

AN ABSTRACT OF THE THESIS OF

Imanga Kaliangile for the degree of Master of Science  
in Crop Science presented on September 26, 1986

Title: PLANT AND SEED DEVELOPMENT IN FOUR CUPHEA SPECIES

*Redacted for Privacy*

Abstract approved: \_

Don F. Grabe \_\_\_\_\_

Cuphea is a potential source of medium-chain triglycerides for manufacture of soaps, pharmaceuticals and nutritional products. Indeterminate flowering and seed shattering are wild-type plant characteristics that make it difficult to determine when to harvest for maximum yield of seed and oil.

The primary objective of this study was to investigate the relationship between stage of Cuphea seed development and yields of seed and oil. A secondary objective was to determine the effect of photoperiod on plant development and flowering.

The flowering patterns of C. laminuligera Koehne, C. leptopoda Hemsley, C. lutea Rose and C. wrightii Gray were determined in the greenhouse. The effect of photoperiod on flowering of the four species was studied under 8/16 and 16/8-h light/dark cycles at 10/25°C in growth chambers. Seed development of C. lutea and C. wrightii was compared in two greenhouse and one field experiment by

harvesting seeds at frequent intervals from tagged flowers and measuring several quality components.

Two opposite branches formed at each of the first two nodes in C. wrightii, the first four in C. lutea, the first five in C. leptopoda and the first six in C. laminuligera. All species formed one branch node<sup>-1</sup> thereafter. The first flowers developed in the axil of the second single branch in all species. All species flowered under 8 and 16-h photoperiods.

The maximum seed dry weight was attained by 18 days after anthesis in both species in the 1985 greenhouse studies. At this time, dry weight was 2.7 mg seed<sup>-1</sup>, moisture content was 36% and oil content was 43% in C. lutea. Measurements for C. wrightii were 1.9 mg seed<sup>-1</sup> dry weight, 37% moisture and 43% oil content. Simple correlation coefficients for oil and dry weight were 0.97 and 0.96 for C. wrightii and C. lutea, respectively. Seed growth rate of C. lutea was 1.4 times that of C. wrightii. Similar relationships between seed quality components were evident in the 1986 greenhouse and 1985 field experiments.

Seeds of C. lutea were mature at the time of emergence from the calyx tube when grown in the field. Waiting beyond this time for green seeds to turn yellow or brown before harvesting does not increase seed yield or oil content. Since the four species flowered under 8 and

16-h photo periods, it appears Cuphea may be grown over a wide range of latitudes.

Plant and Seed Development in Four Cuphea Species

by

Imanga Kaliangile

A THESIS

submitted to

Oregon State University

in partial fulfillment of  
the requirements for the  
degree of

Master of Science

Completed September 26, 1986

Commencement June 1987

APPROVED:

*Redacted for Privacy*

\_\_\_\_\_  
Professor of Agronomy in charge of major

*Redacted for Privacy*

\_\_\_\_\_  
Head of Department of Crop Science

*Redacted for Privacy*

\_\_\_\_\_  
Dean of Graduate School

Date thesis is presented September 26, 1986

## ACKNOWLEDGEMENTS

I express my sincere appreciation to Dr. Don F. Grabe, my major professor, for his guidance throughout my graduate program and for his assistance with this thesis. I thank Dr. Kenton L. Chambers, Dr. Steve Knapp and Dr. Harry J. Mack, my Graduate Council Representative for serving on my graduate committee.

I am also grateful for the help and encouragement that I received from staff members and graduate students in the Department of Crop Science. Special thanks to Carol Garbacik and Thomas G. Chastain for their valuable help.

My special thanks go to my wife, Eunice for her patience and support during my graduate studies and preparation of this thesis.

I am grateful to the Government of The Republic of Zambia, Department of Agriculture (Research), and United States Agency for International Development (USAID) for providing me the opportunity to pursue graduate studies at Oregon State University.

## TABLE OF CONTENTS

	<u>Page</u>
INTRODUCTION	1
LITERATURE REVIEW	3
Vegetative Development and Flowering	3
Maturity	5
MANUSCRIPT: Plant and Seed Development in <u>Cuphea</u>	8
Abstract	9
Introduction	12
Materials and Methods	14
Vegetative Development	14
Flowering	15
Greenhouse studies	15
Growth chamber studies	15
Seed Maturity	16
Greenhouse studies	16
Field study	17
Results and Discussion	18
Vegetative Development	18
Flowering	18
Seed Development and Maturity	19
Greenhouse Studies	19
Field Study	21
References	31
BIBLIOGRAPHY	34
APPENDIX	38

## LIST OF APPENDIX TABLES

<u>Table</u>	<u>Page</u>
1. Effect of temperature, light and seed age on germination of three species of <u>Cuphea</u>	38
2. Effect of harvest date on seed yield, oil content, weight and color of two <u>Cuphea</u> species in the field	41
3. Effect of multiple harvests on seed yield, weight and oil content of two <u>Cuphea</u> species in the field. Yield is total of several harvests and weight and oil content are averages of separate harvests at different dates on the same plot	42
4. Seed yield, weight and oil content of two <u>Cuphea</u> species harvested at different dates in the greenhouse	43
5. Effect of multiple harvests on seed yield and oil content of two <u>Cuphea</u> species grown in the greenhouse in 1986. Yield is total of several harvests and weight and oil content are averages of separate harvests at different dates from the same plants	44
6. Effect of seed color on oil content of two <u>Cuphea</u> species grown in the greenhouse in 1986	45
7. Effect of temperature on early vegetative growth of four <u>Cuphea</u> species in growth chambers. Daylength was 16 h and PPFD was $200 \mu\text{mol m}^{-2} \text{s}^{-1}$	46
8. Height, nodes, branches and flowers of four <u>Cuphea</u> species at different dates in the greenhouse	47

## LIST OF FIGURES

<u>Figure</u>		<u>Page</u>
1.	Dry weight, moisture and oil content of <u>Cuphea wrightii</u> seeds at various stages of maturity. Greenhouse, 1985.	26
2.	Dry weight, moisture and oil content of <u>Cuphea lutea</u> seeds at various stages of maturity. Greenhouse, 1985.	27
3.	Dry weight, moisture and oil content of <u>Cuphea wrightii</u> seeds at various stages of maturity. Greenhouse, 1986.	28
4.	Dry weight, moisture and oil content of <u>Cuphea lutea</u> seeds at various stages of maturity. Greenhouse, 1986.	29
5.	Dry weight, moisture and oil content of <u>Cuphea lutea</u> at various stages of maturity. Field, 1985.	30

LIST OF TABLES

<u>Table</u>		<u>Page</u>
1.	Flowering pattern of four <u>Cuphea</u> species in the greenhouse	24
2.	Effect of daylength on flowering and vegetative characteristics of four <u>Cuphea</u> species in growth chambers at 10/25° C. Measurements were taken 54 days after planting.	25

Plant and Seed Development  
in Four Cuphea Species

INTRODUCTION

Oil from Cuphea seed contains high amounts of medium-chain fatty acids which are used in the manufacture of industrial, medicinal and nutritional products. The present source of these acids is coconut and palm oil of which about a billion pounds are imported annually by the U.S. (Thompson, 1984). Imported supplies are subject to wide fluctuations in quality and price. Since 1983 Oregon State University, USDA and private industry have cooperated in conducting research to study the feasibility of developing Cuphea as an economically viable new oil crop in the Pacific Northwest and other parts of the U.S.

Cuphea species have several agronomic characteristics which impede their domestication. These include indeterminate growth and flowering, seed dormancy and shattering, and viscid glandular hairs on stems, leaves and flowers. Indeterminate flowering leads to seeds ripening over a period of several weeks. At maturity, the ovaries rupture, exposing placental tissue and seeds (Hirsinger and Robbelen, 1980). The exposed seed shatters easily. These factors make it difficult to determine when to harvest for maximum yield of seed and oil. Seed shattering is believed to be the most limiting constraint to domestica-

tion. In addition, seeds are dormant when harvested, making it difficult to establish good plant stands from freshly harvested seed. Viscid glandular hairs make mechanical harvesting and handling difficult because seeds stick to each other and to equipment. Cuphea domestication requires detailed understanding of all aspects of its seed technology.

This study was undertaken to compare plant growth, flowering and seed development characteristics of several Cuphea species. Such information is needed for further work on seed production, timing of seed harvesting, and selection of superior plant types.

## LITERATURE REVIEW

Cuphea is a diverse genus, having about 260 annual and perennial species (Hirsinger and Knowles, 1984; Thompson, 1984). It is native to tropical and subtropical areas of North, South and Central America. Some varieties of C. ignea and C. llavea are popular ornamentals in warm areas of the U.S.A. (Miller et al., 1964). Seed oil from Cuphea species contains high amounts of medium-chain triglycerides (Miwa et al., 1960) which are used in the manufacture of industrial, medicinal and pharmaceutical products (Babayan, 1981). There is strong interest in developing Cuphea as an oil crop in order to have a domestic source of medium-chain triglycerides to replace imported coconut and palm oil.

### Vegetative Development and Flowering

In many crops vegetative development depends on temperature and plant population. Warm temperature enhances vegetative growth (Went, 1953; Seddigh and Jolliff, 1984; Lawn and Hume, 1985). In soybean, warm day temperature compensates for low night temperature so that node formation at 30/12°C was higher than at 20/20°C (Lawn and Hume, 1985). Speed of seedling development at a particular temperature is a rapid technique for evaluating seedling vigor (Cooper et al., 1980). In birdsfoot trefoil,

seedling development was positively correlated with greenhouse yield, field vigor ratings and forage yield (Cooper et al., 1980). One important component of yield is branches plant<sup>-1</sup>. This is affected by plant population. In soybean, nonoptimum plant population reduces branch yield components rather than main stem components (Board, 1985). In beans, interplant competition resulted in fewer branches and pods plant<sup>-1</sup> on types that set the majority of their pods on lower nodes (Lucas and Milbourn, 1976; Kueneman and Wallace, 1979; Mauk et al., 1983).

In Cuphea, flowers are axillary and borne on short lateral shoots. The main stem axis continues vegetative growth and flowering concurrently. This causes seed to mature over a period of several weeks (Hirsinger, 1984). Flowers are singly set rather than combined in inflorescences (Robbelen and Hirsinger, 1982). Two main factors causing changes in flowering time in many crops such as cotton, soybean and subclover are temperature and photoperiod (Aitken, 1955; Board and Hall, 1984; Schweitzer and Harper, 1985). Generally a short photoperiod hastens flowering. There is also a temperature x photoperiod interaction. In sugarbeet, lettuce and subclover, long days hastened flowering of vernalized plants (Aitken, 1955; Dennis, 1984). Removal of leaves and flowers also affects flowering time. In cotton, removing flowers hastens fruit set (Dale, 1959; Kennedy et al., 1986). In

soybean, leaf removal hastens flowering (Goli and Weaver, 1986), while the opposite was found in subclover (Collins and Aitken, 1970). This has important applications in crop management because time of flowering can be controlled, by mowing or grazing, so that seed harvesting is done at a favorable time.

### Maturity

Early efforts to estimate seed maturity dealt with visual indicators such as browning of the plant, leaf or stem senescence, and external appearance of the seed unit (spike, panicle, ear). Later, attention was directed towards stage of seed development. Measurements of moisture content, dry matter accumulation, viability and seedling vigor became common. Studies of this kind have been described for cereals (Harlan, 1920; Collier, 1963; Rajanna and Andrews, 1970), forage crops (Anderson, 1955; Hyde et al., 1959), safflower (Leininger and Urie, 1964) and soybeans (TeKrony et al., 1979).

Maturity was defined as the point in plant growth when maximum grain development is first attained (Aldrich, 1943). The time required to reach maximum dry weight was termed relative maturity. Later, the term physiological maturity was used to refer to the time when a seed reaches its maximum dry weight (Shaw and Loomis, 1950). This is also referred to as functional maturity (Delouche, 1958)

and morphological maturity (Anderson, 1955). Despite differences in the concept of maturity, it is generally accepted that physiological maturity refers to the time when maximum dry weight is first attained in seed development.

Of the many procedures used to estimate maturity, the most common is to compare the relationship of maturity to accumulation of dry weight in the seed, moisture content and germination. Days from anthesis to maximum dry weight is a common estimate of maturity. In Cuphea, Hirsinger and Knowles (1984) reported that seed is fully ripe about a week after the cone-shaped placenta, with attached seeds, breaks through the calyx tube to expose the seed. From the time when the calyx tube opens to expose the seed until the seed is fully ripe, there was no difference in oil content. In Cuphea procumbens, lipid accumulation in developing seeds increases greatly between 11 and 14 days after flowering (Slabas et al., 1982). In subclover, Taylor (1980) found seed development in the cultivar Daliak to be complete 42 days after flowering in the greenhouse. Physiological maturity is attained at 28 days after flowering in safflower (Leininger and Urie, 1964), 27 in birdsfoot trefoil (Anderson, 1955) and 24 in red clover (Hyde et al., 1959). In cereals, a close relationship exists between seed moisture content and maximum dry weight. Moisture content of seeds at the time of maximum

dry weight was reported to be 42% for barley (Harlan, 1920), 45% for oats (Frey et al., 1958), 40% for wheat (Burnett and Bakke, 1930) and 27% for sorghum (Clark et al., 1968).

Dessureaux et al. (1948) reported corn kernel moisture percentages between 31 and 44% at maximum dry weight. Kernel moisture percentage was later found to be inferior to maximum kernel dry weight as an indicator of physiological maturity (Shaw and Thom, 1951).

Seeds of many species are capable of germinating long before physiological maturity (Copeland and McDonald, 1985). Germination increases with days after anthesis (Anderson, 1955; Grabe, 1956; Frey et al., 1958; Rajanna and Andrews, 1970).

In Cuphea, fresh seed is dormant due to a hard seed coat (Hirsinger and Knowles, 1984; Janick and Whipkey, 1986). Fresh seeds can germinate when the testa is removed or scarified (Hirsinger and Knowles, 1984). Excised embryos from dormant seeds of Cuphea wrightii can germinate when grown on a basal medium (Janick and Whipkey, 1984).

MANUSCRIPT

PLANT AND SEED DEVELOPMENT IN CUPHEA

## ABSTRACT

Cuphea is a potential source of medium-chain triglycerides for manufacture of soaps, pharmaceuticals and nutritional products. Indeterminate flowering and seed shattering are wild-type plant characteristics that make it difficult to determine when to harvest for maximum yield of seed and oil.

The primary objective of this study was to investigate the relationship between stage of Cuphea seed development and yields of seed and oil. A secondary objective was to determine the effect of photoperiod on plant development and flowering.

The flowering patterns of C. laminuligera Koehne, C. leptopoda Hemsley, C. lutea Rose and C. wrightii Gray were determined in the greenhouse. The effect of photoperiod on flowering of the four species was studied under 8/16 and 16/8-h light/dark cycles at 10/25°C in growth chambers. Seed development of C. lutea and C. wrightii was compared in two greenhouse and one field experiment by harvesting seeds at frequent intervals from tagged flowers and measuring several quality components.

Two opposite branches formed at each of the first two nodes in C. wrightii, the first four in C. lutea, the first five in C. leptopoda and the first six in C. laminu-

ligera. All species formed one branch node<sup>-1</sup> thereafter. The first flowers developed in the axil of the second single branch in all species. All species flowered under 8 and 16-h photoperiods.

The maximum seed dry weight was attained by 18 days after anthesis in both species in the 1985 greenhouse studies. At this time, dry weight was 2.7 mg seed<sup>-1</sup>, moisture content was 36% and oil content was 43% in C. lutea. Measurements for C. wrightii were 1.9 mg seed<sup>-1</sup> dry weight, 37% moisture and 43% oil content. Simple correlation coefficients for oil and dry weight were 0.97 and 0.96 for C. wrightii and C. lutea, respectively. Seed growth rate of C. lutea was 1.4 times that of C. wrightii. Similar relationships between seed quality components were evident in the 1986 greenhouse and 1985 field experiments.

Seeds of C. lutea were mature at the time of emergence from the calyx tube when grown in the field. Waiting beyond this time for green seeds to turn yellow or brown before harvesting does not increase seed yield or oil content. Since the four species flowered under 8 and 16-h photoperiods it appears Cuphea may be grown over a wide range of latitudes.

---

Additional index words: Cuphea laminuligera Koehne, Cuphea leptopoda Hemsley, Cuphea lutea Rose, Cuphea

wrightii Gray, maturation, dry weight, moisture, flowering, photoperiod.

---

## Plant and Seed Development in Cuphea

### INTRODUCTION

Seeds of Cuphea species are potential sources of medium-chain triglycerides for manufacture of soaps, detergents and pharmaceutical products (Miller et al., 1964; Babayan, 1981; Thompson, 1984). All known species of Cuphea, however, have several wild-plant characteristics which must be eliminated before a suitable agronomic plant type can be developed for agronomic purposes (Robbelen and Hirsinger, 1982; Hirsinger and Knowles, 1984).

The flowering habit of Cuphea is indeterminate, with flowers produced over a period of 1 to 2 months (Hirsinger, 1984). Development of the seeds takes place within the calyx tube. As seeds enlarge, the calyx tube bursts. The placenta, with attached seeds, becomes erect and breaks through the pericarp and calyx tube. The seeds shatter soon after (Hirsinger and Knowles, 1984). To obtain reasonable seed yields, seeds are harvested several times during the season with a non-destructive vacuum-harvester (Hirsinger, 1984). The vacuum-harvested seed contains a mixture of green, yellow and brown seeds, but the relationship of seed color to maturity and oil content is not known. Indexes of seed maturity have been determined for cereals (Harlan, 1920; Bartel, 1941; Hallauer

and Russell, 1962; Clark et al., 1968) and many other species (Grabe, 1956; Frey et al., 1958; Leininger and Urie, 1964; Browne, 1978; TeKrony et al., 1979), but rarely for indeterminate species such as Cuphea.

This study was undertaken to compare plant growth, flowering and seed development characteristics of several Cuphea species. Such information is needed for further work on seed production, timing of seed harvesting and selection of superior plant types.

## MATERIALS AND METHODS

Species selected for study were Cuphea lutea Rose, C. wrightii Gray, C. laminuligera Koehne and C. leptopoda Hemsley. The first two are self- and the last two are cross-pollinated. Vegetative development studies were conducted in the greenhouse, flowering patterns were studied in the greenhouse and growth chambers, while seed maturity studies were carried out in the greenhouse and field.

### Vegetative Development

Vegetative development of the four species was studied in the greenhouse during the 1984-85 and 1985-86 seasons. Seeds were planted in soil in 15-cm plastic pots on 12 February 1985 and 17 December 1985. Fertilizer (8-10-10) and lime were added to a soil mixture of 1 part soil, 1 part peat and 1 part sand at the rate of 4 and 8 g kg<sup>-1</sup> of soil, respectively. Seedlings were thinned to 4 plants pot<sup>-1</sup>. Temperature was controlled to approximately 25°C during daytime and 18°C at night. Light from F48T12/CW/VHO fluorescent tubes was used to extend day-length to 16 h. The experimental design was a randomized complete block with three replications.

The branching pattern of each species was determined at several stages of development beginning 18 days after planting in 1985 and 19 days in 1986.

## Flowering

### Greenhouse Studies

The normal flowering habit of the four species under 16-h daylengths was observed on the same plants used for the vegetative development studies. Observations were made of the number of days to beginning of flowering and the node at which the first flower formed.

### Growth Chamber Studies

The effect of daylength on flowering was studied in 1986 in two growth chambers (Controlled Environments, Model E7). Seeds were planted in soil in 10 x 10 x 10 cm plastic pots on 7 March 1986. Seedlings were thinned to four per pot. Soil, fertilizer and lime were as for the greenhouse studies. The experimental design was a randomized complete block with four replications.

The temperature in both chambers alternated daily between 10°C for 16 h and 25°C for 8 h. One chamber was set at an 8/16-h light/dark cycle and the other at a 16/8-h cycle. Four 25-watt incandescent bulbs and eight 40-watt F48T12/HO/CW fluorescent tubes provided a photosynthetic photon flux density (PPFD) of approximately 200 mol m<sup>-2</sup> S<sup>-1</sup>. PPFD was measured with a Li-COR Solar Monitor, Model Li-1776 with a quantum sensor Li-190SB.

Main stem height, the number of main stem nodes and

primary branches, and the node at which the first flower was formed were determined for each species 54 days after planting.

### Seed Maturity

Seed development and maturation of C. lutea and C. wrightii were studied in the greenhouse and field.

#### Greenhouse Studies

Seed maturation of both species was studied during the 1984-85 and 1985-86 seasons. Seeds were planted in 15-cm plastic pots on 12 February 1985 and 17 December 1985 and seedlings were thinned to 4 plants pot<sup>-1</sup>. Temperature was controlled to approximately 25°C during daytime and 18°C at night. Light from F48T12/CW/VHO fluorescent tubes was used to extend daylength to 16 h. The experiment design was a randomized complete block with three replications. Approximately 1000 flowers of C. lutea and C. wrightii were tagged with cotton thread at anthesis, different colored thread indicating different times of anthesis. Tagging was done on 26 and 27 April 1985 and 7 and 8 March 1986.

Seeds from 25 flowers per replication were collected approximately every 3 days beginning 5 days after anthesis in both years. Harvesting continued for 21 days in 1985 and 20 days in 1986. Seed dry weight was determined after drying for 24 h at 100°C. Percentage moisture content was

determined on a wet-weight basis. Oil content was determined on a dry weight basis according to the method of Comstock and Culbertson (1958). A 0.2-g sample of seeds was compressed at 103.5 MPa, each sample was put in a 50 x 50 mm envelope made from No. 1 Whatman filter paper, and the oil was extracted in hexane at 25°C for 72 h.

### Field Study

C. lutea was planted 17 May 1985 near Corvallis, OR on Woodburn silt loam (fine-silty, mixed, mesic, Aquultic Argixeroll) soil. Fertilizer (16-20-0) was incorporated into the seed bed before planting at 145.6 kg ha<sup>-1</sup>. Seeds were drilled with an Ojyord plot drill 0.5 cm deep in rows 0.76 m apart at the rate of 9.1 kg ha<sup>-1</sup>. Plot size was 9.1 x 30.8 m. The experimental design was a randomized complete block with 4 replications.

About 1200 flowers were tagged on 23 and 24 September 1985. Collection of seeds began 5 days later. Tagging of flowers, collection of seed and analyses of seed dry weight and moisture and oil content were performed as in the greenhouse experiment. Seeds were harvested at approximately 2-day intervals beginning 29 September and harvesting continued until 24 days after anthesis.

Data were analyzed by analysis of variance procedures and separation of treatment means was done by Fisher's protected least significant different (LSD) values.

## RESULTS AND DISCUSSION

### Vegetative Development

Each species had a different but consistent branching pattern. In Cuphea wrightii, two opposite branches formed at each of the first two nodes, with only one branch per node thereafter. Two opposite branches formed at the first four nodes in C. lutea, the first five nodes in C. leptopoda and the first six nodes in C. laminuligera. All species formed one branch per node thereafter. Branches became progressively shorter at each succeeding node until no branches, but only leaves and flowers, were formed near the apex. Secondary and tertiary branches developed in the lower part of the plant canopy, but no additional branching orders were observed.

### Flowering

The first flowers formed in the axil of the second single branch in each species, with C. wrightii invariably flowering at the fourth node and C. laminuligera at the eighth node. C. wrightii was the earliest to flower at 30 and 26 days, and C. leptopoda was the latest at 46 and 41 days in 1985 and 1986, respectively (Table 1). The first flowers produced seed in the self-pollinated species (C. wrightii and C. lutea).

In the growth chamber photoperiod study, all species

flowered under short and long days (Table 2). The node at which the first flower formed was not as consistent in growth chambers as in the greenhouse. The first flower always appeared in the axil of the second single branch, but the number of opposite branches at the lower nodes varied somewhat between plants. The first flowers developed earlier at lower nodes under 8-h days, and the plants were shorter with fewer primary branches and main stem nodes under short days (Table 2).

According to Wareing and Phillips (1981), species with an indeterminate flowering habit flower under both long and short daylengths. Short daylengths generally hasten flowering (Board and Hall, 1984; Schweitzer and Harper, 1985).

### Seed Development and Maturity

#### Greenhouse Studies

Changes in moisture and oil percentages and weight of seed dry matter and oil during seed development in 1985 are shown in Figures 2 and 3. Maximum seed dry weight in both species was reached by 18 days after anthesis. At this time seeds of C. wrightii and C. lutea averaged 1.90 and 2.70 mg seed<sup>-1</sup>. Since the seed-filling duration for both species was identical, the seed growth rate of C. lutea was 1.4 times that of C. wrightii.

When maximum dry weight was first attained, the moisture content was about 36%. Moisture percentage remained at this level for the next 8 days, in contrast to the continued decline in moisture that occurs in many species after maximum dry weight is reached (Harlan, 1920; Frey et al., 1958; Anderson, 1955; Leininger and Urie, 1964; Browne, 1978; TeKrony et al, 1979).

The rate of oil accumulation paralleled that of dry matter accumulation in both species. The attainment of maximum oil content coincided with the attainment of maximum dry weight at 18 days after anthesis. Oil contents were 0.83 mg seed<sup>-1</sup> in C. wrightii and 1.17 mg seed<sup>-1</sup> in C. lutea. The simple correlation coefficients for oil and dry weight were 0.97 and 0.96 for C. wrightii and C. lutea, respectively, indicating that seed dry weight is also an accurate index of oil content.

The maximum percentage of oil was established somewhat earlier than total oil content, reaching 44.9% in C. wrightii in 13 days and 42.2% in C. lutea in 11 days. These oil percentages compare with those reported for other Cuphea species of 36 for C. painteri, 34 for C. ignea and 33 for C. carthagenensis (Miller et al., 1964).

The developing seeds ruptured the calyxes of C. wrightii 13 days after anthesis and those of C. lutea in 14 days. After the calyx opens the seed is not protected and, under field conditions, shatters readily from rain

and wind movements. The seeds did not shatter in the greenhouse, however, making it possible to study seed development for another 11 or 12 days after opening of the calyx. At the time the seeds ruptured the calyx they were still green, but they gradually turned yellow and then dark brown.

C. wrightii averaged four seeds per flower, while C. lutea averaged five. These numbers contrast to previous reports of three and five seeds per flower (Hirsinger and Knowles, 1984).

The relative rates of development of each quality component were similar in experiments repeated in 1986, although the absolute values varied somewhat (Figures 3 and 4). The values at maturity for each seed component in C. wrightii were as follows (with 1985 values in parentheses): number of days to maximum dry weight, 19 (18); seed dry weight, 1.89 mg seed<sup>-1</sup> (1.90); moisture content, 32% (37%); oil weight, 0.59 mg seed<sup>-1</sup> (0.83 mg seed<sup>-1</sup>) and oil percentage, 31% (43%). For C. lutea the values were: seed dry weight, 2.62 mg seed<sup>-1</sup> (2.70 mg seed<sup>-1</sup>); moisture content, 42% (36%); oil weight, 0.64 mg seed<sup>-1</sup> (1.17 mg seed<sup>-1</sup>) and oil percentage, 24% (43%).

### Field Study

The time course of seed development of C. lutea seeds in the field in 1985 is shown in Figure 5. Relationships

between quality components were similar to those in the greenhouse. Maximum dry weight was reached in 21 days after anthesis compared to 18 days in the greenhouse. Moisture contents at maximum dry weight were 45 and 35% in the field and greenhouse, respectively. Oil contents at maximum dry weight were 34 and 24% in the field and greenhouse, respectively. At maturity, seeds produced in the field were smaller than those produced in the greenhouse, weighing  $1.97 \text{ mg seed}^{-1}$  compared to  $2.36 \text{ mg seed}^{-1}$  in the greenhouse. Calices opened 20 days after anthesis compared to 14 days in the greenhouse. The generally slower development in the field could be attributed partially to cooler temperatures, especially at night.

Seed color at maximum dry weight was still green in both field and greenhouse. Shattering occurred about 4 days after maturity in the field, but the seeds remained on the plants indefinitely in the greenhouse. These studies show that Cuphea seeds in the field are mature at the time they rupture the calyx tube and oil content is not increased by delaying harvest until the green seeds turn yellow or dark brown.

The 21-day seed maturation period for C. lutea is relatively short compared to most crop species. Examples of seed maturation periods in other species include 17-18 days in smooth brome grass (Grabe, 1956); 24-26 days in wheat (Bartel, 1941); 36 days in sorghum (Clark et al.,

1968) and 60-63 days in corn (Hallauer and Russell, 1962).

Based on the information gained from these studies, seeds of C. lutea and C. wrightii are mature at the time of emergence from the calyx tube. Waiting beyond this time for green seeds to turn yellow or brown before harvesting does not increase seed yield or oil content. Since the four species flowered under 8 and 16-h photoperiods, it appears Cuphea may be grown over a wide range of latitudes.

Table 1. Flowering pattern of four Cuphea species in the greenhouse

Species	<u>Appearance of first flower</u>	
	<u>Days after planting</u>	<u>Node number<sup>+</sup></u>
	<u>1985</u>	
<u>C. laminuligera</u>	40	8
<u>C. leptopoda</u>	46	7
<u>C. lutea</u>	35	6
<u>C. wrightii</u>	30	4
LSD <sub>0.05</sub>	3.8	0.9
	<u>1986</u>	
<u>C. laminuligera</u>	30	8
<u>C. leptopoda</u>	41	7
<u>C. lutea</u>	39	6
<u>C. wrightii</u>	26	4
LSC <sub>0.05</sub>	2.9	0.6

<sup>+</sup>Nodes on each main stem were numbered acropetally beginning at the cotyledonary node.

Table 2. Effect of daylength on flowering and vegetative characteristics of four *Cuphea* species in growth chambers at 10/25°C. Measurements were taken 54 days after planting.

Species	Main stem height		Primary branches		Main stem nodes		Nodes with first flower <sup>+</sup>	
	8 h	16 h	8h	16 h	8 h	16 h	8 h	16 h
	cm		no. plant <sup>-1</sup>		no. stem <sup>-1</sup>		no.	
<i>C. laminuligera</i>	26	31	3.2	7.2	8.6	15.7	4.5	4.9
<i>C. leptopoda</i>	15	24	0.4	5.7	8.5	12.1	5.4	8.1
<i>C. lutea</i>	17	16	2.8	7.7	7.4	10.3	3.4	5.6
<i>C. wrightii</i>	13	30	0.2	5.2	4.7	12.7	2.7	3.9
Daylength mean	18	25	1.7	6.4	7.3	12.7	4.0	5.6
LSD <sub>0.05</sub>								
Species		4		1.1		1.6		0.4
Daylength		2		0.8		1.1		0.3
Species x daylength		5		NS		2.2		0.6

<sup>+</sup> Nodes on each main stem were numbered acropetally beginning at the cotyledonary node.

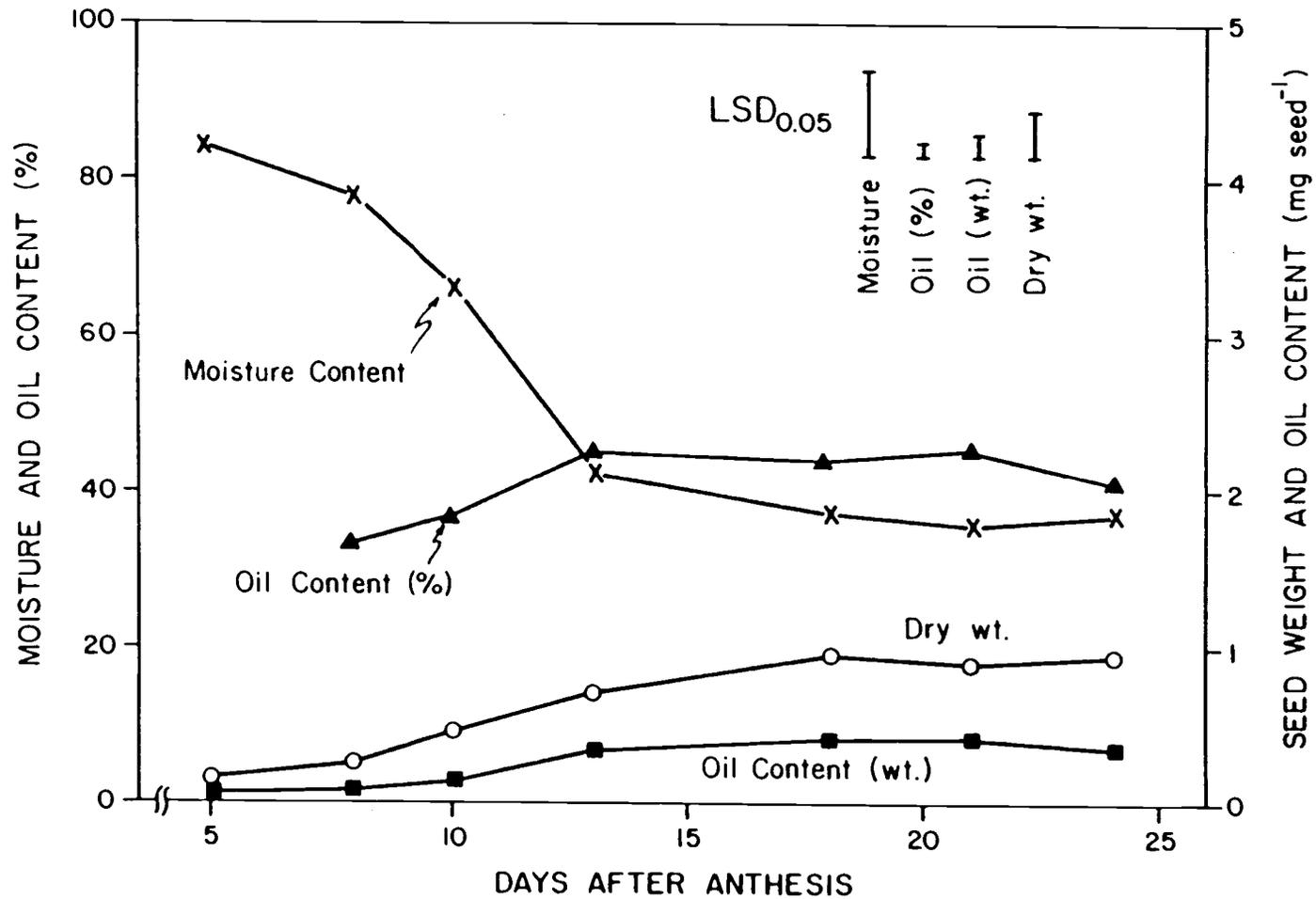


Figure 1. Dry weight, moisture and oil content of *Cuphea wrightii* seeds at various stages of maturity. Greenhouse, 1985.

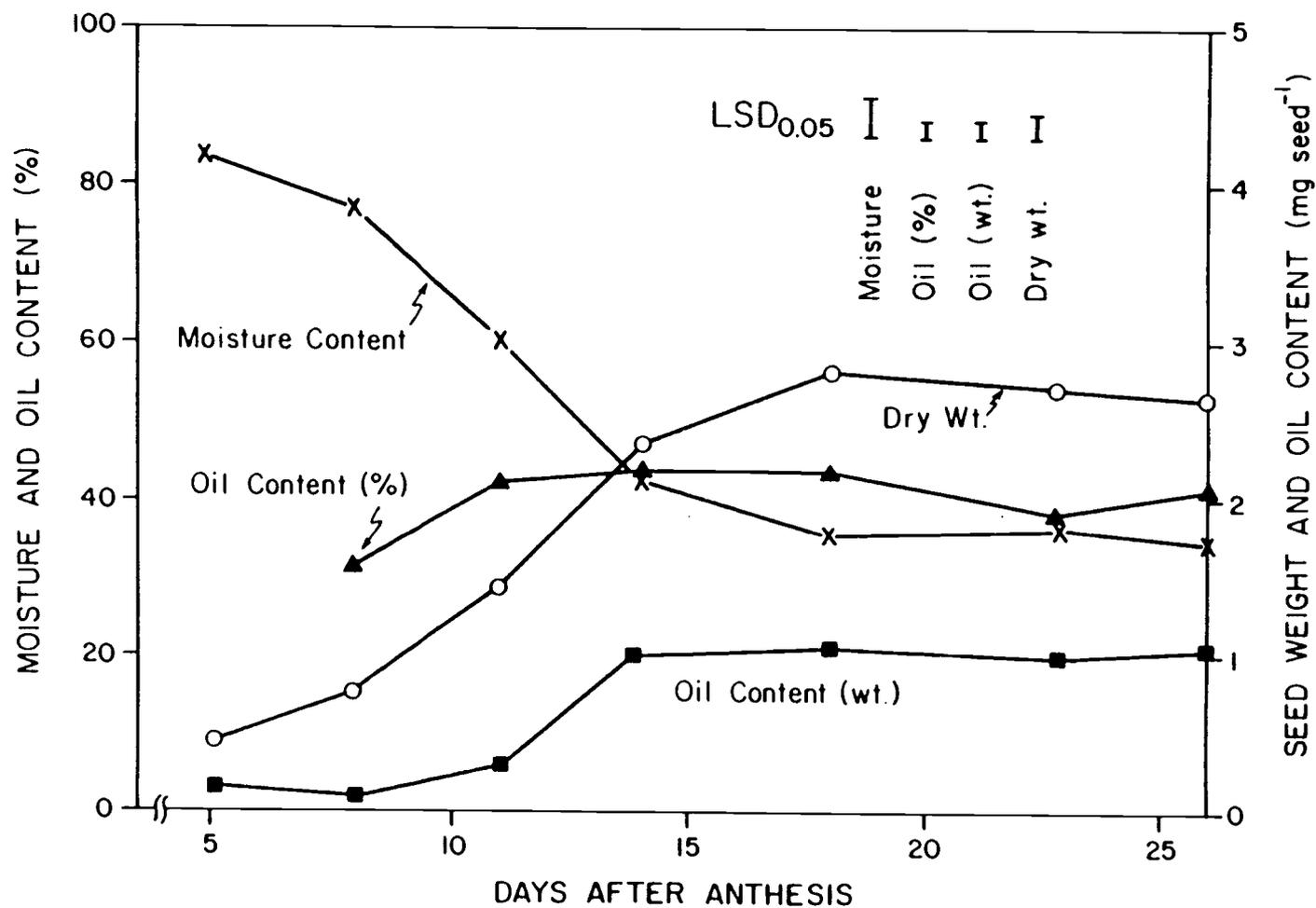


Figure 2. Dry weight, moisture and oil content of *Cuphea lutea* seeds at various stages of maturity. Greenhouse, 1985.

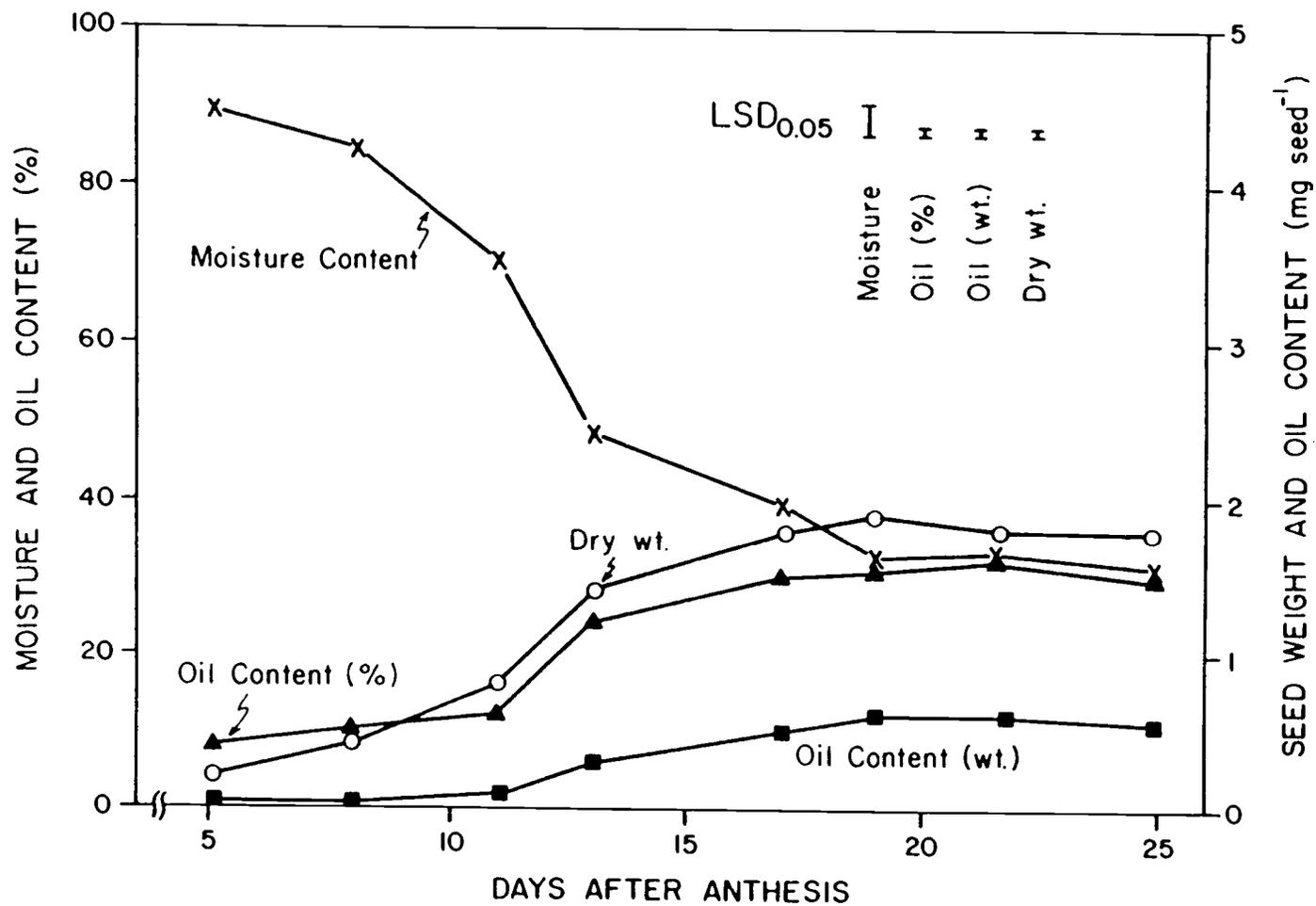


Figure 3. Dry weight, moisture and oil content of *Cuphea wrightii* seeds at various stages of maturity. Greenhouse, 1986.

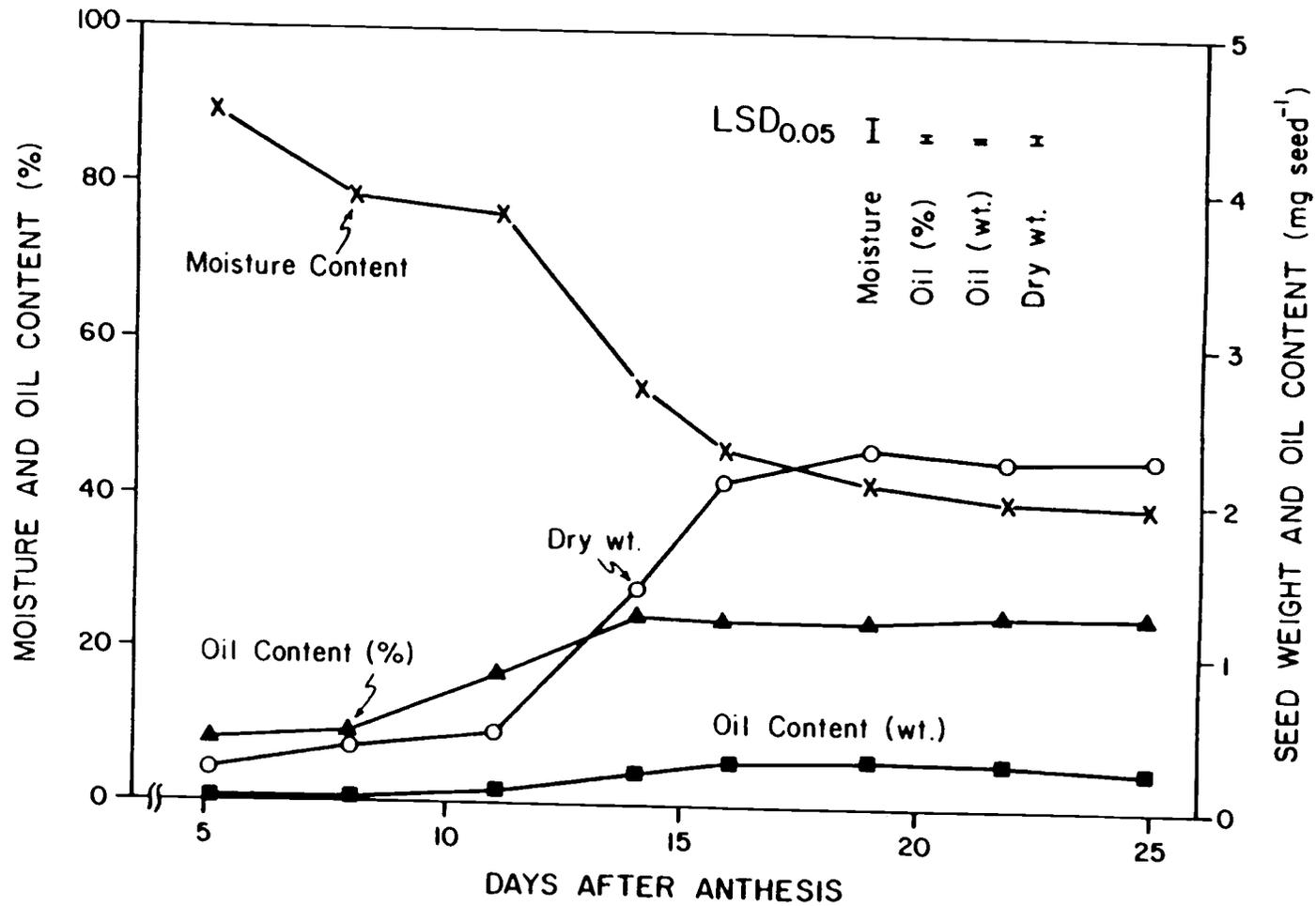


Figure 4. Dry weight, moisture and oil content of *Cuphea lutea* seeds at various stages of maturity. Greenhouse, 1986.

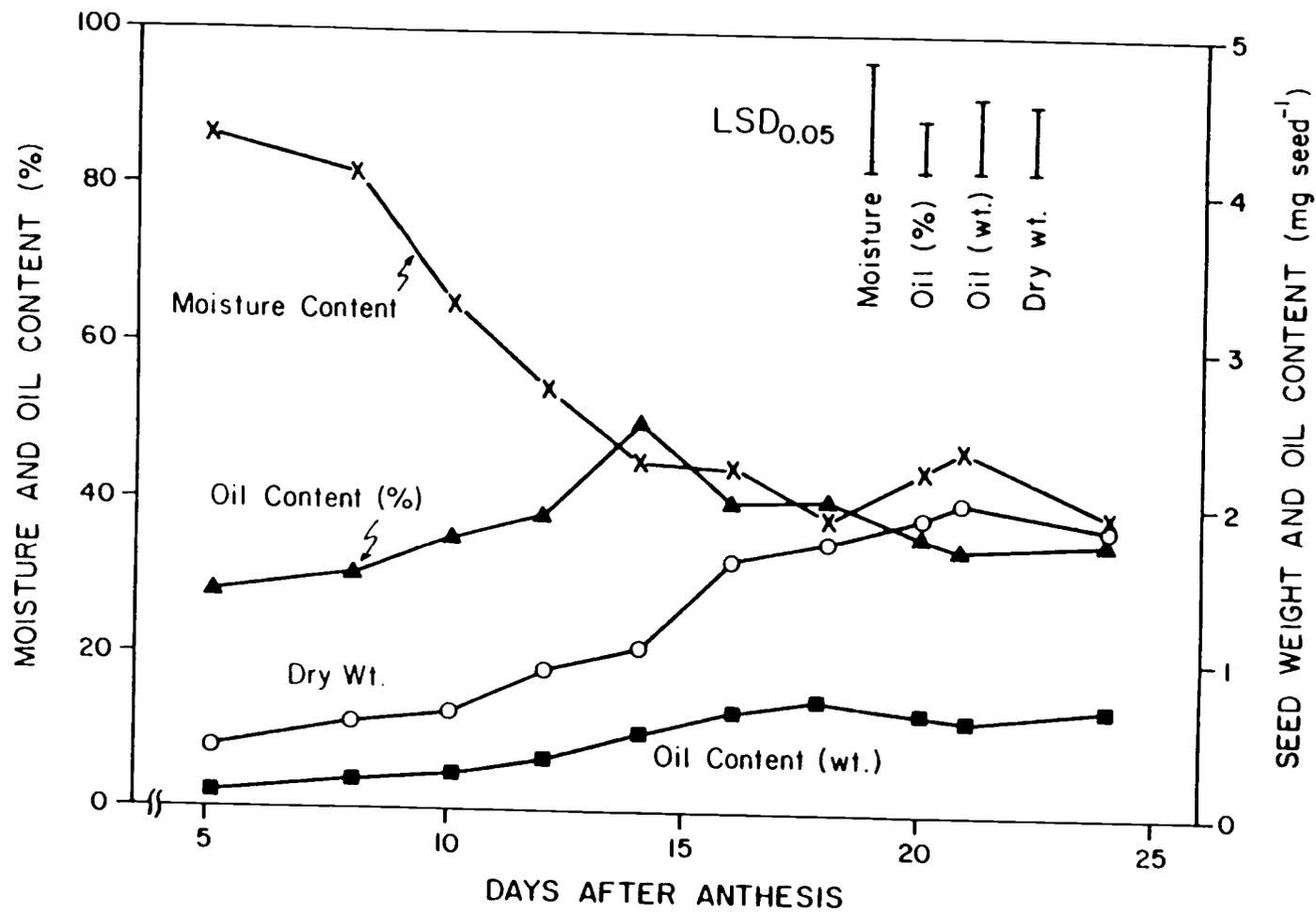


Figure 5. Dry weight, moisture and oil content of *Cuphea lutea* seeds at various stages of maturity. Field, 1985.

## REFERENCES

- Anderson, S. R. 1955. Cultural and harvest practices affecting seed yields of birdsfoot trefoil, Lotus corniculatus L. Agron. J. 47:483-487.
- Babayan, V. K. 1981. Medium chain length fatty acid esters and their medicinal and nutritional application. J. Am. Oil Chem. Soc. 58:49-51A.
- Bartel, A. T. 1941. Green seeds in immature small grains and their relation to germination. J. Am. Soc. Agron. 33:732-738.
- Board, J. E., and W. Hall. 1984. Premature flowering in soybean yield reduction at nonoptimal planting dates as influenced by temperature and photoperiod. Agron. J. 76:700-704.
- Browne, C. L. 1978. Identification of physiological maturity in sunflowers (Helianthus annuus). Aust. J. Exp. Agric. Anim. Husb. 18:282-286.
- Clark, L. E., J. W. Collier, and R. Langston. 1968. Dormancy in Sorghum bicolor (L.) Moench. II. Effect of pericarp and testa. Crop Sci. 8:155-158.
- Comstock, V. E., and J. O. Culbertson, 1958. A rapid method of determining the oil content of the seed and iodine values of the oil from small samples of Flaxseed. Agron. J. 50:113-114.
- Copeland, L. O., and M. B. McDonald. 1985. Principles of seed science and technology. 2nd ed. Burgess Publishing Co., Minneapolis, MN.
- Delouche, J. C. 1958. Germination of Kentucky bluegrass harvested at different stages of maturity. Proc. Assoc. Off. Seed Anal. 48:81-84.
- Flint, E. P., D. T. Patterson, and J. L. Beyers. 1983. Interference and temperature effects on growth of cotton (Gossypium hirsutum), spurred anoda (Anoda cristata) and velvet leaf (Abutilon theophrasti). Weed Sci. 31:892-898.
- Frey, K. J., E. Ruan, and S. C. Wiggans. 1958. Dry weights and germination of developing oat seeds. Agron. J. 50:248-250.

- Grabe, D. F. 1956. Maturity in smooth bromegrass. *Agron. J.* 48:253-256.
- Hallauer, A. R., and C. M. Harrison. 1962. Estimates of maturity and its inheritance in maize. *Crop Sci.* 2:289-294.
- Harlan, H. V. 1920. Daily development of kernels of Hannchen barley from flowering to maturity at Aberdeen, Idaho. *J. Agric. Res.* 19:393-429.
- Hirsinger, F. 1984. Yield potential of Cuphea, a new crop for lauric and capric seed oils. *J. Am. Oil Chem. Soc.* 62:76-80.
- Hirsinger, F., and P. F. Knowles. 1984. Morphological and agronomic descriptions of selected Cuphea germ-plasm. *Econ. Bot.* 38:439-451.
- Lawn, R. J., and D. J. Hume. 1985. Response of tropical and temperate soybean genotypes to temperature during early reproductive growth. *Crop Sci.* 25:137-142.
- Leininger, L. N., and L. A. Urie. 1964. Development of safflower seed from flowering to maturity. *Crop Sci.* 4:83-87.
- Miller, R. W., F. R. Earle, and I. A. Wolff. 1964. Search for new industrial oils. IX. Cuphea, a versatile source of fatty acids. *J. Am. Oil Chem. Soc.* 41:279-280.
- Robbelen, G., and F. Hirsinger. 1982. Cuphea, the first annual oil crop for the production of medium-chain triglycerides (MCT), p. 161-170. In: Improvement of oil-seed and industrial crops by induced mutations. Panel Proceedings Series. International Atomic Energy Agency, Vienna.
- Schweitzer, L. E., and J. E. Harper. 1985. Effect of hastened flowering on seed yield and dry matter partitioning in diverse soybean genotypes. *Crop Sci.* 25:995-998.
- Shaw, R. H., and W. E. Loomis. 1950. Basis for the prediction of corn yields. *Plant Physiol.* 25:225-244.
- TeKrony, D. M., D. B. Egli, J. Balles, T. Pfeiffer, and R. J. Fellows. 1979. Physiological maturity in soybean. *Agron. J.* 71:771-795.

- Thompson, A. E. 1984. Cuphea -- A potential new crop. Hort. Sci. 19:352-354.
- Wareing, P. F., and I. D. J. Phillips. 1981. Growth and differentiation in plants. 3rd ed. Pergamon Press, New York.
- Went, F. W. 1953. The effect of temperature on plant growth. Ann. Rev. Plant Physiol. 4:347-362.

## BIBLIOGRAPHY

- Aitken, Y. 1955. Flower initiation pasture in legumes. I. Factors affecting flower initiation in Trifolium subterraneum L. Aust. J. Agric. Res. 6:212-246.
- Aldrich, S. R. 1943. Maturity measurements in corn and an indication that grain development continues after premature cutting. J. Am. Soc. Agron. 35:667-680.
- Anderson, S. R. 1955. Cultural and harvesting practices affecting seed yields of birdsfoot trefoil, Lotus corniculatus L. Agron. J. 47:483-487.
- Babayán, V. K. 1981. Medium chain length fatty acid esters and their medicinal and nutritional applications. J. Am. Oil Chem. Soc. 58:49-51A.
- Board, J. E. 1985. Yield components associated with soybean yield reductions at nonoptimal planting dates. Agron. J. 77:135-140.
- Board, J. E., and W. Hall. 1984. Premature flowering in soybean yield reduction at nonoptimal planting dates as influenced by temperature and photoperiod. Agron. J. 76:700-704.
- Browne, C. L. 1978. Identification of physiological maturity in sunflowers (Helianthus annuus). Aust. J. Exp. Agric. Anim. Husb. 18:282-286.
- Burnett, L. C., and A. L. Bakke. 1930. The effect of delayed harvest upon yield of grain. Iowa Agric. Exp. Sta. Res. Bull. 130.
- Clark, L. E., J. W. Collier, and R. Langston. 1968. Dormancy in Sorghum bicolor (L.) Moench. II. Effect of pericarp and testa. Crop Sci. 8:155-158.
- Collier, J. W. 1963. Caryopsis development in several grain sorghum varieties and hybrids. Crop Sci. 3:419-422.
- Collins, W. J., and Y. Aitken. 1970. The effect of leaf removal on flowering time in subterranean clover. Aust. J. Agric. Res. 21:893-903.

- Comstock, V. E., and J. O. Culbertson. 1958. A rapid method of determining the oil content of the seed and iodine values of the oil from small samples of Flaxseed. *Agron. J.* 50:113-114.
- Cooper, C. S., M. A. Hughes, and R. L. Ditterline. 1980. Seedling length day 3 - a simple rapid technique for evaluating seedling vigor of birdsfoot trefoil. *J. Seed Technol.* 5:17-25.
- Copeland, L. O., and M. B. McDonald. 1985. Principles of seed science and technology. 2nd ed. Burgess Publishing Co., Minneapolis, MN.
- Dale, J. E. 1959. Some effects of the continuous removal of floral buds on the growth of the cotton plant. *Ann. Bot.* 23:636-649.
- Delouche, J. C. 1958. Germination of Kentucky bluegrass harvested at different stages of maturity. *Proc. Assoc. Off. Seed Anal.* 48:81-84.
- Dennis, F. G., Jr. 1984. Flowering. p. 237-264. In: M. B. Tesar (ed). *Physiological basis of crop growth and development.* American Society of Agronomy, Crop Science Society of America, Madison, WI.
- Dessureaux, L., N. P. Neal, and R. A. Brink. 1948. Maturation in corn. *J. Am. Soc. Agron.* 40:733-745.
- Frey, K. J., E. Ruan, and S. C. Wiggans. 1958. Dry weights and germination of developing oat seeds. *Agron. J.* 50:248-250.
- Goli, A., and D. B. Weaver. 1986. Defoliation responses of determinate and indeterminate late-planted soybeans. *Crop Sci.* 26:156-159.
- Grabe, D. F. 1956. Maturity in smooth bromegrass. *Agron. J.* 48:253-256.
- Harlan, H. V. 1920. Daily development of kernels of Hannchen barley from flowering to maturity at Aberdeen, Idaho. *J. Agric. Res.* 19:393-429.
- Hirsinger, F. 1984. Yield potential of Cuphea, a new crop for lauric and capric seed oils. *J. Am. Oil Chem. Soc.* 62:76-80.
- Hirsinger, F., and P. F. Knowles. 1984. Morphological and agronomic description of selected Cuphea germplasm. *Econ. Bot.* 38:439-451.

- Hyde, E. O. C., M. A. McLeavy, and G. S. Harris. 1959. Seed development in ryegrass, and in red and white clover. *N. Z. J. Agric. Res.* 2:947-952.
- Janick, J., and A. Whipkey. 1986. In vitro propagation of Cuphea wrightii. *Hort. Sci.* 21:135-137.
- Kennedy, C. W., W. C. Smith, Jr., and J. E. Jones. 1986. Effect of early season square removal on three leaf types of cotton. *Crop Sci.* 26:139-145.
- Kueneman, E. A., and D. H. Wallace. 1979. Simplified growth analysis of non-climbing dry beans at three spacings in the tropics. *Exp. Agric.* 15:273-284.
- Lawn, R. J., and D. J. Hume. 1985. Response of tropical and temperate soybean genotypes to temperature during early reproductive growth. *Crop Sci.* 25:137-142.
- Leininger, L. N., and L. A. Urie. 1964. Development of safflower seed from flowering to maturity. *Crop Sci.* 4:83-87.
- Lucas, E. O., and G. M. Milbourn. 1976. The effect of planting density on the growth of two Phaseolus vulgaris varieties in England. *J. Agric. Sci.* 87:89-99.
- Mauk, C. S., P. J. Breen and H. J. Mack 1983. Yield response of major pod-bearing nodes in bush snap beans to irrigation and plant population. *J. Am. Soc. Hort. Sci.* 108:935-939.
- Miller, R. W., F. R. Earle, and I. A. Wolff. 1964. Search for new industrial oils. IX. Cuphea, a new versatile source of fatty acids. *J. Am. Oil Chem. Soc.* 41:279-280.
- Miwa, T. K., C. R. Smith, Jr., and T. L. Wilson. 1960. Preparation of decanoic acid from Cuphea llavea seed. U. S. Patent 2 964 546. Date issued 13 December.
- Rajanna, B., and C. H. Andrews. 1970. Trends in seed maturation of rice (Oryza sativa L.). *Proc. Assoc. Off. Seed Anal.* 60:188-196.
- Robbelen, G., and F. Hirsinger. 1982. Cuphea, the first annual crop for the production of medium-chain triglycerides (MCT), p. 161-170. In: Improvement of oil-seed and industrial crops by induced mutations. Panel Proceedings Series. Internl. Atomic Energy Agency, Vienna.

- Schweitzer, L. E., and J. E. Harper. 1985. Effect of hastened flowering on seed yield and dry matter partitioning in diverse soybean genotypes. *Crop Sci.* 25:995-998.
- Seddigh, M., and G. D. Jolliff. 1984. Effect of night temperature on dry matter partitioning and seed growth of indeterminate field-grown soybean. *Crop Sci.* 24:704-710.
- Shaw, R. H., and W. E. Loomis. 1950. Basis for the prediction of corn yields. *Plant Physiol.* 25:225-244.
- Shaw, R. H., and H. C. S. Thom. 1951. On the phenology of field corn, silking to maturity. *Agron. J.* 43:541-546.
- Slabas, A. R., P. A. Roberts, J. Ormesher, and E. W. Hammond. 1982. A model system for studying the mechanism of medium-chain fatty acid biosynthesis in plants. *Biochem. Biophys. Acta.* 711:411-420.
- Taylor, G. B. 1980. Interrelations between seeds and associated fruiting structures during their development in Trifolium subterraneum L. *Ann. Bot.* 46:323-331.
- TeKrony, D. M., D. B. Egli, J. Balles, T. Pfeiffer, and R. J. Fellows. 1979. Physiological maturity in soybean. *Agron. J.* 71:771-795.
- Thompson, A. E. 1984. Cuphea -- A potential new crop. *Hort. Sci.* 19:352-354.
- Went, F. 1953. The effect of temperature on plant growth. *Ann. Rev. Plant Physiol.* 4:347-362.

APPENDIX

Appendix Table 1. Effect of temperature, light and seed age on germination of three species of Cuphea

Temperature	Species	Age of seed (months)						Temperature mean	
		6		12		18		Light	Dark
		Light	Dark	Light	Dark	Light	Dark		
°C		%							
10	<u>C. laminuligera</u>	4	9	11	19	21	28		
	<u>C. lutea</u>	7	35	67	69	71	68		
	<u>C. wrightii</u>	0	0	0	0	3	0	20	25
15	<u>C. laminuligera</u>	39	36	88	77	77	68		
	<u>C. lutea</u>	87	76	96	93	91	87		
	<u>C. wrightii</u>	31	40	88	88	56	17	62	61
20	<u>C. laminuligera</u>	59	29	81	81	73	76		
	<u>C. lutea</u>	91	75	100	92	96	92		
	<u>C. wrightii</u>	37	12	91	88	60	31	76	64
25	<u>C. laminuligera</u>	37	44	79	88	87	77		
	<u>C. lutea</u>	84	29	92	52	92	83		
	<u>C. wrightii</u>	31	11	96	87	57	52	73	58
30	<u>C. laminuligera</u>	36	27	88	85	84	64		
	<u>C. lutea</u>	37	13	63	27	79	13		
	<u>C. wrightii</u>	19	53	88	92	45	35	60	36
10/20	<u>C. laminuligera</u>	55	37	83	88	75	56		
	<u>C. lutea</u>	95	89	99	97	95	85		
	<u>C. wrightii</u>	15	13	84	75	43	21	71	63
15/25	<u>C. laminuligera</u>	67	55	80	92	77	68		
	<u>C. lutea</u>	92	95	100	95	95	97		
	<u>C. wrightii</u>	44	13	89	89	80	41	80	72
20/30	<u>C. laminuligera</u>	43	44	95	94	75	74		
	<u>C. lutea</u>	91	89	92	96	95	97		
	<u>C. wrightii</u>	25	25	91	90	63	63	75	75
Age means		47	36	81	76	70	58		
LSD <sub>0.05</sub>		20	15	20	15	20	15		

Appendix Table 1. (Cont.)

Species x age means (averaged over temperature)

Species	6		12		18		Species mean		LSD <sub>0.05</sub>	
	Light	Dark	Light	Dark	Light	Dark	Light	Dark	Light	Dark
	§									
<i>C. laminuligera</i>	42	35	76	78	71	64	63	59	6	5
<i>C. lutea</i>	73	61	89	75	89	78	84	71		
<i>C. wrightii</i>	25	11	78	76	51	33	51	40		
LSD <sub>0.05</sub>	Light 6	Dark 5								

Age x temperature means (averaged over species)

Temperature °C	6		12		18		Species mean		LSD <sub>0.05</sub>	
	Light	Dark	Light	Dark	Light	Dark	Light	Dark	Light	Dark
	§									
10	4	15	80	29	32	32	20	25	10	8
15	62	39	91	87	76	66	76	64		
25	51	28	91	77	79	71	73	58		
30	31	11	26	60	69	37	60	36		
10/20	47	47	92	87	71	54	71	63		
15/25	55	54	90	92	84	69	80	72		
20/30	68	53	88	94	78	78	75	75		
LSD <sub>0.05</sub>	Light 12	Dark 8								

Appendix Table 1. (Cont.)

Temperature x species means (averaged over age)

Temperature	<u>C. laminuligera</u>		<u>C. lutea</u>		<u>C. wrightii</u>		<u>Temperature mean</u>		LSD <sub>0.05</sub>	
	Light	Dark	Light	Dark	Light	Dark	Light	Dark	Light	Dark
°C	§									
10	12	19	48	57	1	0	20	25	11	8
15	68	61	91	85	58	36	72	61		
20	71	62	96	86	63	44	76	64		
25	68	70	89	55	61	50	73	58		
30	69	59	60	58	51	44	60	36		
10/20	71	60	96	91	47	36	71	63		
15/25	75	72	96	96	71	48	80	72		
20/30	71	71	93	94	60	60	75	75		
LSD <sub>0.05</sub>	Light	Dark								
	13	9								
LSD <sub>0.05</sub>			<u>Light</u>		<u>Dark</u>					
Age			9		7					
Species			4		3					
Temperature			4		3					
Temperature x species x age			17		14					

Appendix Table 2. Effect of harvesting date on seed yield, oil content, weight and color of two Cuphea species in the field.

Days after planting	Seed Produced			1000-seed weight	Oil content <sup>+</sup>	Green seed
	Total	Recovered	Shattered			
	kg ha <sup>-1</sup>					
<u>Cuphea wrightii</u> (1984)						
105		10.40		1.586	37.9	
111		7.59		1.413	39.2	51.5
118		30.23		1.426	38.0	62.0
127		85.60		1.637	36.4	52.0
134		30.28		1.613	37.1	70.8
141		37.41		1.518	38.7	88.6
LSD <sub>0.05</sub>		15.61		NS	NS	3.9
<u>Cuphea lutea</u> (1985)						
114	22.40	15.22	7.18	2.608	36.8	37.8
121	16.01	1.30	14.71	2.240	35.9	77.3
130	4.65	2.71	1.94	2.435	33.7	78.3
138	24.77	21.63	3.14	2.377	30.8	30.9
145	50.47	33.54	16.93	2.673	31.1	20.5
152	44.66	12.02	32.64	2.272	32.6	82.9
163	48.95	2.18	46.77	1.629	33.1	96.4
LSD <sub>0.05</sub>	8.50	6.72	6.95	0.091	NS	11.2

<sup>+</sup> Calculated on dry matter basis.

Appendix Table 3. Effect of multiple harvests on seed yield, weight and oil content of two Cuphea species in the field. Yield is total of several harvests and weight and oil content are averages of separate harvests at different dates on the same plot.

Number of harvests	Recovered yield	1000-seed weight	Oil <sup>+</sup> content
	kg ha <sup>-1</sup>	g	%
<u>Cuphea wrightii</u> (1984)			
1	107.41	1.482	35.6
2	156.23	1.514	34.8
3	204.03	1.609	35.2
LSD <sub>0.05</sub>	32.91	NS	NS
<u>Cuphea lutea</u> (1985)			
1	21.63	2.377	30.8
2	30.86	2.442	32.7
3	32.98	2.283	31.2
4	39.94	2.279	34.3
LSD <sub>0.05</sub>	10.56	0.098	NS

<sup>+</sup>Calculated on dry matter basis.

Appendix Table 4. Seed yield, weight and oil content of two Cuphea species harvested at different dates in the greenhouse.

Days after Planting	Yield		1000-seed weight		Oil Content <sup>*</sup>		
	<u>C. lutea</u>	<u>C. wrightii</u>	<u>C. lutea</u>	<u>C. wrightii</u>	<u>C. lutea</u>	<u>C. wrightii</u>	
	— g plant <sup>-1</sup> —		— g —		— % —		
			<u>1985</u>				
69	0.6	1.8	2.623	1.700	30.7	37.5	
77	1.6	4.3	2.835	2.137	29.7	35.9	
104	9.3	9.5	3.183	2.254	36.5	35.3	
	Species mean	3.8	5.2	2.880	2.030	31.5	36.6
LSD <sub>0.05</sub>	Species	0.4		0.142		0.5	
	Harvest dates	0.5		NS		0.7	
	Species x har- vest dates	NS		NS		NS	
			<u>1986</u>				
86	0.9	1.5	2.895	1.995	32.3	35.4	
106	2.3	2.5	2.827	1.843	33.4	37.3	
126	2.8	2.8	2.717	1.937	34.7	36.9	
146	3.4	4.3	2.646	1.793	34.8	37.4	
	Species mean	2.3	2.8	2.771	1.892	33.8	36.8
LSD <sub>0.05</sub>	Species	0.2		0.136		0.6	
	Harvest dates	0.3		NS		0.9	
	Species x har- vest dates	NS		NS		NS	

<sup>\*</sup> Calculated on dry matter basis.

Appendix Table 5. Effect of multiple harvests on seed yield, weight and oil content of two *Cuphea* species grown in the greenhouse in 1986. Yield is total of several harvests and weight and oil content are averages of separate harvests at different dates from the same plants.

Number of harvests	Yield		1000-seed weight		Oil Content <sup>+</sup>	
	<i>C. lutea</i>	<i>S. wrightii</i>	<i>C. lutea</i>	<i>C. wrightii</i>	<i>C. lutea</i>	<i>C. wrightii</i>
	g plant <sup>-1</sup>		g		%	
1	1.165	1.646	2.791	2.002	34.3	37.3
2	2.318	3.393	2.821	1.954	34.2	37.6
3	3.415	4.563	2.701	1.920	33.8	36.7
4	3.660	3.896	2.743	1.933	33.5	37.2
Species mean	2.640	3.375	2.764	1.952	34.0	37.2
LSD <sub>0.05</sub> Species		0.286		0.367		0.8
No. of harvests		0.405		NS		NS
Species x harvests		NS		NS		NS

<sup>+</sup> Calculated on dry matter basis.

Appendix Table 6. Effect of seed color on oil content<sup>+</sup> of two Cuphea species grown in the greenhouse in 1986

Seed color	Species		Seed color mean
	<u>C. lutea</u>	<u>C. wrightii</u>	
	%		
Green	32.2	36.2	34.2
Yellow <sup>‡</sup>	33.7	35.3	34.5
Dark brown	33.3	35.7	34.5
Species mean	33.1	35.7	
LSD <sub>0.05</sub>			
Species		0.9	
Seed Color		NS	
Species x seed color		NS	

<sup>+</sup>Calculated on dry matter basis.

<sup>‡</sup>In C. lutea the seed is purple.

Appendix Table 7. Effect of temperature on early vegetative growth of four *Cuphea* species in growth chambers. Daylength was 16 h and PPFD was 200  $\mu\text{mol m}^{-2} \text{s}^{-1}$ .

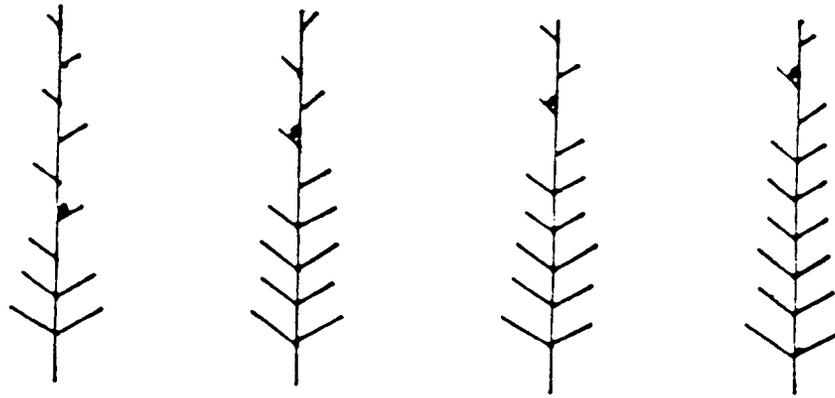
Days after Planting	Temperature	Plant height				Temp. mean	Number of main stem nodes				Temp mean
		<u>laminu-igera</u>	<u>lepto-poda</u>	<u>lutea</u>	<u>wrightii</u>		<u>laminu-ligera</u>	<u>lepto-poda</u>	<u>lutea</u>	<u>wrightii</u>	
		cm					no.				
15	20/30	53	54	48	70	56	3.1	2.4	2.8	3.0	2.8
	10/25	30	30	24	25	27	2.1	2.0	2.1	2.0	2.1
	Species mean	42	42	36	48		2.6	2.2	2.5	2.5	
	LSD <sub>0.05</sub>										
	Species			6					0.1		
	Temperature			4					0.2		
	Species x temperature			8					0.2		
24	20/30	113	100	114	130	114	5.6	5.0	4.5	4.9	5.0
	10/25	68	46	66	63	61	3.9	4.0	3.3	3.5	3.7
	Species mean	91	73	90	97		4.8	4.5	3.9	4.2	
	LSD <sub>0.05</sub>										
	Species			10					0.4		
	Temperature			7					0.2		
	Species x temperature			NS					NS		

Appendix Table 8. Height, nodes, branches and flowers of four Cuphea species at different dates in the greenhouse.

Characteristic	Days after planting	Species				LSD <sub>0.05</sub>
		<u>laminuligera</u>	<u>leptopoda</u>	<u>lutea</u>	<u>wrightii</u>	
<u>1985</u>						
Branches						
Primary, no. plant <sup>-1</sup>	35	12.8	11.6	9.0	8.2	NS
Secondary <sub>1</sub> no. primary branch	47	0.1	1.7	2.2	3.9	0.2
	61	1.7	2.4	6.1	5.9	0.8
Main stem <sub>1</sub> nodes, no. plant <sup>-1</sup>	18	3.8	2.8	2.9	3.4	NS
	37	11.9	8.6	8.9	14.8	1.2
	47	20.8	18.7	13.9	22.8	2.6
	61	26.9	31.0	17.3	27.7	1.8
<u>1986</u>						
Height, mm	19	53	52	35	86	14
	26	87	79	73	147	15
	35	199	150	178	322	33
	44	314	241	267	423	22
	72	788	633	442	487	57
	128	973	991	769	567	44

Appendix Table 8. (Cont.)

Branches						
Primary, no. plant <sup>-1</sup>	19	0.5	0.3	0.8	1.2	NS
	26	1.1	0.7	2.7	4.2	NS
	35	4.8	3.2	8.2	6.3	NS
	44	9.6	7.4	9.9	9.5	NS
	72	11.3	12.7	18.7	10.3	2.5
	128	33.7	21.6	24.3	15.3	2.6
Secondary, no. primary branch <sup>-1</sup>						
	72	6.7	7.2	12.0	6.2	0.8
	128	1.9	2.3	3.7	2.2	0.3
Main stem nodes, no.						
	19	4.1	3.4	3.3	4.0	NS
	62	5.7	4.6	5.1	5.6	NS
	35	10.5	7.2	8.3	12.3	1.2
	44	14.0	9.2	9.7	16.2	2.2
	72	33.0	43.0	21.7	18.7	1.2
	128	60.0	63.0	27.1	28.4	8.9
Flowers						
no. main stem <sup>-1</sup>	72	24.2	26.0	12.3	20.7	1.4
	128	76.0	97.1	21.7	74.3	3.5
no. primary branch <sup>-1</sup>	72	62.0	33.7	33.0	58.7	11.4
	128	139.0	278.3	50.3	108.7	3.5



C. wrightii   C. lutea   C. leptopoda   C. laminuligera

Appendix Figure 1. Branching and flowering habits of four Cuphea species under greenhouse conditions. First flowers form in axil of second single branch in each species.