007929 1.490223 0.005321 0.013765 1.600195 0.008602 X Coefficient(s) 0.538766 0.016682 1.726624 0.009662 Std Err of Coef. 0.019780 0.020124 1.823130 0.011038 Q = 1.856090

b = 799.9947

Ce q Ce/q Constant 0.0004   mmol/l mmol/g Std Err of Y Est 0.0003   0.001167 1.112431 0.001049 R Squared 0.9923   0.001795 1.215669 0.001476 No. of Observations 0.9923   0.003583 1.329381 0.002695 Degrees of Freedom 0.473988   0.004997 1.488727 0.003356 0.473988   0.014662 1.567278 0.004243 X Coefficient(s) 0.473988   0.014662 1.824626 0.008036 Std Err of Coef. 0.018170   0.018852 1.999682 0.009427 Q = 2.109756 D = 574.4293		40 C		Regressi	on Output	:
mmol/lmmol/gStd Err of Y Est0.00030.0011671.1124310.001049 R Squared0.99230.0017951.2156690.001476 No. of Observations0.09230.0035831.3293810.002695 Degrees of Freedom0.0049970.0049971.4887270.0033560.0066500.0066501.5672780.004243 X Coefficient(s)0.4739880.0146621.8246260.008036Std Err of Coef.0.0181700.0188521.9996820.009427Q = 2.109756b = 574.42931.42931.42931.4293	Ce	q	Ce/q	Constant	-	0.000825
0.001167 1.112431 0.001049 R Squared 0.992 0.001795 1.215669 0.001476 No. of Observations 0.003583 1.329381 0.002695 Degrees of Freedom 0.004997 1.488727 0.003356 0.006650 1.567278 0.004243 X Coefficient(s) 0.473988 0.014662 1.824626 0.008036 Std Err of Coef. 0.018170 0.018852 1.999682 0.009427 Q = 2.109756 b = 574.4293	mmol/l	mmol/g	-	Std Err of Y Est		0.000301
0.001795 1.215669 0.001476 No. of Observations 0.003583 1.329381 0.002695 Degrees of Freedom 0.004997 1.488727 0.003356 0.006650 1.567278 0.004243 X Coefficient(s) 0.473988 0.014662 1.824626 0.008036 Std Err of Coef. 0.018170 0.018852 1.999682 0.009427 Q = 2.109756 b = 574.4293	0.001167	1.112431	0.001049	R Squared		0.992705
0.003583 1.329381 0.002695 Degrees of Freedom 0.004997 1.488727 0.003356 0.006650 1.567278 0.004243 X Coefficient(s) 0.473988 0.014662 1.824626 0.008036 Std Err of Coef. 0.018170 0.018852 1.999682 0.009427 Q = 2.109756 b = 574.4293	0.001795	1.215669	0.001476	No. of Observatio	ns	7
0.004997 1.488727 0.003356 0.006650 1.567278 0.004243 X Coefficient(s) 0.473988 0.014662 1.824626 0.008036 Std Err of Coef. 0.018170 0.018852 1.999682 0.009427 Q = 2.109756 b = 574.4293	0.003583	1.329381	0.002695	Degrees of Freedo	m	5
0.006650 1.567278 0.004243 X Coefficient(s) 0.473988 0.014662 1.824626 0.008036 Std Err of Coef. 0.018170 0.018852 1.999682 0.009427 Q = 2.109756 b = 574.4293	0.004997	1.488727	0.003356	-		
0.014662 1.824626 0.008036 Std Err of Coef. 0.018170 0.018852 1.999682 0.009427 Q = 2.109756 b = 574.4293	0.006650	1.567278	0.004243	X Coefficient(s)	0.473988	
$\begin{array}{rcl} 0.018852 & 1.999682 & 0.009427 \\ Q &= & 2.109756 \\ b &= & 574.4293 \end{array}$	0.014662	1.824626	0.008036	Std Err of Coef.	0.018170	
Q = 2.109756 b = 574.4293	0.018852	1.999682	0.009427			
b = 574.4293				0 =	2.109756	
				<b>b</b> =	574.4293	
				-	0,101000	

T(K)	Q	LOG Q	Regressi	on Output	:
296.2	1.6468	0.216640	Constant	. –	2.190101
303.2	1.8561	0.268601	Std Err of Y Est		0.005038
313.2	2.1098	0.324241	R Squared		0.995617
			No. of Observatio	ns	3
			Degrees of Freedo	m .	1
			X Coefficient(s)	-583.841	
			Std Err of Coef.	38.73691	
			Reaction Heat =	2671.694	cal/mol

Reproducibility of 2,4-DCP Isotherm Measurements

(1) Regression Output: Ce q Ce/q Constant 0.000495 1.203545 0.000411 Std Err of Y Est 0.000341 0.000119 0.000938 1.371687 0.000683 R Squared 0.998561 0.001268 1.512101 0.000838 No. of Observations 9 0.002218 1.696173 0.001307 Degrees of Freedom 0.003840 1.844846 0.002081 7 0.004725 1.943961 0.002430 X Coefficient(s) 0.426636 0.007374 2.060776 0.003578 Std Err of Coef. 0.006120 0.016106 2.191750 0.007348 0.019351 2.298535 0.008418 Q = 2.343913b = 1249.300

	(2)		Regression Output:	
Ce	q	Ce/q	Constant	0.000355
0.000535	1.214754	0.000440	Std Err of Y Est	0.000133
0.000896	1.360478	0.000659	R Squared	0.998029
0.001185	1.512691	0.000783	No. of Observations	9
0.002053	1.696763	0.001210	Degrees of Freedom	7
0.003887	1.844846	0.002107		
0.005646	1.942782	0.002906	X Coefficient(s) 0.424376	
0.009085	2.057826	0.004415	Std Err of Coef. 0.007127	
0.011091	2.200600	0.005040		
0.020649	2.296176	0.008992	Q = 2.356400	
			$\bar{b} = 1192.977$	

	(3)		Regression Output:	•
Ce	q	Ce/q	Constant	0.000376
0.000584	1.217704	0.000480	Std Err of Y Est	0.000119
0.000737	1.375227	0.000536	R Squared	0.998806
0.002047	1.511511	0.001354	No. of Observations	.9
0.002123	1.696173	0.001252	Degrees of Freedom	7
0.003651	1.838356	0.001986	-	
0.005787	1.942192	0.002979	X Coefficient(s) 0.426841	
0.008082	2.059596	0.003924	Std Err of Coef. 0.005576	
0.013982	2.185851	0.006396		
0.023421	2.280246	0.010271	Q = 2.342790	
			$\bar{b} = 1132.337$	

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	(4)		Regression Output	:
Ce	q	Ce/q	Constant	0.000348
0.000491	1.213575	0.000404	Std Err of Y Est	0.000126
0.000825	1.345729	0.000613	R Squared	0.998311
0.001280	1.512101	0.000846	No. of Observations	9
0.002230	1.702072	0.001310	Degrees of Freedom	7
0.003657	1.844846	0.001982	-	
0.005415	1.935702	0.002797	X Coefficient(s) 0.426972	
0.009085	2.057826	0.004415	Std Err of Coef. 0.006637	
0.013038	2.197060	0.005934		
0.020295	2.286146	0.008877	Q = 2.342069	
			b = 1225.393	

	(5)		Regressio	on Output:	:
Ce	q	Ce/q	Constant	-	0.000372
0.000666	1.217704	0.000547	Std Err of Y Est		0.000103
0.001002	1.359298	0.000737	R Squared		0.998990
0.001823	1.523900	0.001196	No. of Observation	ns	8
0.003109	1.710922	0.001817	Degrees of Freedom	n	6
0.004790	1.857825	0.002578	-		
0.006725	1.977000	0.003401	X Coefficient(s)	0.444969	
0.012802	2.063726	0.006203	Std Err of Coef.	0.005775	
0.020000	2.185261	0.009152			
			Q =	2.247345	

b = 1195.702

# Appendix E Adsorption Kinetics Data

Effect of Stirring Speed on Adsorption Rate

Carbon dosage: 100 mg/l

	2,4,6-TCP	concentra	tion (mg/l)
Time,hr	200 rpm	400 rpm	800 rpm
0	22.44	22.42	22.44
0.3333	21.02	20.58	20.58
0.6666	19.75	18.97	18.91
1	17.84	16.57	16.36
1.5		13.6 <b>9</b>	13.62
2	12.64	11.5	11.47
3	10.07	8.69	8.6
5	6.62	5.45	5.69
8.25	3.75	2.87	3.12

Effect of pH on Adsorption Rate

Carbon dosage: 100 mg/l

2,4,5-TCP concentration (mg/l)

Time,hr	рН 3 <b>.9</b> 5	рН 7.0	pH 10.0
0	19.4	19.4	19.4
0.1666	18.21	18.6	18.6
0.3333	17.37	17.9	18
0.6666		16.33	
1	13.58		14.1
1.5	11.29	11.74	12.23
2	9.71	9.9	10.94
3	7.36	7.71	9.14
5	4.78	4.87	7.4
8.5	2.09	2.33	5.78

# 2,4-DCP Kinetics Data

	2,4-DCP cc	ncentrati	on $(mmol/l)$
Carbon	0.1 g/l	0.2 g/l	0.1 g/l
Time, hr			
0	0.1154	0.1154	0.02865
0.1666	0.1098	0.1037	0.02675
0.3333	0.1043	0.0945	0.0255 <b>9</b>
0.6666		0.0736	0.02307
1	0.0816	0.062	0.02135
1.5	0.0712	0.044	0.01792
2	0.062	0.0334	0.01528
3	0.0465	0.0194	0.01172
5	0.0282	0.0072	0.00589
8.25	0.0144	0.0022	0.00301

# 2,4,6-TCP Kinetics Data

	2,4,6-TCP c	oncentrat	ion (mmol/l)
Carbon	0.03 g/l	0.1 g/l	0.1 g/l
Time, hr			
0	0.1136	0.1136	0.02376
0.3333	0.1100	0.1042	0.02208
0.6666	0.1064	0.0952	0.02011
1	0.1037	0.079	0.01778
1.5	0.0939	0.0693	0.01514
2	0.0916	0.0582	0.01302
3	0.0844	0.044	0.01003
5	0.0752	0.0276	0.00618
8.25	0.0670	0.0145	0.00294

# 4-CP Kinetics Data

Carbon dosage: 100 mg/l

Time, hr	4-CP concentration	(mmol/l)
0	0.1257	
0.3333	0.1061	
0.6666	0.1039	
1	0.0896	
1.5	0.078	
2	0.0685	
3	0.0528	
5	0.0414	
8.25	0.0242	

2,4-DCP Kinetics Data (High Initial Concentration)

Carbon dosage: 400 mg/l

Time, hr	2,4-DCP	concentration	(mmol/l)
0		0.5	
0.1666		0.411	
0.3333		0.34	
0.6666		0.231	
1		0.163	
1.666		0.0906	
2.666		0.042	
4		0.0214	
7		0.0111	
12		0.0073	

2,4,6-TCP Kinetics Data (Long Term)

Carbon do**s**age: 30 mg/l

Time, hr	2,4,6-TCP conce	ntration	(mmol/l)
0	0.11	36	
0.3333	0.3	11	
0.6666	0.100	54	
1	0.103	37	
1.5	0.093	39	
2	0.093	16	
3	0.084	14	
5	0.075	52	
8.166	0.065	74	
16.5	0.05	76	
29.16	0.048	33	
42	0.04	14	

PCP Kinetics Data (Long Term)

Carbon dosage: 160 mg/l

Time, hr	PCP concentration	(mmol/l)
0	0.2951	
2	0.2534	
6	0.1945	
20	0.1333	
51.6	0.1104	
91.6	0.1036	
120	0.0965	

# Interruption Test

Carbon dosage: 100 mg/l

Time,hr	2,4,6-TCP concentration (mmol/l)
0	0 1136
0.3333	0.1049
0.6666	0.0965
1	0.0873
1.5	0.0751
2	0.0656
3	0.0513 (interrupted)
3.5	0.0444
4	0.0398
5	0.0309
8.25	0.0165

# Irreversibility Test

Carbon dosage: 20 mg/l

	concentra	ation (mmol/l)
Time, hr	2,4-DCP	2,4,6-TCP
0	0.04	0.04
1	0.0364	0.0366
4	0.0302	0.0311
10	0.0241	0.0258
24	0.0205	0.0227
48	0.0176	0.0204
72	0.0162	0.0193
96	0.0149	0.0182
120	0.0138	0.0175
144	0.0133	0.0169

# Irreversibility Test (Continued)

# Carbon dosage: 20 mg/l

concentr 2,4-DCP	ration (mmol/l) · 2,4,6-TCP
	0.04
	0.0365
	0.0308
0.04	0.0232
0.0264	0.0191
0.0211	0.0176
0.0183	0.0166
0.017	0.0158
0.0162	0.0153
0.0153	0.0149
	concentr 2,4-DCP 0.04 0.0264 0.0211 0.0183 0.017 0.0162 0.0153

Carbon dosage: 20 mg/l

Time,hr	concentrat 2,4-DCP	ion (mmol/l) 2,4,6-TCP
0	0.04	
1	0.037	
4	0.0302	
10	0.0219	0.04
13	0.0197	0.0354
23	0.0176	0.029
48	0.0159	0.0249
72	0.0144	0.023
96	0.0133	0.0219
123.5	0.0126	0.021
144	0.012	0.0204

# Appendix F Multicomponent Adsorption Equilibrium Data

Single-Component Adsorption Isotherm Constants for PCP/2,4,6-TCP System

PCP

Ce mmol/1	q mmol/a		Ce/q
0.000904 0.001536 0.002082 0.003379 0.005802 0.013243	0.380935 0.645815 0.809999 0.955752 1.068735 1.192983		0.002374 0.002378 0.00257 0.003535 0.005429 0.011101
Regressi	on Output	:	
Std Err of Y Est R Squared		0.001264 0.000280 0.994586	
Degrees of Freedo	ons om	6 4	
X Coefficient(s) Std Err of Coef.	0.734929 0.027110		
Q =	1.3607 581 43	mmol/g l/mmol	
= U	JOT . 13	-,	
= d	2,4,6-TC	₽	
D =	2,4,6-TC	₽	Ce/q
Ce mmol/1 0.000276	2,4,6-TC q mmol/g 1.101426	₽	Ce/q 0.000250
Ce mmol/l 0.000276 0.000437 0.000782	2,4,6-TC q mmol/g 1.101426 1.225750 1.498805	₽	Ce/q 0.000250 0.000356 0.000522
Ce mmol/l 0.000276 0.000437 0.000782 0.000888	2,4,6-TC q mmol/g 1.101426 1.225750 1.498805 1.582609	P	Ce/q 0.000250 0.000356 0.000522 0.000561
Ce mmol/l 0.000276 0.000437 0.000782 0.000888 0.002841 0.005111	q mmol/g 1.101426 1.225750 1.498805 1.582609 1.830798 2.016825	₽	Ce/q 0.000250 0.000356 0.000522 0.000561 0.001551 0.002534
Ce mmol/l 0.000276 0.000437 0.000782 0.000888 0.002841 0.005111 0.014366	2,4,6-TC q mmol/g 1.101426 1.225750 1.498805 1.582609 1.830798 2.016825 2.344214	₽	Ce/q 0.000250 0.000356 0.000522 0.000561 0.001551 0.002534 0.006128
Ce mmol/l 0.000276 0.000437 0.000782 0.000888 0.002841 0.005111 0.014366 Regressi	2,4,6-TC q mmol/g 1.101426 1.225750 1.498805 1.582609 1.830798 2.016825 2.344214 on Output:	P	Ce/q 0.000250 0.000356 0.000522 0.000561 0.001551 0.002534 0.006128
Ce mmol/l 0.000276 0.000437 0.000782 0.000888 0.002841 0.005111 0.014366 Regressi Constant Std Err of Y Est R Squared	2,4,6-TC q mmol/g 1.101426 1.225750 1.498805 1.582609 1.830798 2.016825 2.344214 on Output:	P 0.000232 0.000119 0.997334	Ce/q 0.000250 0.000356 0.000522 0.000561 0.001551 0.002534 0.006128
Ce mmol/l 0.000276 0.000437 0.000782 0.000888 0.002841 0.005111 0.014366 Regressi Constant Std Err of Y Est R Squared No. of Observation Degrees of Freedom	2,4,6-TC q mmol/g 1.101426 1.225750 1.498805 1.582609 1.830798 2.016825 2.344214 on Outputs	P 0.000232 0.000119 0.997334 7 5	Ce/q 0.000250 0.000356 0.000522 0.000561 0.001551 0.002534 0.006128

Q = 2.4035 mmol/g b = 1793.4 l/mmol Single-Component Adsorption Isotherm Constants for 2,4,6-TCP/2,4-DCP System

#### 2,4,6-TCP

Ce	q	Ce/q
mmol/l	mmol/g	
0.000143	0.790932	0.000181
0.000344	0.968210	0.000356
0.000441	1.189808	0.000370
0.000691	1.420172	0.000486
0.002435	1.724077	0.001412
0.005065	1.857523	0.002726
0.005503	1.954441	0.002815
0.008376	2.069867	0.004047

#### Regression Output:

Constant	0.000185
Std Err of Y Est	0.000085
R Squared	0.997085
No. of Observations	8
Degrees of Freedom	6

X Coefficient(s) 0.474411 Std Err of Coef. 0.010471

Q	=	2.1079	mmol/g
b	=	2564.4	l/mmol

#### 2,4-DCP

Ce	q	Ce/q
mmol/l	mmol/g	
0.000371	0.965786	0.000384
0.000890	1.235994	0.000720
0.001356	1.453104	0.000933
0.003433	1.649565	0.002081
0.008731	1.812397	0.004817

# Regression Output:Constant0.000228Std Err of Y Est0.000037R Squared0.999676No. of Observations5Degrees of Freedom3

X Coefficient(s) 0.527446 Std Err of Coef. 0.005474

> Q = 1.8959 mmol/g b = 2313.4 l/mmol

Single-Component Adsorption Isotherm Constants for 2,4-DCP/4-CP System

#### 2, 4 - DCP

Regression Output: Ce q mmol/lmmol/g Constant 0.000268 0.000371 0.965786 Std Err of Y Est 0.000080 0.00089 1.235994 R Squared 0.999447 0.001356 1.453104 No. of Observations 0.003433 1.649565 Degrees of Freedom 6 4 0.008731 1.812397 0.015523 1.915572 X Coefficient(s) 0.509170 Std Err of Coef. 0.005986

> Q = 1.963977 mmol/g b = 1898.014 l/mmol

#### 4 - CP

Ce	q		
mmol/l	mmol/g		
0.002057	0.789250	Regression Output	:
0.003418	0.927649	Constant	0.002523
0.005782	1.092232	Std Err of Y Est	0.000662
0.008079	1.25 <b>756</b> 3	R Squared	0.99 <b>7</b> 204
0.014737	1.443093	No. of Observations	9
0.023415	1.609172	Degrees of Freedom	7
0.032841	1.722884	-	
0.049973	1.850061	X Coefficient(s) 0.483625	
0.072491	1.971254	Std Err of Coef. 0.009678	

Q	=	2.067715	mmol/	g
b	=	191.6 <b>73</b> 0	l/mmo	ī

# 2,4,5-TCP/2,4-DCP Two-Component Adsorption at Various pH Values

Initial concentrations: 2,4,5-T 2,4,5-TCP, 1.019 mmol/l; 2,4-DCP, 1.026 mmol/l. Carbon dosage, 24 mg/40 ml.

2,4,5-TCP

2,4-DCP

pН	Ce	q mmol/a	Ce mmol/(	q mmol/a
	mmotra	mmol/g	IIIIIOUI	mmol/g
3.66	0.0494	1.604	0.2019	1.354
4.49	0.0533	1.603	0.2044	1.363
7.37	0.1637	1.426	0.2359	1.317
8.19	0.2786	1.229	0.2595	1.262
8.85	0.3413	1.139	0.3088	1.205
9.79	0.4493	0.942	0.4311	0.983
9.91	0.4759	0.909	0.4583	0.95
10.14	0.492	0.868	0.4821	0.895
10.22	0.5016	0.859	0.4987	0.875
10.3	0.5188	0.827	0.5273	0.824

# Appendix G Desorption Equilibrium and Kinetics Data

PCP Adsorption and Desorption Isotherms

# Adsorption

Ce	q	Ce/q	LOG Ce	LOG q
mmol/l	mmol/g			-
0.000574	0.452012	0.001269	-3.24105	-0.34484
0.000891	0.799326	0.001115	-3.04975	-0.09727
0.001505	0.914495	0.001646	-2.82231	-0.03881
0.003216	1.067212	0.003014	-2.49257	0.028251
0.006643	1.258921	0.005276	-2.17763	0.099998
0.013249	1.406583	0.009419	-1.87778	0.148165
0.024441	1.501535	0.016277	-1.61186	0.176535

#### Langmuir Constants

Regression Output:

Constant	0.000830
Std Err of Y Est	0.000203
R Squared	0.998908
No. of Observations	7
Degrees of Freedom	5

X Co	peffi	Lcie	ent (s)	0.637544
Std	Err	of	Coef.	0.009426

Q = 1.568518 mmol/g b = 768.1136 l/mmol

Freundlich Constants

Regression Output:Constant0.664858Std Err of Y Est0.077470R Squared0.845072No. of Observations7Degrees of Freedom5

X Coefficient(s) 0.271059 Std Err of Coef. 0.051903

> K = 4.622305n = 0.271059

### Desorption

Ce	q	Ce/q	LOG Ce	LOG q
mmol/l	mmol/g			-
0.000122	0.452012	0.000271	-3.91097	-0.34484
0.000176	0.624947	0.000283	-3.75225	-0.20415
0.000364	0.798965	0.000456	-3.43813	-0.09747
0.000436	0.914134	0.000477	-3.35966	-0.03898
0.001018	1.065768	0.000955	-2.99220	0.027662
0.002112	1.255671	0.001682	-2.67529	0.098876
0.003935	1.399362	0.002812	-2.40502	0.145930
0.007256	1.487454	0.004878	-2.13925	0.172443
0.012888	1.623564	0.007938	-1.88978	0.210469
0.018232	1.719237	0.010604	-1.73916	0.235335

# Langmuir Constants

Regression Output:

Constant	0.000344
Std Err of Y Est	0.000205
R Squared	0.997182
No. of Observations	10
Degrees of Freedom	8

X Coefficient(s) 0.578264 Std Err of Coef. 0.010868

> Q = 1.729311 mmol/g b = 1678.327 l/mmol

### Freundlich Constants

# Regression Output:

Constant	0.688989
Std Err of Y Est	0.055398
R Squared	0.925297
No. of Observations	10
Degrees of Freedom	8

X Coefficient(s) 0.236191 Std Err of Coef. 0.023727

> K = 4.886406n = 0.236191

# 2,4-DCP Adsorption and Desorption Isotherms

# Adsorption

Ce	q	Ce/q	LOG Ce	LOG q
mmol/l	mmol/g	_		-
0.000385	1.004417	0.000384	-3.41348	0.001914
0.000926	1.285433	0.000720	-3.03315	0.109049
0.001411	1.511228	0.000933	-2.85040	0.179330
0.003570	1.715547	0.002081	-2.44721	0.234402
0.009080	1.884893	0.004817	-2.04187	0.275286
0.027181	2.189225	0.012415	-1.56573	0.340290
0.039268	2.281261	0.017213	-1.40595	0.358175

### Langmuir Constants

Regression	Output:
Constant	0.000448
Std Err of Y Est	0.000275
R Squared	0.998574
No. of Observations	7
Degrees of Freedom	5

X Coefficient(s) 0.432961 Std Err of Coef. 0.007314

Q=	2.309673	mmol/g
b=	965.1070	1/mmol

#### Freundlich Constants

Regression Output:

Constant	0.607820
Std Err of Y Est	0.031305
R Squared	0.950052
No. of Observations	7
Degrees of Freedom	5

X Coefficient(s) 0.164478 Std Err of Coef. 0.016865

> K = 4.053407n = 0.164478

# Desorption

Ce	q	Ce/q	LOG Ce	LOG q
mmol/l	mmol/g	_		-
0.000182	1.285433	0.000141	-3.73937	0.109049
0.000511	1.510614	0.000338	-3.29096	0.179153
0.001325	1.713707	0.000773	-2.87768	0.233936
0.003810	1.954546	0.001949	-2.41904	0.291046
0.010860	2.257013	0.004811	-1.96416	0.353534
0.016259	2.306417	0.007049	-1.78888	0.362938
0.022947	2.388023	0.009609	-1.63926	0.378038
0.030187	2.435881	0.012392	-1.52016	0.386656

### Langmuir Constants

Regression Output:

Constant	0.000245
Std Err of Y Est	0.000154
R Squared	0.999054
No. of Observations	8
Degrees of Freedom	6

X Coefficient(s) 0.407814 Std Err of Coef. 0.005123

Q=	2.452093	mmol/g
b=	1664.044	l/mmol

## Freundlich Constants

Output:
0.587661
0.008905
0.993616
8
6

X Coefficient(s) 0.125103 Std Err of Coef. 0.004093

> K = 3.869563 n = 0.125103

# Adsorption

Ce	q	Ce/q	LOG Ce	LOG q
mmol/l	mmol/g	_		-
0.002057	0.789250	0.002606	-2.68670	-0.10278
0.003418	0.927649	0.003685	-2.46612	-0.03261
0.005782	1.092232	0.005294	-2.23785	0.038315
0.008079	1.257563	0.006424	-2.09261	0.099529
0.014737	1.443093	0.010212	-1.83157	0.159294
0.023415	1.609172	0.014551	-1.63049	0.206602
0.032841	1.722884	0.019062	-1.48357	0.236256
0.049973	1.850061	0.027011	-1.30126	0.267186

# Langmuir Constants

Regression Out	put:
Constant	0.002234
Std Err of Y Est	0.000473
R Squared	0.997367
No. of Observations	8
Degrees of Freedom	6

X Coefficient(s) 0.505869 Std Err of Coef. 0.010609

> Q= 1.976796 mmol/g b= 226.4387 l/mmol

#### Freundlich Constants

# Regression Output:

Constant	0.640400
Std Err of Y Est	0.016656
R Squared	0.986495
No. of Observations	8
Degrees of Freedom	6

X Coefficient(s) 0.270271 Std Err of Coef. 0.012909

K	=	4.369185
n	=	0.270271

# Desorption

Ce	q	Ce/q	LOG Ce	LOG q
mmol/l	mmol/g	. —		-
0.003158	1.132031	0.002790	-2.50047	0.053858
0.003913	1.302419	0.003004	-2.40743	0.114750
0.006955	1.490702	0.004665	-2.15766	0.173390
0.011125	1.654866	0.006723	-1.95366	0.218762
0.014549	1.765346	0.008241	-1.83716	0.246829
0.019217	1.885941	0.010189	-1.71630	0.275528
0.025363	1.995643	0.012709	-1.59578	0.300082
0.036956	2.056329	0.017972	-1.43231	0.313092

Langmuir Constants

**Regression** Output:

Constant	0.001502
Std Err of Y Est	0.000193
R Squared	0.998826
No. of Observations	8
Degrees of Freedom	6

X Coefficient(s) 0.447687 Std Err of Coef. 0.006264

> Q= 2.233700 mmol/g b= 297.9786 l/mmol

#### Freundlich Constants

Regression	Output:
Constant	0.676124
Std Err of Y Est	0.016712
R Squared	0.971706
No. of Observations	8
Degrees of Freedom	6

X Coefficient(s) 0.237980 Std Err of Coef. 0.016578

K	=	4.743777
n	=	0.237980

### Desorption Kinetics Data

PCP (Initial loading: 1.781 mmol/g) Time, hr Conc., mmol/l 0 0 0.25 0.002119 0.666666 0.003646 1.25 0.005018 2 0.006390 3.75 0.008051 6.75 0.009783 11.75 0.011228 23 0.012094 36 0.012961

2,4-DCP (Initial loading: 2.805 mmol/g)

Time, hr	Conc., mmol/l
0	0
0.166666	0.005583
0.333333	0.008651
0.666666	0.013437
1	0.016566
2	0.022763
3.5	0.027794
6	0.032028
10	0.03626
22.5	0.03866

4-CP (Initial loading: 2.336 mmol/g)

Time, hr	Conc., mmol/1
0	0
0.166666	0.006747
0.333333	0.010397
0.666666	0.014924
1.166666	0.018814
2	0.023482
3.5	0.027089
6	0.030130
9.5	0.031616