



AN ABSTRACT OF THE DISSERTATION OF

Liang Lu for the degree of Doctor of Philosophy in Chemistry presented on August 24, 2009.

Title: Part I: Synthetic Studies Toward the Southern Portion of Azaspiracid-1; Part II: Total Synthesis of Amphidinolide B<sub>1</sub> and the Proposed Structure of Amphidinolide B<sub>2</sub>

Abstract approved : \_\_\_\_\_

Rich G. Carter

The structural architecture present in marine toxin azaspiracid - 20 stereocenters, 9 rings, 3 separated spirocenters - has attracted considerable synthetic attention. Our efforts toward the synthesis of azaspiracid have led to the completion of both C<sub>1</sub>-C<sub>26</sub> northern and C<sub>27</sub>-C<sub>40</sub> southern halves. Herein, the synthesis of southern FGHI ring system is described. The key steps included an Andrus anti-aldol coupling to furnish the C<sub>32</sub>, C<sub>33</sub> stereocenters, an acid-catalyzed ketalization to furnish FG rings, and a Yb(OTf)<sub>3</sub>-mediated spiroaminal formation to generate I ring.

The first total synthesis of cytotoxic macrolides amphidinolide B<sub>1</sub> and the

proposed structure of amphidinolide B<sub>2</sub> have been accomplished. The key developed protocols include a metal catalyst-free sequence for the synthesis of the diene subunit, a non-chelation-controlled aldol coupling to install the C<sub>18</sub> stereocenter, an efficient macrocyclization of the 26-membered lactone ring, and the incorporation of the labile allylic epoxide moiety.

The unique structure of the highly substituted diene functionality represents significant synthetic challenges. A Wittig / HWE reaction sequence yielded the C<sub>13</sub>-C<sub>15</sub> diene moiety in good yield in excellent diastereoselectivity. Subsequent Sharpless epoxidation and Red-Al-mediated regioselective epoxide opening gave the C<sub>16</sub> tertiary alcohol.

The protecting groups on C<sub>21</sub> were discovered to have significant effects on the aldol reaction between C<sub>9</sub>-C<sub>18</sub> aldehyde and C<sub>19</sub>-C<sub>25</sub> methyl ketone. Although chelating groups such as PMB, Bn afforded 18*S* isomer as a single diastereomer, the removal of these groups has proven problematic. Non-chelating silyl group generated 18*R* isomer in 8:1 dr at -100°C, while the 18*S* stereomer was obtained at -40°C in 1.2:1 dr.

A spontaneous intramolecular Wadsworth–Emmons olefination established the 26-membered macrocycle. The oxidation and *in situ* elimination of a selenide moiety proceeded smoothly in the presence of free alcohols using TMSOOTMS. The first total synthesis of amphidinolide B<sub>1</sub> and the proposed structure of amphidinolide B<sub>2</sub> were accomplished in 29 linear steps. Additionally, We

discovered that the initially proposed structure of amphidinolide B<sub>2</sub> was incorrect.



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Part I: Synthetic Studies Toward the Southern Portion of Azaspiracid-1;  
Part II: Total Synthesis of Amphidinolide B<sub>1</sub> and the Proposed Structure of  
Amphidinolide B<sub>2</sub>

by  
Liang Lu

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Doctor of Philosophy dissertation of Liang Lu presented on August 24, 2009

APPROVED:

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Major Professor, representing Chemistry

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Chair of the Department of Chemistry

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Dean of the Graduate School

I understand that my dissertation will become part of the permanent collection of Oregon State University libraries. My signature below authorizes release of my dissertation to any reader upon request.

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Liang Lu, Author

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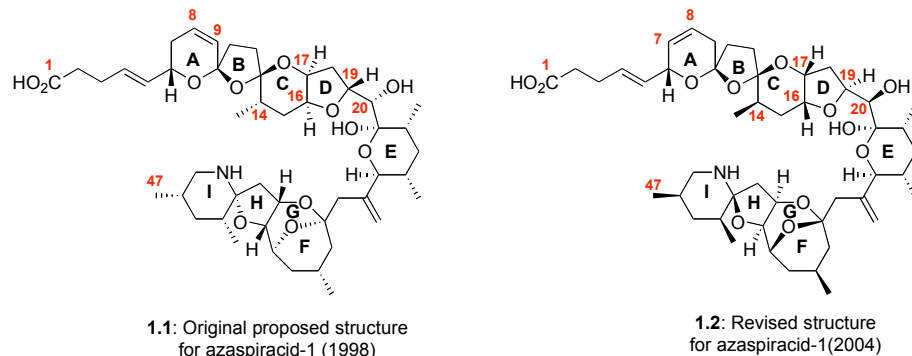
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## **PART I: SYNTHETIC STUDIES TOWARD THE SOUTHERN PORTION OF AZASPIRACID-1**

### **CHAPTER 1. BACKGROUND OF AZASPIRACID**

#### **1.1 Discovery and Bioactivities of Azaspiracid-1**

Azaspiracid poisoning is a recent toxic syndrome first reported in 1995, when several individuals became ill after consuming mussels harvested from Killary Harbor in Ireland.<sup>1</sup> An active search for the causative toxin led to the isolation of azaspiracid-1 by the Satake group in 1998.<sup>2</sup> The initial structure of azaspiracid-1 was proposed based on extensive 2D NMR studies;<sup>2</sup> however, this original structure has been recently discredited and was revised by Nicolaou and co-workers in 2004.<sup>3</sup> Independently and concurrently, our laboratory had converged on the same stereochemical conclusion.<sup>4</sup> The major stereochemical errors were believed to be in the ABCDE northern portion of the molecule. In addition to the inverted stereochemical configurations of C<sub>14</sub>, C<sub>16</sub>, C<sub>17</sub>, C<sub>19</sub> and C<sub>20</sub>, the southern FGHI ring system was found to be enantiomeric to the proposed structure and the C<sub>8,9</sub> olefin in the A ring proved to be actually in C<sub>7,8</sub> position. After structure elucidation of azaspiracid-1, a total of more than 30 azaspiracid analogues differing slightly in their methylation and hydroxylation patterns have subsequently been described and their structure was determined using tandem mass spectrometry and NMR spectroscopy.<sup>5</sup>



**Figure 1.1.** Originally Proposed and Revised Structures of Azaspiracid-1

A marine dinoflagellate was proposed to be the origin of azaspiracids<sup>6</sup> and they have been discovered in multiple shellfish species including mussels, oysters, scallops, clams, *etc.*<sup>7</sup> Human consumption of azaspiracid-contaminated shellfish can result in severe acute symptoms such as nausea, vomiting, diarrhea, and stomach cramps.<sup>1</sup> Although there is no information about toxicity of these analogues to humans, azaspiracid-1 is known to possess toxicity *in vitro* with a lethal dose in mice of 0.2 mg / kg.<sup>2</sup> The mechanism by which azaspiracids induce their toxic effects and their biological target/s is still unknown;<sup>8</sup> however, several effects on *in vitro* cell cultures have been revealed for azaspiracid-1 including cytoskeletal alterations,<sup>9</sup> caspase activation,<sup>10</sup> cytotoxicity,<sup>11</sup> cytosolic calcium levels modulation,<sup>12</sup> and alteration of neuronal network.<sup>13</sup> The considerable toxicity and the mechanistic elusiveness have made azaspiracids a significant threat to the shellfish industry and human health. This situation is further complicated by the scarce amount of azaspiracids obtained from natural sources.<sup>6</sup>

## 1.2 Synthetic Efforts Toward Azaspiracid-1

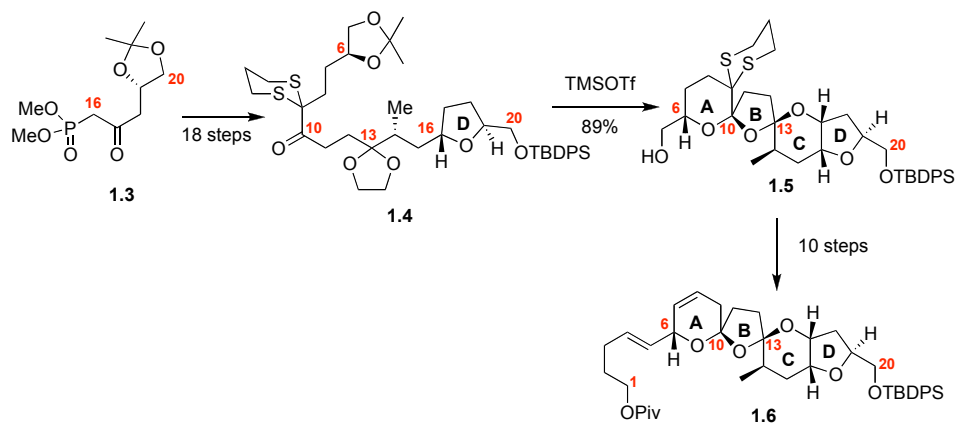
The intriguing structural architecture (20 stereocenters, 9 rings, 3 spirocenters) of azaspiracid-1 has attracted considerable attention from the synthetic community, in particular by the research groups of Carter,<sup>4, 14</sup> Nicolaou,<sup>3, 15</sup> Evans,<sup>16</sup> Forsyth,<sup>17</sup> Sasaki,<sup>18a, 18e</sup> and Mootoo.<sup>18h</sup> The extensive efforts led to the first total synthesis of (-)-azaspiracid-1 and the correction of its structural assignment by the Nicolaou group in 2004.<sup>3</sup> In 2006, Nicolaou and co-workers reported an improved synthesis of (-)-azaspiracid-1.<sup>15g</sup> Besides Nicolaou's landmark work, several partial synthetic studies<sup>4, 14, 17, 18</sup> and Evans' total synthesis of (+)-azaspiracid-1 have also been communicated.<sup>16</sup>

### 1.2.1 Nicolaou's First-Generation Total Synthesis of (-)-azaspiracid-1

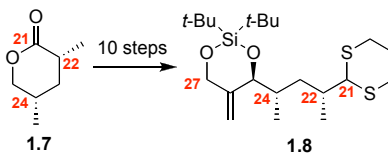
In 2004, Nicolaou and co-workers reported the conquest of (-)-azaspiracid-1 as well as the correction of its originally proposed structure (Scheme 1.1).<sup>3</sup> Nicolaou's approach disconnected the complex molecule into three key building blocks: C<sub>1</sub>-C<sub>20</sub> ABCD ring domain, C<sub>21</sub>-C<sub>27</sub> E ring fragment and C<sub>28</sub>-C<sub>40</sub> FGHI ring system. The ABCD ring system found in compound **1.5** was accessed via TMSOTf catalyzed polycyclization, whereas the the C<sub>22</sub> and C<sub>24</sub> stereocenters in C<sub>21</sub>-C<sub>27</sub> fragment **1.8** were obtained from the known lactone **1.7**.<sup>19</sup> The key steps in the synthesis of FGHI ring system included a Yb(OTf)<sub>3</sub>- or Nd(OTf)<sub>3</sub>-mediated highly stereoselective spiroaminal formation to afford compound **1.15**.



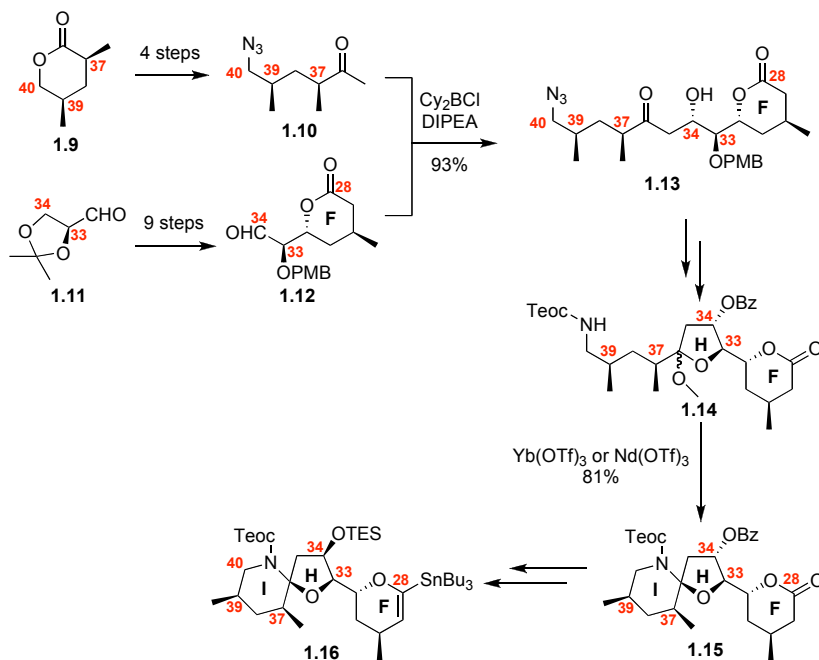
Synthesis of ABCD ring motif:



Synthesis of C<sub>21</sub>-C<sub>27</sub> fragment:

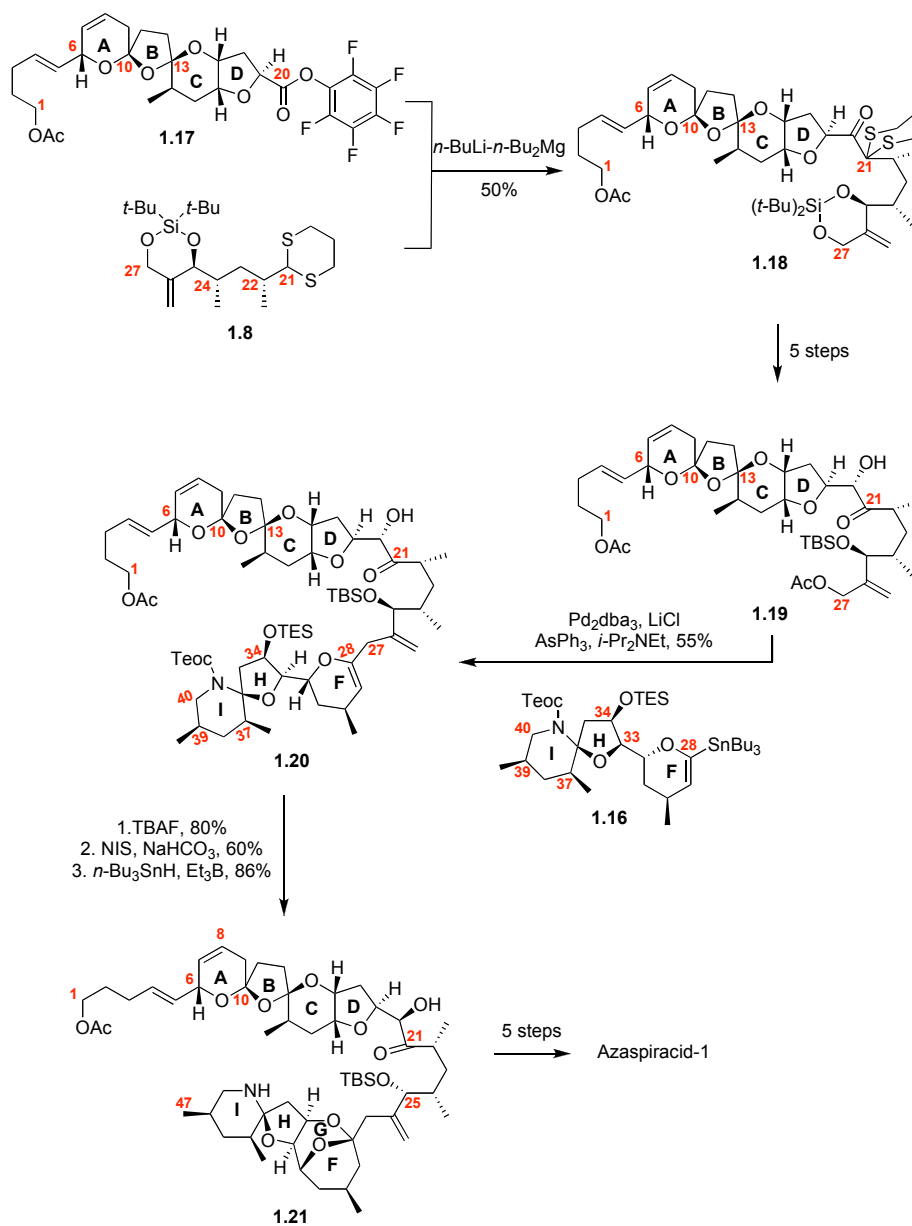


Synthesis of C<sub>28</sub>-C<sub>40</sub> southern portion:



**Scheme 1.1.** Nicolaou's Strategy for the Synthesis of Three Major Fragments

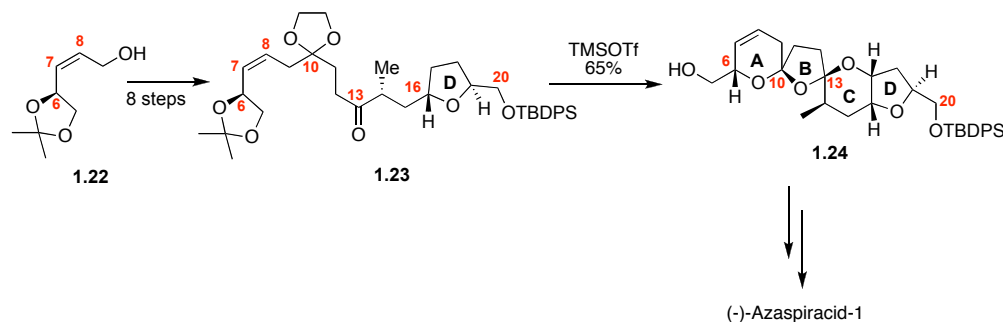
The coupling of these fragments and the completion of the synthesis is shown in Scheme 1.2. The addition of the stabilized dithiane anion to pentafluorophenol ester **1.17** formed C<sub>21</sub>-C<sub>20</sub> bond. The following Stille coupling between allylic acetate **1.19** and stannane **1.16** furnished compound **1.20**, which contains all carbon atoms needed for the azaspiracid-1 structure. In the presence of NIS, G ring was produced via an intramolecular iodoetherification. After the spontaneous formation of E ring during the global desilylation, (-)-azaspiracid-1 was obtained in sequence of 50 longest linear steps.



**Scheme 1.2.** Nicolaou's First-Generation Total Synthesis of (-)-Azaspiracid-1

### 1.2.2 Nicolaou's Second-Generation Total Synthesis of (-)-Azaspiracid-1

In 2006, the Nicolaou group reported their second-generation total synthesis of (-)-azaspiracid-1.<sup>15g</sup> The major improvement of the modified synthesis rest on the construction of ABCD ring fragment. Instead of a dithiane functionality at C<sub>9</sub>, the key TMSOTf-mediated ring-closing cascade was conducted with C<sub>7,8</sub> alkene in place. After obtaining ABCD ring fragment **1.23**, (-)-azaspiracid-1 was synthesized via the analogous sequence used in Nicolaou's first-generation synthesis. The new strategy afforded the natural product in 39 linear steps.



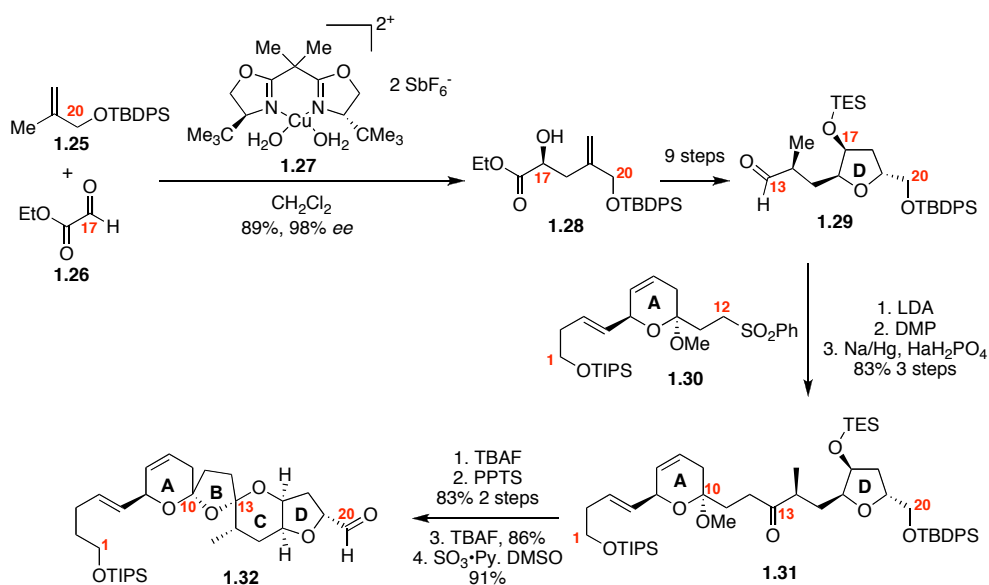
**Scheme 1.3.** Nicolaou's Second-Generation Total Synthesis of (-)-Azaspiracid-1

### 1.2.3 Evans' Total Synthesis of (+)-Azaspiracid-1

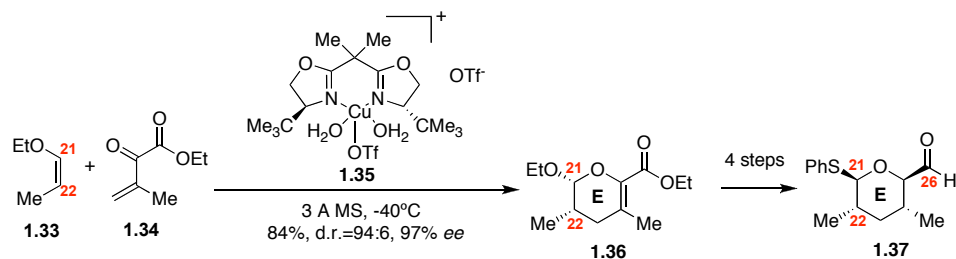
In 2007, the Evans group accomplished the total synthesis of (+)-azaspiracid-1.<sup>16</sup> Sharing the similar disconnection with Nicolaou's approach, Evans' strategy disassembled (+)-azaspiracid-1 into three portions: C<sub>1</sub>-C<sub>20</sub> ABCD ring moiety, C<sub>21</sub>-C<sub>26</sub> E ring fragment and C<sub>27</sub>-C<sub>40</sub> linear motif. The stereocenter at C<sub>17</sub> existed in compound **1.28** was generated from a highly

enantioselective  $\text{Cu}^{2+}$ -catalyzed glyoxylate-ene reaction, whereas the similar catalyst was also found effective in the Diels-Alder cycloaddition to construct E ring fragment **1.37**. Treatment of ketone **1.31** with TBAF then PPTS in nonpolar solvent initiated a stereoselective polycyclization cascade to yield the desired ABCD ring system.

Synthesis of ABCD ring system:

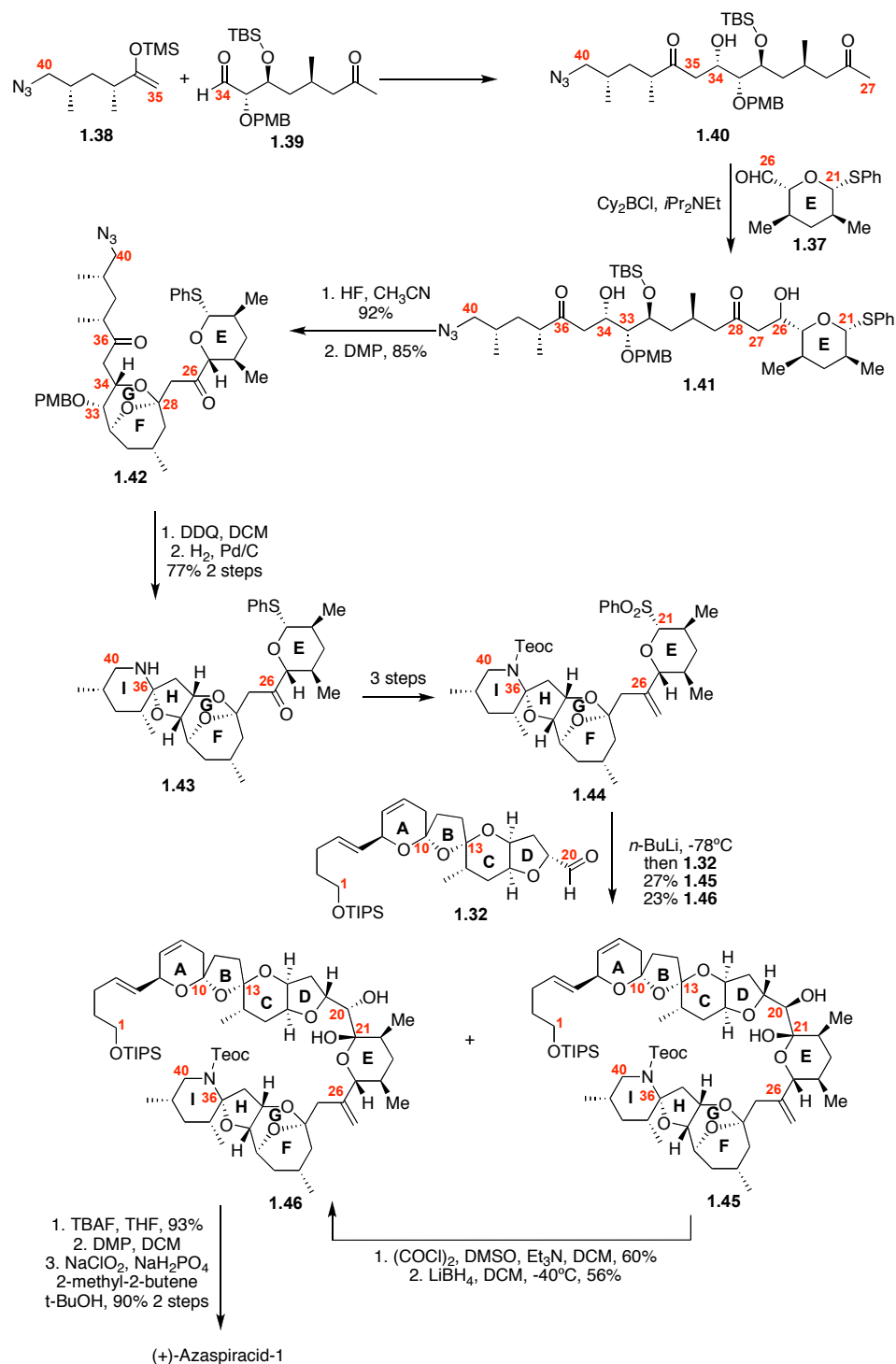


Synthesis of E ring fragment:



**Scheme 1.3.** Evans' Synthesis of ABCD Ring and E Ring Fragments

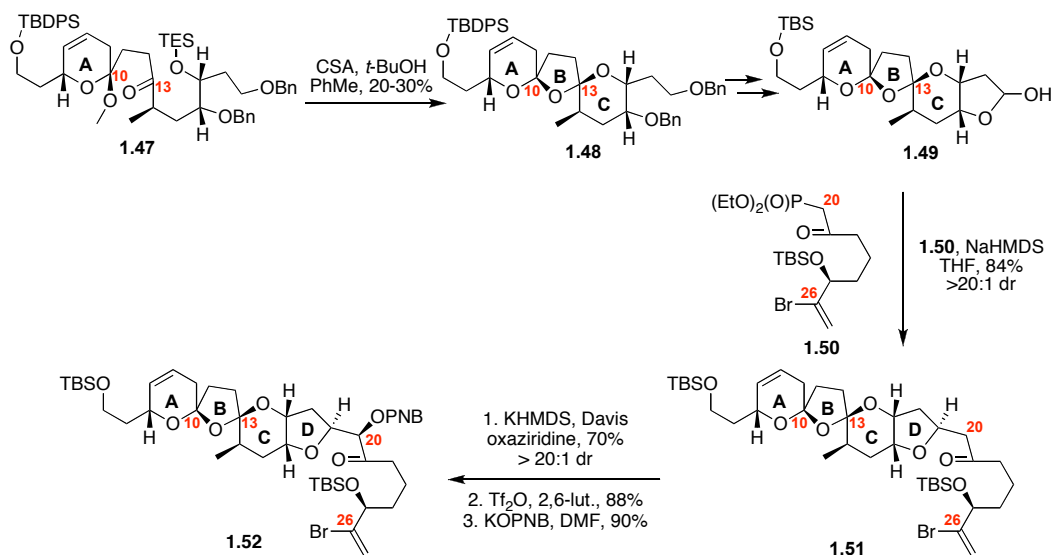
Unlike Nicolaou's synthesis, Evans' approach combined E ring fragment with C<sub>27</sub>-C<sub>40</sub> motif prior to the formation of FGHI ring system (Scheme 1.4). A chelate-controlled Mukaiyama aldol reaction was used to build C<sub>34</sub> stereocenter, while a boron-mediated aldol coupling between methyl ketone **1.40** and aldehyde **1.37** constructed C<sub>26</sub>-C<sub>27</sub> bond. The FGHI ring system **1.43** was constructed via spontaneous ketalization and a spiroaminal formation. Addition of sulfone **1.44** to aldehyde **1.32** followed by a quench at -78°C with pH5 buffer afforded two diastereomers. The undesired alcohol **1.45** was then converted to the desired C<sub>20</sub>-diastereomer **1.46** via a Swern oxidation / LiBH<sub>4</sub> reduction sequence. Further elaboration including the desilylation and a Lindgren-Kraus oxidation (Pinnick oxidation)<sup>20</sup> yielded (+)-azaspiracid-1 in only 26 linear steps.



**Scheme 1.4.** Evans' Total Synthesis of (+)-Azaspiracid-1

## 1.2.4 The Carter Group

Since our first publication in 2000,<sup>14a</sup> our group have made significant contribution to the synthesis of azaspiracid-1.<sup>4, 14</sup> Our conclusion that the correct structure contained the epimeric stereochemistries at C<sub>14</sub>, C<sub>16</sub>, C<sub>17</sub> and C<sub>20</sub> was reported in 2004<sup>4</sup> – independently and concurrently to Nicolaou's efforts.<sup>3</sup> In 2006, we completed the C<sub>1</sub>-C<sub>26</sub> northern half of azaspiracid-1 (Scheme 1.5).<sup>14f</sup> When compound **1.47** was treated with CSA, *t*-BuOH / PhMe, the de-silylation and ketalization proceeded smoothly to give the transoidal bispiroketal **1.48**. Two other highlights of our work are the highly diastereoselective tandem HWE reaction / intramolecular heteratom Michael addition to give compound **1.51** and the highly diastereoselective hydroxylation at C<sub>20</sub> of ketone **1.51**. Our synthesis of C<sub>27</sub>-C<sub>40</sub> southern portion will be discussed in the following chapter.



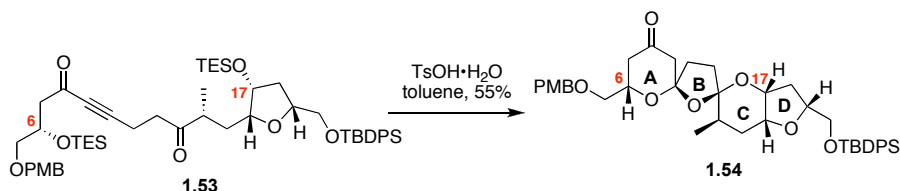
**Scheme 1.5.** Carter's Synthesis of ABCD Ring Fragment



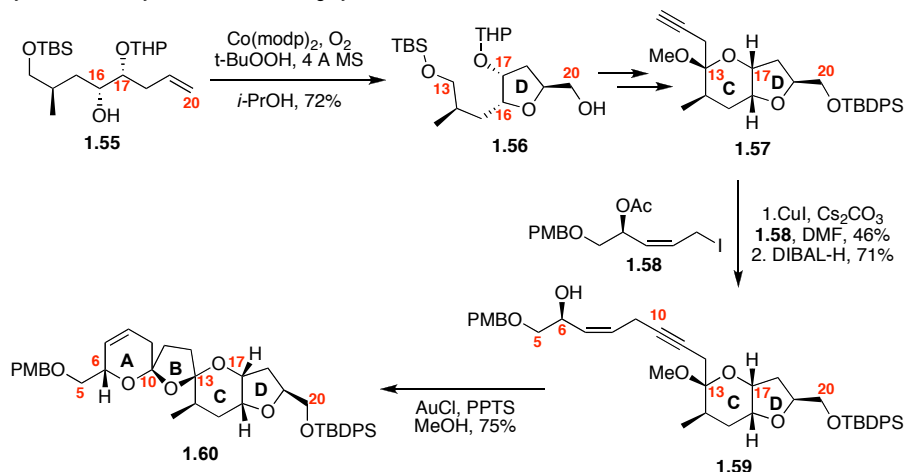
### 1.2.5. The Forsyth Group

Shortly after the elucidation of the structure of azaspiracid-1, Forsyth and co-workers reported a strategy for the synthesis of the ABCD ring trioxadispiroketal (Scheme 1.5).<sup>17f</sup> When ynedione **1.53** was treated with TsOH, selective cleavage of the C<sub>6</sub> and C<sub>17</sub> TES group and the following trioxadispiroketal formation afforded the desired ABCD ring system in a highly diastereoselective manner. Later, a modified synthesis of C<sub>5</sub>-C<sub>20</sub> ABCD ring motif was developed.<sup>17i</sup> In the new strategy, the D ring was obtained from a cobalt-catalyzed oxyetherification. Exposure of enyne **1.59** to Au(I) catalyst led to the bis-spiroketal formation to yield the AB ring moiety.

Forsyth's synthesis of ABCD ring moiety:

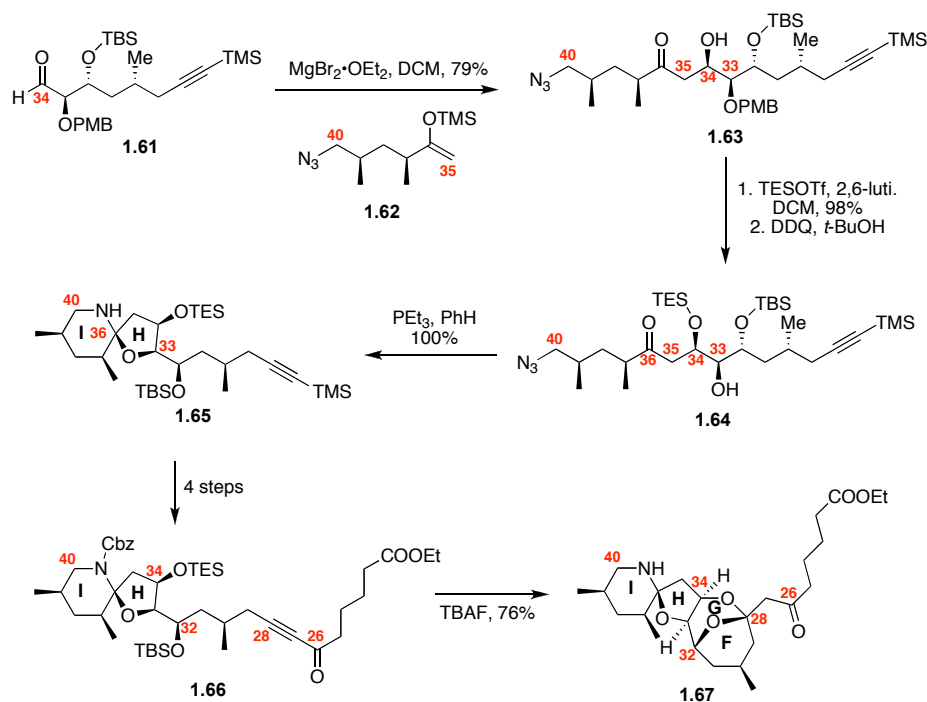


Forsyth's modified synthesis of ABCD ring system:



**Scheme 1.6.** Forsyth's Strategy for the Synthesis of ABCD ring

In 2006, the Forsyth group reported the synthesis of C<sub>26</sub>-C<sub>40</sub> FGHI ring system (Scheme 1.7). The C<sub>34</sub>-C<sub>35</sub> bond was built from a Mukiyama type aldol coupling between **1.61** and **1.62**. PEt<sub>3</sub> mediated azide reduction also induced the spontaneous spiroaminal formation to afford HI ring. Finally, FG ring was installed via a fluoride initiated bis-conjugate addition of C<sub>32</sub> and C<sub>34</sub> hydroxyl groups upon the C<sub>28</sub> Michael acceptor.<sup>17h</sup>

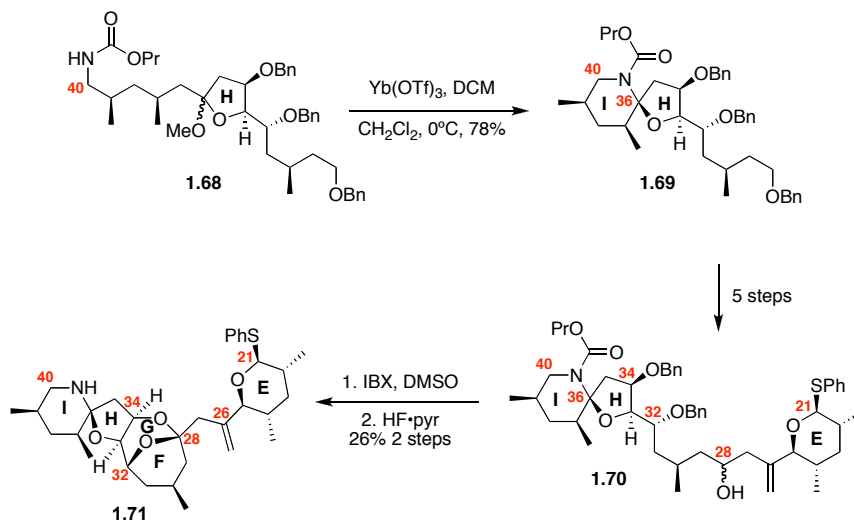


**Scheme 1.7.** Forsyth's Synthesis of C<sub>26</sub>-C<sub>40</sub> FGHI Ring Fragment

### 1.2.6 The Sasaki Group

In 2006, the Sasaki group published the synthesis of C<sub>21</sub>-C<sub>40</sub> EFGHI ring fragment (Scheme 1.8).<sup>18e</sup> The key steps included a Yb(OTf)<sub>3</sub>-catalyzed

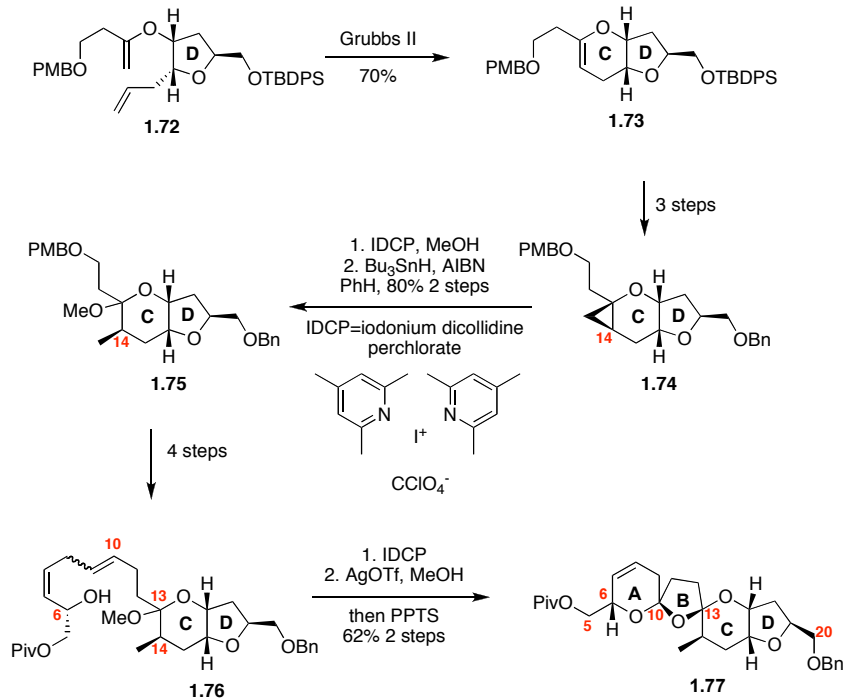
spiroaminal formation to give HI ring and a HF•Pyridine-mediated intramolecular ketalization to afford FG ring. Unfortunately, the C<sub>21</sub>-C<sub>40</sub> portion **1.71** was synthesized in only 0.025% overall yield.



**Scheme 1.8.** Sasaki's Approach for the Synthesis of C<sub>21</sub>-C<sub>40</sub> Portion

### 1.2.7 The Mootoo Group

More recently, the Mootoo group reported their approach for the synthesis of C<sub>5</sub>-C<sub>20</sub> ABCD ring motif (Scheme 1.9).<sup>18h</sup> After the formation of C ring via RCM, subsequent diastereoselective cyclopropanation and opening of the cyclopropane ring afforded C<sub>14</sub> stereocenter. The ketalization initiated by iodonium dicollidine perchlorate (IDCP) and AgOTf gave the desired trioxadispiroketal **1.77**.



**Scheme 1.9.** Mootoo's Synthesis of ABCD Ring System

### 1.3 Conclusion

In summary, the intriguing structure and the unique bioactivity of marine toxin azaspiracid-1 have spurred considerable interests from the synthetic community. These efforts led to the correction of the originally proposed structure in 2004.<sup>3</sup> Subsequent studies resulted in Nicolaou's first-generation and second-generation total syntheses of (-)-azaspiracid-1 with longest linear sequence of 50 and 39 steps, respectively. The enantiomer of (-)-azaspiracid-1, (+)-azaspiracid-1, was later synthesized by Evans and co-workers in only 26 linear steps. Several partial syntheses from the research groups including Carter, Forsyth, Sasaki,

Mootoo, *etc.* have also been reported. Despite all these achievements, there are still several problems such as understanding the controlling features in the formation of the polycyclic systems, more efficient combination of the northern and southern halves, *etc.* deserving more attention from the synthetic chemists. Herein, our endeavors toward the southern portion of azaspiracid-1 are described in the following chapter.

#### 1.4 References

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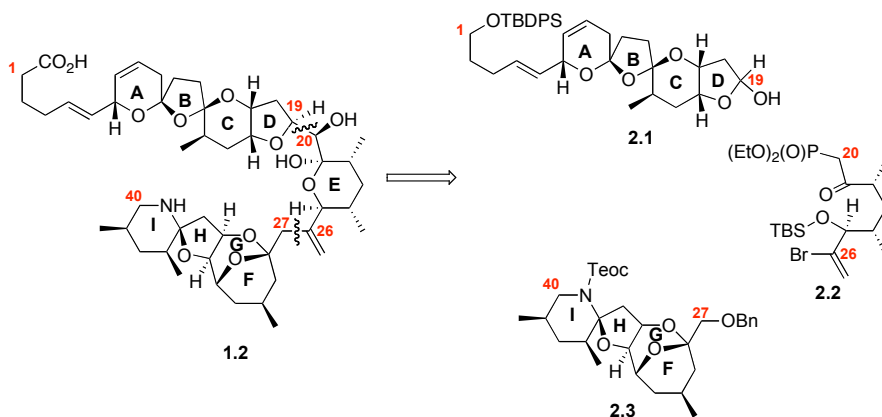
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## CHAPTER 2. STUDIES TOWARD THE SYNTHESIS OF C<sub>27</sub>-C<sub>40</sub> SOUTHERN PORTION OF AZASPIRACID-1

### 2.1 Retrosynthesis of Azaspiracid-1

As was shown in the previous chapter, the unique structural architecture present in azaspiracid-1 (20 stereocenters, 9 rings, 3 spirocenters) has attracted considerable synthetic attention. Our group were particularly drawn to this molecule by the unusual bispiroketal ABCD ring moiety as well as the FGHI ring system containing the spiroaminal and ketal. Our retrosynthesis disconnected azaspiracid-1 into C<sub>1</sub>-C<sub>19</sub> ABCD ring northern fragment **2.1**, C<sub>20</sub>-C<sub>26</sub> motif **2.2**, and C<sub>27</sub>-C<sub>40</sub> FGHI ring southern portion **2.3** (Scheme 2.1). Our endeavors have led to the completion of both C<sub>1</sub>-C<sub>19</sub> and C<sub>20</sub>-C<sub>26</sub> subunits.<sup>1</sup> We also coupled these two substrates successfully to afford the C<sub>1</sub>-C<sub>26</sub> northern halves.<sup>1g</sup> Herein, the studies toward the synthesis of C<sub>27</sub>-C<sub>40</sub> southern portion will be discussed.<sup>2</sup>



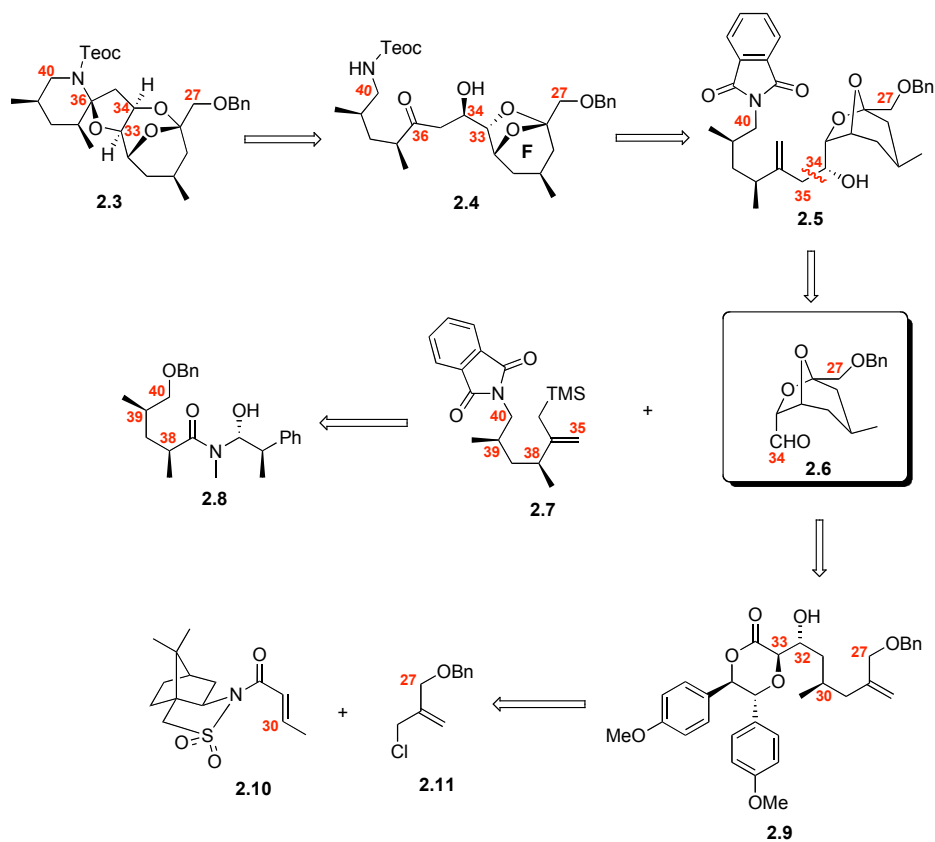
**Scheme 2.1.** Retrosynthetic Analysis of Azaspiracid-1



## 2.2 First-Generation Synthesis of Southern Portion

### 2.2.1 Retrosynthetic Analysis of C<sub>27</sub>-C<sub>40</sub> Southern Portion

Our initial retrosynthetic strategy for the C<sub>27</sub>-C<sub>40</sub> southern portion of azaspiracid-1 cleaved the FGHI ring system via a tandem ring arrangement and spiroaminal formation cascade (Scheme 2.2). Further disconnection at C<sub>34</sub>-C<sub>35</sub> linkage yielded allyl silane **2.7** and aldehyde **2.6**. To establish the correct C<sub>34</sub> stereochemistry, this key coupling would need to proceed via a Cram-chelated intermediate.<sup>3</sup> The allyl silane portion would be available from the known Myers alkylation product **2.8**.<sup>4</sup> The aldehyde **2.6** could be accessible from the Andrus anti-aldol adduct **2.9**,<sup>5</sup> which in turn could be constructed from the known sultam **2.10**<sup>6</sup> and the chloride **2.11**.

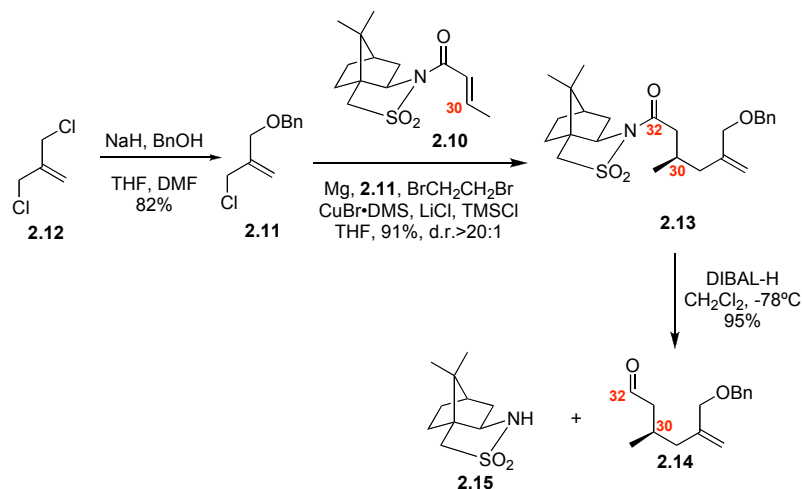


**Scheme 2.2.** Retrosynthesis of Southern Portion **2.3**

### 2.2.2 Synthesis of Aldehyde 2.14

Synthesis of the aldehyde **2.14**, the requisite precursor for the anti-aldol coupling, was accomplished in three steps (Scheme 2.3). Monobenylation of dichloride **2.12**, followed by the cuprate addition on sultam **2.10** under similar conditions described by Paquette and Boulet,<sup>7</sup> generated the stereocenter at C<sub>30</sub> with excellent diastereoselectivity (dr>20:1). It is noteworthy that the preparation of Grignard reagent from allylic chloride **2.11** has extremely low yield (0-10%) due to the undesired Wurtz-type coupling.<sup>8</sup> The side reaction was suppressed by

using activated Mg metal (Dry-stirring under inert atmosphere for 120 hours) and the yield was improved to 50%. When compound **2.13** was treated with DIBAL-H, aldehyde **2.14** was produced with the recovery of the sultam auxiliary **2.15**.

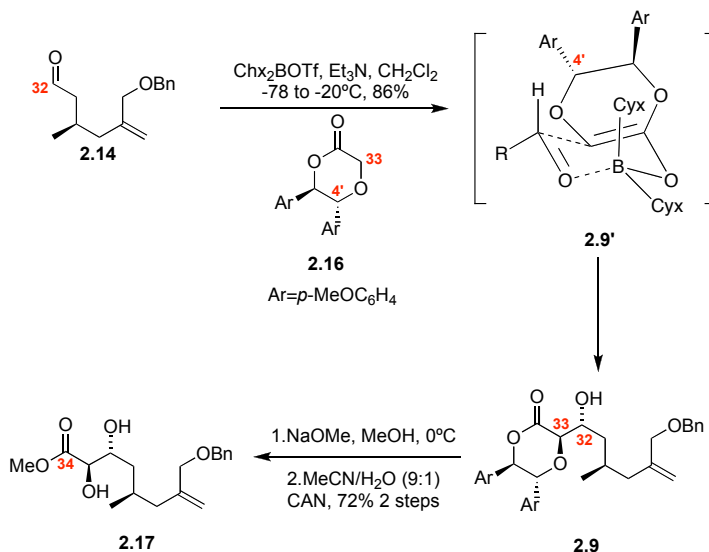


**Scheme 2.3.** Synthesis of Aldehyde **2.14**

### 2.2.3 Anti-aldol Coupling between Aldehyde **2.14** and Dioxane **2.16**

With aldehyde **2.14** and the known dioxane **2.16**<sup>5</sup> in hand, we investigated the anti-aldol coupling (Scheme 2.4). Using the conditions described by Andrus and co-workers,<sup>5</sup> the reaction did not proceed to completion (40-50% conversion). Fortunately, we found that increasing the concentration of the reaction mixture to 0.5 M facilitated complete conversion. A proposed model for the observed stereochemical outcome is shown in transition-state **2.9'**. With the enolate locked in the *E*-configuration, the Zimmerman-Traxler aldol transition state<sup>9</sup> **2.9'** led to the *anti*-aldol adduct. The facial attack on the aldehyde is controlled by the

stereochemistry at C<sub>4'</sub>. The attack at the less hindered face of the enolate generated the corresponding 32*R*, 33*R* stereocenters. Subsequent lactone ring opening and cleavage of the auxiliary with CAN yielded diol **2.17**.

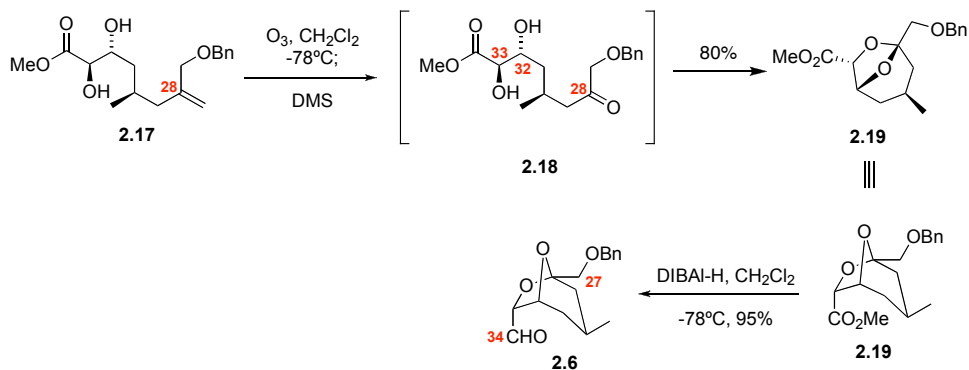


**Scheme 2.4.** Synthesis of Diol **2.17**

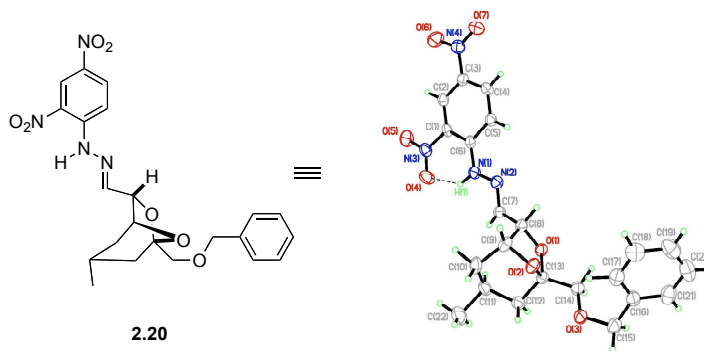
#### 2.2.4 Synthesis of Bicyclic Aldehyde **2.6**

After obtaining diol **2.17**, we shifted our focus to the key ketalization (Scheme 2.5). The [3.2.1] bicyclic ketal moiety was constructed through ozonolysis of **2.17** with DMS workup, which induced spontaneous C<sub>28</sub>-ketal formation (Scheme 2.5). This ketalization process could be driven to completion by the addition of Amberlyst-15. Finally, reduction with DIBAL-H proceeded cleanly to give the aldehyde **2.6**. The stereochemistry of aldehyde **2.6** was

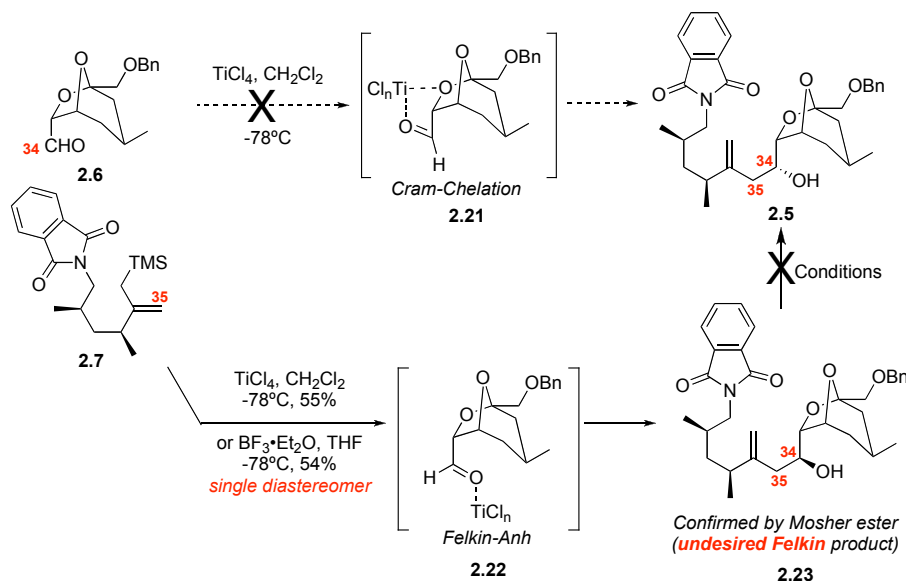
conclusively established through X-ray crystal structure assignment of the 2,4-dinitrohydrazone derivative **2.20** (Figure 2.1).



**Scheme 2.5.** Synthesis of Aldehyde **2.6**



the coupled material as a single diastereomer at C<sub>34</sub> (Scheme 2.6). We had hypothesized that chelating Lewis acids such as titanium or tin<sup>10</sup> would proceed via the intermediate **2.21** to give the desired alcohol **2.5**. We were surprised to find, upon conversion of the intermediate into its Mosher ester,<sup>11</sup> that the C<sub>34</sub> stereochemistry was in fact that of the undesired isomer. Further support for this assignment can be found in the fact that treatment of **2.7** with BF<sub>3</sub>·Et<sub>2</sub>O (a Lewis acid incapable of proceeding via intermediate **2.21**)<sup>10</sup> also gave alcohol **2.23**, again as a single diastereomer. Despite our considerable efforts to invert the C<sub>34</sub> stereochemistry by Mitsunobu reaction or by oxidation-reduction sequence, we were unable to devise a viable route to invert the stereochemistry at C<sub>34</sub>.



**Scheme 2.6.** Aldol Reaction between Allyl Silane **2.7** and Aldehyde **2.6**

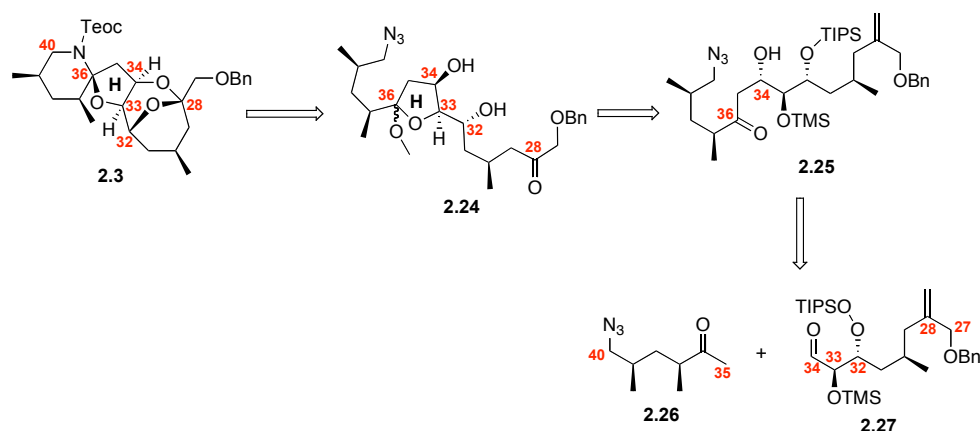
Although still under investigation, one possible explanation for the incapability to chelate might be the highly oxygenated area found in the bicyclic

aldehyde **2.6**. The three O atoms could trap the metal ion and the formation of the desired 5-membered chelation intermediate **2.21** would be prevented. The bulky bicyclic moiety also contributed to the steric congestion at C<sub>34</sub>, which led to the inability to invert the C<sub>34</sub> stereochemistry.

## **2.3 Second-Generation Synthesis of C<sub>27</sub>-C<sub>40</sub> Southern Portion**

### **2.3.1 Modified Retrosynthesis of Southern Portion**

It would appear from our efforts that the encumbered nature of bicyclic moiety made it impossible to properly install the C<sub>34</sub> stereogenic center. On the basis of this setback, we chose to revise our approach and the modified retrosynthesis was shown in Scheme 2.7. Subsequent ketalization and aminial formation were employed to build FGHI ring system. The C<sub>27</sub>-C<sub>40</sub> linear carbon backbone and the C<sub>34</sub> stereochemistry would be constructed prior to the formation of polycyclic ring system. Using the new strategy, we could avoid the complexity caused by the bicyclic structure. Further cleavage at C<sub>34</sub>-C<sub>35</sub> linkage generated two key subunits, methyl ketone **2.26** and aldehyde **2.27**.

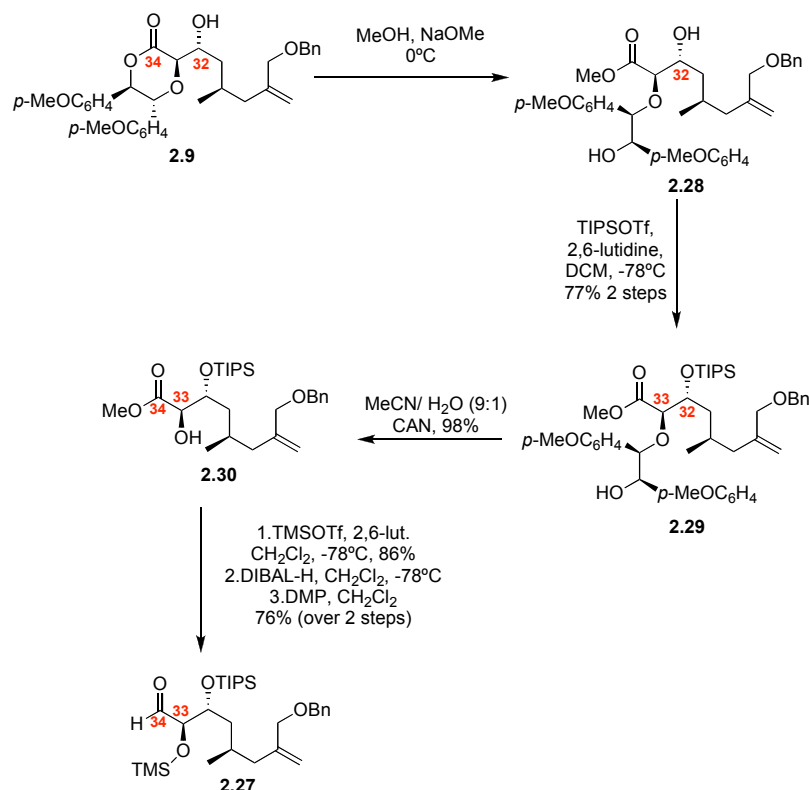


**Scheme 2.7.** Modified Retrosynthesis of C<sub>27</sub>-C<sub>40</sub> Southern Portion

### 2.3.2 Synthesis of Aldehyde 2.27

The synthesis of the aldehyde component **2.27** commenced from the previously made anti-aldol adduct **2.9**. Triisopropylsilylation of compound **2.9** did yield the corresponding silyl ether; however, methanolysis of the lactone proved unsuccessful. The TIPS ether decomposed upon treatment with NaH / MeOH. Fortunately, exposure of **2.9** to TIPSOTf and 2,6-lutidine at low temperature gave selectively the C<sub>32</sub>-OTIPS product **2.28**. None of the corresponding benzyl OTIPS ether was observed, presumably due to the decreased electronic reactivity of the hydroxyl group. Finally, removal of auxiliary with CAN, C<sub>33</sub> TMS protection, and subsequent conversion of methyl ester to aldehyde yielded the desired fragment **2.27**.



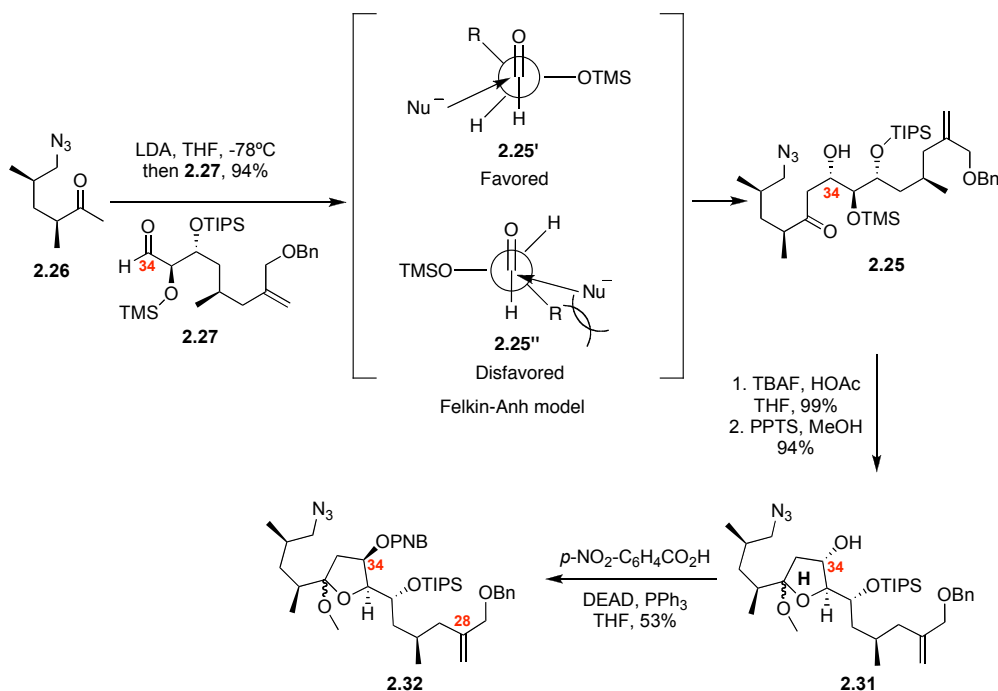


**Scheme 2.8.** Synthesis of Aldehyde **2.27**

### 2.3.3 Completion of C<sub>27</sub>-C<sub>40</sub> Southern Fragment

With the key intermediate aldehyde **2.27** in hands, we explored the aldol reaction to install C<sub>34</sub> stereochemistry (Scheme 2.9).<sup>2</sup> The LDA-mediated aldol coupling between aldehyde **2.27** and the previously made methyl ketone **2.26**<sup>2</sup> generated undesired C<sub>34</sub> stereocenter as a single diastereomer. The stereochemical outcome of the aldol reaction could be explained via Felkin-Anh model in which the  $\alpha$  OTMS group is perpendicular to the carbonyl bond.<sup>12</sup> In this way, the  $\sigma^*_{C-O}$  orbital is aligned parallel with the  $\pi$  orbital of the carbonyl group, allowing the

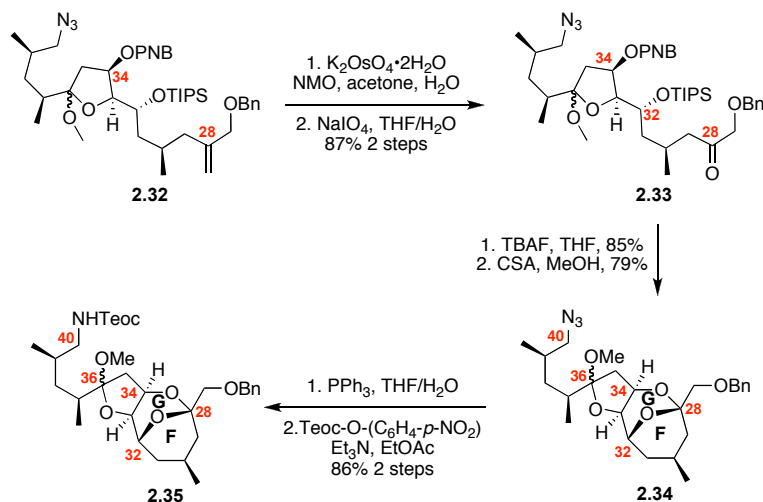
stabilization of the substrate through hyperconjugation. An attack by the enolate on the carbonyl center, in a Bürgi-Dunitz angle (*ca.* 107° relative to the oxygen-carbon double bond<sup>13</sup>) from the side of H (**2.25'**), resulted in the expected 34*S* stereocenter. In contrast, the nucleophilic addition from the side of the more bulky R (**2.25''**) is disfavored due to the increased steric interaction between the enolate and R. After the acid-catalyzed formation of H ring, we were gratified to find that C<sub>34</sub> stereochemistry was inverted successfully using Martin's modified Mitsunobu conditions.<sup>14</sup>



**Scheme 2.9.** Installation of the C<sub>34</sub> Stereocenter

With the setting of the correct C<sub>34</sub> stereochemistry, we were able to construct the FG rings (Scheme 2.10). Since the PNB group is base labile, the basicity of TBAF was harnessed to simultaneously remove the TIPS at C<sub>32</sub> and the

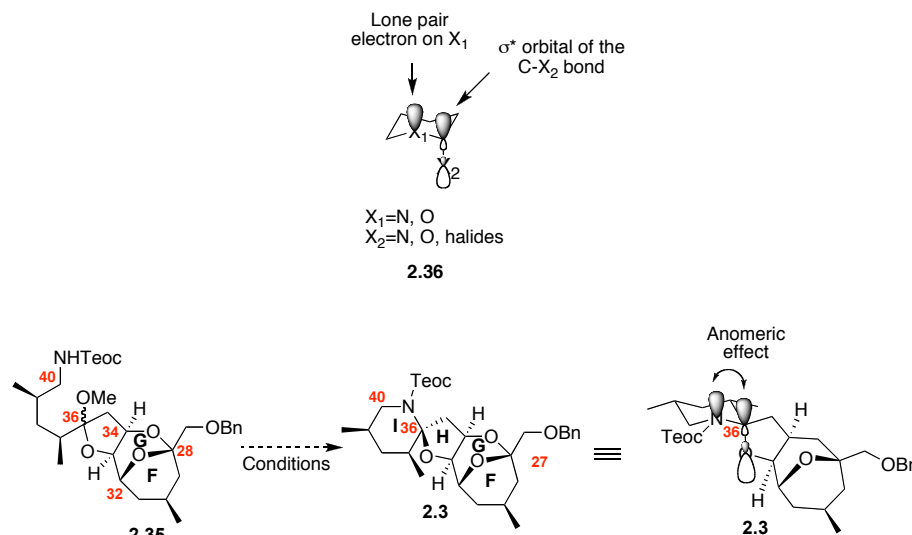
PNB group at C<sub>34</sub>. The following acid-catalyzed ketalization afforded the desired FG rings. Subsequent azide reduction and Teoc protection yielded Teoc protected amine **2.35**.



**Scheme 2.10.** Formation of FG Rings

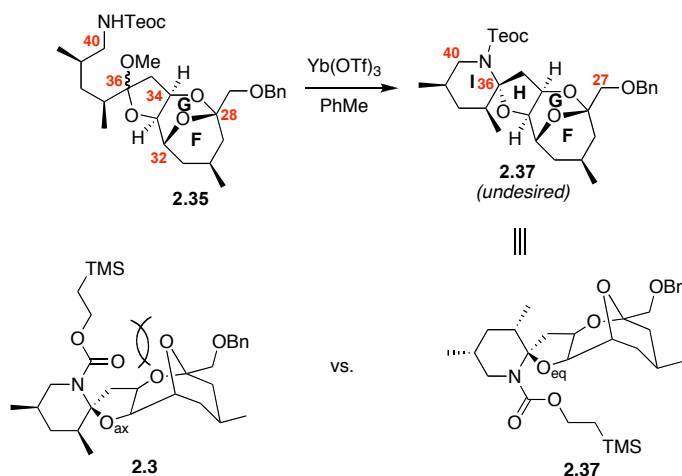
With the FGH rings now in place the next challenge was the formation of the spiroaminal functionality. We had envisioned the desired spiroaminal **2.3** to be the favored product (Scheme 2.11). Our postulation was primarily based on the anomeric effect,<sup>15</sup> a stereoelectronic effect that describes the tendency of heteroatomic substituents adjacent to a heteratom within a cyclohexane ring to prefer the *axial* orientation instead of the less hindered *equatorial* orientation that would be expected from steric considerations. The origins of the anomeric effect are proposed to be the hyperconjugation effects. When the C-X<sub>2</sub> bond is axial, an interaction between the axial lone pair electron on the heteratom and the  $\sigma^*$  orbital of the C-X<sub>2</sub> bond is possible. This interaction leads to

the delocalization of the unshared electrons and would help stabilizing the substrate. In spiroaminal **2.3**, the axial orientation of the C-O bond would be stabilized by the overlap between the axial lone pair electron on the N atom and the  $\sigma^*$  orbital of the C-O bond.



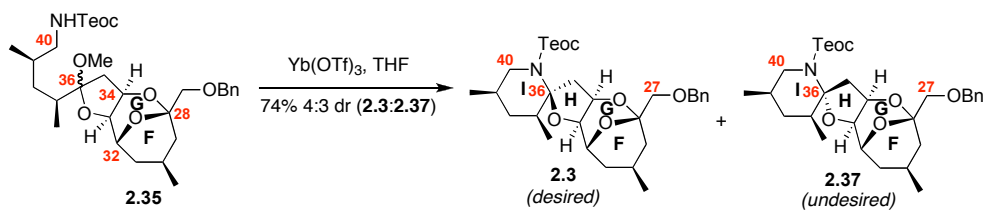
**Scheme 2.11.** Proposed Spiroaminal Formation

Interestingly, treatment of **2.35** with  $\text{Yb}(\text{OTf})_3$  in PhMe led to the rapid formation (30 min, room temperature) of a kinetic product **2.37** (Scheme 2.12). Careful analysis by 2D NMR spectroscopy revealed that **2.37** possessed the undesired stereochemistry at  $\text{C}_{36}$ . We did find that the formation of the non-anomeric **2.37** as the kinetic product to be surprising, as the anomERICALLY stabilized axial orientation is typically kinetically favored as a result of a presumed lower transition-state energy. We attribute this unusual behavior to a severe steric interaction between the NTeoc group and the fused GH ring system.



**Scheme 2.12.** Formation of the Undesired Kinetic Product **2.37**

We next investigated the conditions that would lead to the thermodynamic product **2.3** (Scheme 2.13). Use of extended reaction times in  $\text{PhMe}$  resulted in the formation of a second compound, the desired anomeric diastereomer; however, decomposition was a competitive pathway under these conditions. Fortunately, use of an alternate solvent (THF) led to spiroaminal **2.3** as the major product (74% yield, **2.3/2.37** 4:3 ratio). The minor undesired compound could be recycled by resubmission to the  $\text{Yb}(\text{OTf})_3$  / THF conditions to generate the diastereomers in the same thermodynamic 4:3 ratio.



**Scheme 2.13.** Completion of the Synthesis of FGHI Ring System

## 2.4 Conclusion

In summary, we have successfully synthesized C<sub>27</sub>-C<sub>40</sub> FGHI ring fragment with a longest linear sequence of 21 steps. Although our 1<sup>st</sup> generation strategy led to the key [3.2.1] bicyclic ketal moiety via a spontaneous ketalization, the encumbered nature of bicyclic structure made it impossible to properly install the C<sub>34</sub> stereogenic center. Our modified approach solved this problem by generating C<sub>34</sub> stereocenter prior to the formation of polycyclic system and resulted in the completion of C<sub>27</sub>-C<sub>40</sub> southern half of azaspiracid. The key steps included a highly regioselective C<sub>32</sub> TIPS protection, a Mitsunobu reaction to install the desired C<sub>34</sub> stereocenter, and a Yb(OTf)<sub>3</sub>-catalyzed spiroaminal formation.

## 2.5 References

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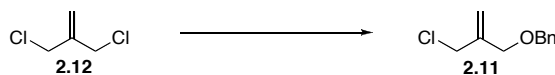
## 2.6 Experimental

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**General.** Infrared spectra were recorded neat, unless otherwise indicated and are reported in  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR spectra were recorded in deuterated solvents and are reported in ppm relative to tetramethylsilane and referenced internally to the residually protonated solvent.  $^{13}\text{C}$  NMR spectra were recorded in deuterated solvents and are reported in ppm relative to tetramethylsilane and referenced internally to the residually protonated solvent.

Routine monitoring of reactions was performed using EM Science DC-Alufolien silica gel, aluminum-backed TLC plates. Flash chromatography was performed with the indicated eluents on EM Science Gedurian 230-400 mesh silica gel.

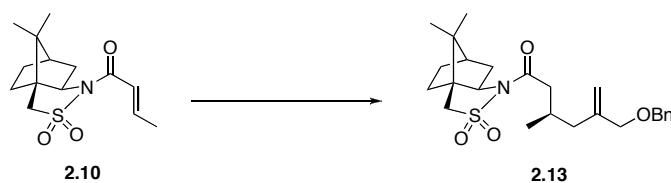
Air and / or moisture sensitive reactions were performed under usual inert atmosphere conditions. Reactions requiring anhydrous conditions were performed under a blanket of argon, in glassware dried in an oven at  $120^\circ\text{C}$  or by a Bunsen flame, then cooled under argon. Solvents and commercial reagents were purified in accord with Perrin and Armarego<sup>1</sup> or used without further purification.



**Allyl chloride 2.11:** To a stirred slurry of pentane-washed NaH (2.05 g, 51.3 mmol, 60% in mineral oil) in THF (70 mL) was added BnOH (5.64 g, 5.4



mL, 52.5 mmol). After 30 min, DMF (15 mL) was added and the reaction mixture was warmed up to reflux. After 30 min, the reaction was allowed to cool to rt. The resulted mixture was then added dropwise to a solution of 3-chloro-2-chloromethyl-1-propene (3.77 g, 3.5 mL, 30.2 mmol) in THF (20 mL) over 1 h at rt. After another 16 h, the reaction mixture was quenched with H<sub>2</sub>O (50 mL) and extracted with ether-pentane (1:1, 3 X 50 mL). The organic phase was washed with water (50 mL) and sat. aq. NaCl (50 mL). The dried extract (MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 2-5% Et<sub>2</sub>O / Pentane, to give the known allyl chloride **2.11**<sup>2</sup> (4.86 g, 24.8 mmol, 82%) as a colorless oil: IR (neat) 3087, 3064, 3031, 2924, 2855, 1496, 1453, 1097, 1075, 1028, 924, 736, 697 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 7.28-7.36 (m, 5H), 5.33 (d, *J* = 0.7 Hz, 1H), 5.28 (d, *J* = 1.2 Hz, 1H), 4.53 (s, 2H), 4.14 (d, *J* = 0.7 Hz, 2H), 4.13 (s, 2H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 142.4, 138.4, 128.8, 128.1(2), 117.3, 72.8, 70.7, 45.6.

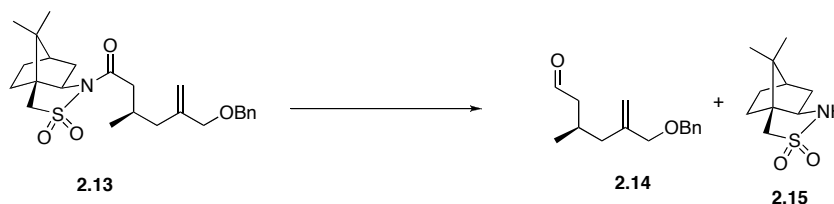


**Sultam 2.13:** Following the similar procedure described by Paquette,<sup>3</sup> Mg (8.0 g, 333 mmol) was stirred vigorously at rt in a dry flask under Ar. After 120 h, when black coating formed inside the flask, THF (100 mL) and 1,2-dibromoethane (1.30 g, 0.6 mL, 6.9 mmol) were added sequentially. After 30 min, a solution of allyl chloride **2.11** (6.5 g, 33.2 mmol) in THF (25 mL) was added slowly to the Mg

slurry over 5 h. The resulted mixture was stirred overnight at rt to give 130 mL Grignard reagent (0.126 M, 50%) as gray solution. The concentration of the Grignard reagent was determined by the titration using menthol in the presence of 1,10-phenanthroline.<sup>4</sup>

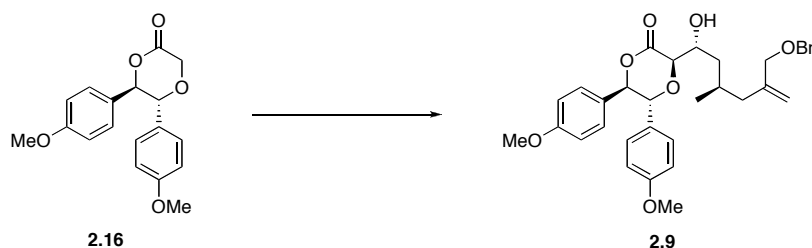
Separately, CuBr•SMe<sub>2</sub> (3.39 g, 16.5 mmol) and LiCl (0.75 g, 17.7 mmol) were dissolved in THF (25 mL) and added to the Grignard solution at -78°C *via* syringe. TMSCl (1.81 g, 2.1 mL, 16.7 mmol) was then added followed by a solution of sultam **2.10**<sup>5</sup> (3.2 g, 11.3 mmol) in THF (25 mL). After another 90 min, the reaction was quenched with aq. NH<sub>4</sub>Cl-NH<sub>4</sub>OH (9:1, pH 9, 20 mL), warmed to rt and partitioned between ether (200 mL) and water (100 mL). The aqueous layer was extracted with EtOAc (3 X 100 mL). The organic phase was washed with sat. aq. NaCl (100 mL). The dried extract (MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 20% EtOAc / Hexanes, to give the sultam **2.13** (4.57 g, 10.3 mmol, 91%) as a colorless oil:  $[\alpha]_D^{23} = -29.7$  (*c* 1.2, CHCl<sub>3</sub>); IR (neat) 2959, 2881, 1695, 1455, 1330, 1217, 1134, 1116, 1058 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 7.33-7.44 (m, 5H), 5.15 (s, 1H), 4.99 (s, 1H), 4.53 (dd, *J* = 13.4, 12.1 Hz, 2H), 4.00 (dd, *J* = 17.1, 12.9 Hz, 2H), 3.90 (t, *J* = 6.3 Hz, 1H), 3.48 (dd, *J* = 26.0, 13.9 Hz, 2H), 2.79 (dd, *J* = 16.1, 5.8 Hz, 1H), 2.53 (dd, *J* = 16.1, 7.6 Hz, 1H), 2.30-2.40 (m, 1H), 2.02-2.17 (m, 4H), 1.86-1.98 (m, 3H), 1.35-1.46 (m, 2H), 1.00 (d, *J* = 6.3 Hz, 3H), 0.99 (s, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 171.8, 144.4, 138.9, 128.7, 128.0, 127.9, 114.1, 73.1, 72.4, 65.6, 53.4,

48.7, 48.1, 45.0, 42.9, 41.2, 39.0, 33.3, 28.4, 26.9, 21.2, 20.3; HRMS (ES<sup>+</sup>) calcd. for C<sub>25</sub>H<sub>36</sub>NO<sub>4</sub>S (M+H) 446.2365, found 446.2337.



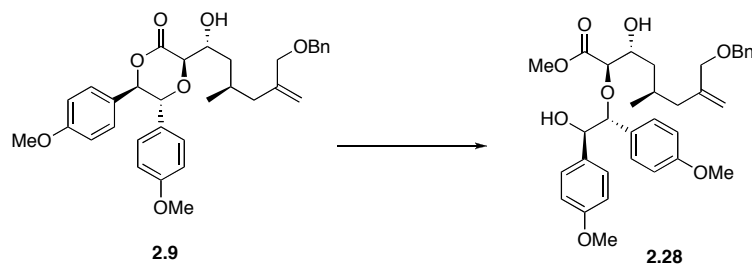
**Aldehyde 2.14:** To a stirred solution of sultam **2.13** (12.50 g, 28.1 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (146 mL) at -78°C was added DIBAL-H (58 mL, 58.0 mmol, 1.0 M in CH<sub>2</sub>Cl<sub>2</sub>) dropwise over 25 min. After 2 h, the reaction was carefully quenched with methanol (2.0 mL) and poured into aq. sodium potassium tartrate (250 mL, 10%) at rt. The reaction flask was rinsed with an addition portion of CH<sub>2</sub>Cl<sub>2</sub> (150 mL). After 3.5 h, the aqueous layer was extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 X 100 mL). The dried extract (MgSO<sub>4</sub>) was concentrated *in vacuo*. The oil was dissolved in a solution of 10% EtOAc / Hexanes solution (40 mL) and placed in the refrigerator to induce crystallization. After 16 h, the crystals were filtered (5% EtOAc / Hexanes rinse) to yield the recovered auxiliary **2.15** (4.38 g, 20.4 mmol) and the mother liquor was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 10-30% EtOAc / Hexanes, to give the aldehyde **2.14** (5.81 g, 26.7 mmol, 95%). Further elution with 75% EtOAc / Hexanes gave additional auxiliary **2.15** (1.00 g, 4.65 mmol, 89% combined yield). **2.14**: [α]<sub>D</sub><sup>23</sup> = +3.9 (c 1.0, CHCl<sub>3</sub>); IR (neat) 2956, 2926, 2851, 2719, 1723, 1455, 1095, 1073 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 9.74 (dd, *J* = 2.4, 1.6 Hz, 1H), 7.28-7.40 (m, 5H), 5.14 (s, 1H), 4.94 (s, 1H), 4.50 (s, 2H), 3.95 (s, 2H), 2.53 (ddd, *J* = 15.3, 4.0 and 1.7

Hz, 1H), 2.16-2.32 (m, 2H), 2.00-2.13 (m, 2H), 0.96 (d,  $J = 6.4$  Hz, 3H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  203.0, 144.2, 138.7, 128.8, 128.1, 128.0, 114.5, 73.1, 72.5, 51.0, 41.4, 26.7, 20.5; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{15}\text{H}_{20}\text{O}_2\text{Na}$  ( $\text{M}+\text{Na}$ ) 255.1361, found 255.1366.



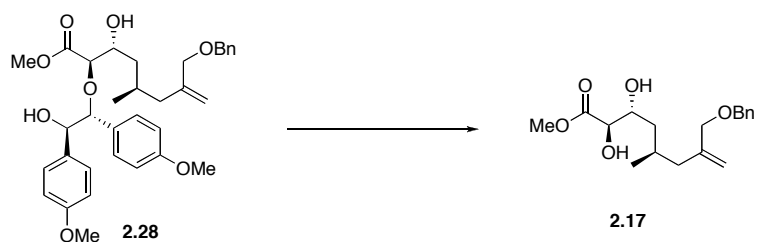
**Andrus aldol adduct 2.9:** To a solution of dioxalone **2.16**<sup>6</sup> (6.25 g, 19.90 mmol) in  $\text{CH}_2\text{Cl}_2$  (20 mL) at  $-78^\circ\text{C}$  was added  $\text{Et}_3\text{N}$  (3.19 g, 4.4 mL, 31.58 mmol). After 3 min, a solution of  $\text{Chx}_2\text{BOTf}$ <sup>7</sup> (28.0 mL, 28.00 mmol, 1.0 M in Hexanes) was added dropwise over 15 min. After 140 min, a solution of the aldehyde **2.14** (5.04 g, 23.10 mmol) in  $\text{CH}_2\text{Cl}_2$  (5 mL, precooled) was added *via* cannula. The aldehyde flask was rinsed with an additional portion of  $\text{CH}_2\text{Cl}_2$  (2 X 0.75 mL, precooled). After 10 min, the reaction flask was transferred to the freezer (approximately  $-30^\circ\text{C}$ ). After 14 h, the reaction was quenched by the addition of MeOH (15 mL). The solution was then poured into a stirring solution of aq. pH 7 phosphate buffer (100 mL) at rt. The reaction flask was rinsed with an additional portion of  $\text{CH}_2\text{Cl}_2$  (75 mL). To the stirring solution was then added  $\text{H}_2\text{O}_2$  (20 mL, 30% aqueous). After 90 min, the reaction mixture was diluted with sat. aq. NaCl (100 mL) and  $\text{CH}_2\text{Cl}_2$  (100 mL) and extracted with  $\text{CH}_2\text{Cl}_2$  (3 X 100 mL). The organic layer was then washed with NaCl (200 mL) and the aqueous layer was

back extracted with  $\text{CH}_2\text{Cl}_2$  (2 X 100 mL). The dried extract ( $\text{MgSO}_4$ ) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 25-50% EtOAc / Hexanes to give **2.9** (9.10 g, 17.11 mmol, 86%) as a colorless oil:  $[\alpha]_D^{23} = +28.4$  ( $c$  1.0,  $\text{CH}_3\text{CN}$ ); IR (neat) 3426, 2956, 2930, 2838, 1740, 1614, 1515, 1455, 1249, 1177, 1029  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.27-7.36 (m, 5H), 6.96 (dd,  $J = 8.7, 6.9$  Hz, 4H), 6.75 (d,  $J = 8.1$  Hz, 4H), 5.35 (d,  $J = 9.3$  Hz, 1H), 5.07 (s, 1H), 4.96 (d,  $J = 9.3$  Hz, 1H), 4.93 (s, 1H), 4.47 (m, 3H), 4.26 (br s, 1H), 3.94 (dd,  $J = 22.0, 12.6$  Hz, 2H), 3.75 (s, 3H), 3.74 (s, 3H), 3.54-3.62 (m, 1H), 3.26 (br s, 1H), 2.30 (dd,  $J = 13.6, 4.6$  Hz, 1H), 1.66-1.98 (m, 6H), 1.49-1.58 (m, 1H), 1.21-1.31 (m, 2H), 0.94 (d,  $J = 6.6$  Hz, 3H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  170.3, 160.4, 160.2, 145.0, 138.7, 129.2, 129.0, 128.8, 128.4, 128.1, 128.0, 127.1, 114.2, 114.1, 113.8, 85.6, 78.2, 76.9, 73.3, 72.5, 72.0, 55.6, 41.4, 40.7, 27.8, 21.1; HRMS ( $\text{FAB}^+$ ) calcd. for  $\text{C}_{33}\text{H}_{38}\text{O}_7$  ( $\text{M}^+$ ) 546.2618, found 546.2641.



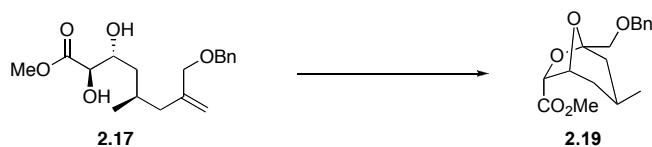
**Methyl ester 2.28:** To a stirred solution of aldol product **2.9** (9.10 g, 17.1 mmol) in dry MeOH (160 mL) at  $0^\circ\text{C}$  was added NaH (72 mg, 1.80 mmol, 60% in mineral oil). After 25 min, the reaction was quenched with sat. aq.  $\text{NH}_4\text{Cl}$  (10 mL). The MeOH was then removed *in vacuo* and the residue was diluted with sat. aq. NaCl (200 mL) and extracted with EtOAc (3 X 150 mL). The dried extract

(MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 20-66% EtOAc / Hexanes to give **2.28** (17.1 mmol) as a glassy semi-solid. The highly glassy nature of **2.28** made effective removal of all residual solvent impossible on large scale. A small amount of **2.28** (50 mg) was placed under high vacuum overnight to provide an analytically pure sample for characterization, but the glassy semi-solid was used in the subsequent step without complete removal of solvents. **2.28**:  $[\alpha]_D^{23} = +55.7$  (*c* 1.5, CHCl<sub>3</sub>); IR (neat) 3423, 2954, 2927, 2837, 1734, 1613, 1514, 1455, 1250, 1176, 1031 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  7.40-7.30(m, 5H), 7.01-6.96 (m, 4H), 6.80-6.76 (m, 4H), 5.10 (s, 1H), 4.95 (s, 1H), 4.94 (d, *J* = 9.3 Hz, 1H), 4.50-4.48 (m, 3H), 4.30-4.26 (m, 1H), 3.96 (dd, *J* = 19.2, 12.6 Hz, 2H), 3.79 (s, 3H), 3.78 (s, 3H), 3.66 (d, *J* = 1.7 Hz, 1H), 3.52 (s, 3H), 3.01 (d, *J* = 4.8 Hz, 1H), 2.32 (dd, *J* = 13.8, 4.8 Hz, 1H), 2.05-1.65 (m, 4H), 0.97 (d, *J* = 6.6 Hz, 3H); <sup>13</sup>C (75 MHz, CDCl<sub>3</sub>):  $\delta$  171.3, 159.7, 159.5, 145.0, 138.6, 132.0, 129.7, 129.6, 128.8, 128.7, 128.2, 128.1, 114.0, 113.8, 113.7, 89.7, 82.2, 78.7, 73.3, 72.5, 70.8, 55.6, 52.1, 40.9, 40.0, 28.0, 21.3; HRMS (FAB<sup>+</sup>) calcd. for C<sub>34</sub>H<sub>42</sub>O<sub>8</sub>Na (M+Na) 601.2777, found 601.2801.



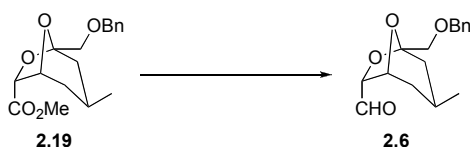
**Diol 2.17**: To a vigorously stirred solution of methyl ester **2.28** (0.82 g, 1.42 mmol) in MeCN-H<sub>2</sub>O (10:1, 40 mL) at 0°C was added CAN (1.00 g, 1.82

mmol) portionwise over 90 min. After a further 30 min, the reaction mixture was diluted with EtOAc (100 mL), the organic phase washed with sat. aq. NaCl (100 mL), and the aqueous layer re-extracted with EtOAc (3 X 100 mL). The dried extract (MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 25-50% EtOAc / Hexanes, to give diol **2.17** (330 mg, 1.02 mmol, 72% 2 steps) as a colorless oil:  $[\alpha]_D^{23} = -3.6$  (*c* 1.7, CHCl<sub>3</sub>); IR (neat) 3419, 2923, 2852, 1738, 1454, 1199, 1096 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.29-7.41 (m, 5H), 5.13 (s, 1H), 4.97 (s, 1H), 4.53 (d, *J*=1.8 Hz, 2H), 4.23 (dd, *J*=5.6, 3.6 Hz, 1H), 3.94-4.05 (m, 3H), 3.82 (s, 3H), 2.40 (d, *J*=6.5 Hz, 1H), 2.20-2.28 (m, 1H), 1.83-1.94 (m, 2H), 1.50 (ddd, *J*=14.7, 9.3 and 5.5 Hz, 1H), 1.40 (ddd, *J*=13.0, 7.3 and 4.2 Hz, 1H), 0.97 (d, *J*=6.3 Hz, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  173.5, 144.9, 138.7, 128.8, 128.2, 128.1, 114.0, 74.5, 73.3, 72.4, 71.8, 53.0, 41.0, 39.4, 28.1, 21.1; HRMS (FAB<sup>+</sup>) calcd. for C<sub>18</sub>H<sub>27</sub>O<sub>5</sub> (M+H) 323.1856, found 323.1854.



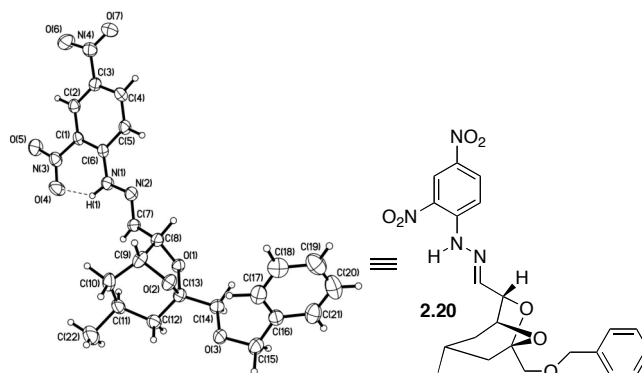
**Bicyclic ester 2.19:** To a stirred solution of methyl ester **2.17** (78 mg, 0.24 mmol) in CH<sub>2</sub>Cl<sub>2</sub>-MeOH (1:1, 4 mL) at -78°C was bubbled ozone until a faint blue color was observed (4 min). At this point, the reaction mixture was briefly degassed with argon. Next, DMS (0.34 g, 0.40 mL, 5.4 mmol) was added. After 10 min, the cold bath was removed and the solution was allowed to warm to rt. The

solvents were removed *in vacuo* followed by the addition of CH<sub>2</sub>Cl<sub>2</sub> (4 mL). The resulted solution was stirred with amberlyst-15 resin (*ca.* 100 mg) until the reaction was complete by TLC (1-3 h). The reaction mixture was filtered through Celite, concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 25-50% EtOAc / Hexanes, to give the bicyclic ketal **2.19** (59 mg, 0.193 mmol, 80%) as a colorless oil:  $[\alpha]_D^{23} = +47.2$  (*c* 0.25, CHCl<sub>3</sub>); IR (neat) 2954, 2922, 2870, 2851, 1762, 1732, 1454, 1438, 1206, 1110, 1073 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.27-7.38 (m, 5H), 4.73-4.77 (m, 1H), 4.63 (dd, *J* = 20.6, 12.2 Hz, 2H) overlaps with 4.63 (d, *J* = 6.0 Hz, 1H), 3.81 (s, 3H), 3.58 (dd, *J*<sub>I</sub> = *J*<sub>2</sub> = 10.3 Hz, 2H), 2.08-2.21 (m, 1H), 1.90 (dd, *J* = 13.6, 5.7 Hz, 1H), 1.71 (dd, *J* = 14.1, 5.5 Hz, 1H), 1.45 (dd, *J* = 13.4, 11.4 Hz, 1H) overlaps with 1.43-1.55 (m, 1H), 0.90 (d, *J* = 6.4 Hz, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  170.0, 138.3, 128.8, 128.2, 128.1, 109.8, 78.4, 74.1, 72.9, 52.6, 39.7, 34.5, 23.5, 22.1; HRMS (FAB<sup>+</sup>) calcd. for C<sub>17</sub>H<sub>25</sub>O<sub>5</sub> (M+H) 307.1545, found 307.1541.

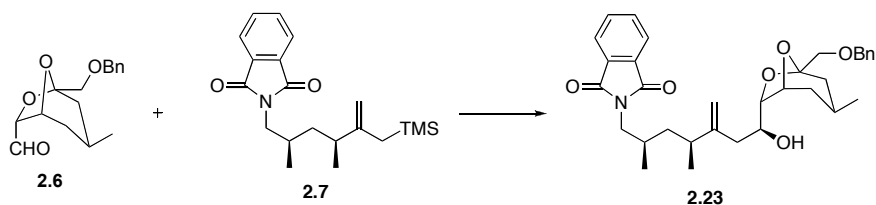




After 2 h, the reaction mixture was extracted with ether (3 X 25 mL) and the organic phase washed with water (25 mL) and sat. aq. NaCl (25 mL). The dried extract (MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 25-50% EtOAc / Hexanes, to give aldehyde **2.6** (47.5 mg, 0.172 mmol, 95%) as a colorless oil:  $[\alpha]_D^{23} = +23.3$  (*c* 1.6, CHCl<sub>3</sub>); IR (neat) 2957, 2922, 2851, 1731, 1455, 1260, 1104, 1070, 1027 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  10.03 (d, *J* = 1.1 Hz, 1H), 7.27-7.37 (m, 5H), 4.74-4.78 (m, 1H), 4.65 (s, 2H), 4.42 (d, *J* = 5.5 Hz, 1H), 3.59 (dd, *J* = 13.2, 10.6 Hz, 2H), 1.90 (dd, *J* = 13.3, 5.5 Hz, 1H), 1.73-1.86 (m, 1H), 1.66 (dd, *J* = 14.3, 5.7 Hz, 1H), 1.45 (dd, *J* = 13.5, 10.4 Hz, 1H) overlaps with 1.52 (dd, *J* = 12.2, 3.0 Hz, 1H), 0.94 (d, *J* = 6.4 Hz, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  200.9, 138.2, 128.8, 128.2(2), 109.7, 84.4, 74.2, 73.0, 39.6, 33.9, 25.1, 21.9; HRMS (CI<sup>+</sup>) calcd. for C<sub>16</sub>H<sub>21</sub>O<sub>4</sub> (M+H) 277.1440, found 277.1437.

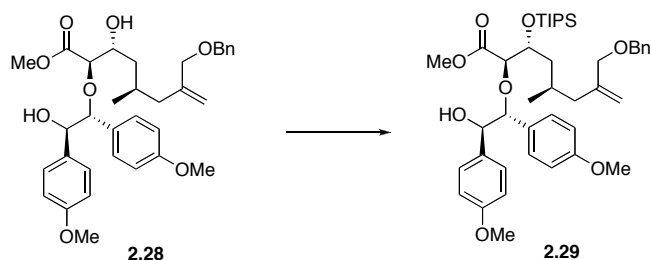


**Figure 1.** ORTEP Representation of 2,4-Dinitrohydrazone **2.20**<sup>8</sup>

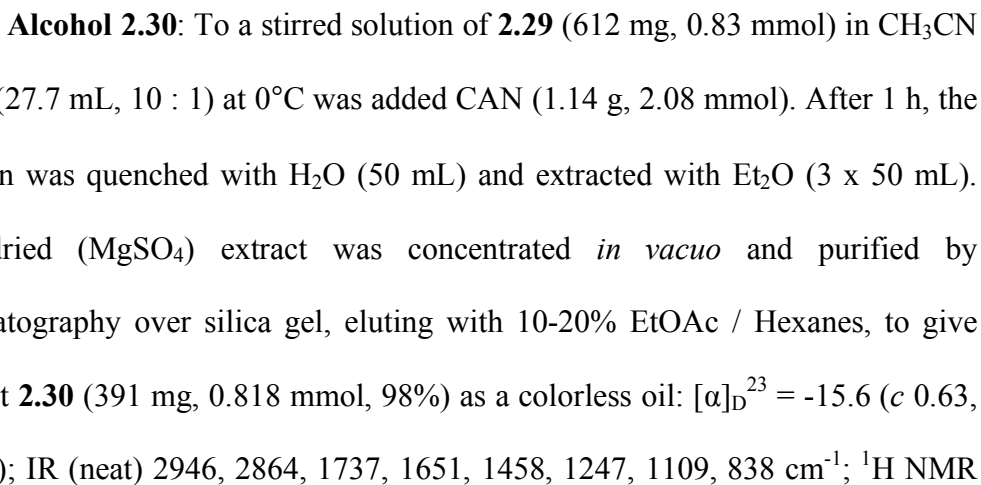


**Coupled product 2.23:** To a stirred solution of aldehyde **2.6** (42.6 mg, 0.154 mmol) in  $\text{CH}_2\text{Cl}_2$  (3.0 mL) at  $-78^\circ\text{C}$  was added  $\text{BF}_3 \cdot \text{Et}_2\text{O}$  (23.8 mL, 0.184 mmol). The resulted faintly pink solution was stirred for 5 min before the addition of allyl silane **2.7** (163 mg, 0.475 mmol) in  $\text{CH}_2\text{Cl}_2$  (0.5 mL). After 10 min, the reaction was quenched with aq. pH 7 buffer (3 mL) and extracted with ether (3 x 5 mL). The dried ( $\text{MgSO}_4$ ) extract was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 10 - 30% EtOAc / Hexanes, to give product **2.23** (45.6 mg, 0.083 mmol, 54%) as colorless oil:  $[\alpha]_{\text{D}}^{23} = +37.3$  (*c* 4.2,  $\text{CHCl}_3$ ); IR (neat) 3488, 2963, 2933, 2860, 1767, 1711, 1449, 1393, 1109, 1066, 1015, 911, 756, 722  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.84 – 7.87 (m, 2H), 7.72 – 7.75 (m, 2H), 7.30 – 7.39 (m, 5H), 4.94 (s, 1H), 4.93 (s, 1H), 4.68 (d,  $J = 12.0$  Hz, 1H), 4.64 (d,  $J = 12.4$  Hz, 1H), 4.55 (m, 1H), 3.99 (t,  $J = 10.0$  Hz, 1H), 3.79 (dd,  $J = 9.2, 4.0$  Hz, 1H), 3.48 – 3.59 (m, 4H), 2.71 (d,  $J = 14.4$  Hz, 1H), 2.37 – 2.43 (m, 1H), 1.97 – 2.20 (m, 5H), 1.84 (dd,  $J = 13.2, 6.0$  Hz, 1H), 1.42 – 1.59 (m, 3H), 1.19 – 1.25 (m, 1H), 1.13 (d,  $J = 6.8$  Hz, 3H), 0.96 (d,  $J = 6.8$  Hz, 3H), 0.93 (d,  $J = 6.8$  Hz, 3H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  168.7, 150.1, 138.1, 133.9, 131.9, 128.3, 127.7, 127.5, 123.2, 112.0, 107.6, 82.7, 76.6, 73.6, 73.3, 67.8, 44.0, 40.9, 40.6, 39.4, 37.7, 33.1, 30.4, 24.4, 22.1, 20.8, 18.0; HRMS ( $\text{FAB}^+$ ) calcd. for  $\text{C}_{33}\text{H}_{42}\text{NO}_6$  ( $\text{M}+\text{H}$ ) 548.3012, found 548.3027.

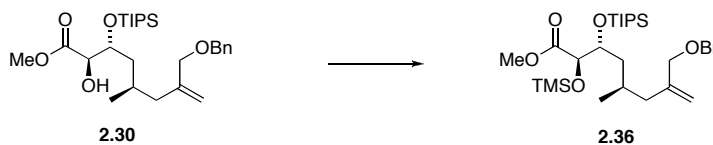
**MTPA esters:** To a solution of **2.23** (20 mg, 0.058 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (1 mL) was sequentially added DMAP (71.2 mg, 0.58 mmol) and (*R*) or (*S*)-(+)- $\alpha$ -methoxy- $\alpha$ -trifluoromethylphenylacetyl chloride (73.2 mg, 54.3  $\mu$ L, 0.29 mmol). After 10 min, the solution was evaporated and the residue was loaded directly onto silica gel and purified by chromatography, eluting with 2 - 10% EtOAc / Hexanes, to give product (*S*)- or (*R*)- **MTPA esters** (68-72%) as colorless oils. <sup>1</sup>H NMR Difference in ppm [(*S*)-Mosher Ester – (*R*)-Mosher ester, CDCl<sub>3</sub>, CDCl<sub>3</sub>, 300 MHz NMR] H<sub>32</sub>: 4.030 – 3.910 = **+0.120**, H<sub>33</sub>: 4.031 – 4.210 = **+0.179**, H<sub>35</sub>: 2.7215 – 2.7655 = **-0.044**, H<sub>36</sub>: 4.792 – 4.877 = **-0.085**, H<sub>36</sub>: 4.779 – 4.877 = **-0.098**.



**TIPS ether 2.29:** To a stirred solution of methyl ester **2.28** (9.87 g, 17.1 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (150 mL) at -78°C was sequentially added 2,6-lutidine (4.06 g, 4.4 mL, 37.9 mmol) and TIPSOTf (5.93 g, 5.2 mL, 19.3 mmol). An additional portion of TIPSOTf (343 mg, 0.3 mL, 1.11 mmol) was added after 25 min. After an additional 10 min, the reaction was quenched with MeOH (1 mL) followed by the addition of sat. aq. NaHCO<sub>3</sub> (200 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 X 100 mL). The dried (MgSO<sub>4</sub>) extract was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 10-25% EtOAc / Hexanes, to give

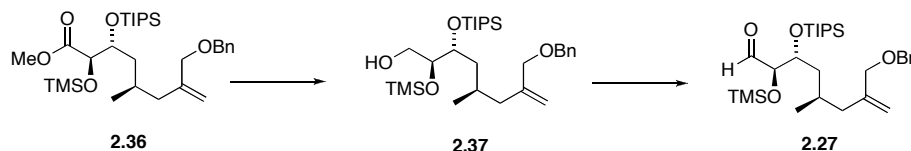


(300 MHz, CDCl<sub>3</sub>)  $\delta$  7.28-7.38 (m 5H), 5.14 (s, 1H), 4.97 (s, 1H), 4.52 (s, 2H), 4.24-4.26 (m, 2H), 3.97-3.99 (m, 2H), 3.70 (s, 3H), 1.74-2.05 (m, 4H), 1.27-1.31 (m, 1H), 1.09 (m, 21H), 0.91 (d,  $J$  = 6.6 Hz, 3H), 0.14 (s, 9H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  172.3, 144.2, 138.5, 128.3, 127.6, 127.4, 113.0, 76.0, 73.7, 72.9, 71.9, 51.5, 42.1, 41.5, 27.4, 19.7, 18.1, 12.6, -0.11; HRMS (Cl<sup>+</sup>) calcd. for C<sub>27</sub>H<sub>47</sub>O<sub>5</sub>Si (M+H) 479.3192, found 479.3224.



**TMS ether 2.36:** To a stirred solution of **2.30** (260 mg, 0.54 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (5.4 mL) at -78°C was sequentially added 2,6-lutidine (0.23 g, 0.25 mL, 2.16 mmol) and TMSOTf (0.24 g, 0.20 mL, 1.08 mmol). After 30 min, the reaction was quenched with sat. aq. NH<sub>4</sub>Cl (5 mL) and extracted with Et<sub>2</sub>O (3 x 10 mL). The dried (MgSO<sub>4</sub>) extract was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 10-20% EtOAc / Hexanes, to give product **2.36** (256 mg, 0.465 mmol, 86%) as a colorless oil:  $[\alpha]_D^{23}$  = -15.6 ( $c$  0.63, CHCl<sub>3</sub>); IR (neat) 2946, 2864, 1737, 1651, 1458, 1247, 1109, 838 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.28-7.38 (m, 5H), 5.14 (s, 1H), 4.97 (s, 1H), 4.52 (s, 2H), 4.24-4.26 (m, 2H), 3.97-3.99 (m, 2H), 3.70 (s, 3H), 1.74-2.05 (m, 4H), 1.27-1.31 (m, 1H), 1.09 (m, 21H), 0.91 (d,  $J$  = 6.6 Hz, 3H), 0.14 (s, 9H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  172.3, 144.2, 138.5, 128.3, 127.6, 127.4, 113.0, 76.0, 73.7, 72.9, 71.9,

51.5, 42.1, 41.5, 27.4, 19.7, 18.1, 12.6, -0.11; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{30}\text{H}_{54}\text{O}_5\text{Si}_2\text{Na}$  ( $\text{M} + \text{Na}$ ) 573.3408, found 573.3403.



**Aldehyde 2.27:** To a stirred solution of **2.36** (250 mg, 0.45 mmol) in  $\text{CH}_2\text{Cl}_2$  (4.5 mL) at  $-78^\circ\text{C}$  was added DIBAL-H (1.08 mL, 1.08 mmol, 1.0 M in  $\text{CH}_2\text{Cl}_2$ ). After 1 h, the reaction was allowed to warm to  $0^\circ\text{C}$ , quenched with methanol (0.2 mL) and poured into aq. sodium potassium tartrate (10 mL, 10%) at rt. The reaction flask was rinsed with an additional portion of  $\text{CH}_2\text{Cl}_2$  (5 mL). After another 3 h, the reaction mixture was extracted with EtOAc (3 x 10 mL). The dried ( $\text{MgSO}_4$ ) extract was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 10-40% EtOAc / Hexanes, to give alcohol **2.37** (138 mg, 0.26 mmol, 59%) and partial aldehyde **2.27** (69 mg, 0.13 mmol, 29%) as colorless oils. Data for **2.37**:  $[\alpha]_{\text{D}}^{23} = -23.8$  ( $c$  1.20,  $\text{CHCl}_3$ ); IR (neat) 3557, 2956, 2863, 1646, 1463, 1390, 1252, 1106, 842, 675  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.29-7.38 (m, 5H), 5.16 (s, 1H), 4.96 (s, 1H), 4.54 (d,  $J = 11.6$  Hz, 1H), 4.51 (d,  $J = 12.0$  Hz, 1H), 4.07-4.09 (m, 1H), 3.95 (s, 2H), 3.81-3.86 (m, 1H), 3.61-3.66 (m, 2H), 2.56-2.58 (m, 1H), 2.08 (dd,  $J = 13.6, 6.0$  Hz, 1H), 1.98 (dd,  $J = 13.6, 6.0$  Hz, 1H), 1.68-1.75 (m, 2H), 1.31-1.38 (m, 1H), 1.11-1.16 (m, 2H), 0.95 (d,  $J = 6.4$  Hz, 3H), 0.17 (s, 9H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  144.0, 138.3, 128.4, 127.7,

127.6, 113.6, 75.1, 73.8, 72.9, 71.9, 63.4, 42.6, 42.1, 27.8, 19.5, 18.2, 12.6, 0.27; HRMS (FAB<sup>+</sup>) calcd. for C<sub>29</sub>H<sub>55</sub>O<sub>4</sub>Si<sub>2</sub> (M+H) 523.3639, found 523.3641.

To a stirred solution of **2.37** (138 mg, 0.26 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (2.6 mL) at room temperature was added DMP (330 mg, 0.78 mmol) and solid NaHCO<sub>3</sub> (*ca.* 50 mg). After 1 h, the reaction was quenched with sat. aq. NaHCO<sub>3</sub> (10 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 x 10 mL). The dried (MgSO<sub>4</sub>) extract was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 10-40% EtOAc / Hexanes, to give product **2.27** (108 mg, 0.21 mmol, 80%) as colorless oil:  $[\alpha]_D^{23} = -28.2$  (*c* 0.68, CHCl<sub>3</sub>); IR (neat) 2954, 2864, 1733, 1251, 1088, 873, 838, 679 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  9.64 (s, 1H), 7.29-7.39 (m, 5H), 5.16 (s, 1H), 4.98 (s, 1H), 4.54 (d, *J* = 12.0 Hz, 1H), 4.51 (d, *J* = 12.0 Hz, 1H), 4.22 (dd, *J* = 10.4, 3.2 Hz, 1H), 3.95-4.06 (m, 3H), 2.06, (d, *J* = 7.2 Hz, 2H), 1.84-1.91 (1H), 1.62-1.66 (m, 1H), 1.23-1.30 (m, 1H), 1.13-1.23 (m, 21H), 0.90 (d, *J* = 6.8 Hz, 3H), 0.17 (s, 9H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  204.0, 144.0, 138.4, 128.3, 127.6, 127.5, 80.4, 74.9, 72.8, 71.9, 42.1, 41.0, 27.1, 19.5, 18.1, 12.5, 0.06; HRMS (FAB<sup>+</sup>) calcd. for C<sub>29</sub>H<sub>51</sub>O<sub>4</sub>Si<sub>2</sub> (M-H) 519.3326, found 519.3319.

## References

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1. Perrin, D. D.; Armarego, W. L. F. *Purification of Laboratory Chemicals: Third Edition*; Pergamon Press: New York, 1993.
  2. Kelly, D. R.; Mahdi, J. G. *Tetrahedron Lett.* **2002**, 43, 511.

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3. Boulet, S. L.; Paquette, L. A. *Synthesis* **2002**, 895.

4. Titration of Grignard reagent: To a stirred solution of menthol (15.6 mg, 0.1 mmol) and 1,10-phenanthroline (2 mg) in THF (0.5 mL) was added prepared Grignard reagent solution until a burgundy color persists. The concentration was calculated using the formula:

$$[\text{RMgX}] = 0.1 \text{ mmol} / \text{volume of added RMgX in mL}$$

*For the references, see:* (a) Lin, H, -S; Paquette, L. A. *Synth. Comm.* **1994**, 24, 2503. (b) Watson, S. C.; Eastham, J. F. *J. Organomet. Chem.* **1967**, 9, 165.

5. Vanderwalle, M.; Van der Eycken, J.; Oppolzer, W.; Vullioud, C. *Tetrahedron* **1986**, 42, 4035.

6. Dioxalone **2.16** was prepared via a modified procedure to originally described by Andrus (*J. Org. Chem.* **2003**, 68, 8162-69). To a vigorously stirred heterogeneous solution of 4,4-dimethoxy-stilbene (9.525 g, 39.6 mmol) in *t*-BuOH (16 mL) and NMO (12 mL, 51.2 mmol, 50% w/v in H<sub>2</sub>O) was added (DHQD)<sub>2</sub>PHAL (86.2 mg, 0.11 mmol, 0.3 mol%) and K<sub>2</sub>OsO<sub>4</sub>•2H<sub>2</sub>O (28.7 mg, 0.078 mmol, 0.2 mol%). After 16 h, sat. aq. Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> (75 ml) was added. After 20 min, the mixture was filtered to isolate (1*R*,2*R*)-Bis-(4-methoxy-phenyl)-ethane-1,2-diol (washed sequentially with H<sub>2</sub>O and Hexanes). The diol was dried on paper then under high vacuum to give the product (9.095 g, 33.2 mmol, 84%). To a stirred solution of the diol (9.095 g, 33.2 mmol) in PhMe (500 mL) was added Bu<sub>2</sub>SnO (9.00 g, 36.2 mmol) and equipped with a Dean-Stark apparatus. The solution was heated to reflux. After 18 h, the solution was allowed to cool. Next,



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TBAI (19.28 g, 52.2 mmol) and *t*-butylbromoacetate (12.95 g, 9.8 mL, 66.4 mmol) were sequentially added. The reaction was again heated to reflux. After 4 h, the reaction was cooled to rt, quenched with aq. Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> (300 mL, 10%), diluted with Et<sub>2</sub>O (200 mL) and sat. aq. NaCl (75 mL) and extracted with Et<sub>2</sub>O (3 X 250 mL). The dried (MgSO<sub>4</sub>) extract was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 20-75% EtOAc / Hexanes, followed by recrystallization from EtOAc / Hexanes to give the dioxalone **2.16** (6.60 g, 21.0 mmol, 63%).

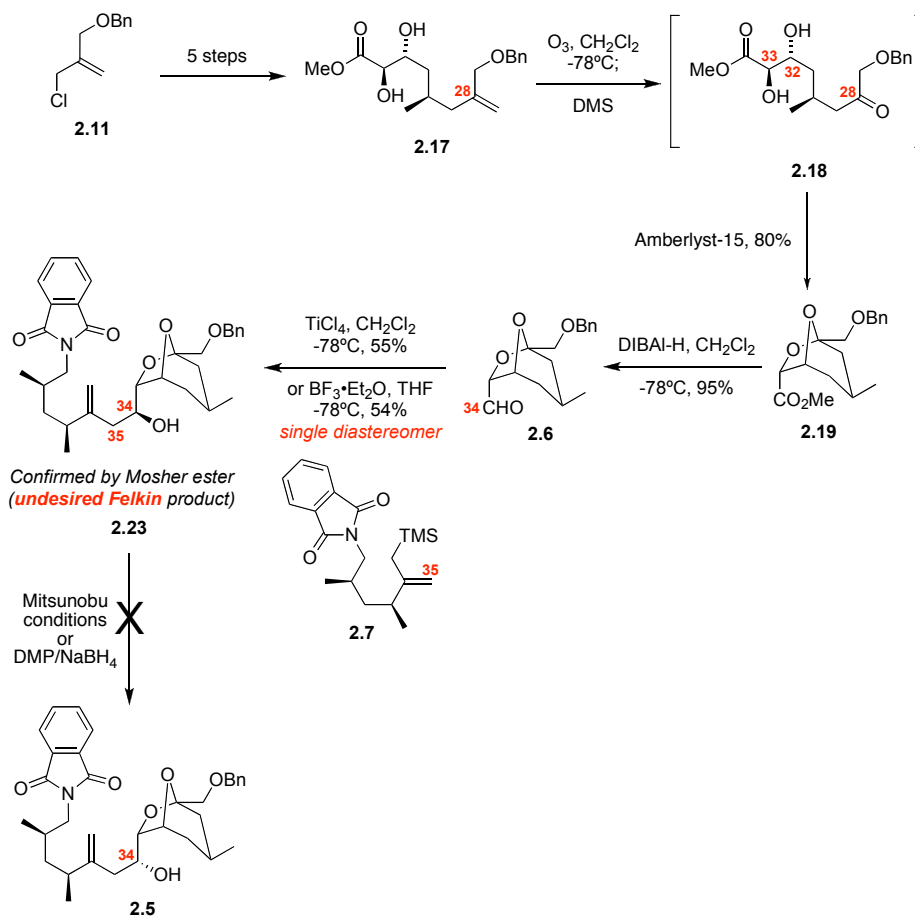
7. Brown, H. C.; Ganesan, K.; Dhar, R. K. *J. Org. Chem.* **1993**, 58, 147.

8. CCDC-605,862 contains the supplementary crystallographic data of compound **2.20** from this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via [www.ccdc.cam.ac.uk/data\\_request/cif](http://www.ccdc.cam.ac.uk/data_request/cif).

## CHAPTER 3. CONCLUSION AND PROPOSED FUTURE WORK

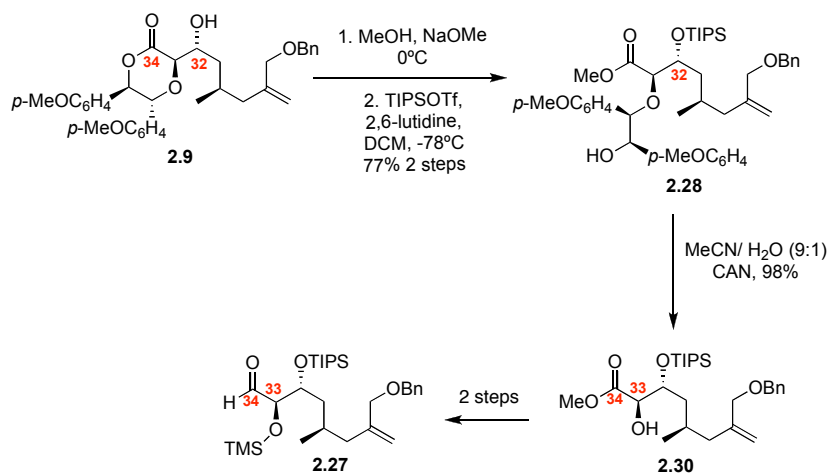
### 3.1 Conclusion

In summary, we have successfully synthesized C<sub>27</sub>-C<sub>40</sub> FGHI ring fragment with a longest linear sequence of 22 steps. Our 1<sup>st</sup> generation strategy features a spontaneous ketalization to afford the key [3.2.1] bicyclic ketal moiety. Unfortunately, we were unable to install the desired C<sub>34</sub> stereocenter through the chelation-controlled aldol coupling and our attempts to invert the C<sub>34</sub> stereochemistry also proved problematic.



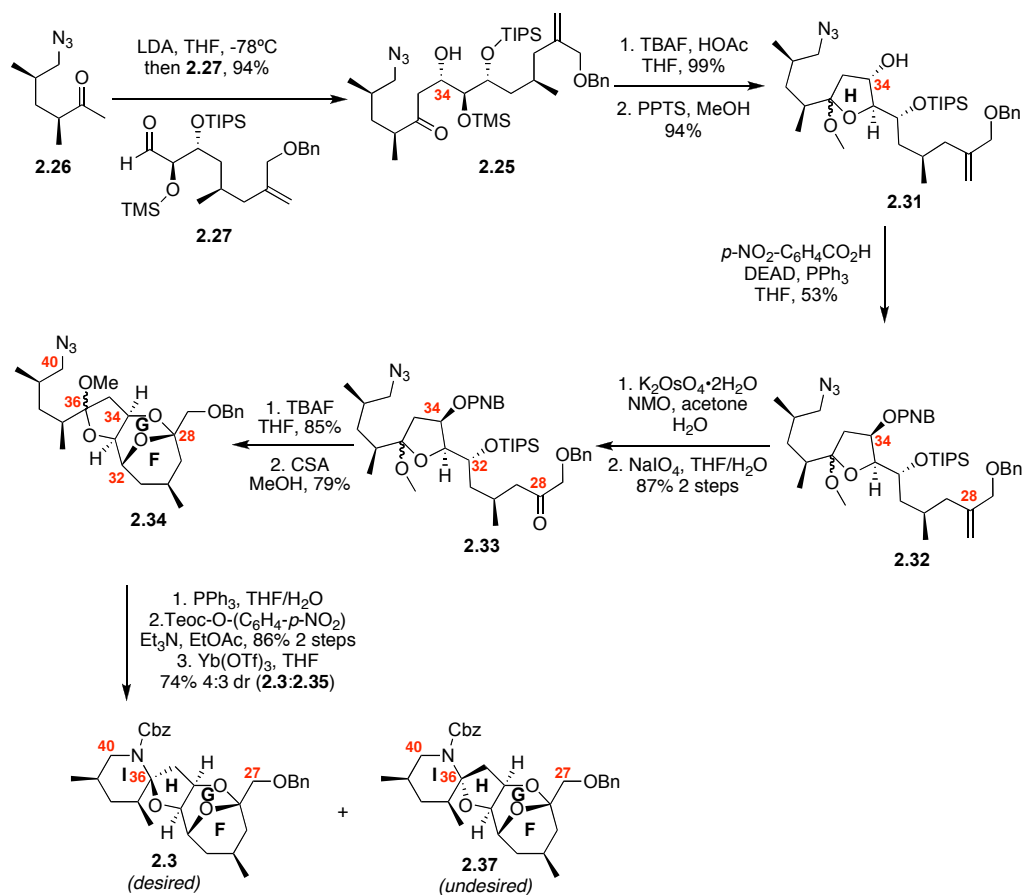
**Scheme 3.1.** Our 1<sup>st</sup> Generation Approach for the Synthesis of Southern Portion

Faced with these roadblocks, we were forced to revise the synthetic strategy. Our 2<sup>nd</sup> generation strategy required the synthesis of aldehyde **2.27** (Scheme 3.2). Commenced from the anti-aldol adduct **2.9**, aldehyde **2.27** was prepared in 5 steps. Key steps include a highly regioselective C<sub>32</sub> TIPS protection.



**Scheme 3.2.** Synthesis of Aldehyde **2.27**

The parallel research from our group led to the other fragment, methyl ketone **2.26**. With both fragments in hand, we were finally able to complete the synthesis of C<sub>27</sub>-C<sub>40</sub> southern portion (Scheme 3.3). The key steps include a highly stereoselective aldol coupling between aldehyde **2.26** and methyl ketone **2.27**, a Mitsunobu reaction to install the desired C<sub>34</sub> stereocenter, and a Yb(OTf)<sub>3</sub>-catalyzed spiroaminal formation.

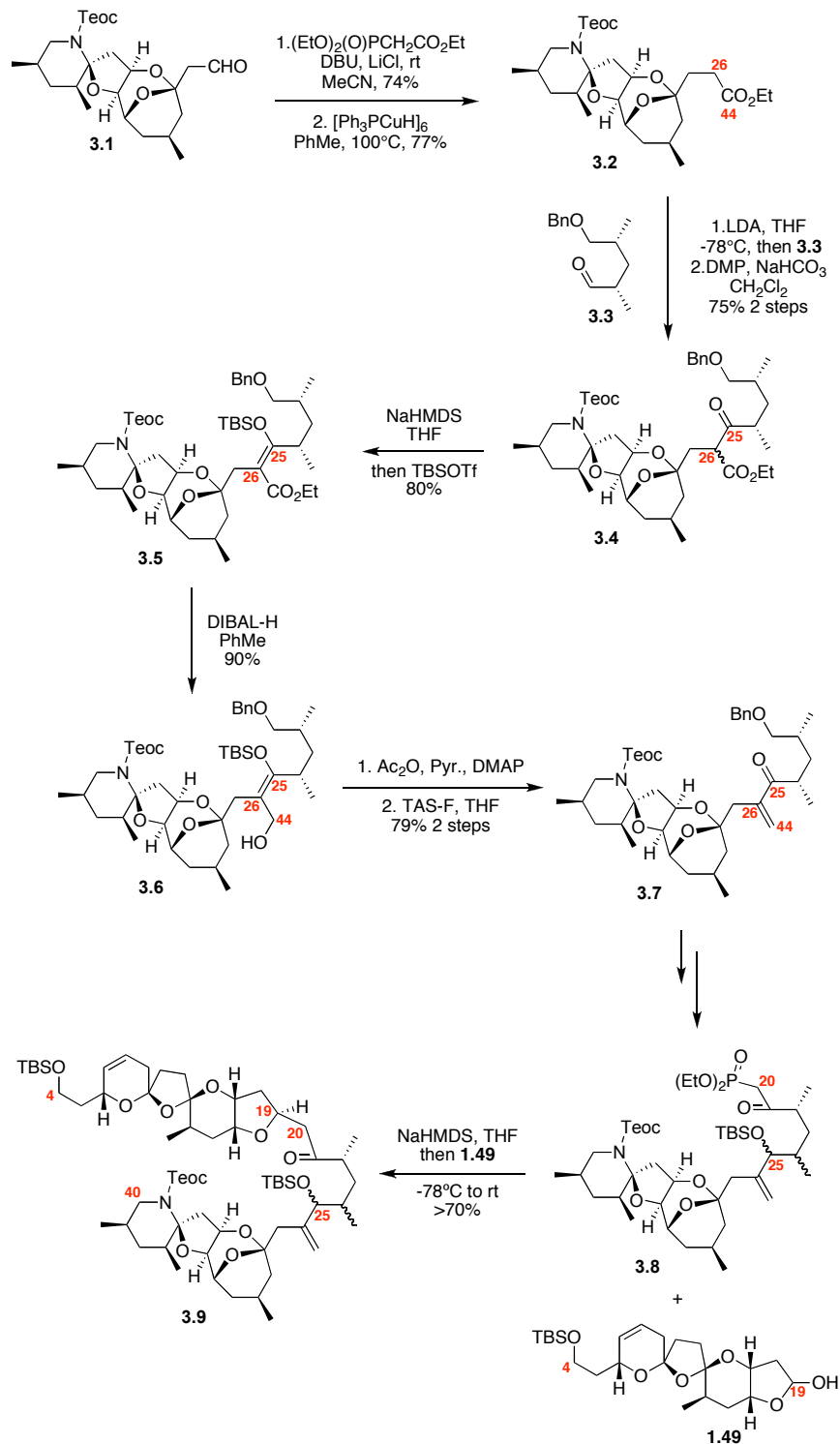


**Scheme 3.3.** Completion of the Synthesis of C<sub>27</sub>-C<sub>40</sub> Southern Portion **2.3**

### 3.2 Proposed Future Work

With efficient routes to both southern and northern portions, we are in excellent position to complete the total synthesis of azaspiracid-1. Our group have recently advanced to the key intermediate enone **3.7** using the sequence shown in Scheme 3.4. Ester **3.2** was obtained from aldehyde **3.1** via HWE reaction and reduction of the double bond with Stryker's reagent. LDA-mediated aldol coupling

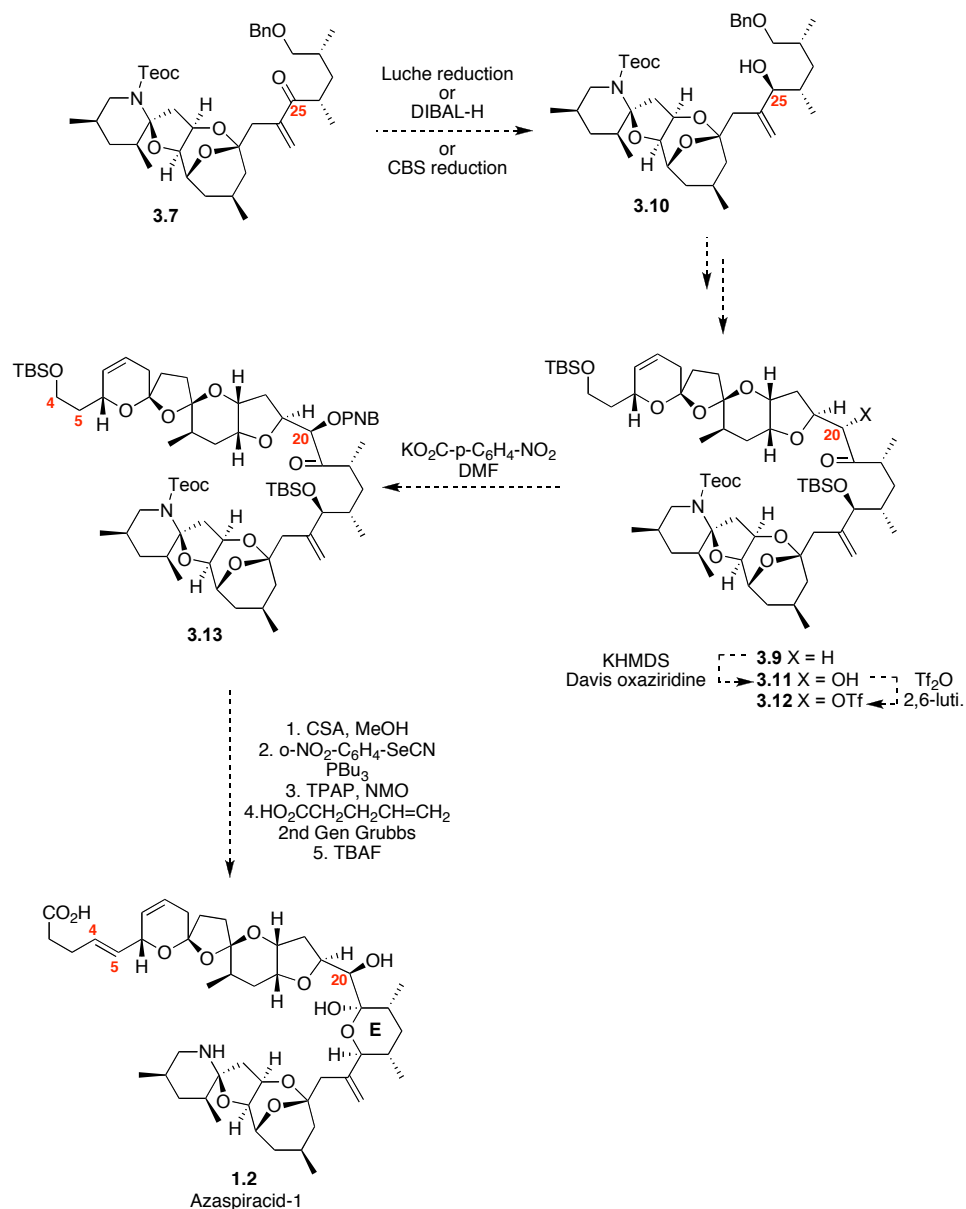
proceeded smoothly to give an inconsequential mixture of diastereomers. The following Dess-Martin oxidation afforded the ketone **3.4** as tautomers. Treatment of compound **3.4** with NaHMDS and TBSOTf led to enol ether **3.5**. Sequential DIBAL-H reduction, C<sub>44</sub> acylation, and TASF-induced desilylation / elimination finally yielded the key intermediate, enone **3.7**. More recently, our preliminary results showed that enone **3.7** was converted to phosphonate **3.8** in moderate diastereoselectivity via an un-optimized protocol. We next investigated the key combination of southern and northern halves. Gratifyingly, the HWE reaction between phosphonate **3.8** and the previously made lactol **1.49**<sup>1</sup>, followed by the *in situ* Michael addition, afforded the desired coupling product **3.9** in greater than 70% yield.



**Scheme 3.4.** Our Recent Progress on the Synthesis of Azaspiracid-1

With the encouraging results obtained from our recent research, the next target would be the development of a distereoselective route to phosphonate **3.8** and accomplish the total synthesis of azaspiracid-1 (Scheme 3.5). Luche reduction, DIBAL-H reduction, or CBS reduction are the potential options for the diastereoselective installation of C<sub>25</sub> stereocenter. After the combination of southern and northern halves, the remaining steps will follow closely our previously reported procedure.<sup>1</sup> Diastereoselective incorporation of the C<sub>20</sub> hydroxyl functionality using Davis oxaziridine followed by triflation and inversion with KOPNB will afford **3.13**. Removal of the primary TBS group under acidic conditions followed by selenation, elimination and cross metathesis will yield the protected version of azaspiracid-1. Finally, removal of the secondary TBS group should lead to the spontaneous ketalization to give azaspiracid-1 in approximately 40 linear steps.





**Scheme 3.5.** Proposed Route to the Total Synthesis of Azaspiracid-1

### 3.3 References

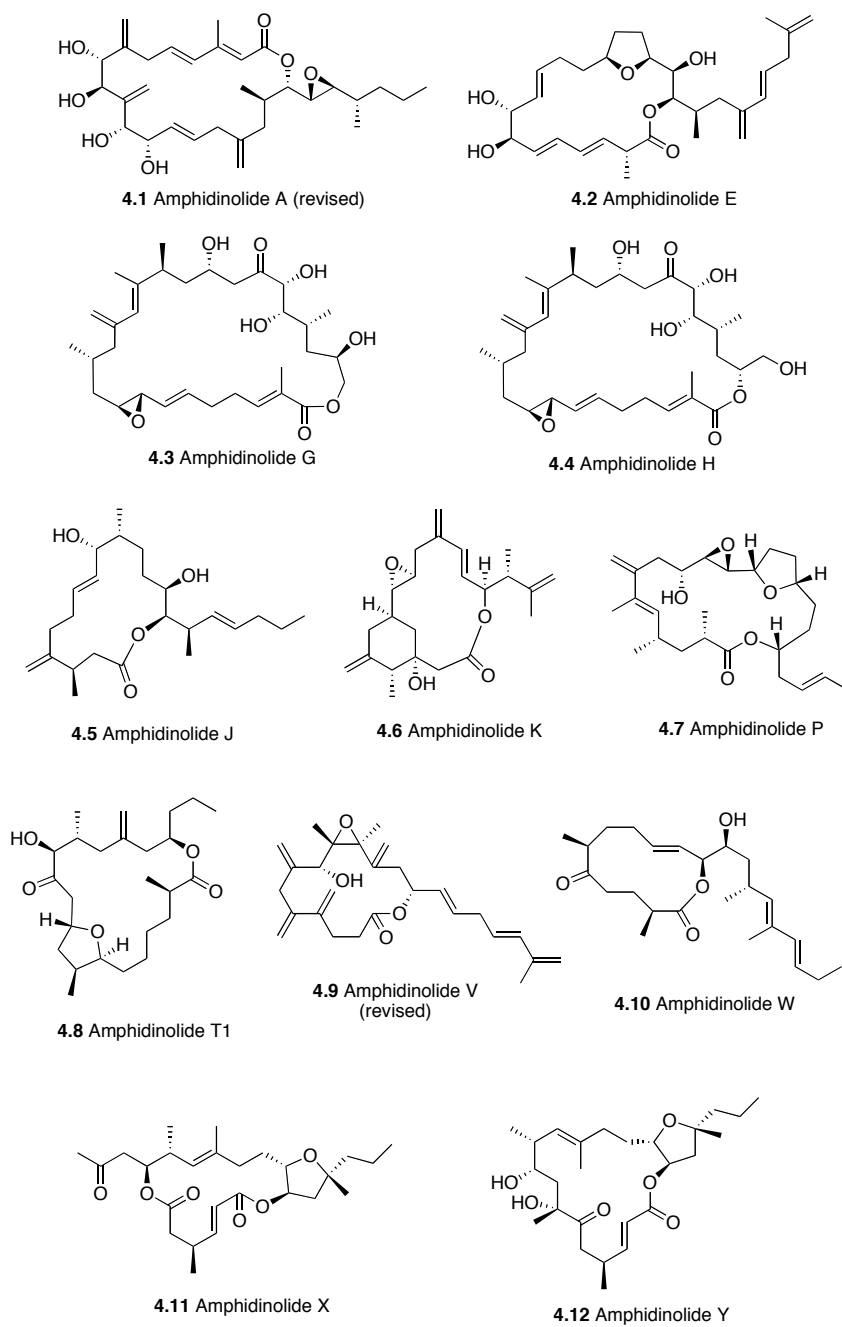
1. Zhou, X.-T.; Carter, R. G. *Angew. Chem., Int. Ed.* **2006**, 45, 1787.

**PART II: TOTAL SYNTHESIS OF AMPHIDINOLIDE B<sub>1</sub> AND THE  
PROPOSED STRUCTURE OF AMPHIDINOLIDE B<sub>2</sub>**

**CHAPTER 4: BACKGROUND OF AMPHIDINOLIDE B**

**4.1 Introduction of Amphidinolide Family**

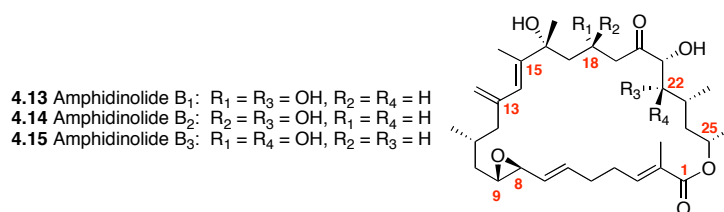
As the producers of substances with novel structures and appealing bioactivities, dinoflagellates have been investigated worldwide by natural product chemists.<sup>1</sup> Since Kobayashi and co-workers discovered the first amphidinolide, amphidinolide A, from the cultures of the dinoflagellates *Amphidinium sp.* in 1986,<sup>2</sup> amphidinolides have extended to a family of more than 30 macrolides consisting 12–29 membered macrocycles.<sup>3</sup> Most amphidinolides exhibit potent cytotoxicity against a series of human cancer cell lines.<sup>3</sup> Intrigued by their structural features and significant bioactivity, the synthetic community has devoted much attention to the synthesis of amphidinolides in the past two decades. Total syntheses of many amphidinolides including amphidinolide A<sup>4</sup>, E<sup>5</sup>, G and H<sup>6</sup>, J<sup>7</sup>, K<sup>8</sup>, P<sup>9</sup>, T<sup>10</sup>, V<sup>11</sup>, W<sup>12</sup>, X<sup>13</sup> and Y<sup>14</sup> have been accomplished, with several resulting in stereochemistry reassignment.



**Figure 4.1.** Synthesized Amphidinolides

## 4.2 Isolation and Bioactivity of Amphidinolide B

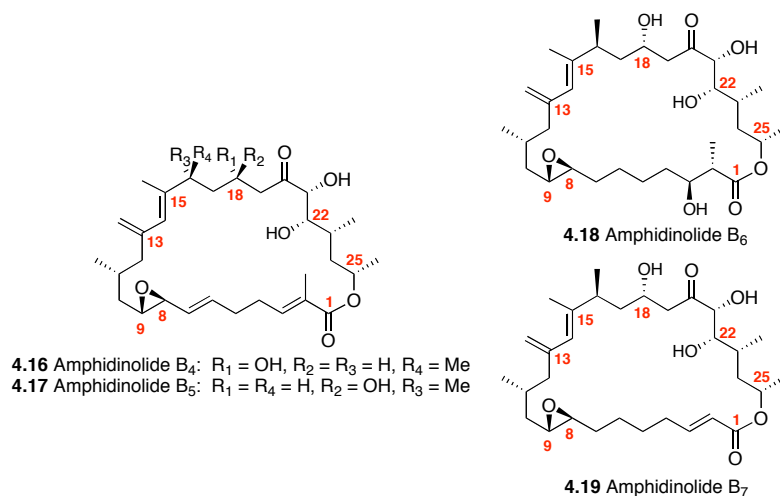
In 1987, the Kobayashi group discovered Amphidinolide B from the dinoflagellate *Amphidinium* sp., which was isolated from the Okinawan flatworm *Amphiscolops* sp..<sup>15</sup> Later, three amphidinolide B congeners, namely amphidinolides B<sub>1</sub> (**4.13**), B<sub>2</sub> (**4.14**) and B<sub>3</sub> (**4.15**), were isolated by Shimizu and co-workers from a free-swimming dinoflagellate *Amphidinium operculatum* ver nov *Gibbosum*.<sup>16</sup> In accord with the isolation of amphidinolide B, structural investigations by both the Kobayashi and Shimizu groups led to the determination of the relative stereochemistry of amphidinolide B<sub>1</sub> with the use of X-ray crystallography.<sup>15, 16</sup> Subsequently, the absolute stereochemistry was established via chemical degradation.<sup>17</sup> NMR spectra data analysis indicated that amphidinolide B<sub>2</sub> was the C<sub>18</sub> epimer of amphidinolide B<sub>1</sub> and the structure of amphidinolide B<sub>3</sub> was 22-*epi*-amphidinolide B<sub>1</sub>.<sup>16</sup>



**Figure 4.2.** Structure of Amphidinolide B<sub>1</sub>, B<sub>2</sub> and B<sub>3</sub>

Amphidinolide B is among the most cytotoxic molecules in the family of amphidinolides. Amphidinolide B<sub>1</sub> displays significant IC<sub>50</sub> values against a series of human cancer cell lines: the L1210 murine leukemia cell line (0.14 ng/mL); the

human colon tumor HCT 116 cell line (0.122  $\mu\text{g/mL}$ ); and the KB cancer cell line (4.2 ng/mL).<sup>18</sup> In addition to its potent cytotoxicity, amphidinolide B<sub>1</sub> was also used as a powerful activator of actomyosin ATPase to enhance skeletal muscle contraction.<sup>19</sup>



**Figure 4.3.** Structures of Amphidinolide B<sub>4</sub>, B<sub>5</sub>, B<sub>6</sub> and B<sub>7</sub>

More recently, several other amphidinolide B macrolides, namely amphidinolide B<sub>4</sub>, B<sub>5</sub>, B<sub>6</sub> and B<sub>7</sub>, were isolated from the marine acoel flatworms of the genus *Amphiscolops*.<sup>20</sup> Sharing similar structural features with amphidinolide B<sub>1</sub>, amphidinolide B<sub>4</sub> and B<sub>5</sub> showed potent cytotoxicity against the L1210 murine leukemia cell line (IC<sub>50</sub>: 0.12 ng/mL and 1.4 ng/mL, respectively) and the KB cancer cell line (IC<sub>50</sub>: 1.0 ng/mL and 4.0  $\mu\text{g/mL}$ , respectively), whereas amphidinolide B<sub>6</sub> and B<sub>7</sub> exhibited cytotoxicity against human B lymphocyte DG-75 cells (IC<sub>50</sub>: 0.02  $\mu\text{g/mL}$  and 0.4  $\mu\text{g/mL}$ , respectively).

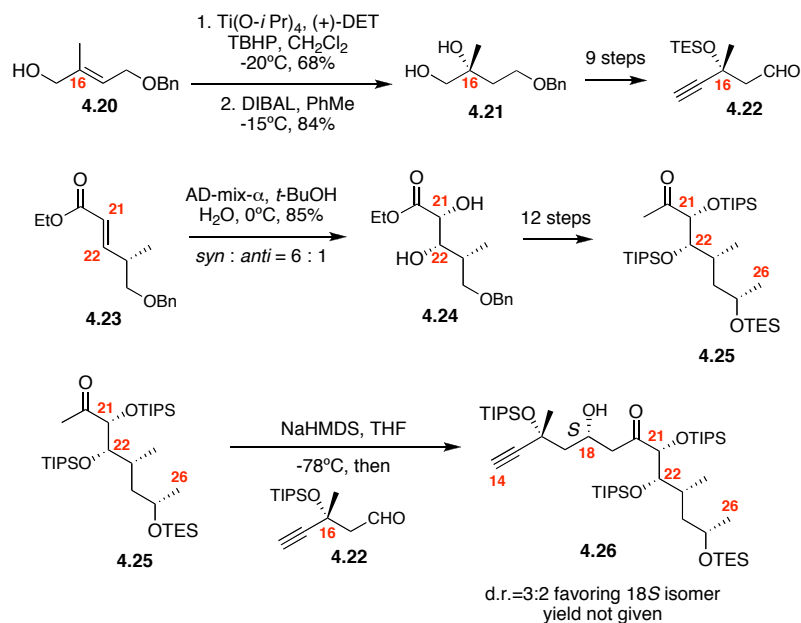
### 4.3 Synthetic Efforts toward Amphidinolide B<sub>1</sub>

Amphidinolide B<sub>1</sub> possesses unique structural features including a highly substituted C<sub>13</sub>-C<sub>15</sub> diene, the C<sub>21</sub>-C<sub>25</sub> domain with dense area of stereocenters, an unusual vinyl epoxide motif and a 26-membered macrolactone. In addition to the intriguing structure of amphidinolide B<sub>1</sub>, the highly potent cytotoxicity and the sparse amounts available from natural sources have made it an attractive synthetic target. Since the first synthetic efforts reported by Chakraborty and co-workers in 1997,<sup>21</sup> numerous research groups have been working on the synthesis of amphidinolide B<sub>1</sub>.<sup>21-26, 28, 29</sup> Despite all these efforts, no total synthesis had been accomplished prior to our efforts.

#### 4.3.1 The Chakraborty Group

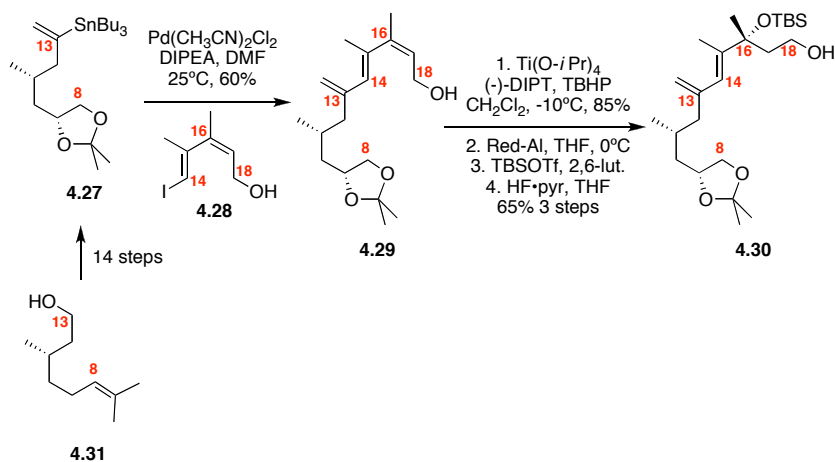
In 1997, Chakraborty and co-workers reported the first approach towards amphidinolide B<sub>1</sub>.<sup>21a-b</sup> In the synthesis of C<sub>14</sub>-C<sub>26</sub> moiety **4.26**, the C<sub>16</sub> tertiary alcohol was accessed from allylic alcohol **4.20** via Sharpless asymmetric epoxidation and a regioselective epoxide opening (Scheme 4.1). To set the *cis*-diol relationship across the C<sub>21</sub>-C<sub>22</sub> bond, the Sharpless dihydroxylation of unsaturated ester **4.23** (*cis:trans* = 6:1) was utilized. An aldol reaction between aldehyde **4.22** and methyl ketone **4.25** was employed to construct the C<sub>18</sub> stereochemistry in 3:2

dr, favoring the 18*S* isomer. The lengthy sequences and the poor diastereoselectivity of the aldol coupling made this method not practical.



**Scheme 4.1.** Chakraborty's Synthesis of C<sub>14</sub>-C<sub>26</sub> Portion

Later, the Chakraborty group published their revised approach to the synthesis of the C<sub>8</sub>-C<sub>18</sub> fragment (Scheme 4.2).<sup>21c</sup> A palladium-catalyzed Stille coupling was used to construct C<sub>13</sub>-C<sub>14</sub> bond. Subsequent Sharpless asymmetric epoxidation and a regioselective epoxide opening afforded the desired compound **4.30**. Although **4.30** was successfully made, the number of steps (14 steps from commercially available material **4.31**) required for synthesis of coupling precursor **4.27** diminished the efficiency of this approach.

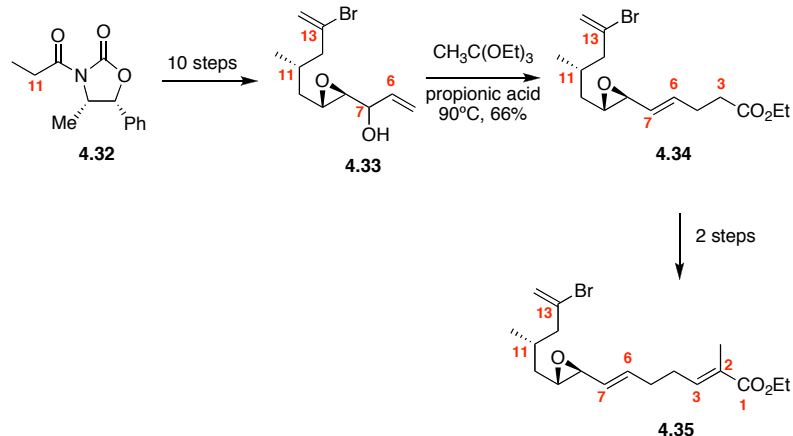


**Scheme 4.2.** Chakraborty's Revised Synthesis of C<sub>8</sub>-C<sub>18</sub> Motif.

### 4.3.2 The Lee Group

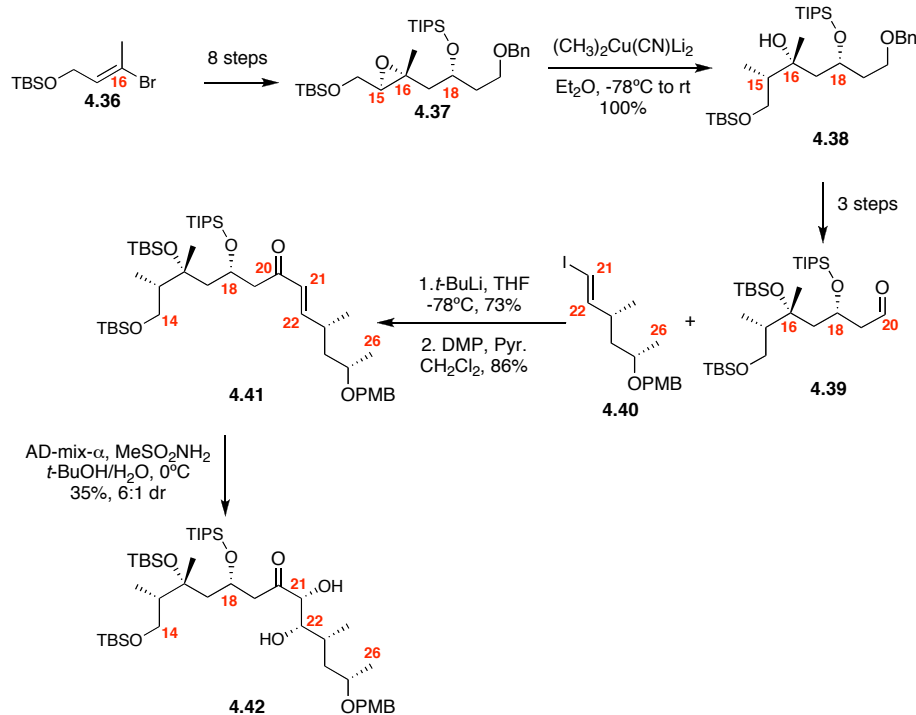
In 1997, the Lee group synthesized C<sub>1</sub>-C<sub>13</sub> portion **4.35** of amphidinolide B<sub>1</sub> via a 13-step sequence, starting from propionyl oxazolidinone **4.32** (Scheme 4.3).<sup>22a</sup> The orthoester Claisen rearrangement reaction was employed to form the C<sub>6</sub>-C<sub>7</sub> *trans* double bond. Further elaboration produced compound **4.35** in only 3.5% overall yield.





**Scheme 4.3.** Lee's Approach For the Synthesis of  $\text{C}_1\text{-C}_{13}$  Fragment

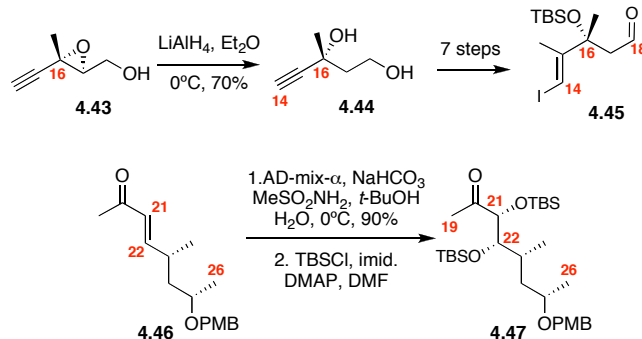
In a later 2000 publication, Lee and co-workers revealed their approach to the synthesis of the  $\text{C}_{14}\text{-C}_{25}$  fragment **4.42** (Scheme 4.4).<sup>22b</sup> A Sharpless asymmetric epoxidation, followed by the regioselective epoxide opening with methyl cuprate, yielded  $\text{C}_{16}$  tertiary alcohol. The  $\text{C}_{21}$  and  $\text{C}_{22}$  stereocenters were produced via Sharpless dihydroxylation in 35% yield in 6:1 dr.



**Scheme 4.4.** Lee's Efforts to the Synthesis of  $\text{C}_{14}\text{-C}_{26}$  Portion

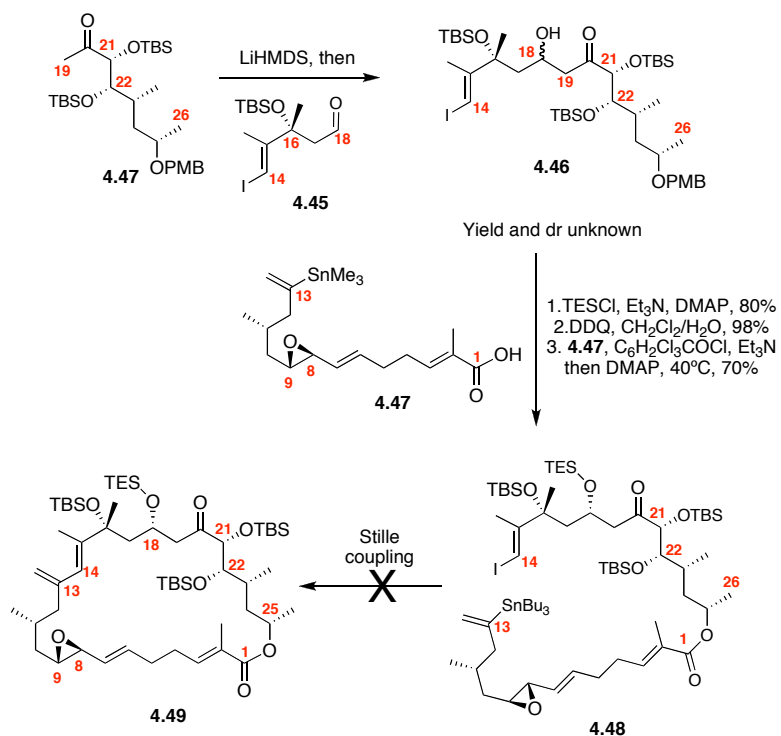
### 4.3.3 The Pattenden Group

In 1998, the Pattenden group synthesized the aldehyde **4.45** and methyl ketone **4.47**, the precursors required for the synthesis of  $\text{C}_{14}\text{-C}_{26}$  portion of amphidinolide B (Scheme 4.5).<sup>23a</sup> Similarly, the regioselective epoxide opening generated  $\text{C}_{16}$  tertiary alcohol while a Sharpless dihydroxylation was used to construct the  $\text{C}_{21}$  and  $\text{C}_{22}$  stereocenters. The conversion of alkyne **4.44** to the corresponding *E*-trisubstituted vinyl iodide **4.45** required seven steps.



**Scheme 4.5.** Pattenden's Synthesis of C<sub>14</sub>-C<sub>18</sub> and C<sub>19</sub>-C<sub>26</sub> Fragments

In Pattenden's subsequent research, the aldol reaction between **4.47** and **4.48** yielded the C<sub>18</sub> stereocenter; however, the yield and the dr were not specified by the authors (Scheme 4.6).<sup>23b</sup> An intermolecular Yamaguchi esterification linked the C<sub>1</sub>-C<sub>13</sub> intermediate to the C<sub>14</sub>-C<sub>26</sub> fragment. Unfortunately, efforts to affect an intramolecular Stille reaction for the construction of the C<sub>13</sub>-C<sub>14</sub> bond in the sterically demanding system were unsuccessful. Only a dimer species and a destannylated compound were formed.

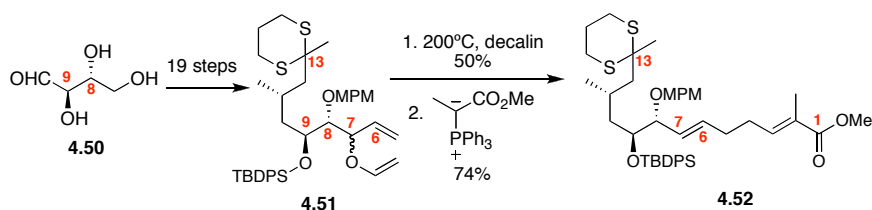


**Scheme 4.6.** Pattenden's Attempts on the Intramolecular Stille Reaction

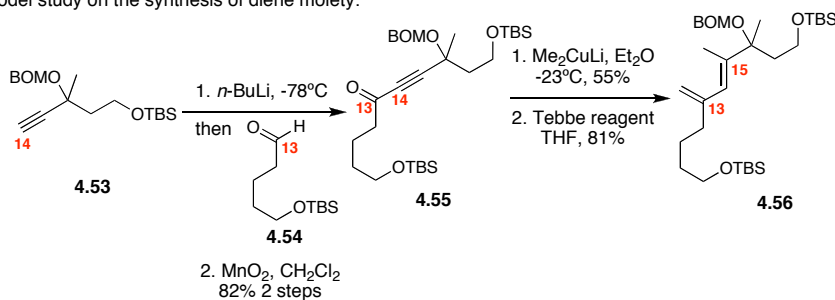
#### 4.3.4 The Nishiyama Group

At about the same time as Pattenden's research was published, the Nishiyama group reported their synthesis of the C<sub>1</sub>-C<sub>13</sub> subunit **4.52** (Scheme 4.7).<sup>24a</sup> A Claisen rearrangement was used to generate the C<sub>6</sub>-C<sub>7</sub> alkene, whereas a (*D*)-erythrose-derived diol **4.50** served as the source for the C<sub>8</sub> and C<sub>9</sub> stereocenters. In a following model study, the same group successfully synthesized the racemic form of diene **4.56** via a Michael addition of methyl group to compound **4.55** and the subsequent methylenation of the resulted enone with Tebbe reagent.<sup>24b</sup>

Synthesis of C<sub>1</sub>-C<sub>13</sub> fragment:

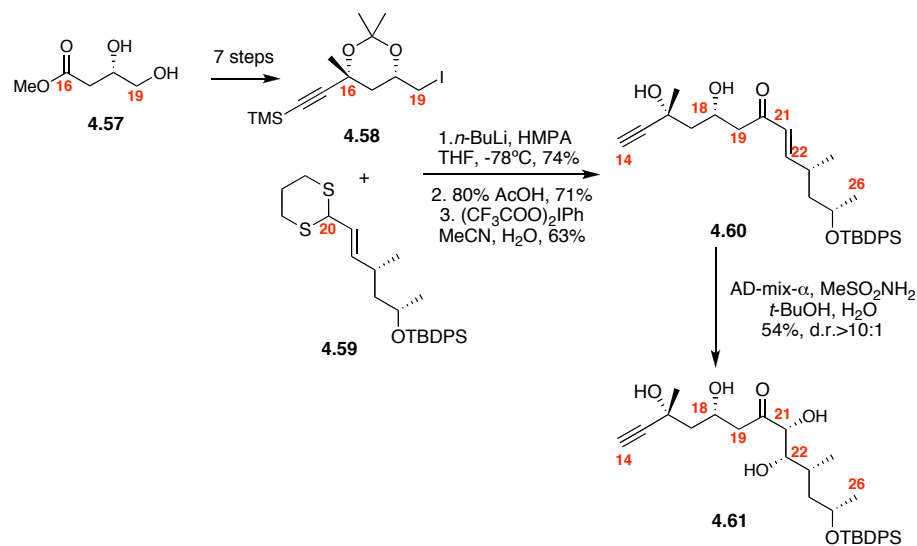


Model study on the synthesis of diene moiety:



**Scheme 4.7.** Nishiyama's Strategy for the Synthesis of C<sub>1</sub>-C<sub>13</sub> Fragment and the Model Study on the Synthesis of C<sub>13</sub>-C<sub>15</sub> diene

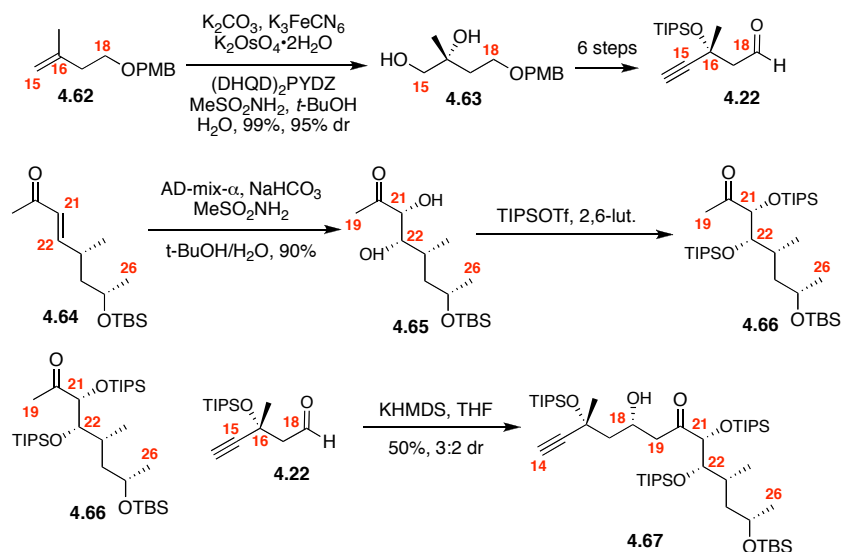
Encouraged by the results obtained from the model study, Nishiyama and co-workers then reported their strategy for the synthesis of the C<sub>14</sub>-C<sub>26</sub> fragment (Scheme 4.8).<sup>24c</sup> The key steps included the addition of the anion of dithiane **4.59** to iodide **4.58** and the Sharpless dihydroxylation of enone **4.60** to yield the C<sub>21</sub> and C<sub>22</sub> stereocenters in good diastereoselectivity. To date, the chemistry from the model study has not been applied to the real substrates.



**Scheme 4.8.** Nishiyama's Synthesis of C<sub>14</sub>-C<sub>26</sub> Motif

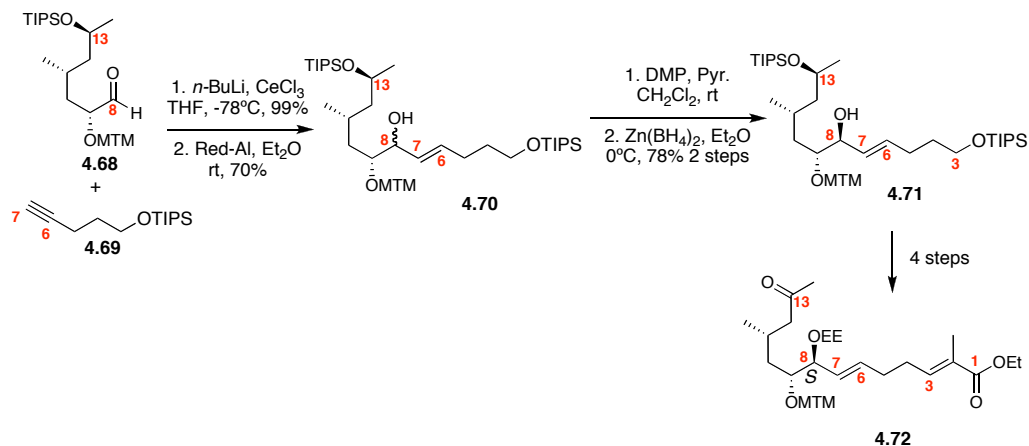
#### 4.3.5 The Kobayashi Group

Similar to Chakraborty's and Pattenden's syntheses, Kobayashi's approach to the synthesis of the C<sub>14</sub>-C<sub>26</sub> fragment **4.67** involves an aldol disconnection (Scheme 4.9).<sup>25a</sup> The C<sub>16</sub> tertiary alcohol stereocenter in aldehyde **4.22** was set using Sharpless' asymmetric dihydroxylation, as was the stereochemistry at C<sub>21</sub> and C<sub>22</sub>. The diastereoselectivity of the aldol coupling was poor, only 3:2 favoring the desired 18*S* diastereomer.



**Scheme 4.9.** Kobayashi's Synthesis of C<sub>14</sub>-C<sub>26</sub> Portion

After the preparation of C<sub>14</sub>-C<sub>26</sub> fragment, Kobayashi and co-workers reported the synthesis of the lower C<sub>1</sub>-C<sub>13</sub> fragment **4.72** (Scheme 4.10). The addition of acetylene **4.69** to aldehyde **4.68**, followed by the Red-Al-mediated reduction, generated the C<sub>6</sub>-C<sub>7</sub> double bond.<sup>25b</sup> A consecutive oxidation / reduction sequence produced the C<sub>8</sub> stereocenter. Further elaboration afforded the desired C<sub>3</sub>-C<sub>13</sub> fragment.

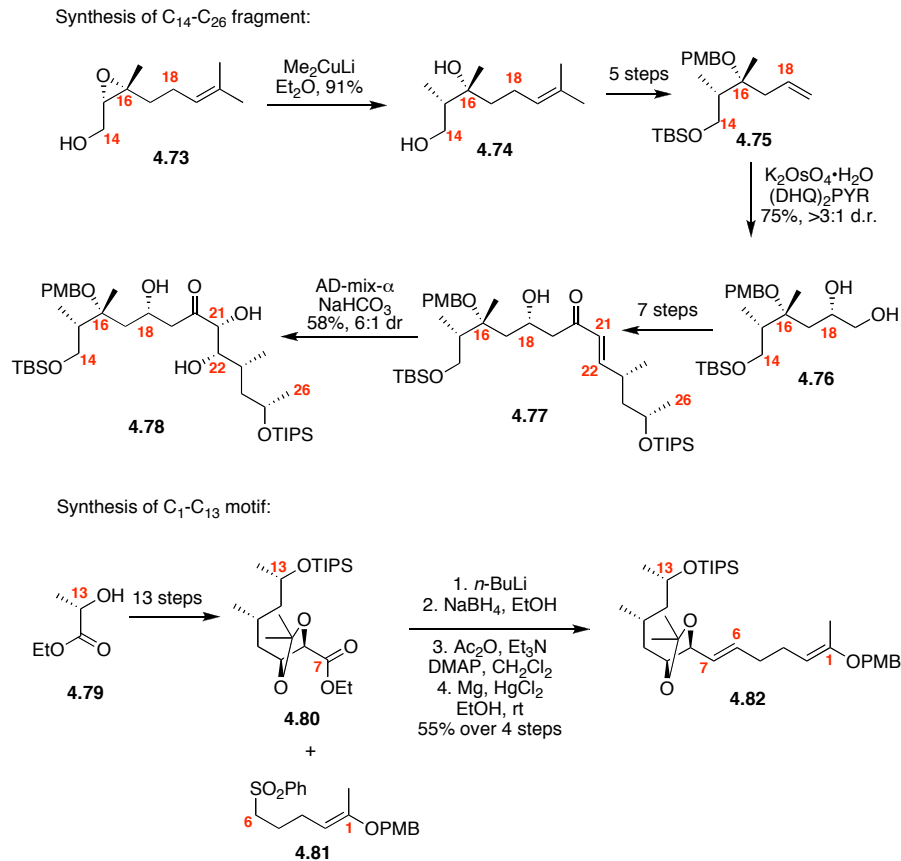


**Scheme 4.10.** Kobayashi's Approach to the Synthesis of  $\text{C}_1\text{-C}_{13}$  Intermediate

#### 4.3.6 The Myles Group

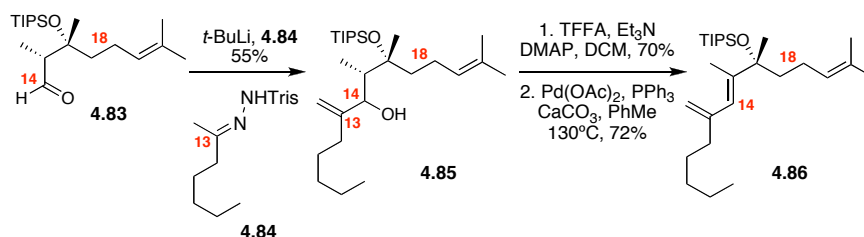
In 1999, the Myles group reported the synthesis of the  $\text{C}_1\text{-C}_{13}$  and the  $\text{C}_{14}\text{-C}_{26}$  fragments (Scheme 4.11).<sup>26a</sup> The  $\text{C}_6\text{-C}_7$  alkene found in  $\text{C}_1\text{-C}_{13}$  subunit was furnished through a Julia coupling. The strategy to  $\text{C}_{14}\text{-C}_{26}$  fragment featured an epoxide opening to build  $\text{C}_{16}$  tertiary alcohol, and a Sharpless dihydroxylation to generate the  $\text{C}_{21}$  and  $\text{C}_{22}$  stereocenters.





**Scheme 4.11.** Myles' Strategy for the Synthesis of C<sub>1</sub>-C<sub>13</sub> and C<sub>14</sub>-C<sub>26</sub> Fragments

In Myles' model study,<sup>26b</sup> a Shapiro reaction between aldehyde **4.83** and trisylhydrazone **4.84** was utilized to form the C<sub>13</sub>-C<sub>14</sub> bond. The diene motif **4.86** was successfully synthesized through a sequence involving the conversion of alcohol **4.85** to the corresponding trifluoroacetate ester and the Pd-mediated Hauser-type elimination (Scheme 4.12).<sup>27</sup> To date, there have been no reports that discuss applying the developed approach on the authentic substrates.

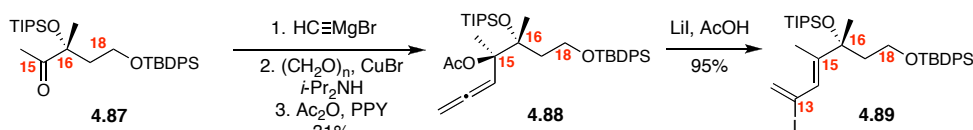


**Scheme 4.12.** Myles' Model Study on the Synthesis of the Diene Motif

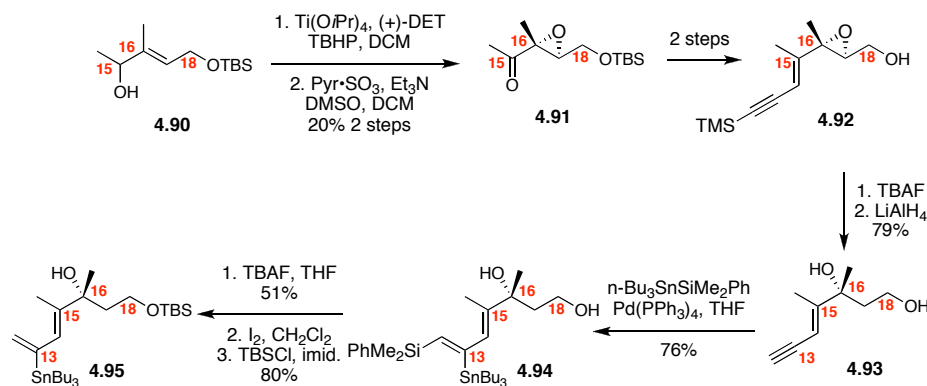
### 4.3.7 The Crews Group

In their 2005 publication, Crews and co-workers revealed their first-generation strategy for the synthesis of the C<sub>1</sub>-C<sub>12</sub>, C<sub>13</sub>-C<sub>18</sub> and C<sub>19</sub>-C<sub>25</sub> fragments (Scheme 4.13).<sup>28a</sup> Notable steps included an iodide-mediated S<sub>N</sub><sup>2'</sup> reaction on the allenic acetate **4.88** to generate the diene motif **4.89**; however, the yield of the synthesis of the allenic acetate **4.88** was moderate. One year later, the second-generation approach for the synthesis of the three fragments was reported (Scheme 4.13).<sup>28b</sup> Stereochemistry at C<sub>16</sub> was installed via Sharpless asymmetric epoxidation / regioselective epoxide opening. The triple bond of enyne **4.93** was silylstannylated regio- and stereoselectively employing *n*-Bu<sub>3</sub>SnSiMe<sub>2</sub>Ph/Pd(PPh<sub>3</sub>)<sub>4</sub> to ultimately furnish the functionalized diene **4.94**. After the removal of PhMe<sub>2</sub>Si group with TBAF, sequential iodization and selective TBS protection gave the C<sub>13</sub>-C<sub>18</sub> fragment **4.95**.

First-generation synthesis of C<sub>13</sub>-C<sub>18</sub> fragment:



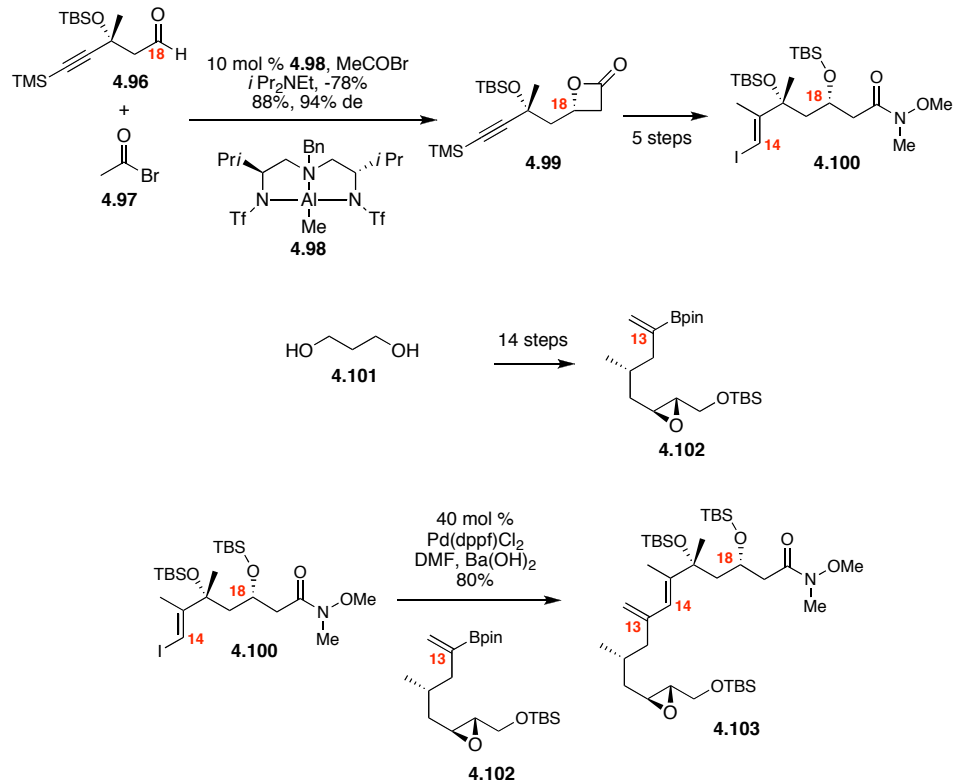
Second-generation synthesis of C<sub>13</sub>-C<sub>18</sub> fragment:



**Scheme 4.13.** Crews' Efforts on the Synthesis of Amphidinolide B<sub>1</sub>

### 4.3.8 The Nelson Group

In 2006, Nelson and co-workers reported a successful synthesis of the C<sub>7</sub>-C<sub>20</sub> fragment (Scheme 4.14).<sup>29</sup> Nelson's approach featured with an asymmetric acyl halide-aldehyde cyclocondensation to furnish the C<sub>18</sub> stereochemistry and a Pd-catalyzed Suzuki coupling between iodide **4.100** and boronic ester **4.102** to yield diene motif **4.103**. Unfortunately, the synthesis of Suzuki coupling precursor **4.102** required 14 steps from the commercially available starting material and 40 mol % palladium catalyst was used in the key Suzuki coupling.

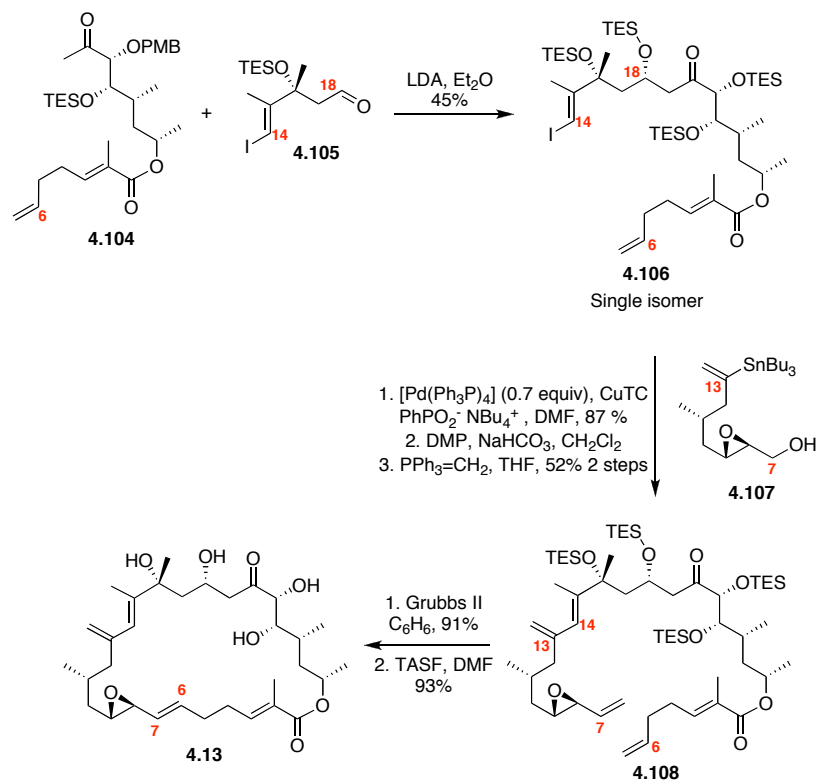


**Scheme 4.14.** Nelson's Synthesis of C<sub>7</sub>-C<sub>20</sub> Fragment

### 4.3.9 Fürstner's Total Synthesis of Amphidinolide B<sub>1</sub>

In 2008, our group reported the first conquest of amphidinolide B<sub>1</sub> and the proposed structure of amphidinolide B<sub>2</sub>.<sup>30</sup> More recently, the second total synthesis of amphidinolide B<sub>1</sub> by Fürstner and co-workers has followed.<sup>31</sup> The C<sub>18</sub> stereocenter was established via a chelation controlled aldol coupling developed by our group.<sup>32</sup> The successful route hinged upon a highly productive Stille–Migita cross-coupling reaction to construct C<sub>13</sub>-C<sub>15</sub> diene motif, which required 70 mol % palladium catalyst and the development of a chloride- and fluoride-free

protocol. The macrocycle was furnished via ring-closing metathesis engaging a vinyl epoxide unit as one of the reaction partners.



**Scheme 4.15.** Fuestner's Total Synthesis of Amphidinolide B<sub>1</sub>

#### 4.4 Conclusion

Although extensive efforts have been made toward the synthesis of amphidinolide B<sub>1</sub> since 1997, questions still arise as to how to efficiently prepare the C<sub>13</sub>-C<sub>15</sub> diene motif, cyclize the 26-membered macrocycle, and incorporate the labile allylic epoxide moiety by the date we initiated our synthetic study. In 2008, our group accomplished the first total synthesis of amphidinolide B<sub>1</sub> and the

proposed structure of amphidinolide B<sub>2</sub>.<sup>30</sup> One year later, another total synthesis of amphidinolide B<sub>1</sub> was reported by Fürstner and co-workers.<sup>31</sup> Herein, our synthetic studies toward amphidinolide B will be detailed in the next several chapters.

#### 4.5 References

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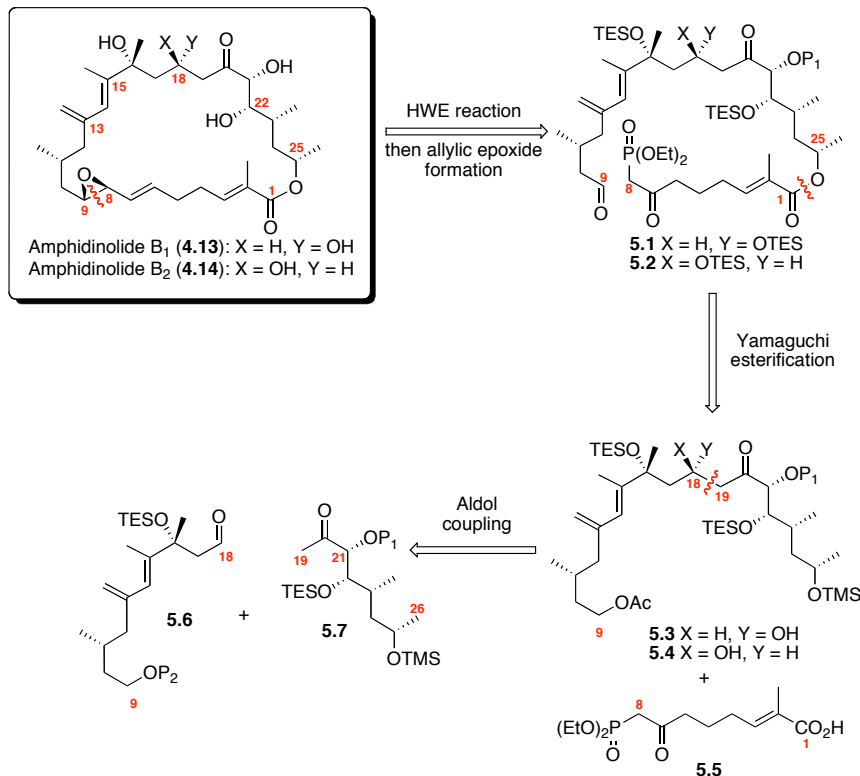
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## CHAPTER 5. SYNTHESIS OF DIENE SUBUNITS

### 5.1 Retrosynthesis

As discussed in the previous chapter, amphidinolide B has shown potent cytotoxicity against several cancer cell lines and its structural architecture contains an unusual highly substituted diene, a dense area of stereocenters, a 26-membered macrocycle and a labile allyl epoxide moiety. After more than ten years of extensive efforts toward the synthesis of amphidinolide B, most synthetic problems were still not successfully addressed. Attracted by the unique structure and the potent cytotoxicity of amphidinolide B, we initiated our synthetic research with the intention of synthesizing the key motifs and ultimately complete the total synthesis of amphidinolide B. Our retrosynthesis commences with a disconnection at C<sub>8,9</sub> via intramolecular HWE reaction to reveal aldehyde **5.1** and **5.2** (Scheme 2.1). Further cleavage at C<sub>18,19</sub> via aldol coupling and C<sub>1</sub> C-O bond via Yamaguchi esterification resulted in three key intermediates: aldehyde **5.6**, methyl ketone **5.7** and phosphonate **5.5**. Our idea was to control the stereoselectivity of the aldol reaction between aldehyde **5.6** and methyl ketone **5.7** by employing different protecting group for the C<sub>21</sub> hydroxyl group.

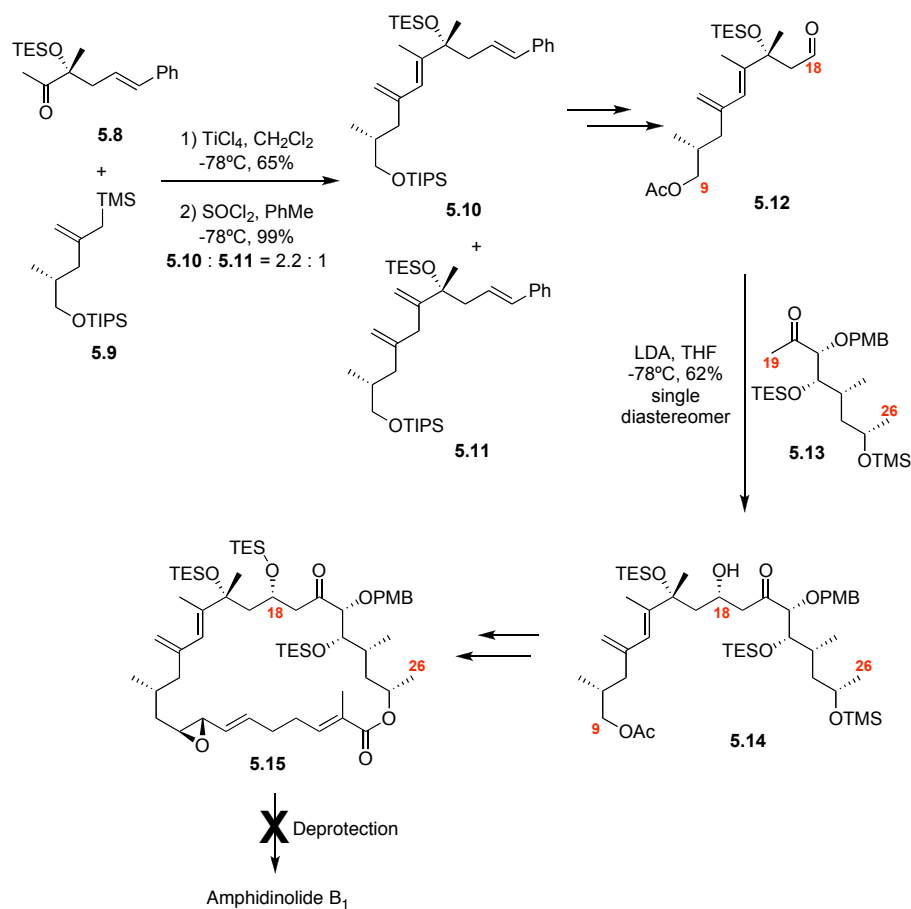


**Scheme 5.1.** Retrosynthetic Study of Amphidinolide B<sub>1</sub> and B<sub>2</sub>

## 5.2 Our Previous Progress toward the Synthesis of Amphidinolide B<sub>1</sub>

After several years of extensive efforts, our group had made significant progress toward the total synthesis of amphidinolide B<sub>1</sub> (Scheme 5.2).<sup>1</sup> We have previously reported the 1<sup>st</sup> generation synthesis of C<sub>9</sub>-C<sub>26</sub> fragment of amphidinolide B<sub>1</sub>, which featured a chelation controlled aldol reaction between aldehyde **5.12** and methyl ketone **5.13** to build the C<sub>18</sub> stereocenter. The other key steps included the Fleming-type coupling of two readily available subunits, methyl ketone **5.8** and allyl silane **5.9**, and the following SOCl<sub>2</sub> dehydration to furnish the

highly substituted C<sub>13</sub>-C<sub>15</sub> diene. Further elaboration afforded the fully functionalized amphidinolide B<sub>1</sub> **5.15** with the hydroxyl groups protected with TES groups and a PMB group. Unfortunately, all the attempts to remove protecting groups resulted in decomposition of the substrate. Similar results have been reported by Fürstner in his recent synthesis of amphidinolide G and H.<sup>2</sup> Besides the difficulties associated with the deprotection step, the inability to scale up the coupling between methyl ketone **5.8** and allyl silane **5.9**, as well as the tedious separation of diene **5.10** and its isomer **5.11** presented additional obstacles in our efforts to finish the total synthesis of amphidinolide B<sub>1</sub>.



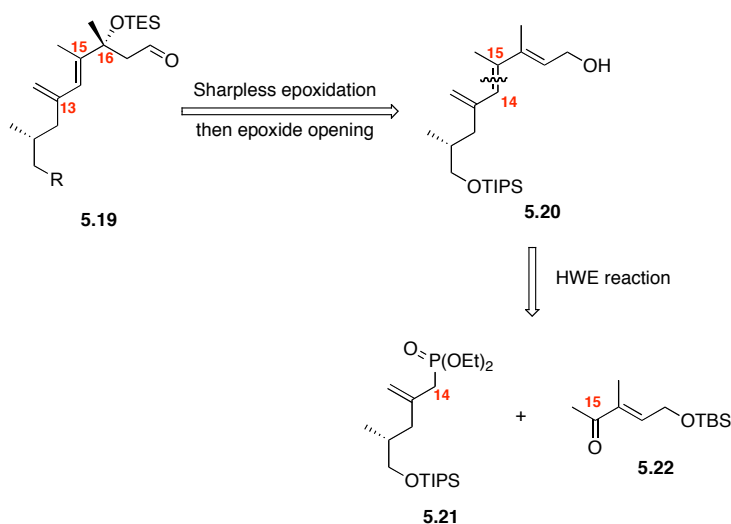
**Scheme 5.2.** Our Previous Progress toward the Synthesis of Amphidinolide B<sub>1</sub>

### 5.3 Modified Strategy for the Synthesis of C<sub>13</sub>-C<sub>15</sub> Diene Motif

#### 5.3.1 Retrosynthetic Analysis

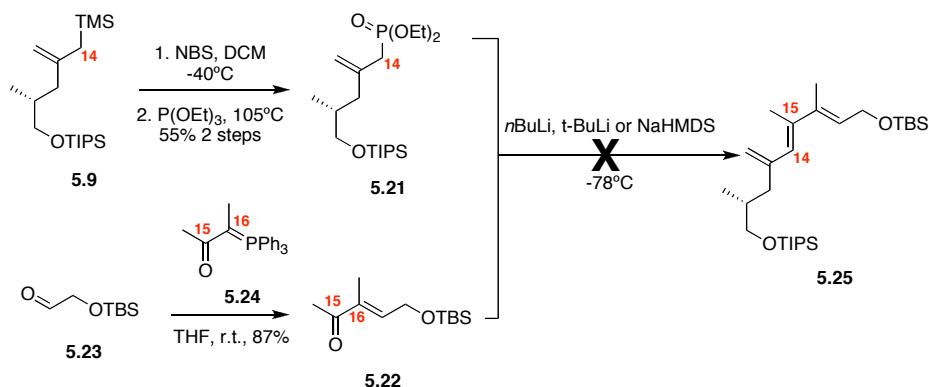
Faced with the roadblocks mentioned previously, we were forced to reconsider the strategy for the synthesis of the C<sub>13</sub>-C<sub>15</sub> diene moiety. As is discussed in the previous chapter, this diene functionality has frustrated numerous synthetic laboratories during their endeavors toward amphidinolide B. In the prior

syntheses, the disconnection between C<sub>13</sub>-C<sub>14</sub> linkage typically involves a palladium-mediated coupling between a suitably functionalized and stereodefined vinyl halide and its corresponding 1,1-disubstituted coupling partner. As has been demonstrated by Pattenden,<sup>3</sup> this coupling in sterically challenging systems does not perform well. More recently, Nelson and co-workers<sup>4</sup> did demonstrate a successful metal-mediated coupling process; however, it required extremely high catalyst loadings (40 mol%) – a requirement not amendable to total synthesis. The previously reported research and our own experience made us mindful in selecting the approaches for the synthesis of the diene subunit **5.19**. In our modified retrosynthesis, to avoid the steric bulk introduced by the C<sub>16</sub> tertiary alcohol, the subsequent Sharpless epoxidation and the regioselective epoxide opening were employed to install the C<sub>16</sub> stereocenter after the formation of the diene motif. Instead of the C<sub>13</sub>-C<sub>14</sub> cleavage, we chose to disconnect the molecule at the C<sub>14</sub>-C<sub>15</sub> double bond. Using this method, we could envision the formation of two subunits as coming from the methyl ketone **5.22** and an allyl phosphonate **5.21**.<sup>5</sup>



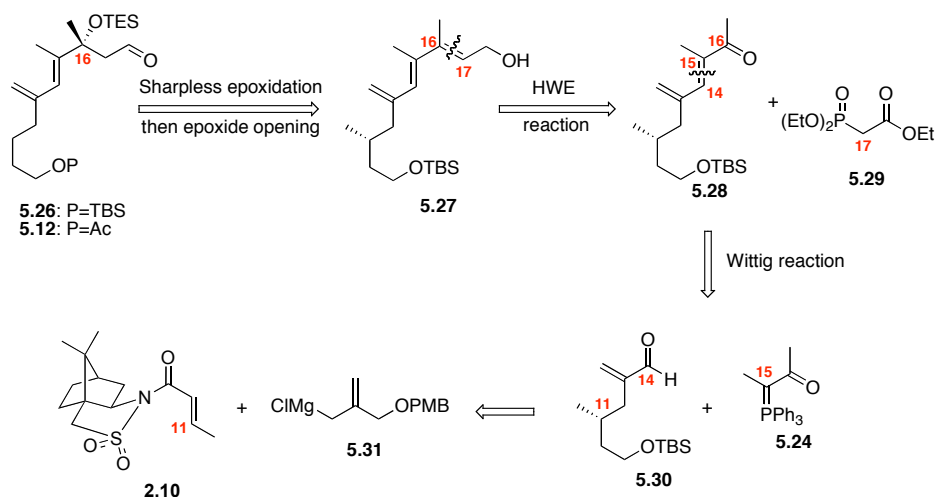
**Scheme 5.4.** Retrosynthesis of Diene **5.19**.

Commenced from previously synthesized allyl silane **5.9**,<sup>1</sup> phosphonate **5.21** was produced in moderate yield after a bromation and subsequent Arbuzov reaction (Scheme 5.5). The Wittig reaction between the known ylide **5.24**<sup>6</sup> and aldehyde **5.23**<sup>7</sup> furnished methyl ketone **5.22**. With these two intermediates in hand, we investigated the Horner-Wadsworth-Emmons olefination. Unfortunately, our repeated attempts proved unsuccessful as phosphonate **5.21** decomposed when treated with strong base (e.g. *n*BuLi, *t*-BuLi, KHMDS) at -78°C.



**Scheme 5.5.** HWE Reaction between Phosphoante **5.21** and Methyl Ketone **5.22**

After the unsuccessful attempts of using HWE olefination, we sought a Wittig reaction<sup>5</sup> as a reasonable substitute to construct the C<sub>14</sub>-C<sub>15</sub> alkene (Scheme 5.6). As similar method was used to produce the C<sub>16</sub> tertiary alcohol from triene alcohol **5.27**, the C<sub>16</sub>-C<sub>17</sub> double bond could be obtained from a HWE reaction. Further disconnection at C<sub>14</sub>-C<sub>15</sub> led to the known ylide **5.24**<sup>6</sup> and the aldehyde **5.30**. The C<sub>11</sub> stereocenter of aldehyde **5.30** could be generated through the cuprate addition to the commercially available Oppolzer sultam derivative **2.10**.<sup>8</sup> Our synthetic plan required the selective removal of C<sub>9</sub> protecting group in the presence of secondary TES and PMB groups. The use of TBS group would be the first option based on our strategy for the synthesis of diene subunit. If the deprotection proved problematic, the previously employed acetate group<sup>1</sup> would be an alternative. Next, our focus shifted to the synthesis of C<sub>9</sub> TBS protected and C<sub>9</sub> acetate protected diene fragments.

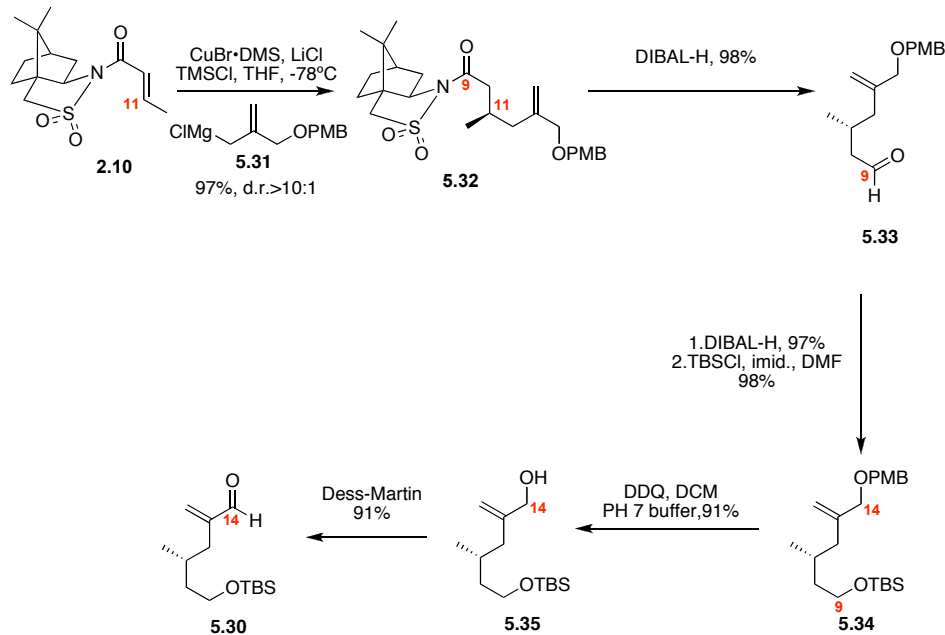


**Scheme 5.6.** Modified Retrosynthesis of Diene Subunit

### 5.3.2 Preparation of Aldehyde 5.30

The required aldehyde **5.30** was prepared in six steps from the known sultam **2.10** (Scheme 5.6).<sup>9</sup> Cuprate addition of Grignard reagent **5.31**<sup>9</sup> to sultam **2.10**, under similar conditions described by Paquette,<sup>9</sup> afforded the stereocenter at C<sub>11</sub> with excellent diastereoselectivity (dr>20:1). The following reductive cleavage of the auxiliary and C<sub>9</sub> TBS protection produced TBS ether **5.34**. The PMB group was then removed using DDQ to yield alcohol **5.35**. Finally, Dess-Martin oxidation<sup>10</sup> revealed the coupling precursor **5.30**.



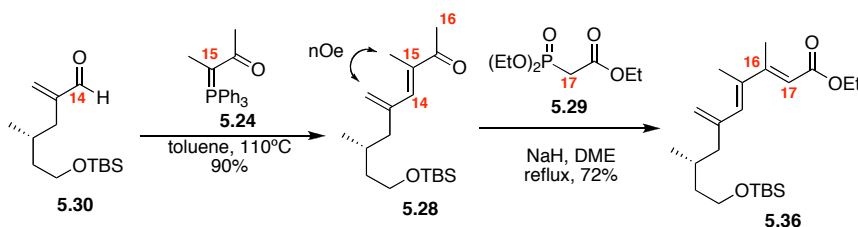


**Scheme 5.7.** Synthesis of Aldehyde **5.30**

### 5.3.3 Wittig / HWE Reaction Sequence

With aldehyde **5.30** in hand, we next explored the key Wittig / HWE reaction sequence (Scheme 5.8). In general, “stabilized” ylides with strongly conjugating substituents (e.g.,  $\text{COOMe}$ ,  $\text{CN}$ , or  $\text{COCH}_3$ ) on the ylidic carbon are known to favor the production of *E* alkenes.<sup>5</sup> We were pleased to find that the Wittig reaction between aldehyde **5.30** and the known ylide **5.24**<sup>6</sup> performed smoothly in good yield and great *E/Z* selectivity. The geometry of  $\text{C}_{14}\text{-C}_{15}$  double bond was confirmed via nOe analysis. Due to the low reactivity of ketone **5.28** with phosphonate **5.29**,<sup>5</sup> the Horner-Wadsworth-Emmons olefination was sluggish

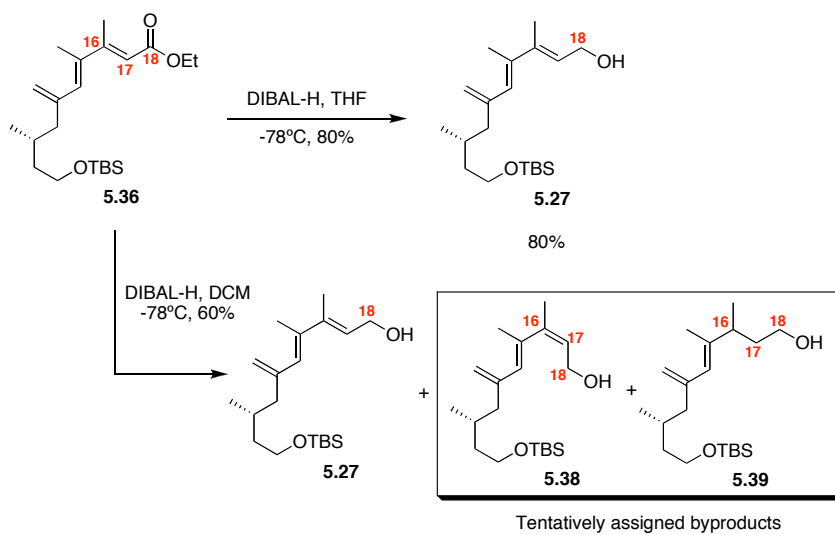
at room temperature. Fortunately, upon refluxing the reaction mixture in DME, we observed significant rate acceleration and the reaction was completed in 4 hours to give C<sub>16</sub>-C<sub>17</sub> alkene.



**Scheme 5.8.**Wittig / HWE Reaction Sequence

#### 5.3.4 Reduction of $\alpha,\beta$ -Unsaturated Ester **5.36**

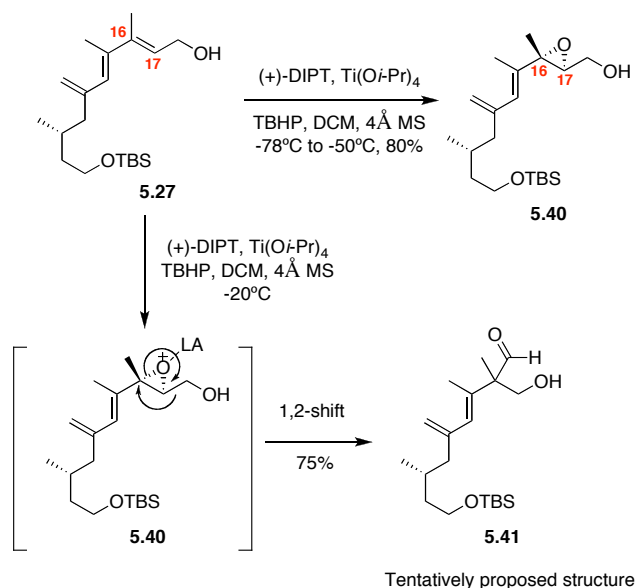
The Wittig / HWE reaction sequence generated the highly substituted C<sub>13</sub>-C<sub>15</sub> diene moiety successfully. The remained challenges were the installation of C<sub>16</sub> tertiary alcohol via sequential epoxidation and regioselective opening of the epoxide. The required reduction of ester **5.36** is shown in Scheme 5.9. Under typical DIBAL-H reduction conditions (DIBAL-H / DCM), moderate yield was obtained with the occurrence of the several undesired compounds. The unidentified by-products appeared to arise from the double bond *E/Z* isomerization and 1,4-reduction of  $\alpha,\beta$ -unsaturated ester **5.36**. Interestingly, using THF as solvent<sup>11</sup> suppressed the formation of the by-products and the yield was improved to 80%.



**Scheme 5.9.** Reduction of Ester **5.36**

### 5.3.5 Epoxidation of C<sub>16</sub>-C<sub>17</sub> Alkene

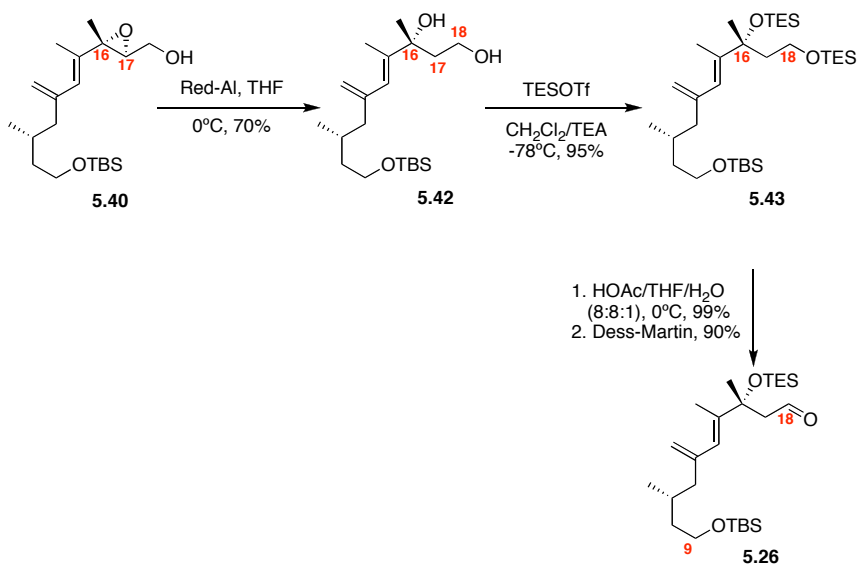
After obtaining triene alcohol **5.27**, we investigated the epoxidation on this highly reactive system. When the epoxidation was conducted at -20°C, we were surprised that the major product was an aldehyde most likely resulting from Yamamoto-type epoxide rearrangement of substrate **5.40**.<sup>12</sup> Lower temperature (-78°C to -50°C) and freshly distilled Ti(O-*i*Pr)<sub>4</sub> suppressed this 1,2-alkyl shift and the desired epoxide **5.40** was obtained in 80% yield.



**Scheme 5.10.** Epoxidation of C<sub>16</sub>-C<sub>17</sub> Alkene

### 5.3.6 Regioselective Opening of the Epoxide

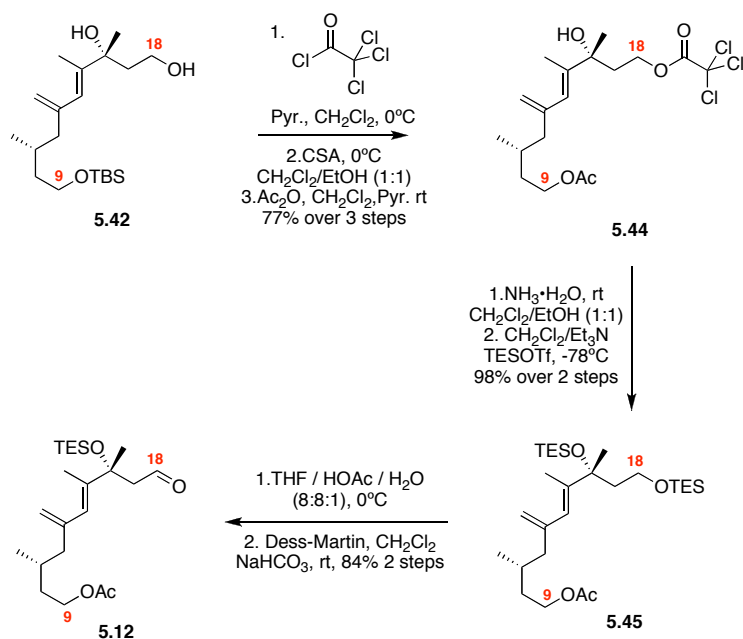
With the epoxidation of C<sub>16</sub>-C<sub>17</sub> alkene, the last challenge was the regioselective epoxide opening (Scheme 5.11). Base-catalyzed epoxide opening in which nucleophile provides the driving force generally involves the break of the C-O bond at the less substituted position.<sup>13</sup> Gratifyingly, treatment of epoxide **5.42** with Red-Al<sup>14</sup> yielded the desired diene diol **5.43**. Subsequent TES protection, selective removal of primary TES group<sup>15</sup> and Dess-Martin oxidation<sup>11</sup> finally afforded diene aldehyde **5.26**.



**Scheme 5.12.** Synthesis of Diene Aldehyde **5.26**

### 5.3.7 Synthesis of C<sub>9</sub> Acetate Protected Diene Motif **5.12**

Synthesis of C<sub>9</sub> acetate-protected diene aldehyde **5.12** required protecting group manipulations on diene diol **5.42**, which was realized by using a trichloroacetate group<sup>16</sup> (Scheme 5.13). Commenced from diene diol **5.42**, protecting group manipulation afforded ester **5.44**. Next, selective removal of the trichloroacetate group in the presence of acetate group,<sup>16</sup> followed by the TES protection, yielded TES ether **5.48**. Finally, consecutive deprotection of the C<sub>18</sub> primary TES and Dess-Martin oxidation give the desired C<sub>9</sub> acetate protected diene aldehyde **5.12**.



**Scheme 5.13.** Synthesis of Diene Aldehyde **5.12**

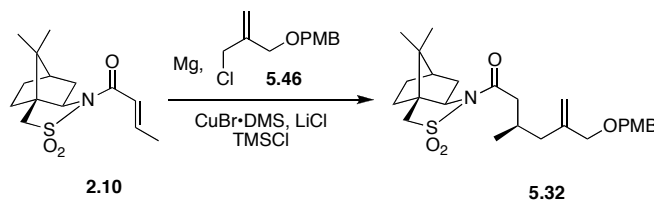
## 5.4 Conclusion

In summary,  $\text{C}_9$  TBS-protected and acetate-protected  $\text{C}_9$ - $\text{C}_{18}$  diene subunits have been synthesized diastereoselectively from commercially available Oppolzer sultam derivative **2.10** in 13 steps in 20% overall yield and in 17 steps in 14% overall yield respectively. The key steps included a Wittig / HWE sequence to yield  $\text{C}_{13}$ - $\text{C}_{15}$  diene moiety and a regioselective epoxide opening to generate the  $\text{C}_{16}$  stereocenter. The new strategy has proven to be much more conducive to scale up than our 1<sup>st</sup> generation synthesis. Both diene aldehydes have been prepared on grams scale, which provided a solid base for the completion of the total syntheses of amphidinolide  $\text{B}_1$  and  $\text{B}_2$ .

## 5.5 References

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## 5.6 Experimental

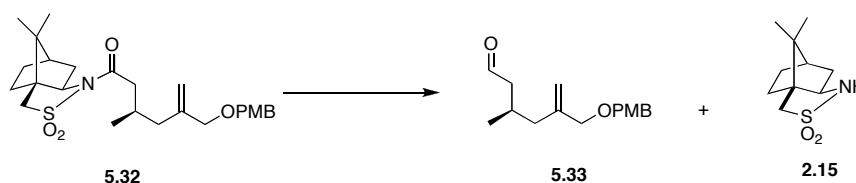


**Sultam 5.32:** Following the similar procedure described by Paquette,<sup>1</sup> Mg (36.0 g, 1.5 mol) was stirred vigorously at rt in a dry flask under Ar. After 120 h, when black coating formed inside the flask, THF (200 mL) and 1,2-dibromoethane (2.60 g, 1.2 mL, 13.9 mmol) were added sequentially. After 30 min, a solution of allyl chloride **5.46** (17.0 g, 75.0 mmol) in THF (80 mL) was added slowly to the Mg slurry over 5 h. The resulted mixture was stirred overnight at rt to give 300 mL Grignard reagent (0.12 M, 47%) as gray solution. The concentration of the Grignard reagent was determined by the titration using menthol in the presence of 1,10-phenanthroline.<sup>2</sup>

Separately, CuBr•SMe<sub>2</sub> (7.29 g, 35.5 mmol) and LiCl (1.61 g, 37.9 mmol) were dissolved in THF (80 mL) and added to the solution of Grignard reagent (263 mL, 31.5 mmol) at -78°C *via* syringe. TMSCl (3.96 g, 4.5 mL, 36.5 mmol) was then added followed by a solution of known sultam **2.10**<sup>3</sup> (6.9 g, 24.3 mmol) in THF (60 mL). After another 90 min, the reaction was quenched with NH<sub>4</sub>Cl-NH<sub>4</sub>OH (9:1, pH 9, 60 mL), warmed to rt. The aqueous layer was extracted with EtOAc (3 X 200 mL). The organic phase was washed with sat. aq. NaCl (100 mL). The dried extract (MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by

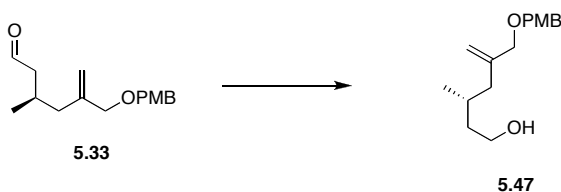


chromatography over silica gel, eluting with 8-15% EtOAc / Hexanes, to give the product **5.32** (11.2 g, 34.4 mmol, 97%) as a colorless oil:  $[\alpha]_D^{23} = -68.0$  ( $c$  0.51,  $\text{CHCl}_3$ ); IR (neat) 2959, 2927, 2851, 1693, 1512, 1454, 1328, 1246, 1032  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.28 (d,  $J = 8.7$  Hz, 2H), 6.88 (d,  $J = 8.7$  Hz, 2H), 5.12 (s, 1H), 4.96 (s, 1H), 4.44 (t,  $J = 11.8$  Hz, 2H), 3.94 (t,  $J = 11.8$  Hz, 2H), 4.00 (dd,  $J = 14.0$  Hz, 2H), 3.88 (t,  $J = 6.2$  Hz, 1H), 3.82 (s, 3H), 3.46 (dd,  $J = 23.0, 13.8$  Hz, 2H), 2.78 (dd,  $J = 16.1, 5.7$  Hz, 1H), 2.51 (dd,  $J = 16.1, 7.6$  Hz, 1H), 2.30-2.40 (m, 1H), 2.02-2.15 (m, 4H), 1.82-1.96 (m, 3H), 1.28-1.45 (m, 3H), 1.15 (s, 3H), 0.99 (d,  $J = 6.4$  Hz, 3H), 0.98 (s, 3H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  171.4, 159.1, 144.1, 130.6, 129.3, 113.7, 113.6, 72.4, 71.7, 65.2, 55.3, 53.0, 48.3, 47.7, 44.7, 42.5, 40.8, 38.6, 32.9, 28.0, 26.5, 20.8, 19.9; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{26}\text{H}_{37}\text{NO}_5\text{SNa}$  ( $\text{M}+\text{Na}$ ) 498.2290, found 498.2271.

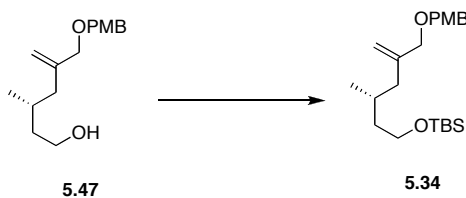


**Aldehyde 5.33:** To a stirred solution of sultam **5.32** (11.0 g, 23.1 mmol) in  $\text{CH}_2\text{Cl}_2$  (115 mL) at  $-78^\circ\text{C}$  was added DIBAL-H (50.8 mL, 50.8 mmol, 1.0 M in  $\text{CH}_2\text{Cl}_2$ ). After 2 h, the reaction was carefully quenched with methanol (2.0 mL) and poured into aq. sodium potassium tartrate (250 mL, 10% aq.) at rt. The reaction flask was rinsed with an additional portion of  $\text{CH}_2\text{Cl}_2$  (150 mL). After 3 h,

the aqueous layer was extracted with  $\text{CH}_2\text{Cl}_2$  (3 X 150 mL). The dried extract ( $\text{MgSO}_4$ ) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 10% EtOAc / Hexanes, to give the aldehyde **5.33** (5.9 g, 22.6 mmol, 98%) as colorless oil. Further elution with 5% MeOH / EtOAc gave recovered auxiliary **2.15** (4.9 g, 22.4 mmol, 97%). **5.33**:  $[\alpha]_{\text{D}}^{23} = +5.93$  ( $c$  0.91,  $\text{CHCl}_3$ ); IR (neat) 2956, 2929, 2837, 1723, 1612, 1513, 1247, 1077, 1034  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  9.75 (t,  $J = 1.5$  Hz, 1H), 7.29 (d,  $J = 8.7$  Hz, 2H), 6.90 (d,  $J = 8.7$  Hz, 2H), 5.14 (d,  $J = 1.2$  Hz, 1H), 4.95 (s, 1H), 4.44 (s, 2H), 3.93 (s, 2H), 3.83 (s, 3H), 2.47 (ddd,  $J = 14.0, 4.0$  and  $1.3$  Hz, 1H), 2.17-2.34 (m, 2H), 2.01-2.11 (m, 2H), 0.98 (d,  $J = 6.6$  Hz, 3H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  202.6, 159.2, 143.9, 130.3, 129.3, 114.1, 113.8, 72.4, 71.7, 55.3, 50.6, 41.0, 26.3, 20.1; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{16}\text{H}_{22}\text{O}_3\text{Na}$  ( $\text{M}+\text{Na}$ ) 285.1467, found 285.1494.

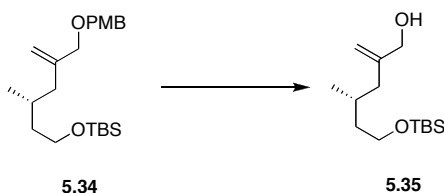


**Aldohol 5.47**: To a stirred solution of aldehyde **5.33** (5.6 g, 21.4 mmol) in  $\text{CH}_2\text{Cl}_2$  (110 mL) at  $-78^\circ\text{C}$  was added DIBAL-H (28.3 mL, 28.3 mmol, 1.0 M in  $\text{CH}_2\text{Cl}_2$ ). After 1 h, the reaction was carefully quenched with methanol (2.0 mL) and poured into aq. sodium potassium tartrate (250 mL, 10% aq.) at rt. The reaction flask was rinsed with an additional portion of  $\text{CH}_2\text{Cl}_2$  (150 mL). After 3 h, the aqueous layer was extracted with  $\text{CH}_2\text{Cl}_2$  (3 X 150 mL). The dried extract

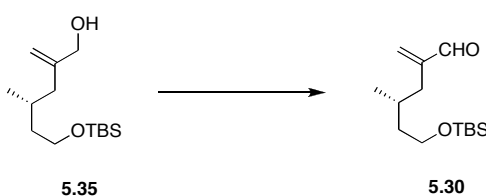


**TBS ether 5.34:** To a stirred solution of alcohol **5.47** (5.5 g, 20.8 mmol) in DMF (50 mL) at rt was sequentially added imidazole (3.4 g, 50.0 mmol) and TBSCl (3.8 g, 25.2 mmol). After 1 h, the reaction was quenched with sat. aq. NH<sub>4</sub>Cl (50 mL) and extracted with Et<sub>2</sub>O (3 x 150 mL). The dried (MgSO<sub>4</sub>) extract was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 5% EtOAc / Hexanes, to give TBS ether **5.34** (7.7 g, 20.3 mmol, 97%) as a colorless oil:  $[\alpha]_D^{23} = -3.27$  (*c* 1.31, CHCl<sub>3</sub>); IR (neat) 2954, 2928, 2856, 1513, 1249, 1094, 835 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.29 (d, *J* = 8.7 Hz, 2H),

6.90 (d,  $J = 8.7$  Hz, 2H), 5.11 (s, 1H), 4.93 (s, 1H), 4.44 (s, 2H), 3.92 (s, 2H), 3.83 (s, 3H), 3.63-3.70 (m, 2H), 2.13 (dd,  $J = 13.8, 6.3$  Hz, 1H), 1.88-1.96 (m, 1H), 1.76-1.86 (m, 1H), 1.57-1.69 (m, 1H), 1.28-1.35 (m, 1H), 0.91 (s, 9H), 0.89 (d,  $J = 6.6$  Hz, 3H), 0.06 (s, 6H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  159.1, 144.7, 130.6, 129.3, 113.8, 112.8, 72.6, 71.6, 61.3, 55.3, 41.4, 39.8, 27.5, 26.0, 19.6, 18.3, -5.3; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{22}\text{H}_{38}\text{O}_3\text{Na}$  ( $\text{M}+\text{Na}$ ) 401.2488, found 401.2489.

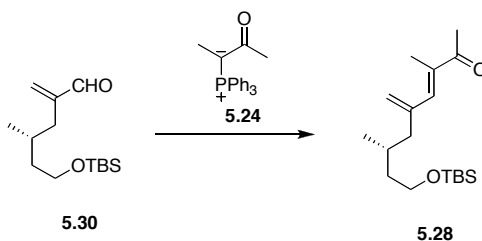


**Alcohol 5.35:** To a stirred solution of TBS ether **5.34** (3.85 g, 10.2 mmol) in  $\text{CH}_2\text{Cl}_2$ /PH 7 buffer (10 : 1, 110 mL) was added DDQ (2.77 g, 12.2 mmol) at rt. After 1 h, the reaction was quenched with sat. aq.  $\text{NaHCO}_3$  (50 mL) and extracted with  $\text{Et}_2\text{O}$  (3 x 100 mL). The dried ( $\text{MgSO}_4$ ) extract was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 8%  $\text{EtOAc}$  / Hexanes, to give a mixture of product **5.35** and 4-methoxybenzaldehyde (3.90 g, 1:1 mole/mole, 9.9 mmol, 97%) as colorless oil. An analytically pure sample was prepared by chromatography over silica gel, eluting with 3%-5%  $\text{EtOAc}$  / Hexanes, for characterization, but the product mixture was used in the subsequent step without complete removal of 4-methoxybenzaldehyde. **5.35:**  $[\alpha]_{\text{D}}^{23} = -6.09$  ( $c$  1.21,  $\text{CHCl}_3$ ); IR (neat) 3338, 2955, 2929, 2858, 1471, 1463, 1255, 1098, 835  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  5.09 (d,  $J = 1.5$  Hz, 1H), 4.89 (s, 1H), 4.07 (d,  $J =$

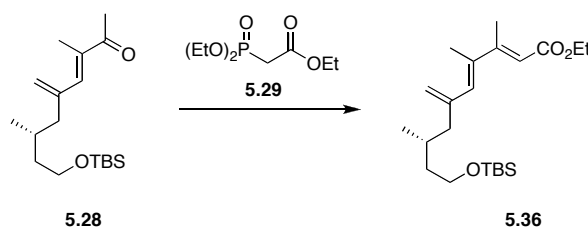


**Aldehyde 5.30:** To a stirred solution of alcohol **5.35** and 4-methoxybenzaldehyde (7.8 g, 1:1 mole/mole, 19.7 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (200 mL) was sequentially added NaHCO<sub>3</sub> (3.0 g, 35.7 mmol) and DMP (10.0 g, 23.7 mmol) at rt. After 1 h, the reaction was quenched with sat. aq. NaHCO<sub>3</sub> (50 mL) and extracted with Et<sub>2</sub>O (3 x 150 mL). The dried (MgSO<sub>4</sub>) extract was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 3% EtOAc / Hexanes, to give aldehyde **5.30** (4.6 g, 17.7 mmol, 90%) as a colorless oil:  $[\alpha]_D^{23} = -8.20$  (*c* 1.21, CHCl<sub>3</sub>); IR (neat) 2956, 2929, 2857, 1698, 1255, 1099, 835 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  9.56 (s, 1H), 6.28 (s, 1H), 6.07 (s, 1H), 3.60-3.73 (m, 2H), 2.28 (dd, *J* = 14.1, 6.9 Hz, 1H), 2.12 (dd, *J* = 13.8, 8.1 Hz, 1H), 1.77-1.86 (m, 1H), 1.53-1.64 (m, 1H), 1.29-1.39 (m, 1H), 0.91 (s, 9H), 0.89 (d, *J* = 6.6 Hz, 3H), 0.07 (s, 6H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  194.7, 149.0, 135.2, 61.1, 39.5, 35.2,

28.4, 25.9, 25.5, 19.4, -5.4; HRMS (EI<sup>+</sup>) calcd. for C<sub>14</sub>H<sub>28</sub>O<sub>2</sub>Si (M) 256.1859, found 256.1861.



**Methyl ketone 5.28:** A solution of aldehyde **5.30** (4.5 g, 17.5 mmol) and ylide **5.24**<sup>4</sup> (10.2 g, 30.7 mmol) in toluene (60 mL) was refluxed in a sealed tube (oil bath 112°C). After 16 h, the reaction mixture was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 5% EtOAc / Hexanes (1% Et<sub>3</sub>N added), to give diene **5.28** (5.0 g, 16.1 mmol, 92%) as a slightly yellow oil:  $[\alpha]_D^{23} = -41.1$  (*c* 0.53, CHCl<sub>3</sub>); IR (neat) 2955, 2928, 2857, 1671, 1255, 1100, 836, 775 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  6.90 (s, 1H), 5.29 (s, 1H), 5.14 (s, 1H), 3.62-3.73 (m, 2H), 2.37 (s, 3H), 2.30 (dd, *J* = 10.2, 4.8 Hz, 1H), 2.05 (dd, *J* = 10.5, 6.3 Hz, 1H), 1.97 (d, *J* = 10.2 Hz, 3H), 1.71-1.78 (m, 1H), 1.59-1.64 (m, 1H), 1.34-1.38 (m, 1H), 0.91 (s, 9H), 0.89 (d, *J* = 6.6 Hz, 3H), 0.07 (s, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  200.3, 143.7, 140.9, 137.8, 118.9, 61.0, 45.1, 39.6, 28.4, 25.9, 25.7, 19.3, 18.3, 13.1, -5.3; HRMS (FAB<sup>+</sup>) calcd. for C<sub>18</sub>H<sub>35</sub>O<sub>2</sub>Si (M+H) 311.2406, found 311.2400.

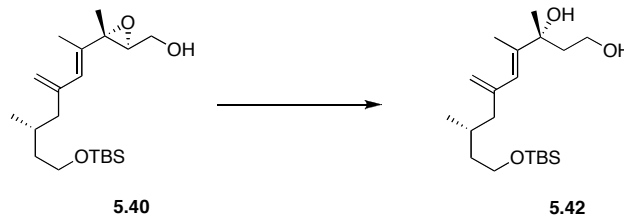


**Triene ester 5.36:** To a stirred slurry of NaH (1.29 g, 32.2 mmol) in DME (50 mL) was added phosphonate **5.29** (6.48 g, 5.74 mL, 28.9 mmol) at rt. After 1 h, a solution of methyl ketone **5.28** (5.00 g, 16.1 mmol) in DME (25 mL) was added. The resulted solution was refluxed for 3 h then quenched with H<sub>2</sub>O (15 mL) and extracted with Et<sub>2</sub>O (3 x 150 mL). The dried extract (MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 2% EtOAc / Hexanes (1% Et<sub>3</sub>N added), to give triene ester **5.36** (4.42 g, 11.6 mmol, 70%) as a colorless oil:  $[\alpha]_D^{23} = -34.7$  (*c* 1.66, CHCl<sub>3</sub>); IR (neat) 2955, 2928, 2857, 1716, 1610, 1255, 1163, 1098, 836, 775 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  6.24 (s, 1H), 5.92 (s, 1H), 5.13 (s, 1H), 4.93 (s, 1H), 4.19 (q, *J* = 7.1 Hz, 2H), 3.61-3.68 (m, 2H), 2.36 (s, 3H), 2.20 (dd, *J* = 13.3, 5.9 Hz, 1H), 1.93-2.00 (m, 4H), 1.53-1.71 (m, 2H), 1.31 (t, *J* = 7.2 Hz, 3H), 0.89 (s, 9H), 0.87 (d, *J* = 6.9 Hz, 3H), 0.05 (s, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  167.4, 156.4, 144.7, 137.8, 132.4, 116.4, 115.9, 61.2, 59.7, 45.7, 39.7, 28.3, 25.9, 19.5, 18.3, 15.8, 15.5, 14.4, -5.3; HRMS (EI<sup>+</sup>) calcd. for C<sub>22</sub>H<sub>40</sub>O<sub>3</sub>Si (M+H) 380.2747, found 380.2732.

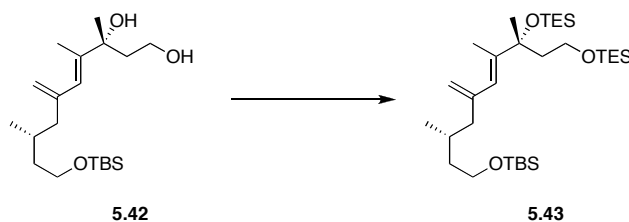
**Allyl alcohol 5.27:** To a stirred solution of triene ester **5.36** (8.61 g, 22.6 mmol) in THF (200 mL) was added DIBAL-H (46 mL, 46.0 mmol, 1 M in toluene) at -78°C. After 1.5 h, the reaction was quenched with MeOH (1.0 mL) and poured into aq. sodium potassium tartrate (350 mL, 10% aq.) at rt. The reaction flask was rinsed with an additional portion of Et<sub>2</sub>O (50 mL). After 3 h, the aqueous layer was extracted with Et<sub>2</sub>O (3 X 200 mL). The dried extract (MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 8% EtOAc / Hexanes, to give allyl alcohol **5.27** (6.20 g, 18.3 mmol, 81%) as a colorless oil:  $[\alpha]_D^{23} = -36.6$  (*c* 1.66, CHCl<sub>3</sub>); IR (neat) 3327, 2954, 2928, 2857, 1471, 1462, 1376, 1255, 1098, 1006, 835, 775 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 5.97 (s, 1H), 5.80 (t, *J* = 6.3 Hz, 1H), 5.08 (s, 1H), 4.87 (s, 1H), 4.34 (d, *J* = 6.4 Hz, 2H), 3.46-3.72 (m, 2H), 2.18 (dd, *J* = 13.6, 6.0 Hz, 1H), 1.99 (d, *J* = 0.8 Hz, 3H), 1.93 (dd, *J* = 13.5, 5.2 Hz, 1H), 1.86 (s, 3H), 1.54-1.73 (m, 2H), 1.28-1.37 (m, 1H), 0.90 (s, 9H), 0.87 (d, *J* = 6.6 Hz, 3H), 0.05 (s, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 145.2, 139.3, 137.6, 128.1, 125.9, 115.4, 61.3, 60.1, 46.0, 39.7, 28.3, 25.9, 19.5, 18.3, 15.6, 14.2, -5.3; HRMS (EI<sup>+</sup>) calcd. for C<sub>20</sub>H<sub>38</sub>O<sub>2</sub>Si (M) 338.2641, found 338.2612.



**Epoxide 5.40:** To a stirred solution of (+)-DIPT (41.5 mg, 0.18 mmol) and 4 Å mol sieves (about 200 mg) in CH<sub>2</sub>Cl<sub>2</sub> (4.0 mL) was sequentially added Ti(O-*i*Pr)<sub>4</sub> (34 mg, 34.6 µL, 0.12 mmol) and TBHP (236 µL, 1.30 mmol, 5.0-6.0 M in decane) at -20°C. After 20 min, the reaction mixture was cooled to -78°C and a pre-cooled solution (-78°C) of allyl alcohol **5.27** (200 mg, 0.59 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (4.0 mL) was added *via* cannula. The resulted solution was warmed up to -50°C. After another 60 min, the reaction was quenched with pH7 phosphate buffer (0.5 mL), filtered over Celite, and extracted with Et<sub>2</sub>O (3 x 20 mL). The dried organic layers (MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 10% EtOAc / Hexanes, to give epoxide **5.40** (167 mg, 0.47 mmol, 80%) as colorless oil:  $[\alpha]_D^{23} = -17.3$  (*c* 1.66, CHCl<sub>3</sub>); IR (neat) 3430, 2954, 2927, 2856, 1471, 1463, 1378, 1255, 1097, 836, 775 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 5.89 (s, 1H), 5.03 (s, 1H), 4.86 (s, 1H), 3.86-3.92 (m, 1H), 3.73-3.81 (m, 1H), 3.62-3.71 (m, 2H), 3.02 (dd, *J* = 10.5, 6.3 Hz, 1H), 2.14 (dd, *J* = 13.5, 5.6 Hz, 1H), 1.91 (dd, *J* = 13.5, 8.4 Hz, 1H), 1.84 (d, *J* = 1.1 Hz, 3H), 1.54-1.66 (m, 2H), 1.45 (s, 3H), 1.27-1.34 (m, 1H), 0.91 (s, 9H), 0.83 (d, *J* = 6.4 Hz, 3H), 0.06 (s, 6H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 144.0, 137.5, 126.2, 115.2, 63.7, 61.3, 45.6, 39.8, 28.2, 26.0, 19.3, 18.3, 16.7, 14.8, -5.3; HRMS (CI<sup>+</sup>) calcd. for C<sub>20</sub>H<sub>39</sub>O<sub>3</sub>Si (M+H) 355.2669, found 355.2666.

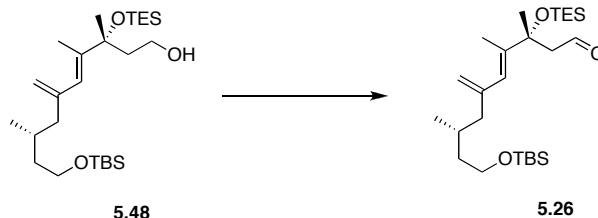


**Diol 5.42:** To a stirred solution of epoxide **5.40** (1.5 g, 4.23 mmol) in THF (30 mL) was added Red-Al (1.5 mL, 9.91 mmol, 65% W/V in toluene) at 0°C. After 1 h, another portion of Red-Al (1.5 mL, 9.91 mmol, 65% W/V in toluene) was added. After another 1.5 h, the reaction was quenched with H<sub>2</sub>O (0.10 mL), and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 X 150 mL). The dried organic layers (MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 6-12% EtOAc / Hexanes (1% Et<sub>3</sub>N added), to give diol **5.42** (1.05 g, 2.96 mmol, 70%) as a colorless oil:  $[\alpha]_D^{23} = -29.4$  (*c* 0.81, CHCl<sub>3</sub>); IR (neat) 3389, 2955, 2928, 2858, 1471, 1462, 1382, 1255, 1097, 836, 775 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  6.10 (s, 1H), 5.04 (d, *J* = 0.9 Hz, 1H), 4.84 (s, 1H), 3.62-3.77 (m, 4H), 3.04 (s, br, 1H), 2.60 (s, br, 1H), 2.16 (dd, *J* = 13.2, 5.1 Hz, 1H), 1.88-1.96 (m, 3H), 1.79 (d, *J* = 0.9 Hz, 3H), 1.55-1.70 (m, 2H), 1.37 (s, 1H), 1.27-1.34 (m, 1H), 0.91 (s, 9H), 0.86 (d, *J* = 6.5 Hz, 3H), 0.07 (s, 6H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  145.1, 141.7, 125.1, 114.4, 77.1, 61.6, 60.4, 46.1, 40.1, 39.7, 28.6, 28.2, 26.0, 19.6, 18.4, 15.1, -5.2; HRMS (ES<sup>+</sup>) calcd. for C<sub>20</sub>H<sub>40</sub>O<sub>3</sub>SiNa (M+Na) 379.2644, found 379.2643



**TES ether 5.43:** To a stirred solution of diol **5.42** (470 mg, 1.32 mmol) in DCM / TEA (6 mL, 1:1) was added freshly distilled TESOTf (1.05 g, 0.89 mL, 3.96 mmol) at  $-78^{\circ}\text{C}$ . After 30 min, the reaction was quenched with sat. aq.  $\text{NaHCO}_3$  (1 mL) and extracted with ether (3 X 20 mL). The dried extract ( $\text{MgSO}_4$ ) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 2-5% EtOAc / Hexanes, to give **5.43** (732 mg, 1.25 mmol, 95%) as a colorless oil:  $[\alpha]_{\text{D}}^{23} = -21.3$  ( $c$  1.37,  $\text{CHCl}_3$ ); IR (neat) 2954, 2929, 2876, 1460, 1254, 1093, 1007, 835, 741  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  5.88 (s, 1H), 4.99 (s, 1H), 4.80 (s, 1H), 3.61-3.71 (m, 3H), 3.47 (dt,  $J = 15.6, 5.4$  Hz, 1H), 2.15 (dd,  $J = 13.5, 5.4$  Hz, 1H), 1.80-1.99 (m, 5H), 1.77 (d,  $J = 0.9$  Hz, 3H), 1.54-1.66 (m, 1H), 0.88-1.00 (m, 27H), 0.84 (d,  $J = 6.3$  Hz, 3H), 0.55-0.66 (m, 12H), 0.063 (s, 6H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  145.2, 142.0, 124.6, 114.1, 77.2, 61.7, 59.3, 46.2, 44.5, 40.0, 28.7, 26, 19.6, 18.4, 14.7, 7.2, 7.0, 6.9, 6.8, 4.4, -5.3; HRMS ( $\text{EI}^+$ ) calcd. for  $\text{C}_{32}\text{H}_{68}\text{O}_3\text{Si}_3$  ( $\text{M}^+$ ) 584.4476, found 584.4500.

**Aldohol 5.48:** TES ether **5.43** (300 mg, 0.51 mmol) was dissolved in a stirred solution of HOAc / THF / H<sub>2</sub>O (8 mL, 8:8:1) at 0°C. After 1.5 h, the reaction was then quenched with solid NaHCO<sub>3</sub> and extracted with ether (3 X 20 mL). The dried extract (MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 10% EtOAc / Hexanes, to give alcohol **5.48** (241 mg, 0.51 mmol, 99%) as a colorless oil:  $[\alpha]_D^{23} = -15.5$  (*c* 0.64, CHCl<sub>3</sub>); IR (neat) 3437, 2954, 2928, 2876, 1461, 1254, 1099, 1008, 835 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.02 (s, 1H), 5.04 (s, 1H), 4.85 (s, 1H), 3.63-3.73(m, 4H), 2.79 (t, *J* = 5.6 Hz, 1H), 2.17 (dd, *J* = 13.6, 5.6 Hz, 1H), 1.88-1.97 (m, 3H), 1.74-1.83 (m, 1H), 1.79 (d, *J* = 1.2 Hz, 3H), 1.59-1.68 (m, 1H), 1.47 (s, 3H), 1.28-1.40 (m, 1H), 0.99-1.03 (t, *J* = 7.6 Hz, 9H), 0.92 (s, 9H), 0.88 (d, *J* = 6.4 Hz, 3H), 0.69 (q, *J* = 7.2 Hz, 6H), 0.078 (s, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  145.0, 141.4, 125.6, 114.6, 80.0, 61.6, 60.1, 46.1, 42.8, 39.9, 29.7, 28.6, 27.6, 26.0, 19.5, 18.4, 14.9, 7.2, 6.9, 6.8, -5.3; HRMS (EI<sup>+</sup>) calcd. for C<sub>26</sub>H<sub>54</sub>O<sub>3</sub>Si<sub>2</sub> (M<sup>+</sup>) 470.3611, found 470.3604.

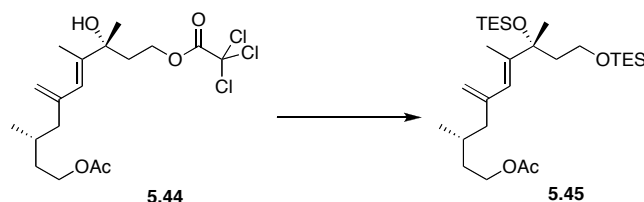


**Aldehyde 5.26:** To a stirred solution of alcohol **5.48** (1.29 g, 2.73 mmol) in DCM (40 mL, 1:1) was sequentially added DMP (2.17 g, 5.12 mmol) and  $\text{NaHCO}_3$  (1.68 g, 20.0 mmol) at rt. After 30 min, the reaction was quenched with sat. aq.  $\text{NaHCO}_3$  (10 mL) and extracted with ether (3 X 40 mL). The dried extract ( $\text{MgSO}_4$ ) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 5% EtOAc / Hexanes, to give aldehyde **5.46** (1.17 g, 2.49 mmol, 91%) as a colorless oil:  $[\alpha]_D^{23} = -12.5$  (*c* 0.56,  $\text{CHCl}_3$ ); IR (neat) 2955, 2929, 2877, 1725, 1255, 1099, 1007, 836, 726  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  9.69 (t,  $J = 3.3$  Hz, 1H), 6.03 (s, 1H), 5.03 (s, 1H), 4.83 (s, 1H), 3.61-3.68(m, 2H), 2.64 (dd,  $J = 15.0, 3.0$  Hz, 1H), 2.45 (dd,  $J = 15.0, 3.0$  Hz, 1H), 2.15 (dd,  $J = 13.5, 6.0$  Hz, 1H), 1.85-1.92 (m, 1H), 1.83 (d,  $J = 1.2$  Hz, 3H), 1.54-1.64 (m, 1H), 1.49 (s, 3H), 1.25-1.35 (m, 2H), 0.95-1.00 (t,  $J = 7.7$  Hz, 9H), 0.91 (s, 9H), 0.85 (d,  $J = 6.5$  Hz, 3H), 0.65 (q,  $J = 7.8$  Hz, 6H), 0.061 (s, 6H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  203.1, 144.7, 141.0, 126.0, 114.9, 76.9, 61.5, 54.2, 46.0, 39.9, 28.5, 27.8, 26.0, 19.5, 18.3, 14.7, 7.1, 6.7, -5.3; HRMS ( $\text{EI}^+$ ) calcd. for  $\text{C}_{26}\text{H}_{52}\text{O}_3\text{Si}_3$  ( $\text{M}^+$ ) 468.3455, found 468.3448

To a stirred solution of crude ester (3.30 g) in CH<sub>2</sub>Cl<sub>2</sub> / EtOH (1:1, 50 mL) was added CSA (2.31 g, 9.88 mmol) at 0°C. After 1.5 h, the reaction was quenched with sat. aq. NaHCO<sub>3</sub> (10 mL) and extracted with ether (3 X 40 mL). The dried extract (MgSO<sub>4</sub>) was concentrated *in vacuo* to give crude diol (2.02 g), which was used in the next step without further purification.

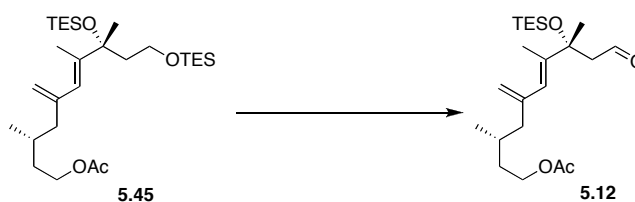
To a stirred solution of crude diol (2.02 g) in CH<sub>2</sub>Cl<sub>2</sub> (50 mL) was sequentially added pyridine (1.53 g, 1.57 mL, 19.5 mmol) and Ac<sub>2</sub>O (0.99 g, 0.92 mL, 9.73 mmol). After 1.5 h, the reaction was quenched with sat. aq. NH<sub>4</sub>Cl (10 mL) and extracted with ether (3 X 40 mL). The dried extract (MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 10% EtOAc / Hexanes, to give **5.44** (1.90 g, 4.42 mmol, 75% over 3 steps) as a colorless oil:  $[\alpha]_D^{23} = -14.1$  (*c* 1.16, CHCl<sub>3</sub>); IR (neat) 3481, 2962, 2928, 1766, 1739, 1720, 1458, 1368, 1247, 828, 682 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  6.01

(s, 1H), 5.04 (s, 1H), 4.85 (s, 1H), 4.47-4.36 (m, 2H), 4.15-4.06 (m, 2H), 2.12-2.02 (m, 3H), 2.03 (s, 3H), 1.97-1.89 (m, 3H), 1.83 (s, 3H), 1.70-1.60 (m, 2H), 1.50-1.38 (m, 1H), 1.41 (s, 3H), 0.89 (d,  $J = 6.3$  Hz, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  171.2, 161.9, 144.1, 141.4, 124.9, 115.3, 74.6, 66.5, 62.3, 45.6, 37.8, 35.2, 28.7, 28.3, 21.0, 19.4, 14.9; HRMS ( $\text{EI}^+$ ) calcd. for  $\text{C}_{18}\text{H}_{27}\text{O}_5\text{Cl}_3$  ( $\text{M}^+$ ) 428.0924, found 428.0932.



**TES ether 5.45:** To a stirred solution of alcohol **5.44** (1.90 g, 4.4 mmol) in  $\text{CH}_2\text{Cl}_2$  / EtOH (1:1, 50 mL) was added  $\text{NH}_3 \cdot \text{H}_2\text{O}$  (15 mL) at rt. After 1 h, the reaction was quenched with sat. aq.  $\text{NH}_4\text{Cl}$  (15 mL) and extracted with ether (3 X 40 mL). The dried extract ( $\text{MgSO}_4$ ) was concentrated *in vacuo* to afford crude diol (1.55 g), which which was used in the next step without further purification.

To a stirred solution of crude diol (1.55 g) in  $\text{CH}_2\text{Cl}_2$  /  $\text{Et}_3\text{N}$  (1:1, 30 mL) was added freshly distilled  $\text{TESOTf}$  (3.52 g, 3.01 mL, 13.1 mmol) at  $-78^\circ\text{C}$ . After 20 min, the reaction was quenched with sat. aq.  $\text{NaHCO}_3$  (10 mL) and extracted with ether (3 X 40 mL). The dried extract ( $\text{MgSO}_4$ ) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 2% EtOAc / Hexanes, to give TES ether **5.45** (2.12 g, 4.09 mmol, 93% over 2 steps) as a colorless oil:  $[\alpha]_{\text{D}}^{23} = -18.4$  ( $c$  1.11,  $\text{CHCl}_3$ ); IR (neat) 29554, 2912, 2876, 1748, 1458, 1238,



To a stirred solution of crude alcohol (2.0 g) in CH<sub>2</sub>Cl<sub>2</sub> (20 mL) was added sequentially solid NaHCO<sub>3</sub> (1.0 g, 11.9 mmol) and DMP (2.16 g, 5.09 mmol) at rt. After 1 h, the reaction was quenched with sat. aq. NaHCO<sub>3</sub> (15 mL) and extracted with ether (3 X 40 mL). The dried extract (MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 5% EtOAc / Hexanes,



to give aldehyde **5.12** (1.36 g, 3.43 mmol, 74% over 2 steps) as a colorless oil:  $[\alpha]_D^{23} = -11.6$  ( $c$  1.83,  $\text{CHCl}_3$ ); IR (neat) 2957, 2877, 1741, 1724, 1458, 1368, 1239, 1050, 1017, 743  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  9.66 (t,  $J = 3.0$  Hz, 1H), 6.02 (s, 1H), 5.03 (s, 1H), 4.83 (s, 1H), 4.14-4.04 (m, 2H), 2.65 (dd,  $J = 15.1$ , 2.9 Hz, 1H), 2.46 (dd,  $J = 15.1$ , 3.1 Hz, 1H), 2.14 (dd,  $J = 13.6$ , 5.8 Hz, 1H), 2.04 (s, 3H), 1.92 (dd,  $J = 13.5$ , 7.8 Hz, 1H), 1.82 (s, 3H), 1.70 -1.60 (m, 2H), 1.48 (s, 3H), 1.45-1.39 (m, 1H), 1.00-0.94 (m, 9H), 0.87 (d,  $J = 6.5$  Hz, 3H), 0.68-0.56 (m, 6H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  203.0, 171.2, 144.3, 141.2, 125.7, 115.1, 62.8, 54.0, 45.8, 35.3, 28.7, 27.9, 20.9, 19.2, 14.7, 7.1, 6.7, 6.6, 5.8; HRMS ( $\text{EI}^+$ ) calcd. for  $\text{C}_{22}\text{H}_{40}\text{O}_4\text{Si}$  ( $\text{M}^+$ ) 396.2696, found 396.2678.

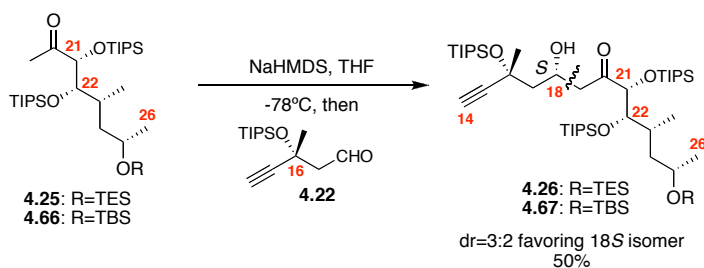
- 
1. Boulet, S. L.; Paquette, L. A. *Synthesis* **2002**, 895.
  2. Titration of Grignard reagent: To a stirred solution of menthol (15.6 mg, 0.1 mmol) and 1,10-phenanthroline (2 mg) in THF (0.5 mL) was added prepared Grignard reagent solution until a burgundy color persists. The concentration was calculated using the formula:  
$$[\text{RMgX}] = 0.1 \text{ mmol} / \text{volume of added RMgX in mL}$$
*For the references, see:* (a) Lin, H, -S; Paquette, L. A. *Synth. Comm.* **1994**, 24, 2503. (b) Watson, S. C.; Eastham, J. F. *J. Organomet. Chem.* **1967**, 9, 165.
  3. Vanderwalle, M.; Van der Eycken, J.; Oppolzer, W.; Vulliod, C. *Tetrahedron* **1986**, 42, 4035.
  4. Aitken, R. A.; Atherton, J. I. *J. Chem. Soc., Perkin Trans. I* **1994**, 1281.

## CHAPTER 6. GAME OF PROTECTING GROUPS AND THE STUDIES OF THE KEY ALDOL COUPLING

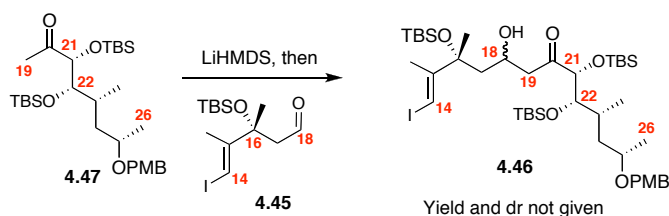
### 6.1 The Chelation-Controlled Aldol Reaction

After the success in the synthesis of the key diene subunits, our priority shifted to the aldol reaction to install the C<sub>18</sub> stereocenter. There have been several attempts from Chakraborty,<sup>1a</sup> Pattenden,<sup>1b</sup> and Kobayashi<sup>1c</sup> to furnish the C<sub>18</sub> stereochemistry utilizing the aldol coupling (Scheme 6.1). Unfortunately, the aldol reaction between an aldehyde and the C<sub>21</sub> TBS- or TIPS-protected methyl ketone led to low yield and poor diastereoselectivity.

Chakraborty and Kobayashi:

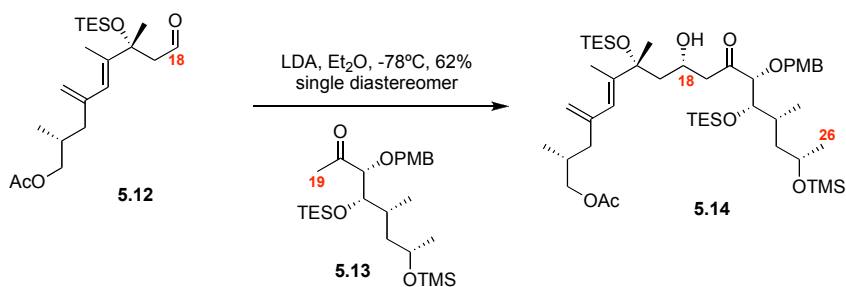


Pattenden:



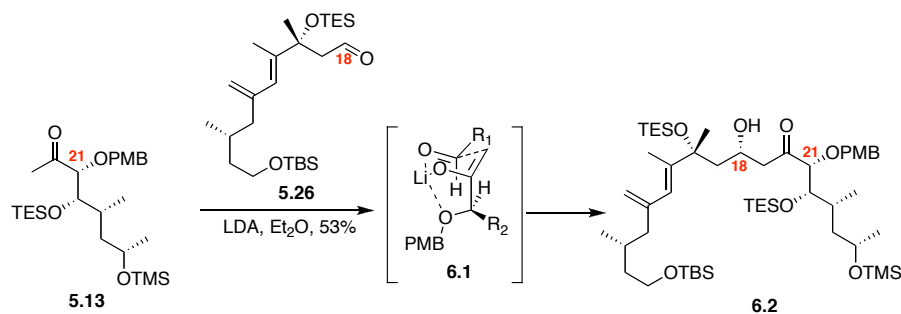
**Scheme 6.1.** Precedents of Aldol Coupling

Our group have reported the first chelation-controlled aldol reaction to give C<sub>18</sub> stereocenter diastereoselectively (Scheme 6.2).<sup>2</sup> The aldol coupling between aldehyde **5.12** and C<sub>21</sub> PMB-protected methyl ketone **5.13** afforded 18*S* aldol adduct as a single diastereomer. This strategy has been used in Fürstner's recent synthesis of amphidinolide G, H and B.<sup>3</sup>



**Scheme 6.2.** Carter's Chelation-Controlled Aldol Coupling

Equipped with the knowledge gained from our successful experience, we next investigated the key aldol reaction between the diene motif and the C<sub>21</sub> PMB-protected methyl ketone **5.13**<sup>2</sup> (Scheme 6.3). We initially chose to explore our proposed chemistry on the C<sub>9</sub> TBS protected diene aldehyde **5.26**. Under our typical conditions (LDA / Et<sub>2</sub>O, -78°C), the aldol coupling proceeded smoothly to yield 18*S* adduct **6.2** in 53% yield as a single diastereomer. Since the PMB protected hydroxyl group is known for its ability to participate in chelation-controlled processes,<sup>4</sup> one possible explanation for the excellent stereoselectivity is depicted in transition state **6.1**. The chelation effect of the PMB protected C<sub>21</sub> hydroxyl group would result in a transition state like **6.1**, which in turn should lead to the 18*S* isomer **6.2** in good diastereoselectivity.

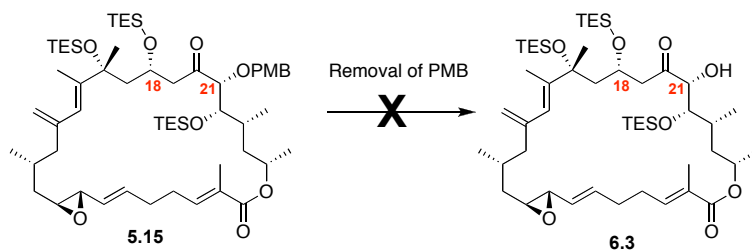


**Scheme 6.3.** Aldol Reaction Between Methyl Ketone **5.13** and Aldehyde **5.26**

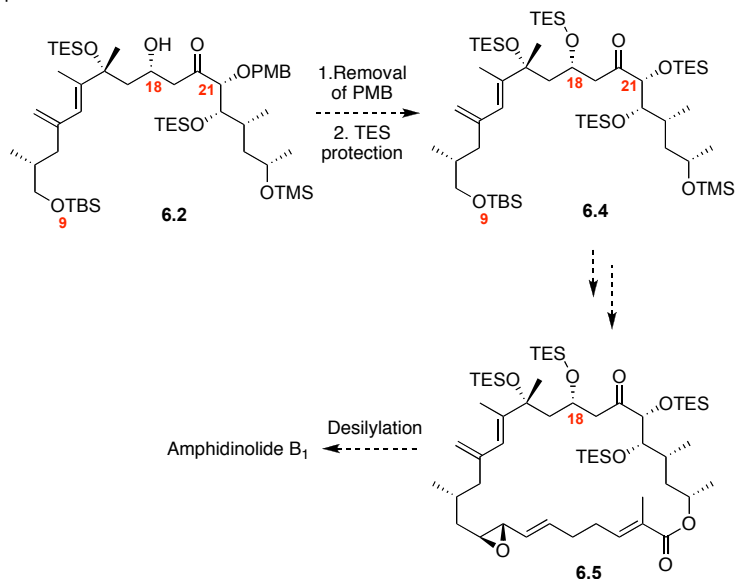
## 6.2 Attempts to Remove the PMB Protecting Group

With adol adduct **6.2** in hand, we next focused on the deprotection of PMB group. Our previous work<sup>2b</sup> indicated that general PMB deprotection conditions (DDQ, CAN, *etc.*) would lead to the decomposition of macrocycle **5.15**. Under the suspicion that the presence of an allylic epoxide moiety might be problematic, we decided to investigate the removal of the PMB group found in compound **6.2**. Cleavage of the PMB protecting group followed by C<sub>21</sub> TES protection should afford TES ether **6.4**. After converting **6.4** to the TES protected version of amphidinolide B, the TES groups could be deprotected under mild conditions in the presence of the labile allylic epoxide moiety.

Previous work:

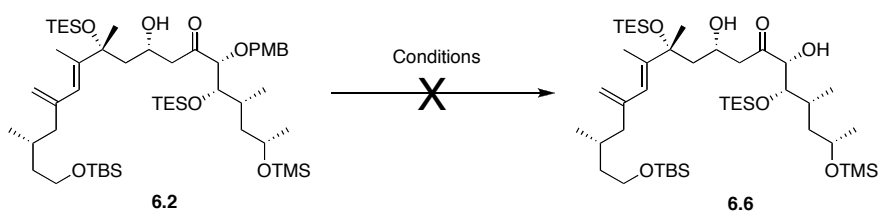


Proposed work:



**Scheme 6.4.** Modified Deprotection Strategy

Our previous difficulties with deprotection<sup>2b</sup> made us mindful in selecting the deprotection conditions we would employ for this transformation. Generally, PMB protecting groups can be removed under oxidative conditions (DDQ, CAN, NBS, *etc.*);<sup>5</sup> reductive conditions (NaBH<sub>3</sub>(CN) / BF<sub>3</sub>•OEt<sub>2</sub>);<sup>6</sup> acidic conditions (TFA, HCl, *etc.*);<sup>7</sup> or Lewis acidic conditions (TMSI, MgBr<sub>2</sub>•Et<sub>2</sub>O, *etc.*).<sup>8</sup> Due to the diene motif and the carbonyl group found in adol adduct **6.2**, only oxidative conditions would be reasonable for this substrate. Consequently, we chose to



Entry	Conditions	Results
1	DDQ, pH 7 buffer, CH <sub>2</sub> Cl <sub>2</sub> , rt	Decomposition
2	DDQ, pH 9 buffer, CH <sub>2</sub> Cl <sub>2</sub> , rt	Decomposition
3	CAN, pH 7 buffer, rt	Decomposition
4	DDQ, aq NaHCO <sub>3</sub> , CH <sub>2</sub> Cl <sub>2</sub> , 0°C	No Reaction
5	MgBr <sub>2</sub> •Et <sub>2</sub> O, THF, 0°C	Decomposition

### Table 6.1. Attempts to Remove PMB Group

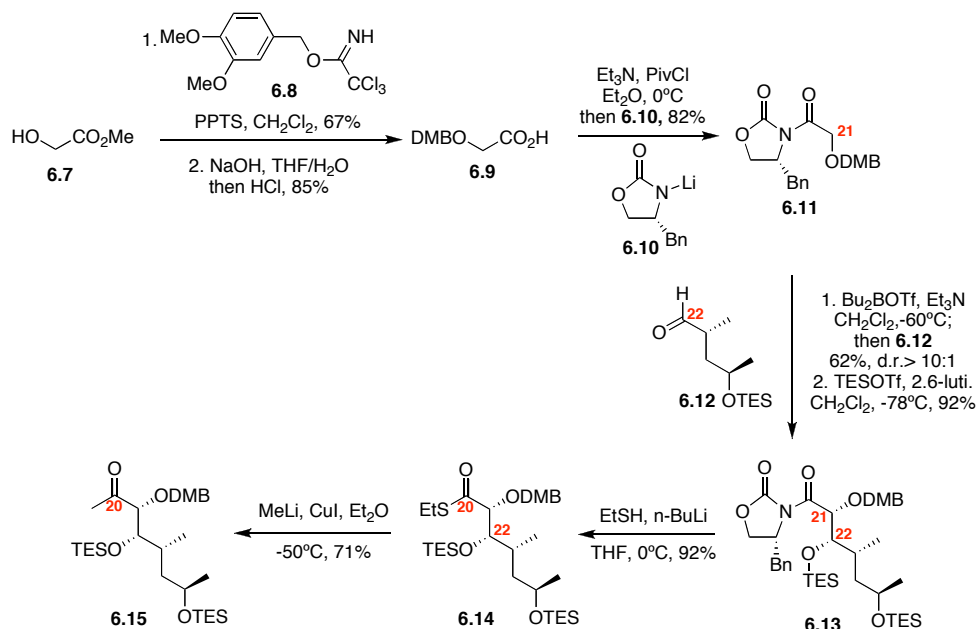
### 6.3 Studies of DMB Group as Protecting Group

#### 6.3.1 Synthesis of C<sub>21</sub> DMB Protected Methyl Ketone 6.13

Given that PMB group could not be removed without the decomposition of the aldol adduct **6.2**, an alternate plan would involve implementing different protecting groups on the C<sub>21</sub> hydroxyl group. First, we investigated the 3,4-dimethoxybenzyl (DMB) group, a moiety that is structurally similar to the PMB group.<sup>9</sup> Utilization of the DMB group would provide the desired chelation control in the aldol coupling. More importantly, the DMB group is much more reactive toward the oxidizing reagents due to its lower oxidation potential than that of PMB group ( $E_{1/2}$ =1.45 V and 1.78 V, respectively).<sup>10</sup> DMB groups have been reported to be successfully removed from an alcohol with DDQ in the presence of PMB group with 98% selectivity.<sup>11</sup> Due to its greater reactivity, we could potentially cleave the DMB group at lower temperature allowing the diene moiety to remain intact.

Our next goal was focused on the synthesis of C<sub>21</sub> DMB-protected methyl ketone (Scheme 6.5). Oxazolidinone **6.11** was prepared via the known procedure described by Roush and co-workers.<sup>12</sup> Boron-mediated aldol reaction<sup>13</sup> between the previously made aldehyde **6.12** and compound **6.11** gave the C<sub>21</sub>-C<sub>23</sub> *syn, syn* adduct in 80% yield with good diastereoselectivity (d.r.>10:1). Subsequent silylation at the C<sub>22</sub> hydroxyl group, the conversion of oxazolidinone **6.13** to the

corresponding thioester **6.14** and cuprate addition of the desired methyl group gave the requisite methyl ketone **6.15**.<sup>14</sup>

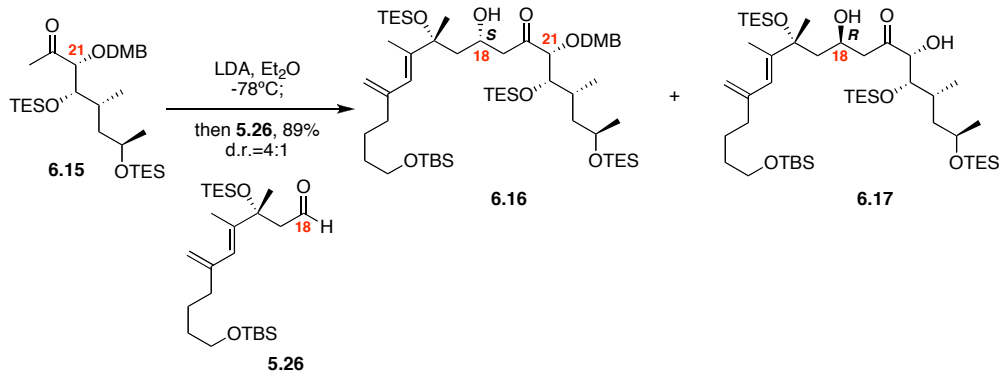


**Scheme 6.5.** Synthesis of Methyl Ketone **6.15**

### 6.3.2 Aldol Coupling Between Methyl ketone **6.15** and Aldehyde **5.26**

With both aldol precursors in hand, the next target became the coupling between aldehyde **5.26** and methyl ketone **6.15** (Scheme 6.6). Using our typical protocol (LDA / Et<sub>2</sub>O, -78°C),<sup>2</sup> the use of DMB-protected methyl ketone led to better overall yield, albeit in moderate diastereoselectivity (4:1, favoring the 18*S* isomer). Fortunately, the two isomers were easily separated via conventional silica gel chromatography.

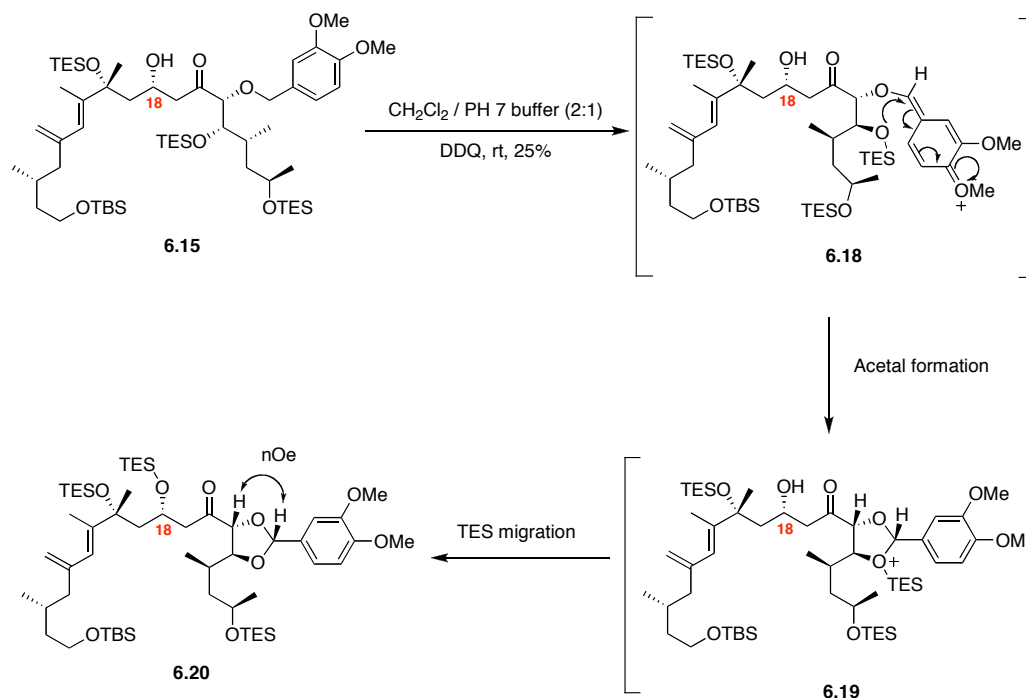




**Scheme 6.6.** Aldol Coupling Between Methyl Ketone **6.15** and Aldehyde **5.26**

### 6.3.3 Attempts to Remove DMB Group

After obtaining the adol adduct **6.16**, our next priority became the deprotection of the DMB group (Scheme 6.7). When DMB ether **6.16** was treated with DDQ in DCM / pH 7 buffer at 0°C or room temperature, an unexpected compound was isolated as a single isomer in 25% yield. The undesired product appeared to be acetal **6.20**, which would arise from the formation of an acetal and the migration of the TES group. The stereochemistry was determined by nOe analysis. Attempts to improve the yield by using anhydrous CH<sub>2</sub>Cl<sub>2</sub> as solvent, lowering the reaction temperature, or utilizing alternative oxidizing reagents (CAN or 2,3,5,6-tetrachlorobenzoquinone<sup>15</sup>) resulted in comparable yields or no reaction.



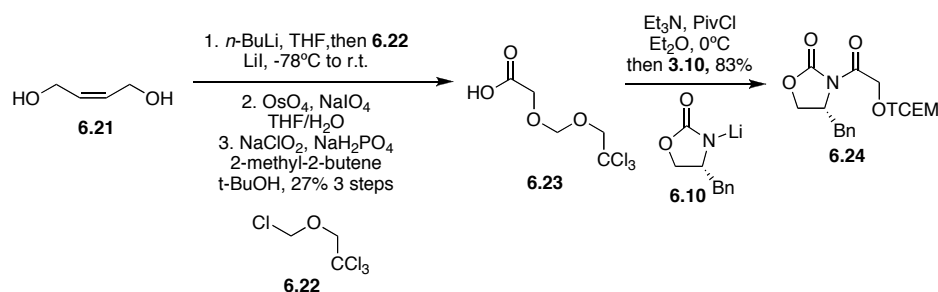
**Scheme 6.7.** Attempt to Remove DMB Group

#### 6.4 Studies of Other Chelation Protecting Groups

Our synthetic efforts demonstrated that a PMB-type protecting group provided satisfactory chelation control during the aldol coupling. Unfortunately, conditions required to remove these groups were not amendable to the diene substrate. A reasonable solution to this problem would be to use another C<sub>21</sub> chelation protecting group, the removal of which would not require harsh conditions. First, we studied 2,2,2-trichloroethoxymethyl (TCEM) group.<sup>16</sup> This MEM group derivative should generate the desired chelation control and could be removed under much milder conditions.<sup>16</sup>

### 6.4.1 Synthesis of Oxazolidinone 6.20

Our new strategy required the synthesis of oxazolidinone **6.24** (Scheme 6.8). Starting from diol **6.21**, acid **6.23** was produced in three steps in moderate yield. After converting acid **6.23** to the corresponding *t*-butyl ester, the addition of anion **6.10** yielded the desired oxazolidinone **6.24**.

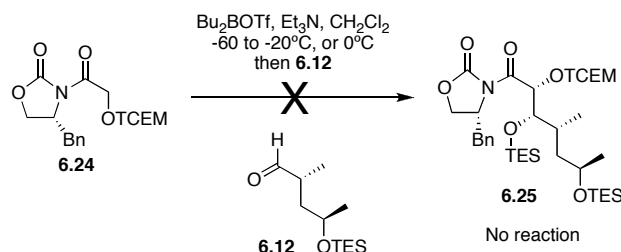


**Scheme 6.8.** Synthesis of Oxazolidinone **6.24**

### 6.4.2 Aldol Coupling Between Oxazolidinone 6.24 and Aldehyde 6.12

With oxazolidinone **6.24** in hand, we explored the aldol coupling between compound **6.24** and aldehyde **6.12** (Scheme 6.9). Unfortunately, the aldol coupling was unsuccessful due to the steric hindrance introduced by the TCEM group. Attempts to accelerate the reaction by raising the temperature and increasing the concentration of the reaction mixture proved unsuccessful. Once we realized the inefficiency of the TCEM group, we investigated several other chelation protecting groups including benzyl group,<sup>17</sup> THP group,<sup>18</sup> acetonide<sup>19</sup> and DMB acetal groups. Again, our efforts were thwarted either by poor diastereoselectivity and

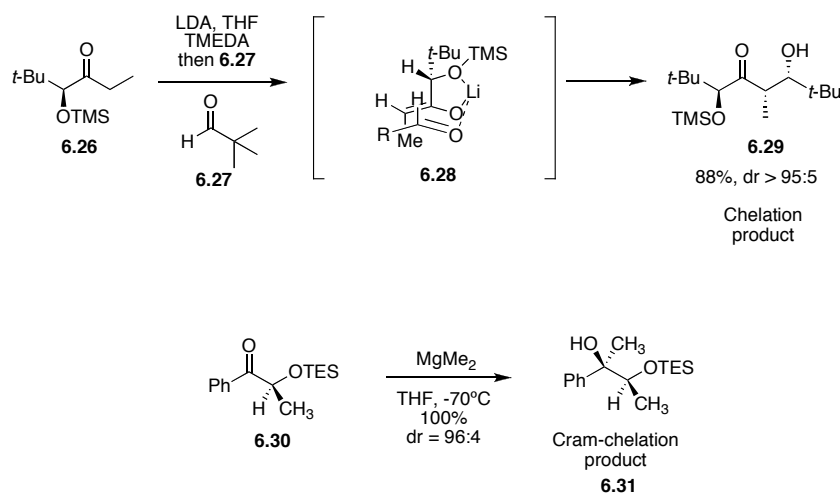
low conversion during the aldol coupling, or by the inability to remove the protecting groups.



**Scheme 6.9.** Aldol Coupling between Oxazolidinone **6.24** and Aldehyde **6.12**

## 6.5 Studies of Silyl Groups as $\text{C}_{21}$ Protecting Group

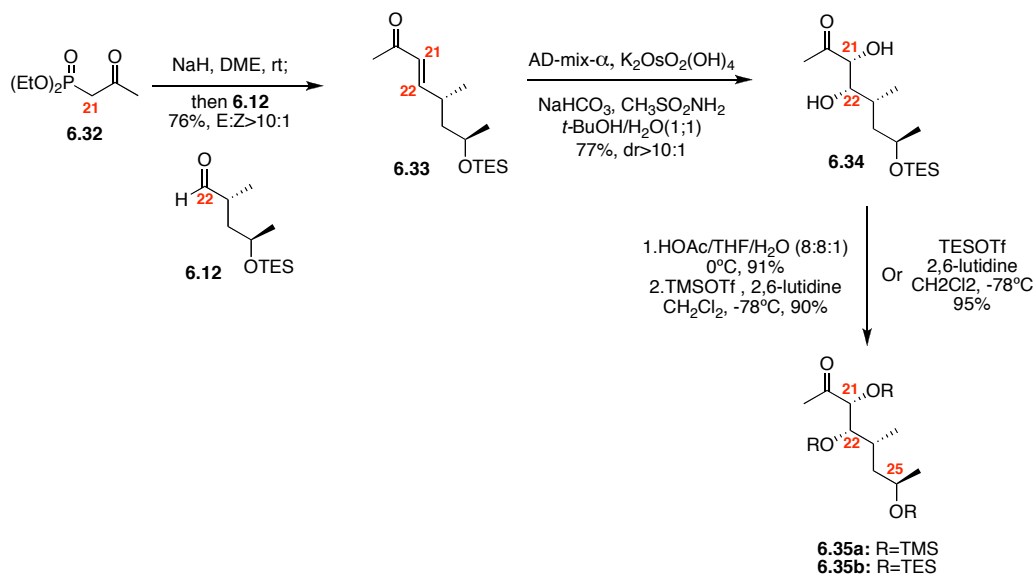
Our strategies employing alkoxy groups proved problematic due to the incompatibility of the diene moiety with the deprotection conditions. In order to circumvent this problem, we shifted our focus on the silyl protecting groups. Conventional wisdom states that hindered silyl protecting groups prevent chelation with most metal ions<sup>20</sup> due to the decreased basicity of O atom,<sup>21</sup> however, the research from Heathcock and Frye groups demonstrated that chelation control is possible for small silyl groups such as TMS or TES (Scheme 6.10).<sup>22</sup> More evidence to support this concept was obtained when a X-ray structure of a dimeric lithium ketone enolate-lithium diisopropylamide complex, where the coordination between TBS ether oxygen and lithium ion was observed.<sup>23</sup> Based on this information, we decided to investigate the use of relatively small silyl groups (TMS and TES) as possible  $\text{C}_{21}$  protecting groups.



**Scheme 6.10.** Examples of Chelation Control of Silyl Groups

### 6.5.1 Synthesis of Methyl Ketone **6.35a/b**

With a modified strategy in hand, we sought to synthesize the  $\text{C}_{21}$  TMS and TES protected methyl ketones (Scheme 6.11). Starting with previously synthesized aldehyde **6.12**,<sup>2a</sup> a Horner–Wadsworth–Emmons olefination and Sharpless dihydroxylation yielded the diol **6.34**. Consecutive de-silylation and the tri-silylation yielded the TMS-protected methyl ketone **6.35a**. Alternatively, TES-protected methyl ketone **6.35b** was generated via the di-silylation of diol **6.34**.

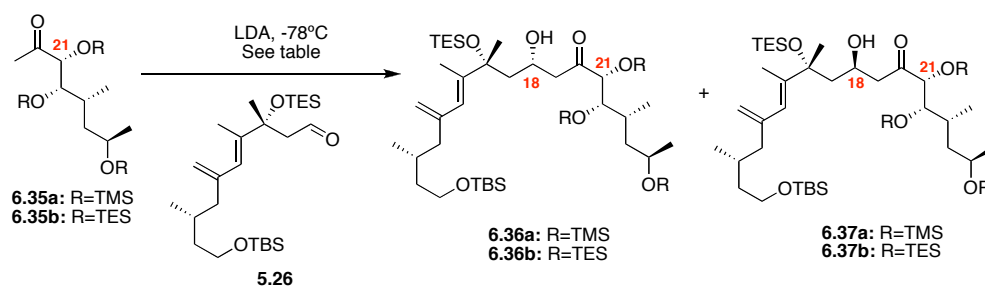


**Scheme 6.11.** Synthesis of Methyl Ketone **6.35a/b**

### 6.5.2 Aldol Coupling between Methyl ketone **6.35a/b** and Aldehyde **5.26**

After obtaining both methyl ketones with the undesired stereochemistry at C<sub>25</sub>, we studied the key aldol reaction on the model system (Table 6.2). Treatment of ketone **6.35a** under our standard LDA / THF conditions resulted in low conversion (~15%) and poor diastereoselectivity favoring the 18*S* stereochemistry [approximately 1.1:1 dr (**6.36a**:**6.37a**)]. Suspecting that aggregation of lithium enolate decreased its reactivity, we added TMEDA to the reaction mixture with the intention to break the aggregation.<sup>24</sup> We were pleased to find that the addition of TMEDA led to dramatic rate acceleration and complete conversion, although the diastereoselectivity was still poor [1.1:1 dr (**6.36a**:**6.37a**)]. Fortunately, the two

diastereomers were easily separable by conventional silica chromatography. When the TES protected methyl ketone was treated under the same conditions, almost identical results were obtained. Switching the solvent to ether resulted in poor conversion with similar diastereoselectivity. The desired chelation-control was not observed under all the conditions we investigated.



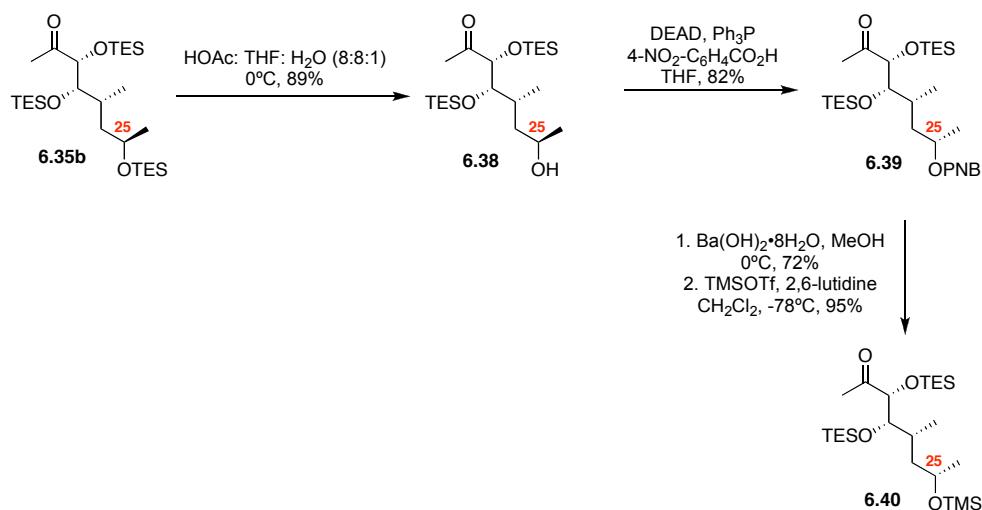
R	Conditions	Yield	dr ( <b>6.36</b> : <b>6.37</b> )
R = TMS	THF	~10% (15% conversion)	1.1:1
	TMEDA, THF	65% (100% conversion)	1.1:1
R = TES	TMEDA, THF	64% (100% conversion)	1.1:1
	TMEDA, ether	<10% (30% conversion)	1.5:1

**Table 6.2.** Aldol Coupling between Methyl ketone **6.35a/b** and Aldehyde **5.26**

### 6.5.3 Synthesis of Methyl Ketone **6.40**

Equipped with the knowledge gained from our previous study, we shifted our focus to the authentic substrate. To avoid the potential problems associated with the lability of TMS group, we chose to use TES to protect the C<sub>21</sub> hydroxyl group. The desired C<sub>25</sub> stereocenter was generated through the sequence shown in

Scheme 6.12. Selective C<sub>25</sub> TES deprotection yielded the free alcohol **6.38**. Subsequent Mitsunobu inversion of the alcohol, followed by saponification of PNB ester and C<sub>25</sub> TMS protection revealed the desired methyl ketone **6.40**.

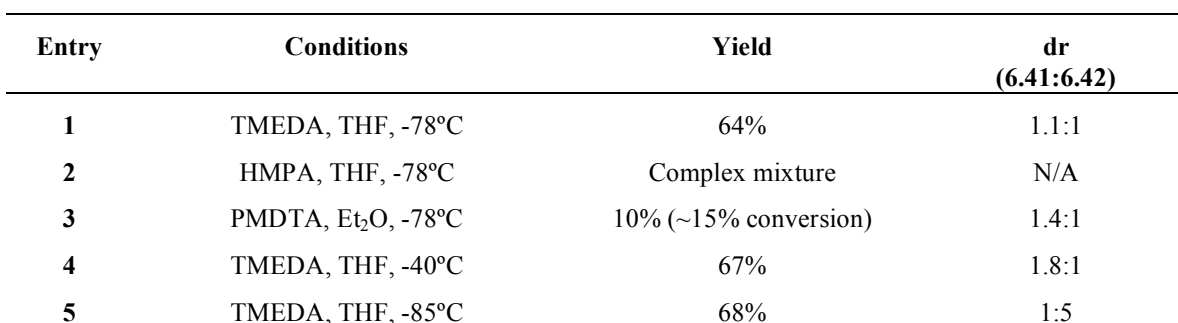


**Scheme 6.12.** Synthesis of Methyl Ketone **6.40**

#### 6.5.4 Aldol Coupling between Methyl Ketone **6.40** and Aldehyde **5.26**

With the ability to efficiently prepare methyl ketone **6.40**, our next target was the key aldol coupling (Table 6.3). Warming of the reaction to  $-40^\circ\text{C}$  led to improved diastereoselectivity toward 18*S* diastereomer [1.8:1 dr (**6.41**:**6.42**)] in reasonable yield (67% overall). We were gratified to find that when the reaction was cooled to  $-85^\circ\text{C}$ , the diastereoselectivity of the 18*R* diastereomer was improved significantly to 1:5 (**6.41**:**6.42**). It should be noted that the addition of other ligands such as HMPA<sup>25</sup> or PMDTA<sup>26</sup> to the aldol reaction resulted in



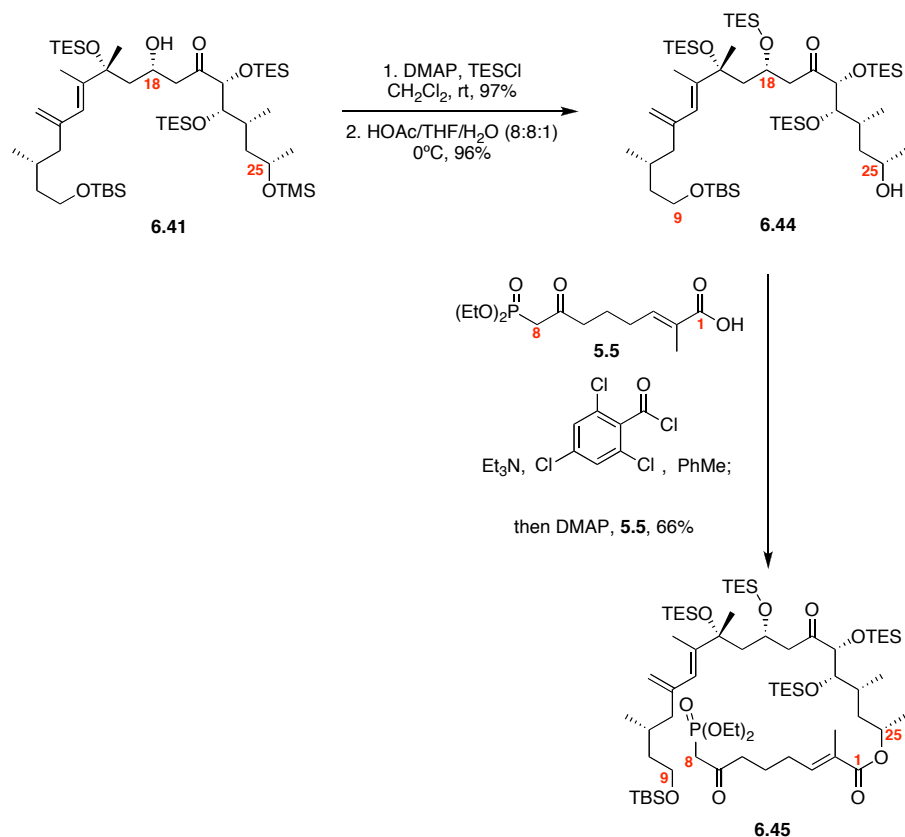


**Table 6.3.** Aldol Coupling Between Methyl Ketone **6.40** and Aldehyde **5.26**



### 6.5.5 Synthesis of Phosphonate 6.45

After successful synthesis of the key aldol adducts, the next priority became the coupling of the C<sub>1</sub>-C<sub>8</sub> phosphonate acid **5.5**<sup>1</sup> with the C<sub>9</sub>-C<sub>25</sub> fragment (Scheme 6.13). We initially used 18*S* isomer **6.41** to explore our proposed chemistry. Our attempts to silylate C<sub>18</sub> hydroxyl group with TES were not successful. When alcohol **6.41** was treated with Et<sub>3</sub>N / DCM / TESOTf at -78°C, a complex mixture was obtained due to the decomposition of the diene moiety. There was no reaction when TESCl / imid. / DMF conditions were employed, presumably because of the steric congestion at C<sub>18</sub> hydroxyl group. After extensive investigation, we discovered that the silylation proceeded smoothly to give TES ether **6.44** when excess DMAP (15 eq.) and TESCl (10 eq.) were used. Next, selective deprotection of the C<sub>25</sub> TMS group, followed by the intermolecular Yamaguchi esterification,<sup>27</sup> revealed the desired phosphonate **6.45**. It should be noted that compound **6.45** contains all the carbon atoms required to complete the synthesis of amphidinolide B.

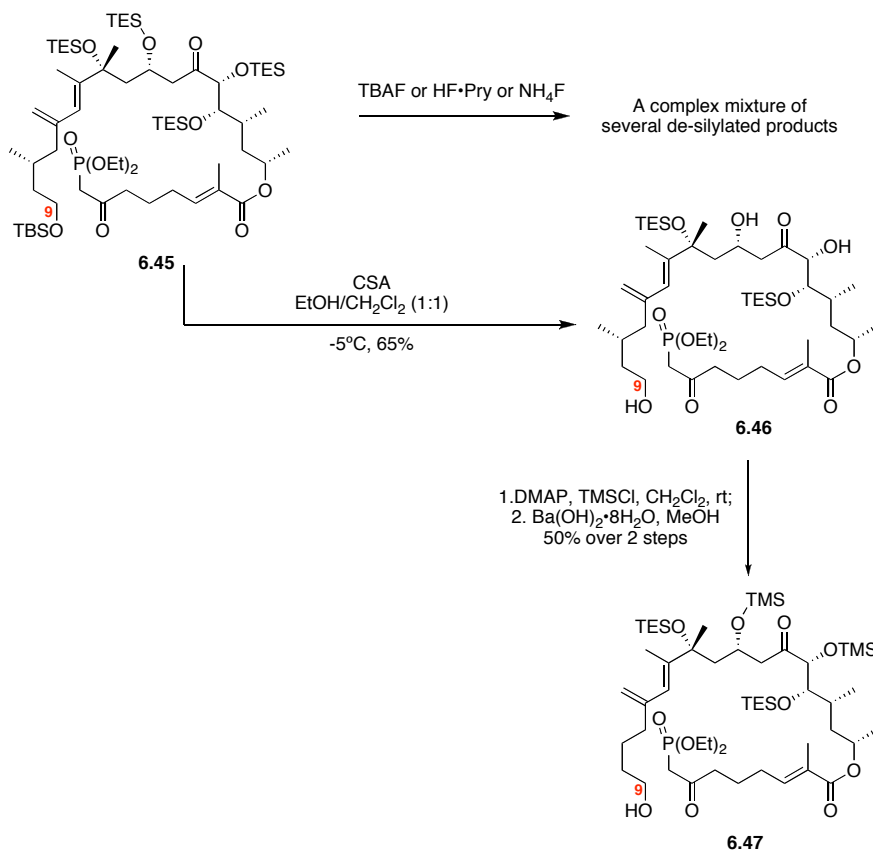


**Scheme 6.13.** Synthesis of Phosphonate **6.45**

### 6.5.6 Attempts to Selectively Remove C<sub>9</sub> TBS Group

With the synthesis of phosphonate **6.45**, the next goal was the selective removal of the C<sub>9</sub> TBS group in the presence of several secondary TES groups (Scheme 6.14). Unfortunately, our efforts were thwarted by the poor selectivity. Under fluoride based conditions, a complex mixture of several de-silylated products were observed. The exposure of TBS ether **6.45** to acidic conditions generated triol **6.46** with the deprotection of TBS group and relatively less hindered C<sub>18</sub>, C<sub>21</sub> TES groups in moderate yield (65%). The desired alcohol **6.47**

was prepared through a sequence involving the silylation of triol **6.46** and the selective deprotection of C<sub>9</sub> primary TMS group. Unfortunately, the low overall yield (32% from phosphonate **6.45**) limited our ability to move forward.

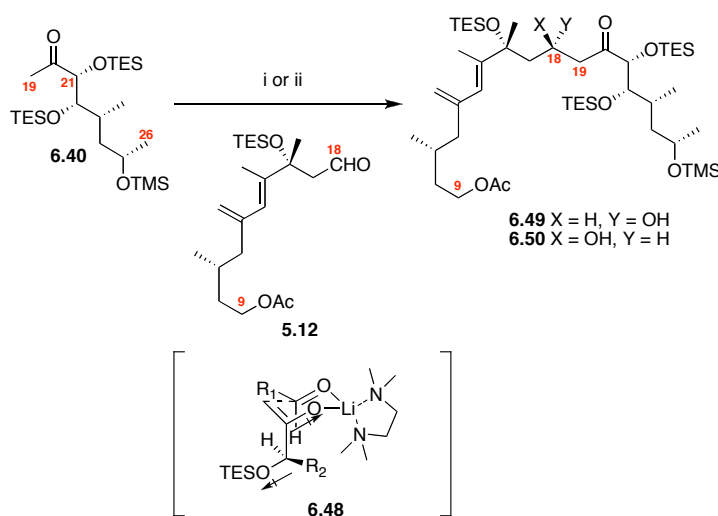


**Scheme 6.14.** Attempts to Selectively Remove TBS Group

### 6.5.7 Aldol Coupling between Methyl Ketone **6.40** and Aldehyde **5.12**

Faced with these synthetic hurdles, we were forced to use the C<sub>9</sub> acetate protected diene aldehyde **5.12** (Scheme 6.15). After conducting more investigation on the key aldol coupling, we were pleased to find the excellent

diastereoselectivity (1:8 dr, favoring 18*R* diastereomer) was obtained at -100°C. The alternate 18*S* diastereomer can be afforded by performing the reaction at higher temperature (-40°C, 1.2:1 dr) and elongation of the reaction time resulted in no ratio change. Although still under investigation, a transition state **6.48** which minimizes the dipoles of the C<sub>21</sub> C–O  $\sigma$  bond and the enolate might lead to the good diastereoselectivity at -100°C.

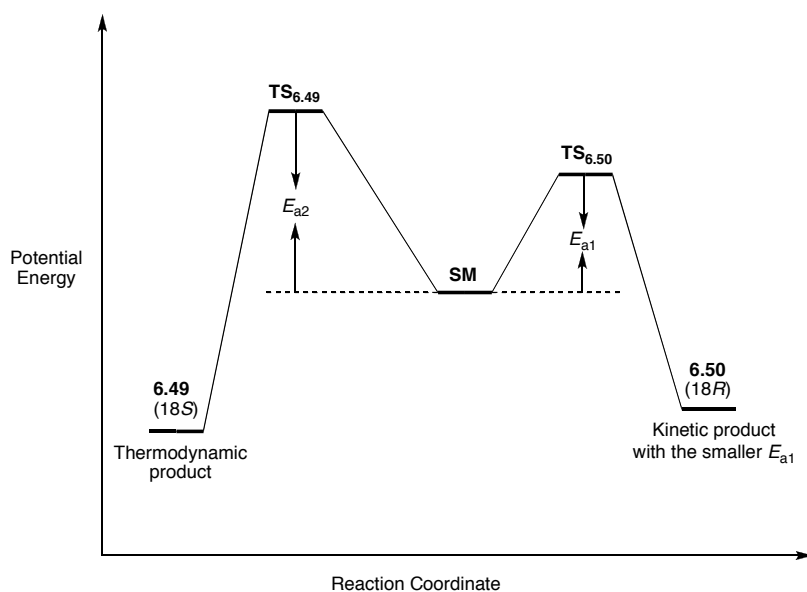


(i) LDA, TMEDA, THF, -100°C then add **5.12**, 65% (1:8 dr, **6.49**:**6.50**); (ii) LDA, TMEDA, THF, -40°C then add **5.12**, 66% (1.2:1 dr, **6.49**:**6.50**)

**Scheme 6.15.** Aldol Coupling Between Methyl Ketone **6.40** and Aldehyde **5.12**

One possible explanation for the observed stereochemical outcome could be a dueling kinetic vs. thermodynamic controlled process, which has been widely reported by the others.<sup>28</sup> The proposed energy diagram of the aldol coupling is depicted in Figure 6.2. This argument would pose that 18*R* diastereomer **6.50** would be the kinetic product as it is generated at low temperature (-100°C) and

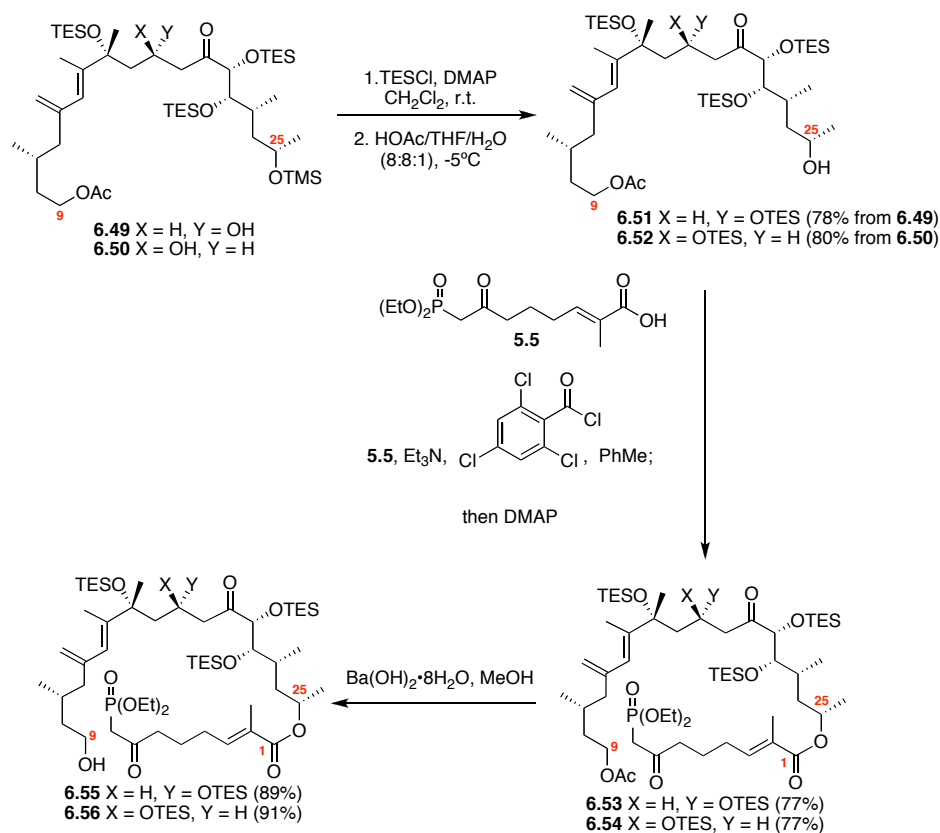
would require lower activation energy. In contrast, the diastereomeric mixture (18*S*:18*R*=1.2:1) observed at a higher temperature (-40°C) would support the presumed energetic similarity between 18*S* & 18*R* diastereomers under reversible (thermodynamic) conditions.



**Figure 6.2.** Proposed Energy Diagram of the Aldol Coupling

### 6.5.8 Synthesis of Phosphonate Alcohol 6.55 and 6.56

The conversion of aldol adducts to phosphonate alcohol **6.55/6.56** was displayed in Scheme 6.16. Silylation, followed by selective removal of C<sub>25</sub> TMS group and Yamaguchi esterification produced phosphonate **6.53/6.54**. When **6.53/6.54** was treated with Ba(OH)<sub>2</sub>•8H<sub>2</sub>O in MeOH, the selective deprotection of acetate group proceeded cleanly to yield phosphonate alcohols **6.55/6.56** in high yield.



**Scheme 6.16:** Synthesis of phosphonate alcohol **6.55/6.56**

## 6.6 Conclusion

The key aldol coupling between the C<sub>9</sub>-C<sub>18</sub> diene moiety and C<sub>19</sub>-C<sub>26</sub> methyl ketone fragment was investigated. The protecting groups on C<sub>21</sub> were discovered to have significant effects on the aldol reaction. Although the PMB and Bn groups provided chelation-control to give great diastereoselectivity, favoring the 18*S* isomer, the attempts to remove these groups proved unsuccessful. The C<sub>21</sub> TES-protected methyl ketone led to the production of the 18*R* isomer in 1:8 dr at  $-100^\circ\text{C}$ , while the 18*S* isomer was yielded at  $-40^\circ\text{C}$  in 1.2:1 dr. Both C<sub>9</sub>-C<sub>26</sub> adol

adducts were successfully coupled with C<sub>1</sub>-C<sub>8</sub> fragment and were converted to the corresponding phosphonate alcohols. With compounds **6.55** and **6.56** in hand, the challenges that remained were the macrocyclization, incorporation of the allylic epoxide moiety and the global de-silylation. Our methods to effect these transformations will be discussed in the next chapter.

## 6.7 References

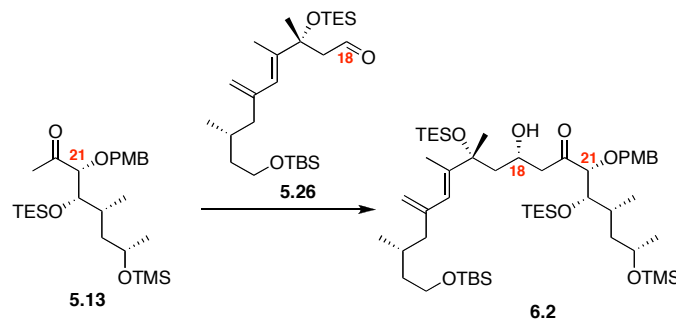
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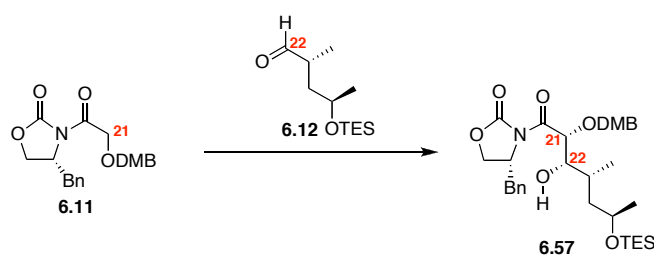
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## 6.8 Experimental



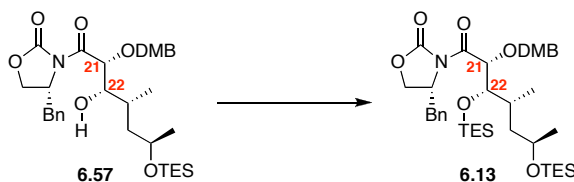
**Aldol adduct 6.2:** To a stirred solution of methyl ketone **5.13**<sup>1</sup> (136 mg, 0.27 mmol) in THF (1.5 mL) at -78°C was added LDA<sup>2</sup> (0.32 mL, 1 M in THF) was added. After 30 min, a pre-cooled (-78°C) solution of aldehyde **5.26** (117 mg, 0.25 mmol) in THF (0.5 mL) was added via *cannula* in one portion. After another 0.5 h, the reaction was quenched with sat. aq. NH<sub>4</sub>Cl (2 mL) at -78°C, warmed up to rt and extracted with ether (3 X 15 mL). The dried extract (MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 3-5% EtOAc / Hexanes, to give aldol adduct **6.2** (128 mg, 0.13 mmol, 53%) as a colorless oil:  $[\alpha]_D^{23} = +11.8$  (*c* 0.61, CHCl<sub>3</sub>); IR (neat) 3513, 2955, 2929, 2877, 1715, 1614, 1515, 1462, 1250, 1091, 1038, 1007, 838, 742 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.30 (d, *J* = 8.7 Hz, 2H), 6.87 (d, *J* = 8.7 Hz, 2H), 5.89 (s, 1H), 5.03 (s, 1H), 4.84 (s, 1H), 4.55 (d, *J* = 11.1 Hz, 1H), 4.42 (br, 1H), 4.37 (d, *J* = 11.4 Hz, 1H), 3.82 (s, 3H), 3.72-3.82 (m, 3H), 3.62-3.69 (m, 2H), 3.10 (dd, *J* = 17.7, 7.2 Hz, 1H), 2.31 (dd, *J* = 17.7, 4.8 Hz, 1H), 2.19 (dd, *J* = 13.5, 5.1 Hz, 1H), 1.80-1.91 (m, 1H), 1.83 (s, 3H), 1.42-1.70 (m, 4H), 1.54 (s, 3H), 1.20-1.37 (m, 4H), 1.13 (d, *J* = 6.0 Hz, 3H), 0.89-1.00 (m, 27H), 0.82-0.87 (m, 6H), 0.54-0.67 (m,

12H), 0.08 (s, 9H), 0.06 (s, 6H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  211.2, 159.2, 144.6, 143.1, 129.9, 129.7, 125.1, 114.9, 113.6, 88.1, 79.5, 77.1, 72.4, 65.9, 64.8, 61.4, 55.2, 48.1, 46.3, 46.0, 44.4, 39.9, 32.1, 29.7, 28.4, 26.0, 24.8, 19.5, 18.3, 14.7, 12.9, 7.2, 7.1, 6.6, 5.3, 0.4, -5.2; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{52}\text{H}_{100}\text{O}_8\text{Si}_4\text{Na}$  ( $\text{M}+\text{Na}$ ) 987.6393, found 987.6396.



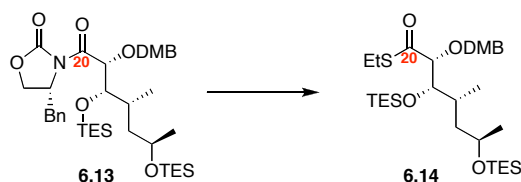
**Aldol adduct 6.57:** To a stirred solution of **6.11** (1.60 g, 0.15 mmol) in  $\text{CH}_2\text{Cl}_2$  (11.2 mL) at  $-60^\circ\text{C}$  was sequentially added  $\text{Et}_3\text{N}$  (0.44 g, 0.60 mL, 4.33 mmol) and  $\text{Bu}_2\text{BOTf}$  (1.19 g, 1.08 mL, 4.33 mmol). After 3 h, the resulted solution was warmed up  $0^\circ\text{C}$  for 30 min and then cooled back to  $-60^\circ\text{C}$ . A solution of aldehyde **6.12**<sup>3</sup> (1.12 g, 4.86 mmol) in DCM (4.8 mL) was transferred to the reaction mixture via *cannula*. After 2 h, the reaction was allowed to warm up to  $0^\circ\text{C}$ . After another 20 min, the reaction was quenched by adding pH 7 phosphate buffer (20 mL) followed by MeOH (15 mL) and 30%  $\text{H}_2\text{O}_2$  (4 mL). After 1 h, the reaction mixture was extracted with EtOAc (4 X 35 mL). The dried ( $\text{MgSO}_4$ ) extract was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 15-20% EtOAc / Hexanes, to give **6.57** (1.58 g, 2.57 mmol, 62%) as a colorless oil:  $[\alpha]_{\text{D}}^{23} = -9.2$  ( $c$  0.77,  $\text{CHCl}_3$ ); IR (neat) 3493, 2957, 2876, 1781, 1709, 1593, 1517, 1455, 1390, 1265, 1240, 1159, 1052, 1028,  $746\text{ cm}^{-1}$ ;  $^1\text{H}$  NMR

(400 MHz)  $\delta$  7.23-7.38 (m, 5H), 6.83-7.03 (m, 3H), 5.36 (d,  $J = 2.0$  Hz, 1H), 4.63-4.71 (m, 2H), 4.51 (d,  $J = 10.8$  Hz, 1H), 4.21-4.29 (m, 2H), 3.93 (s, 3H), 3.84-3.90 (m, 1H), 3.88 (s, 3H), 3.62-3.66 (m, 1H), 3.32 (dd,  $J = 13.6, 3.6$  Hz, 1H), 2.77 (dd,  $J = 13.6, 10.0$  Hz, 1H), 2.32 (d,  $J = 10.0$  Hz, 1H), 1.78-1.80 (m, 1H), 1.58-1.64 (m, 1H), 1.15 (d,  $J = 6.0$  Hz, 3H), 1.03 (d,  $J = 6.8$  Hz, 3H), 0.98 (t,  $J = 8.0$  Hz, 9H), 0.62 (q,  $J = 8.0$  Hz, 6H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  177.2, 153.2, 149.1, 149.0, 135.2, 129.5, 129.4, 129.0, 127.4, 121.4, 112.1, 110.9, 78.1, 76.1, 66.9, 55.9, 55.7, 42.8, 37.7, 34.1, 23.2, 15.7, 6.9, 4.9; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{33}\text{H}_{49}\text{NO}_8\text{SiNa}$  ( $\text{M}+\text{Na}$ ) 638.3125, found 638.3155.



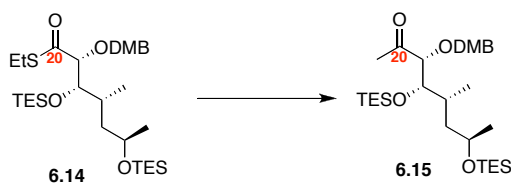
**TES ether 6.13:** To a stirred solution of adol adduct **6.57** (500 mg, 0.81 mmol) in DCM (3.32 mL) at  $0^\circ\text{C}$  was sequentially added 2,6-lutidine (184 mg, 0.20 mL, 1.72 mmol) and TESOTf (287 mg, 0.25 mL, 1.09 mmol). After 1 h, the reaction was quenched with sat. aq.  $\text{NH}_4\text{Cl}$  (10 mL) and extract with  $\text{Et}_2\text{O}$  (3 X 30 mL). The dried ( $\text{MgSO}_4$ ) extract was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 15%  $\text{EtOAc}$  / Hexanes, to give **6.13** (570 mg, 0.78 mmol, 96%) as a colorless oil:  $[\alpha]_{\text{D}}^{23} = -40.7$  ( $c$  0.42,  $\text{CHCl}_3$ ); IR (neat) 2955, 2911, 2876, 1784, 1702, 1517, 1456, 1239, 1084, 740  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz)  $\delta$  7.17-7.32 (m,H), 6.80-7.00 (m, 3H), 5.28 (d,  $J = 6.0$  Hz, 1H), 4.71

(d,  $J = 11.7$  Hz, 1H), 4.54 (d,  $J = 11.7$  Hz, 2H), 4.47-4.49 (m, 1H), 4.00-4.12 (m, 3H), 3.90 (s, 3H), 3.84 (s, 3H), 3.78-3.82 (m, 1H), 3.14 (dd,  $J = 3.0, 13.2$  Hz, 1H), 2.40 (dd,  $J = 10.5, 13.5$  Hz, 1H), 1.42-1.56 (m, 3H), 1.11 (d,  $J = 5.7$  Hz, 3H), 0.89-0.97 (m, 21H), 0.55-0.63 (m, 12H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  171.9, 152.9, 148.7, 148.8, 135.3, 130.3, 129.3, 129.0, 127.3, 121.1, 111.9, 110.7, 73.4, 67.2, 66.4, 56.0, 55.9, 55.8, 44.4, 37.4, 33.5, 23.7, 14.8, 7.1, 6.9, 5.4, 5.3, 5.2, 5.0; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{39}\text{H}_{63}\text{NO}_8\text{Si}_2\text{Na}$  ( $\text{M}+\text{Na}$ ) 752.3990, found 752.3992.



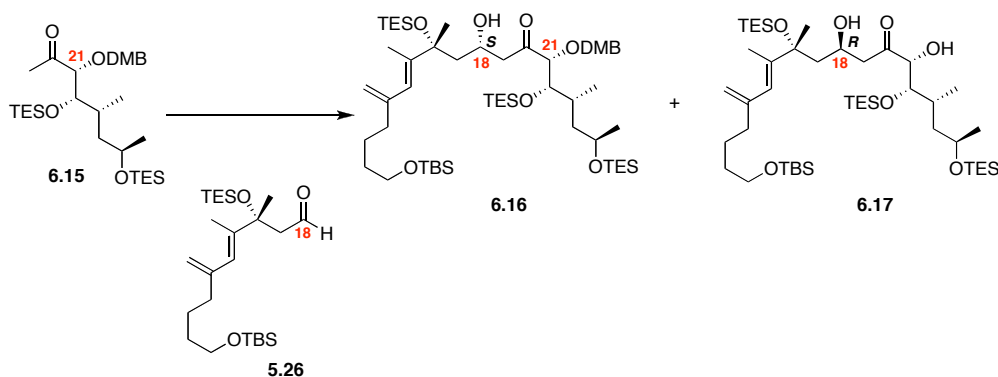
**Thiol ester 6.14:** To a stirred solution of EtSH (90 mg, 0.107 mL, 1.45 mmol) in THF (12.6 mL) at 0°C was added *n*-BuLi (0.51 mL, 1.27 mmol, 2.5 M in Hexanes). After 1 h, a solution of **6.13** (610 mg, 0.83 mmol) in THF (2.7 mL) was added dropwise via *cannula*. After another 1 h, the reaction was quenched with sat. aq.  $\text{NH}_4\text{Cl}$  (10 mL) and extract with  $\text{Et}_2\text{O}$  (3 X 30 mL). The dried ( $\text{MgSO}_4$ ) extract was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 5-8% EtOAc / Hexanes, to give **6.14** (470 mg, 0.76 mmol, 92%) as a colorless oil:  $[\alpha]_{\text{D}}^{23} = +42.0$  ( $c$  0.39,  $\text{CHCl}_3$ ); IR (neat) 2955, 2911, 2876, 1683, 1517, 1458, 1419, 1378, 1266, 1240, 1161, 1079, 1032, 811, 740  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz)  $\delta$  7.04 (dd,  $J = 1.6$  Hz, 1H), 6.94 (dd,  $J = 1.6, 8.0$  Hz, 1H), 6.86 (d,  $J = 8.4$  Hz, 1H), 4.63 (d,  $J = 10.8$  Hz, 1H), 4.43 (d,  $J = 10.8$  Hz, 1H), 3.94 (s, 3H),

3.92-3.93 (m, 4H), 3.81-3.86 (m, 2H), 2.91 (q,  $J = 7.6$  Hz, 2H), 1.45-1.57 (m, 3H), 1.30 (t,  $J = 7.2$  Hz, 3H), 1.14 (d,  $J = 6.0$  Hz, 3H), 0.90-0.98 (m, 21H), 0.53-0.62 (m, 12H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  202.2, 148.8, 148.6, 129.9, 120.6, 110.6, 110.7, 88.2, 72.9, 66.9, 55.9, 55.8, 44.9, 32.7, 23.4, 22.5, 14.6, 13.6, 7.0, 6.9, 5.3, 5.2, 5.0; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{31}\text{H}_{58}\text{O}_6\text{Si}_2\text{SNa}$  ( $\text{M}+\text{Na}$ ) 637.3390, found 637.3407.



**Methyl ketone 6.15:** To a stirred slurry of CuI (859 mg, 4.51 mmol) in  $\text{Et}_2\text{O}$  (8.3 mL) at  $0^\circ\text{C}$  was added MeLi (5.6 mL, 9.6 mmol, 1.6 M in  $\text{Et}_2\text{O}$ ). After 15 min, the colorless solution was cooled to  $-50^\circ\text{C}$  and a solution of **6.14** (450 mg, 0.75 mmol) in  $\text{Et}_2\text{O}$  (4.2 mL) was transferred into the reaction mixture dropwise via *cannula*. After 2 h, the reaction was quenched with sat. aq.  $\text{NH}_4\text{Cl}$  (10 mL) at  $-50^\circ\text{C}$ , warmed to rt and extracted with  $\text{Et}_2\text{O}$  (3 X 30 mL). The dried ( $\text{MgSO}_4$ ) extract was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 2-4% EtOAc / Hexanes, to give **6.15** (296 mg, 0.52 mmol, 71%) as a colorless oil:  $[\alpha]_{\text{D}}^{23} = +22.6$  ( $c$  0.23,  $\text{CHCl}_3$ ); IR (neat) 2955, 2911, 2876, 1716, 1517, 1457, 1267, 1240, 1082, 1031, 1007, 741  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz)  $\delta$  6.82-6.90 (m, 3H), 4.50 (dd,  $J = 11.4, 15.0$  Hz, 2H), 3.90 (s, 6H), 3.80-3.85 (m, 2H), 3.76 (d,  $J = 6.3$  Hz, 1H), 2.13 (s, 3H), 1.50-1.52 (m, 3H), 1.13 (d,  $J = 6.0$  Hz, 3H),

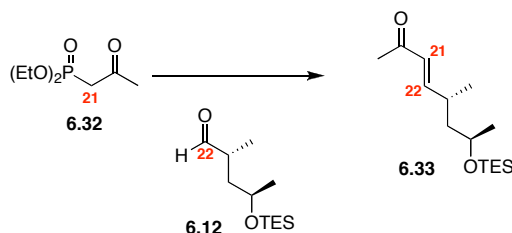
0.88-1.00 (m, 21H), 0.57-0.64 (m, 12H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  210.6, 148.8, 129.9, 120.7, 111.4, 110.8, 88.5, 72.9, 67.0, 55.9, 55.8, 44.5, 33.1, 27.0, 23.5, 14.0, 7.0, 6.9, 5.3, 5.0; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{30}\text{H}_{56}\text{O}_6\text{Si}_2\text{Na}$  ( $\text{M}+\text{Na}$ ) 591.3513, found 591.3527.



**Aldol adduct 6.16:** To a stirred solution of methyl ketone **6.15** (30 mg, 0.0527 mmol) in  $\text{Et}_2\text{O}$  (0.5 mL) at  $-78^\circ\text{C}$  was added  $\text{LDA}^2$  (64  $\mu\text{L}$ , 0.064 mmol, 1 M in THF). After 15 min, a pre-cooled ( $-78^\circ\text{C}$ ) solution of aldehyde **5.26** (50 mg, 0.105 mmol) in THF (0.5 mL) was added via *cannula* in one portion. After another 0.5 h, the reaction was quenched with sat. aq.  $\text{NH}_4\text{Cl}$  (2 mL) at  $-78^\circ\text{C}$ , warmed up to rt and extracted with ether (3 X 10 mL). The dried extract ( $\text{MgSO}_4$ ) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 6-8%  $\text{EtOAc}$  / Hexanes, to sequentially give aldol adduct **6.17** (10 mg, 0.0096 mmol, 18%) and **6.16** (39 mg, 0.0375 mmol, 71%) and as colorless oils. **6.16**:  $[\alpha]_{\text{D}}^{23} = +9.1$  ( $c$  0.58,  $\text{CHCl}_3$ ); IR (neat) 3503, 2955, 2934, 2876, 1715, 1517, 1463, 1265, 1240, 1095, 1007, 741  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  6.83-6.97 (m, 3H), 5.90 (s, 1H), 5.05 (s, 1H), 4.85 (s, 1H), 4.53 (d,  $J = 11.2$  Hz, 1H), 4.44-

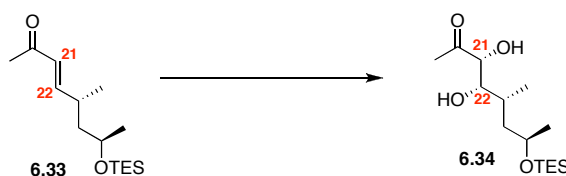


4.50 (m, 1H), 4.39(d,  $J = 10.8$  Hz, 1H), 3.91 (s, 6H), 3.76-3.83 (m, 3H), 3.66-3.70 (m, 2H), 3.05 (dd,  $J = 17.6, 6.8$  Hz, 1H), 2.38 (dd,  $J = 17.6, 5.6$  Hz, 1H), 2.19 (dd,  $J = 13.5, 4.8$  Hz, 1H), 1.80-1.91 (m, 1H), 1.84 (s, 3H), 1.48-1.68 (m, 4H), 1.57 (s, 3H), 1.20-1.42 (m, 4H), 1.10 (d,  $J = 6.0$  Hz, 3H), 0.86-1.00 (m, 42H), 0.57-0.66 (m, 18H), 0.08 (s, 6H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  211.0, 148.8, 148.6, 144.6, 142.9, 130.1, 125.2, 120.7, 114.9, 111.5, 110.7, 88.3, 79.5, 72.6, 66.9, 64.8, 61.4, 55.9, 55.8, 48.0, 46.7, 46.0, 44.8, 39.9, 33.1, 28.4, 23.3, 19.5, 14.7, 13.5, 7.1, 7.0, 6.9, 6.5, 5.3, 5.0, -5.2; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{56}\text{H}_{107}\text{O}_9\text{Si}_4$  ( $\text{M}+\text{H}$ ) 1035.6992, found 1035.7047.



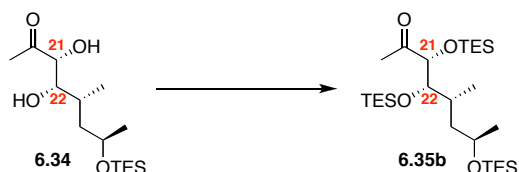
**Enone 36:** To a stirred slurry of NaH (36 mg, 0.90 mmol, 60% W / W in mineral oil) in DME (2 mL) was added phosphonate **6.32** (138 mg, 0.83 mmol) at rt. After 1 h, a solution of aldehyde **6.12** (160 mg, 0.69 mmol) in DME (2 mL) was added via *cannula*. After another 6 h, the reaction was quenched with sat. aq.  $\text{NH}_4\text{Cl}$  (2 mL) and extracted with  $\text{Et}_2\text{O}$  (4 X 10 mL). The dried extract ( $\text{MgSO}_4$ ) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 5%  $\text{EtOAc}$  / Hexanes, to give enone **6.33** (142 mg, 0.52 mmol, 76%) as a colorless oil:  $[\alpha]_{\text{D}}^{23} = -46.0$  ( $c$  1.47,  $\text{CHCl}_3$ ); IR (neat) 2958, 2877, 1700, 1678, 1627, 1458, 1360, 1252, 1139, 1055, 984, 745  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$

6.76 (dd,  $J = 15.9, 7.8$  Hz, 1H), 6.09 (d,  $J = 15.9$  Hz, 1H), 3.86-3.79 (m, 1H), 2.58-2.53 (m, 1H), 2.26 (s, 3H), 1.41-1.55 (m, 2H), 1.18 (d,  $J = 6.0$  Hz, 3H), 1.09 (d,  $J = 6.9$  Hz, 3H), 0.97 (t,  $J = 7.8$  Hz, 9H), 0.64 (q,  $J = 7.5$  Hz, 6H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  198.8, 153.5, 129.6, 66.4, 46.3, 33.4, 27.0, 24.3, 20.3, 6.9, 5.2, 5.0; HRMS ( $\text{EI}^+$ ) calcd. for  $\text{C}_{15}\text{H}_{30}\text{O}_2\text{Si}$  ( $\text{M}^+$ ) 270.2015, found 270.2008.

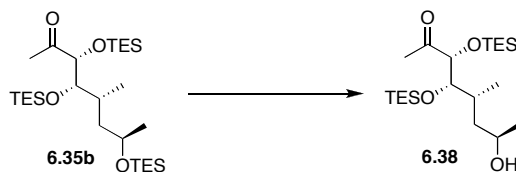


**Diol 6.34:** To a stirred solution of enone **6.33** (142 mg, 0.52 mmol) in  $t\text{BuOH}/\text{H}_2\text{O}$  (5 mL, 1:1) at  $0^\circ\text{C}$  was sequentially added AD-mix- $\alpha$  (0.735 g),  $\text{NaHCO}_3$  (132 mg, 1.57 mmol),  $\text{MeSO}_2\text{NH}_2$  (50.6 mg, 0.53 mmol), and  $\text{K}_2\text{OsO}_2(\text{OH})_4$  (1.9 mg, 0.005 mmol). After 8 h, the reaction was quenched with sat. aq.  $\text{Na}_2\text{SO}_3$  (8 mL) and extracted with EtOAc (4 X 10 mL). The dried extract ( $\text{MgSO}_4$ ) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 15-40% EtOAc / Hexanes, to give diol **6.34** (122 mg, 0.40 mmol, 77%) as a colorless oil:  $[\alpha]_{\text{D}}^{23} = -29.2$  ( $c$  1.5,  $\text{CHCl}_3$ ); IR (neat) 3456, 2957, 2877, 1717, 1380, 1238, 1132, 1048, 1011, 744  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  4.28 (d,  $J = 3.6$  Hz, 1H), 4.04-3.99 (m, 1H), 3.72-3.78 (m, 1H & OH), 2.41 (d,  $J = 10.4$  Hz, 1H), 2.30 (s, 3H), 1.89-1.96 (m, 1H), 1.71-1.77 (m, 1H), 1.40-1.47 (m, 1H), 1.22 (d,  $J = 6.0$  Hz, 3H), 1.10 (d,  $J = 6.8$  Hz, 3H), 1.00 (t,  $J = 7.8$  Hz, 9H), 0.68 (q,  $J = 7.9$  Hz, 6H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  208.9, 77.7, 75.2, 66.9, 42.8, 34.0,

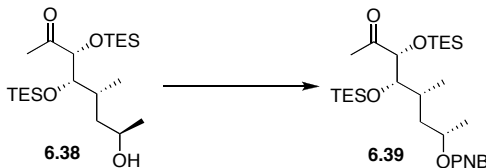
25.3, 23.1, 16.4, 6.9, 4.9; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{15}\text{H}_{32}\text{O}_4\text{SiNa}$  ( $\text{M}+\text{Na}$ ) 327.1968, found 327.1950.



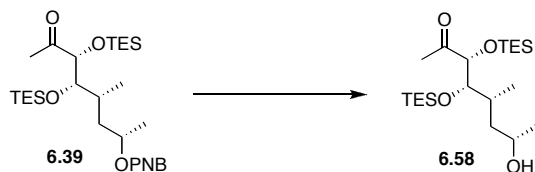
**TES ether 6.35b:** To a stirred solution of diol **6.34** (800 mg, 2.63 mmol) in  $\text{CH}_2\text{Cl}_2$  (10 mL) at  $-78^\circ\text{C}$  was sequentially added 2,6-lutidine (1.41 g, 1.53 mL, 13.1 mmol) and TESOTf (1.74 g, 1.49 mL, 6.58 mmol). After 30 min, the reaction was quenched with sat. aq.  $\text{NH}_4\text{Cl}$  (10 mL) and extracted with  $\text{Et}_2\text{O}$  (4 X 25 mL). The dried extract ( $\text{MgSO}_4$ ) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 10% EtOAc / Hexanes, to give TES ether **6.35b** (1.29 g, 2.42 mmol, 92%) as a colorless oil:  $[\alpha]_{\text{D}}^{23} = -0.83$  (*c* 1.2,  $\text{CHCl}_3$ ); IR (neat) 2957, 2878, 1716, 1458, 1238, 1005  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  4.07 (d,  $J = 6.0$  Hz, 1H), 3.78-3.85 (m, 1H), 3.70 (dd,  $J = 5.9, 2.6$  Hz, 1H), 2.20 (s, 3H), 1.60-1.64 (m, 1H), 1.45-1.51 (m, 2H), 1.13 (d,  $J = 6.0$  Hz, 3H), 0.94-1.00 (m, 27H), 0.83 (d,  $J = 6.6$  Hz, 3H), 0.55-0.70 (m, 18H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  210.0, 81.5, 78.6, 67.1, 45.4, 32.2, 27.3, 23.1, 14.0, 7.0, 6.84, 6.79, 5.2, 4.9, 4.8; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{27}\text{H}_{60}\text{O}_4\text{Si}_3\text{Na}$  ( $\text{M}+\text{Na}$ ) 555.3697, found 555.3683.



**Alcohol 6.38:** TES ether **6.35b** (5.60 g, 10.5 mmol) was dissolved in a stirred solution of HOAc / THF / H<sub>2</sub>O (107 mL, 8:8:1) at 0°C. After 12 h, the reaction was quenched with solid NaHCO<sub>3</sub>, filtered over Celite and extracted with ether (4 X 100 mL). The dried extract (MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 10% EtOAc / Hexanes, to give alcohol **6.38** (3.90 g, 9.31 mmol, 89%) as a colorless oil:  $[\alpha]_D^{23} = -30.5$  (*c* 1.45, CHCl<sub>3</sub>); IR (neat) 3446, 2958, 2878, 1716, 1458, 1239, 1005, 739 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  4.18 (d, *J* = 5.6 Hz, 1H), 3.86-3.88 (m, 2H), 2.24 (s, 3H), 1.90 (d, *J* = 4.0 Hz, 1H), 1.82 (m, 1H), 1.41-1.55 (m, 2H), 1.20 (d, *J* = 6.0 Hz, 3H), 0.97-1.05 (m, 18H), 0.87 (d, *J* = 6.8 Hz, 3H), 0.61-0.73 (m, 12H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  209.3, 81.4, 76.4, 65.6, 43.7, 31.6, 27.8, 23.6, 15.4, 7.0, 6.8, 5.2, 4.8, 4.7; HRMS (ES<sup>+</sup>) calcd. for C<sub>21</sub>H<sub>46</sub>O<sub>4</sub>Si<sub>2</sub>Na (M+Na) 441.2832, found 441.2836.

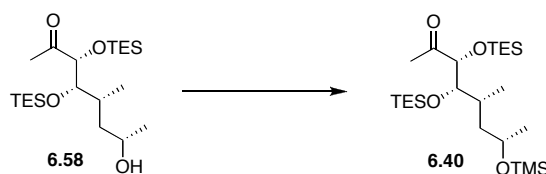


**PNB ester 6.39:** To a stirred solution of alcohol **6.38** (600 mg, 1.43 mmol) in THF (15 mL) at 0°C was sequentially added PPh<sub>3</sub> (1.50 g, 5.72 mmol), 4-nitrobenzoic acid (0.96 g, 5.74 mmol), and DEAD (0.99 g, 0.90 mL, 5.70 mmol). After 1 h, the reaction mixture was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 1-3% EtOAc / Hexanes, to give ester **6.39** (670 mg, 1.18 mmol, 82%) as a colorless oil:  $[\alpha]_D^{23} = +13.6$  (*c* 1.08, CHCl<sub>3</sub>); IR (neat) 2956, 2878, 1723, 1530, 1319, 1275, 1014, 721 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 8.31 (d, *J*=9.0 Hz, 2H), 8.23 (d, *J*=9.0 Hz, 2H), 5.26 (m, 1H), 4.17 (d, *J* = 4.8 Hz, 1H), 3.69 (t, *J* = 4.8 Hz, 1H), 2.18 (s, 3H), 2.02-2.12 (m, 1H), 1.76 (m, 1H), 1.45-1.49 (m, 1H), 1.38 (d, *J* = 6.1 Hz, 3H), 0.93-1.02 (m, 18H), 0.90 (d, *J* = 6.6 Hz, 3H), 0.56-0.70 (m, 12H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 209.3, 164.3, 150.4, 136.1, 130.7, 123.4, 81.5, 78.2, 70.8, 40.2, 32.4, 27.9, 20.9, 14.8, 7.0, 6.8, 5.1, 4.8; HRMS (ES<sup>+</sup>) calcd. for C<sub>28</sub>H<sub>49</sub>NO<sub>7</sub>Si<sub>2</sub>Na (M+Na) 590.2945, found 590.2926.



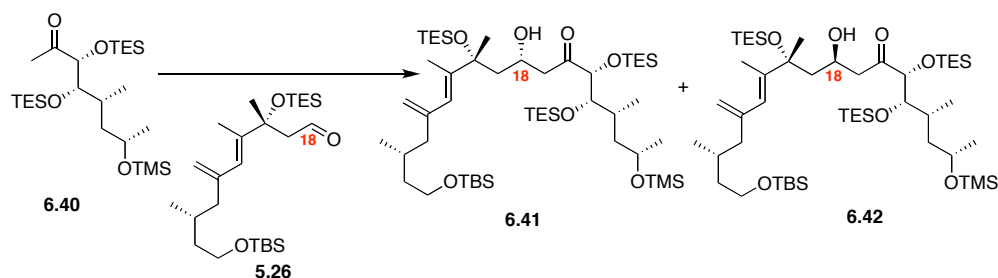
**Alcohol 6.58:** To a stirred solution of ester **6.39** (700 mg, 1.23 mmol) in MeOH (20 mL) at 0°C was added Ba(OH)<sub>2</sub>•8H<sub>2</sub>O (390 mg, 1.24 mmol). After 4 h,

the reaction was quenched with sat. aq.  $\text{NH}_4\text{Cl}$  (10 mL) and extracted with EtOAc (4 X 20 mL). The dried extract ( $\text{MgSO}_4$ ) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 15% EtOAc / Hexanes, to give alcohol **6.58** (369 mg, 0.88 mmol, 72%) as a colorless oil:  $[\alpha]_{\text{D}}^{23} = -7.6$  ( $c$  1.2,  $\text{CHCl}_3$ ); IR (neat) 3434, 2957, 2878, 1716, 1459, 1415, 1239, 1005, 739  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  4.17 (d,  $J = 5.4$  Hz, 1H), 3.81 (m, 1H), 3.69 (dd,  $J = 5.4, 3.9$  Hz, 1H), 2.22 (s, 3H), 1.83 (m, 1H), 1.61-1.69 (m, 1H), 1.24-1.30 (m, 1H), 1.20 (d,  $J = 6.0$  Hz, 3H), 0.94-1.04 (m, 18H), 0.87 (d,  $J = 6.6$  Hz, 3H), 0.58-0.71 (m, 12H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  210.1, 81.5, 78.1, 66.2, 43.8, 32.9, 27.6, 24.4, 15.2, 7.0, 6.8, 5.2, 4.8; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{21}\text{H}_{47}\text{O}_4\text{Si}_2$  ( $\text{M}+\text{H}$ ) 419.3013, found 419.2993.



**TMS ether 6.40:** To a stirred solution of alcohol **6.58** (1.90 g, 4.54 mmol) in  $\text{CH}_2\text{Cl}_2$  (25 mL) at  $-78^\circ\text{C}$  was sequentially added 2,6-lutidine (1.45 g, 1.58 mL, 13.5 mmol) and TMSOTf (1.51 g, 1.23 mL, 6.81 mmol). After 30 min, the reaction was quenched with sat. aq.  $\text{NH}_4\text{Cl}$  (10 mL) and extracted with  $\text{Et}_2\text{O}$  (4 X 20 mL). The dried extract ( $\text{MgSO}_4$ ) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 10% EtOAc / Hexanes, to give TMS ether **6.40** (2.12 g, 4.32 mmol, 95%) as a colorless oil:  $[\alpha]_{\text{D}}^{23} = +14.4$  ( $c$  2.2,  $\text{CHCl}_3$ ); IR (neat) 2957, 2878, 1716, 1459, 1415, 1124, 1006, 841, 741  $\text{cm}^{-1}$ ;  $^1\text{H}$

NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  4.03 (d,  $J = 6.3$  Hz, 1H), 3.79-3.85 (m, 1H), 3.71 (dd,  $J = 6.3, 2.4$  Hz, 1H), 2.19 (s, 3H), 1.73-1.79 (m, 1H), 1.45-1.54 (m, 1H), 1.23-1.32 (m, 1H), 1.16 (d,  $J = 6.0$  Hz, 3H), 0.95-1.03 (m, 18H), 0.83 (d,  $J = 6.6$  Hz, 3H), 0.58-0.70 (m, 12H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  210.2, 81.8, 78.6, 65.9, 45.3, 31.3, 26.8, 24.7, 13.2, 7.0, 6.8, 5.3, 4.8, 0.3; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{24}\text{H}_{54}\text{O}_4\text{Si}_3\text{Na}$  ( $\text{M}+\text{Na}$ ) 513.3224, found 513.3204.

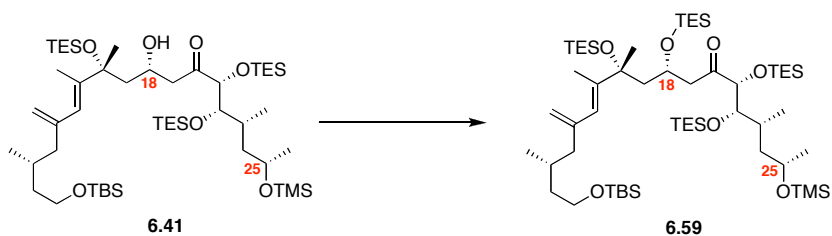


**Aldol adduct 6.41&6.42:** To a stirred solution of methyl ketone **6.40** (312 mg, 0.64 mmol) in THF (5 mL) at  $-78^\circ\text{C}$  was added  $\text{LDA}^2$  (0.765 mL, 1 M in THF). After 15 min, TMEDA (133 mg, 0.172 mL, 1.14 mmol) was added. After 5 min, the reaction was warmed up to  $-40^\circ\text{C}$ , followed by the addition of a pre-cooled ( $-40^\circ\text{C}$ ) solution of aldehyde **5.26** (200 mg, 0.43 mmol) in THF (5 mL) via *cannula* in one portion. After another 0.5 h, the reaction was quenched with sat. aq.  $\text{NH}_4\text{Cl}$  (10 mL)  $-78^\circ\text{C}$ , warmed up to rt and extracted with ether (4 X 20 mL). The dried extract ( $\text{MgSO}_4$ ) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 1-1.5% EtOAc / Hexanes, to give aldol adduct **6.41** (142 mg, 0.15 mmol, 35%) and **6.42** (114 mg, 0.12 mmol, 28%) as colorless oil. **6.41**:  $[\alpha]_{\text{D}}^{23} = -12.0$  ( $c$  1.3,  $\text{CHCl}_3$ ); IR (neat) 3511, 2955, 2929, 2877, 1715, 1460, 1413, 1250, 1092, 1006, 838, 742  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,

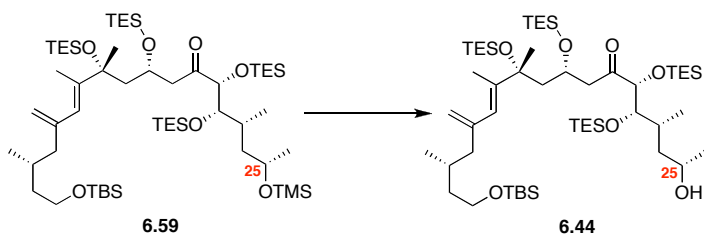
CDCl<sub>3</sub>)  $\delta$  5.89 (s, 1H), 5.02 (s, 1H), 4.83 (s, 1H), 4.33 (m, 1H), 4.10 (d,  $J$  = 5.7 Hz, 1H), 3.82 (m, 1H), 3.74 (s, 1H), 3.64-3.72 (m, 3H), 2.96 (dd,  $J$  = 17.7, 6.2 Hz, 1H), 2.60 (dd,  $J$  = 18.1, 6.4 Hz, 1H), 2.18 (dd,  $J$  = 12.8, 4.0 Hz, 1H), 1.81-1.90 (m, 2H), 1.84 (s, 3H), 1.47-1.67 (m, 4H), 1.56 (s, 3H), 1.22-1.38 (m, 3H), 1.15 (d,  $J$  = 6.0 Hz, 3H), 0.92-1.03 (m, 36H), 0.87 (d,  $J$  = 6.4 Hz, 3H), 0.80 (d,  $J$  = 6.7 Hz, 3H), 0.58-0.70 (m, 18H), 0.10 (s, 9H), 0.06 (s, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  210.7, 144.7, 125.1, 114.7, 81.1, 79.4, 78.4, 65.9, 65.1, 61.5, 48.2, 47.2, 46.0, 45.0, 39.9, 31.1, 28.5, 26.2, 26.0, 24.7, 19.5, 18.3, 14.7, 13.8, 7.2, 7.1, 6.9, 6.6, 5.3, 4.9, 0.4, -5.2; HRMS (ES<sup>+</sup>) calcd. for C<sub>50</sub>H<sub>106</sub>O<sub>7</sub>Si<sub>5</sub>Na (M+Na) 981.6683, found 981.6646.

**MTPA esters:** To a solution of **6.41** (5 mg, 0.005 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (0.5 mL) was sequentially added DMAP (6.4 mg, 0.052 mmol) and (*R*) or (*S*)-(+)- $\alpha$ -methoxy- $\alpha$ -trifluoromethylphenylacetyl chloride (6.6 mg, 4.9  $\mu$ L, 0.026 mmol). After 10 min, the solution was evaporated and the residue was loaded directly onto silica gel and purified by chromatography, eluting with 2 - 10% EtOAc / Hexanes, to give product (*S*)- or (*R*)- **MTPA esters** (52-61%) as colorless oils. <sup>1</sup>H NMR Difference in ppm [(*S*)-Mosher Ester – (*R*)-Mosher ester, CDCl<sub>3</sub>, CDCl<sub>3</sub>, 300 MHz NMR] H<sub>19</sub> = 2.847 – 2.834 = **+0.013**, H<sub>21</sub>: 3.996 – 3.961 = **+0.035**, H<sub>22</sub>: 3.686 – 3.678 = **+0.008**, H<sub>25</sub>: 3.848 – 3.818 = **+0.030**, H<sub>31</sub>: 0.881 – 0.876 = **+0.005**, H<sub>29</sub>: 1.888 – 1.895 = **-0.007**, H<sub>14</sub>: 5.723 – 5.824 = **-0.101**, H<sub>28</sub>: 4.905 – 4.925 = **-0.020**, H<sub>12</sub>: 2.319 – 2.334 = **-0.015**, H<sub>279</sub>: 1.130 – 1.137 = **-0.007**.

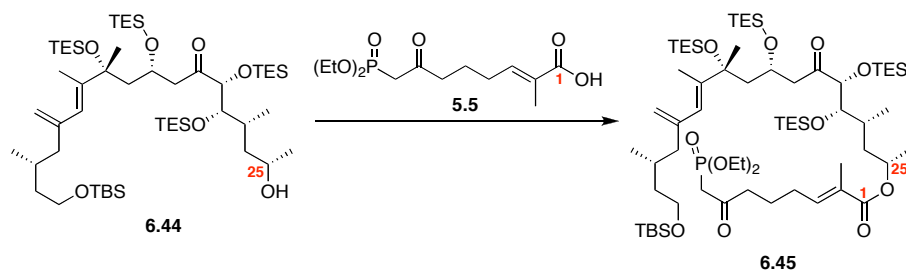




**TES ether 6.59:** To a stirred solution of aldol adduct **6.41** (247 mg, 0.26 mmol) in DCM (5 mL) at rt was sequentially added DMAP (471 mg, 3.86 mmol) and TESCl (194 mg, 0.216 mL, 1.29 mmol). After 1 h, the reaction mixture was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 3% EtOAc / Hexanes, to give TES ether **6.59** (271 mg, 0.25 mmol, 97%) as a colorless oil:  $[\alpha]_D^{23} = -30.6$  (*c* 0.32, CHCl<sub>3</sub>); IR (neat) 2955, 2929, 2877, 1717, 1459, 1414, 1250, 1093, 1006, 838, 741 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  5.78 (s, 1H), 5.02 (s, 1H), 4.91 (s, 1H), 4.19 (m, 1H), 4.04 (d, *J* = 5.7 Hz, 1H), 3.80-3.86 (m, 1H), 3.63-3.72 (m, 3H), 2.93 (dd, *J* = 18.0, 5.7 Hz, 1H), 2.73 (dd, *J* = 18.0, 6.3 Hz, 1H), 2.17 (dd, *J* = 12.8, 5.40 Hz, 1H), 1.79-1.89 (m, 2H), 1.85 (s, 3H), 1.46-1.73 (m, 4H), 1.45 (s, 3H), 1.23-1.39 (m, 3H), 1.15 (d, *J*=5.7 Hz, 3H), 0.91-1.04 (m, 45H), 0.87 (d, *J*=6.3 Hz, 3H), 0.78 (d, *J*=6.9 Hz, 3H), 0.56-0.68 (m, 24H), 0.11 (s, 9H), 0.07 (s, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  208.6, 144.7, 142.7, 125.2, 114.7, 81.0, 78.3, 77.7, 66.1, 65.7, 61.6, 49.9, 48.5, 46.1, 44.9, 40.0, 31.0, 29.7, 28.4, 27.9, 26.0, 24.6, 19.4, 18.3, 14.5, 14.2, 7.2, 7.1, 7.0, 6.9, 6.8, 5.3, 4.9, 0.4, -5.3; HRMS (ES<sup>+</sup>) calcd. for C<sub>56</sub>H<sub>120</sub>O<sub>7</sub>Si<sub>6</sub>Na (M+Na) 1095.7547, found 1095.7495.

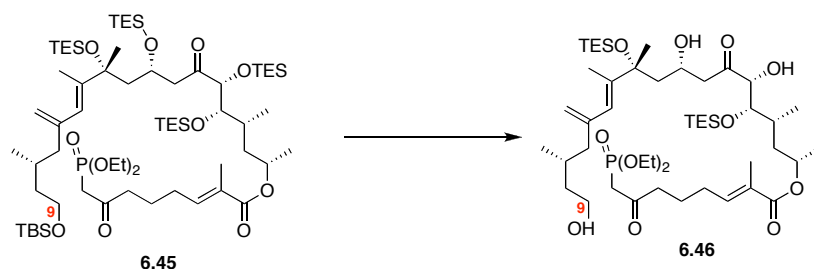


**Alcohol 6.44:** TMS ether **6.59** (543 mg, 0.51 mmol) was dissolved in a stirred solution of HOAc / THF / H<sub>2</sub>O (28 mL, 8:8:1) at 0°C. After 4 h, the reaction was quenched with solid NaHCO<sub>3</sub>, filtered over Celite and extracted with ether (4 X 20 mL). The dried extract (MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 10-20% EtOAc / Hexanes, to give alcohol **6.44** (490 mg, 0.49 mmol, 96%) as a colorless oil:  $[\alpha]_D^{23} = -35.6$  (*c* 0.39, CHCl<sub>3</sub>); IR (neat) 3481, 2955, 2877, 1717, 1459, 1414, 1240, 1098, 1005, 836, 740 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  5.78 (s, 1H), 5.02 (s, 1H), 4.90 (s, 1H), 4.21 (m, 1H), 4.13 (d, *J* = 5.7 Hz, 1H), 3.80 (m, 1H), 3.65-3.69 (m, 3H), 2.92 (dd, *J* = 18.3, 6.0 Hz, 1H), 2.78 (dd, *J* = 18.3, 6.3 Hz, 1H), 2.17 (dd, *J* = 13.2, 5.1 Hz, 1H), 1.56-1.89 (m, 6H), 1.84 (s, 3H), 1.44 (s, 3H), 1.28 (m, 3H), 1.19 (d, *J* = 6.1 Hz, 3H), 0.91-1.05 (m, 45H), 0.81-0.86 (m, 6H), 0.59-0.73 (m, 24H), 0.06 (s, 6H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  208.9, 144.7, 142.3, 125.4, 114.7, 81.4, 78.1, 77.7, 66.3, 65.4, 61.6, 49.7, 49.0, 46.0, 44.1, 40.0, 32.6, 28.4, 28.0, 26.0, 24.4, 19.5, 18.3, 15.6, 14.5, 7.2, 7.0, 6.9, 6.8, 5.3, 5.2, 4.8, -5.3; HRMS (ES<sup>+</sup>) calcd. for C<sub>53</sub>H<sub>112</sub>O<sub>7</sub>Si<sub>5</sub>Na (M+Na) 1023.7152, found 1023.7132.



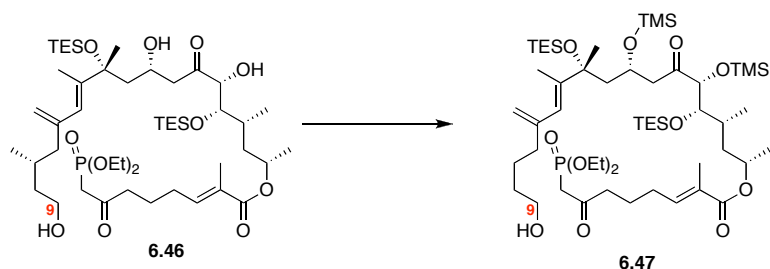
**Phosphonate 6.45:** To a stirred solution of acid **5.5** (421 mg, 1.37 mmol) in PhMe (5.2 mL) at rt was sequentially added Et<sub>3</sub>N (139 mg, 0.191 mL, 1.37 mmol) and 2,4,6-trichlorobenzoyl chloride (323 mg, 0.207 mL, 1.37 mmol). After 12 h, the resulted solution was concentrated *in vacuo*. DMAP (168 mg, 1.37 mmol) was added, followed by the addition of a solution of alcohol **6.44** (260 mg, 0.26 mmol) in PhMe (5.2 mL). After another 12 h, the reaction was quenched with sat. aq. NaHCO<sub>3</sub> (10 mL) and extracted with EtOAc (4 X 20 mL). The dried extract (MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 20-60% EtOAc / Hexanes, to give ester **6.45** (220 mg, 0.17 mmol, 66%) as a colorless oil:  $[\alpha]_D^{23} = -23.8$  (*c* 1.1, CHCl<sub>3</sub>); IR (neat) 2955, 2877, 1715, 1459, 1255, 1096, 1019, 836, 741 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.70 (t, *J* = 8.3 Hz, 1H), 5.79 (s, 1H), 5.05 (m, 1H), 5.03 (s, 1H), 4.90 (s, 1H), 4.21 (m, 1H), 4.14-4.22 (m, 5H), 4.10 (d, *J* = 5.3 Hz, 1H), 3.61-3.73 (m, 3H), 3.12 (d, *J* = 22.8 Hz, 2H), 2.89 (dd, *J* = 18.1, 6.0 Hz, 1H), 2.77 (dd, *J* = 18.3, 6.2 Hz, 1H), 2.68 (t, *J* = 7.3 Hz, 2H), 2.13-2.22 (m, 3H), 1.58-1.89 (m, 8H), 1.84 (s, 3H), 1.83 (s, 3H), 1.46 (s, 3H), 1.36 (t, *J* = 7.1 Hz, 6H), 1.32-1.40 (m, 1H), 1.25 (d, *J* = 6.0 Hz, 3H), 0.92-1.03 (m, 45H), 0.87 (d, *J* = 6.5 Hz, 3H), 0.81 (d, *J* = 6.8 Hz, 3H), 0.56-0.71 (m, 24H), 0.07 (s, 6H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  208.4, 201.5, 167.6, 144.7,

142.5, 140.5, 129.0, 125.3, 114.7, 80.9, 77.8, 77.6, 68.8, 65.6, 62.6, 62.5, 61.6, 49.8, 49.0, 46.1, 43.4, 43.1, 41.9, 40.6, 40.0, 31.4, 28.4, 27.9, 27.7, 26.3, 26.0, 22.3, 20.7, 19.5, 18.3, 16.4, 16.3, 14.7, 14.5, 7.2, 7.0, 6.9, 6.8, 5.2, 5.1, 4.9, -5.3; HRMS ( $ES^+$ ) calcd. for  $C_{66}H_{133}O_{12}Si_5PNa$  ( $M+Na$ ) 1311.8279, found 1311.8269.



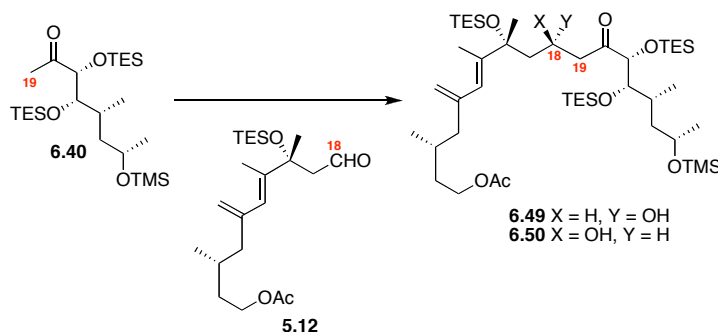
**Triol 6.46:** To a stirred solution of TBS ether **6.45** (129 mg, 0.10 mmol) in DCM / EtOH (6 mL, 1:1) at  $-5^{\circ}C$  was added CSA (35 mg, 0.15 mmol). After 12 h, the reaction was quenched with sat. aq.  $NaHCO_3$  (8 mL) and extracted with EtOAc (4 X 15 mL). The dried extract ( $MgSO_4$ ) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 20-40-80% EtOAc / Hexanes, to give triol **6.46** (62 mg, 0.065 mmol, 65%) as a colorless oil:  $[\alpha]_D^{23} = -1.64$  ( $c$  0.61,  $CHCl_3$ ); IR (neat) 3420, 2955, 2877, 1715, 1458, 1253, 1022, 969, 742  $cm^{-1}$ ;  $^1H$  NMR (400 MHz,  $CDCl_3$ )  $\delta$  6.71 (t,  $J = 6.4$  Hz, 1H), 6.05 (s, 1H), 5.05 (m, 1H), 5.02 (s, 1H), 4.84 (s, 1H), 4.46 (m, 1H), 4.13-4.19 (m, 5H), 3.62-3.74 (m, 3H), 3.13 (d,  $J=22.8$  Hz, 1H), 2.91 (dd,  $J=18.4, 8.6$  Hz, 1H), 2.59-2.75 (m, 3H), 2.11-2.23 (m, 3H), 1.98 (dd,  $J=12.0, 8.0$  Hz, 1H), 1.59-1.91 (m, 13H), 1.43 (s, 3H), 1.31-1.39 (m, 1H), 1.36 (t,  $J=7.1$  Hz, 6H), 1.25 (d,  $J=6.2$  Hz, 3H), 0.96-1.03 (m, 18H), 0.89 (d,  $J=6.6$  Hz, 3H), 0.85 (d,  $J=6.7$  Hz, 3H), 0.59-0.70 (m, 12H);  $^{13}C$

NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  212.2, 201.6, 167.7, 145.0, 143.3, 140.8, 128.9, 124.0, 114.5, 81.2, 78.3, 75.4, 68.7, 65.5, 62.7, 62.6, 61.0, 47.7, 45.9, 45.0, 43.5, 43.4, 43.1, 41.8, 40.5, 39.7, 32.1, 29.7, 28.3, 27.8, 26.3, 26.0, 22.3, 20.9, 19.6, 18.3, 16.4, 16.3, 14.7, 12.4, 7.0, 6.8, 5.4, 5.2, 4.8; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{48}\text{H}_{91}\text{O}_{12}\text{Si}_2\text{PNa}$  ( $\text{M}+\text{Na}$ ) 969.5784, found 969.5720.



**TMS ether 6.47:** To a stirred solution of triol **6.46** (62 mg, 0.065 mmol) in DCM (2 mL) at rt was sequentially added DMAP (155 mg, 1.27 mmol) and  $\text{TMSCl}$  (66 mg, 80  $\mu\text{L}$ , 0.63 mmol). After 1 h, the reaction mixture was concentrated *in vacuo* and then diluted with MeOH (2 mL), followed by the addition of  $\text{Ba}(\text{OH})_2 \cdot 8\text{H}_2\text{O}$  (19 mg, 0.062 mmol). After another 10 min, the reaction was quenched with sat. aq.  $\text{NH}_4\text{Cl}$  (3 mL) and extracted with  $\text{Et}_2\text{O}$  (3 X 10 mL). The dried extract ( $\text{MgSO}_4$ ) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 40-60%  $\text{EtOAc}$  / Hexanes, to give ester **6.47** (37 mg, 0.034 mmol, 52% over 2 steps) as a colorless oil:  $[\alpha]_{\text{D}}^{23} = -29.7$  ( $c$  0.31,  $\text{CHCl}_3$ ); IR (neat) 3447, 2955, 2877, 1716, 1458, 1250, 1021, 841, 744  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  6.69 (t,  $J = 7.4$  Hz, 1H), 5.79 (s, 1H), 5.05 (m,

1H), 5.03 (s, 1H), 4.87 (s, 1H), 4.14-4.21 (m, 5H), 4.07 (d,  $J = 5.4$  Hz, 1H), 3.66-3.74 (m, 3H), 3.12 (d,  $J = 22.8$  Hz, 2H), 3.01 (dd,  $J = 18.5, 6.9$  Hz, 1H), 2.80 (dd,  $J = 18.3, 4.9$  Hz, 1H), 2.68 (t,  $J = 7.2$  Hz, 2H), 2.09-2.22 (m, 3H), 1.64-1.90 (m, 15H), 1.44 (s, 3H), 1.36 (t,  $J = 7.01$  Hz, 6H), 1.32-1.39 (m, 1H), 1.25 (d,  $J = 6.0$  Hz, 3H), 0.92-1.03 (m, 18H), 0.87 (d,  $J = 6.4$  Hz, 3H), 0.81 (d,  $J = 6.8$  Hz, 3H), 0.56-0.71 (m, 12H), 0.13 (s, 9H), 0.08 (s, 9H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  209.1, 201.5, 167.7, 144.9, 142.0, 140.5, 129.0, 125.5, 114.9, 80.7, 78.2, 77.9, 68.9, 65.2, 62.6, 62.5, 61.0, 49.2, 49.0, 46.2, 43.5, 43.4, 41.9, 40.7, 40.0, 31.2, 28.4, 27.8, 22.3, 20.7, 19.4, 16.4, 16.3, 14.9, 14.7, 7.1, 6.9, 5.2, 4.9, 2.4, 0.6; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{54}\text{H}_{107}\text{O}_{12}\text{Si}_4\text{PNa}$  ( $\text{M}+\text{Na}$ ) 1113.6475, found 1113.6493



**Aldol adducts 6.49 & 6.50: Method A (-100°C Conditions)** – To a stirred solution of methyl ketone **6.40** (574 mg, 1.17 mmol) in THF (6 mL) at -78°C was added  $\text{LDA}^2$  (1.38 mL, 1 M in THF). After 15 min, TMEDA (400 mg, 0.310 mL, 3.44 mmol) was added. After 5 min, the reaction was cooled to -100°C, followed by the addition of a pre-cooled (-100°C) solution of aldehyde **5.12** (310 mg, 0.78 mmol) in THF (6 mL) via *cannula* in one portion. After another 0.5 h, the reaction

was quenched with 1 M AcOH in THF (1.5 mL) at -100°C. The reaction mixture was then warmed up to rt, diluted with sat. aq. NH<sub>4</sub>Cl (10 mL) and extracted with ether (4 X 25 mL). The dried extract (MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 50% CH<sub>2</sub>Cl<sub>2</sub> / Hexanes - 2% EtOAc / Hexanes, to give aldol adduct **6.50** (405 mg, 0.45 mmol, 58%) and **6.49** (50 mg, 0.056 mmol, 7%) as colorless oils.

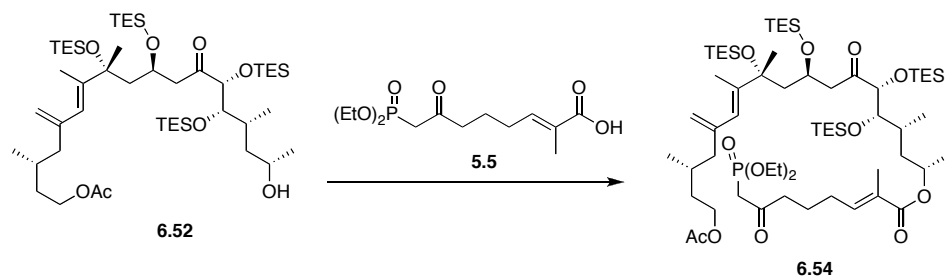
*Method B (-40°C Conditions)* – To a stirred solution of methyl ketone **6.40** (37.2 mg, 0.0758 mmol) in THF (0.4 mL) at -78°C was added LDA<sup>2</sup> (90 µL, 0.09 mmol, 1 M in THF). After 15 min, TMEDA (15.5 mg, 20 µL, 0.133 mmol) was added. After 5 min, the reaction was warmed up to -40°C, followed by the addition of a pre-cooled (-40°C) solution of aldehyde **5.12** (20 mg, 0.0504 mmol) in THF (0.4 mL) via *cannula* in one portion. After another 0.5 h, the reaction was quenched with sat. aq. NH<sub>4</sub>Cl (2 mL) and extracted with ether (4 X 5 mL). The dried extract (MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 50% CH<sub>2</sub>Cl<sub>2</sub> / Hexanes -2% EtOAc / Hexanes, to give aldol adduct **6.49** (16.1 mg, 0.0181 mmol, 36%) and **6.50** (13.4 mg, 0.0151 mmol, 30%) as colorless oils. **6.49**:  $[\alpha]_D^{23} = -9.42$  (*c* 1.21, CHCl<sub>3</sub>); IR (neat) 3516, 2956, 2913, 2877, 1743, 1719, 1458, 1414, 1370, 1249, 1116, 1088, 1008, 841, 742 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 5.89 (s, 1H), 5.02 (s, 1H), 4.83 (s, 1H), 4.38-4.27 (m, 1H), 4.06-4.10 (m, 3H), 3.86-3.75 (m, 1H), 3.72-3.68 (m, 1H), 3.66 (s, 1H, OH), 2.96 (dd, *J* = 17.9, 6.2 Hz, 1H), 2.59 (dd, *J* = 18.0, 6.1 Hz, 1H), 2.15 (dd, *J* = 13.2, 5.7 Hz, 1H), 2.04 (s, 3H), 1.94-1.76 (m, 4H), 1.81 (s, 3H), 1.70-1.63 (m,

2H), 1.54 (s, 3H), 1.48-1.38 (m, 2H), 1.27-1.22 (m, 1H), 1.13 (d,  $J = 5.9$  Hz, 3H), 1.02-0.93 (m, 27H), 0.89 (d,  $J = 6.3$  Hz, 3H), 0.79 (d,  $J = 6.6$  Hz, 3H), 0.59-0.69 (m, 18H), 0.09 (s, 9H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  210.8, 171.1, 144.3, 143.3, 124.8, 115.0, 81.0, 79.3, 78.4, 65.9, 65.0, 62.9, 48.2, 47.2, 45.8, 45.0, 35.3, 31.0, 28.7, 26.3, 24.6, 21.0, 19.3, 14.7, 13.8, 7.1, 7.0, 6.8, 6.6, 5.2, 4.9, 0.3; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{46}\text{H}_{94}\text{O}_8\text{Si}_4\text{Na}$  ( $\text{M}+\text{Na}$ ) 909.5924, found 909.5895. **6.50**:  $[\alpha]_{\text{D}}^{23} = +1.76$  ( $c$  1.25,  $\text{CHCl}_3$ ); IR (neat) 3511, 2956, 2913, 2877, 1743, 1718, 1458, 1369, 1249, 1119, 1088, 1011, 841, 742  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  6.06 (s, 1H), 5.03 (s, 1H), 4.88 (s, 1H), 4.17-4.06 (m, 4H), 3.88 (s, 1H, OH), 3.88-3.81 (m, 1H), 3.73-3.69 (m, 1H), 2.66-2.78 (m, 2H), 2.17 (dd,  $J = 13.5, 6.0$  Hz, 1H), 2.05 (s, 3H), 1.97-1.93 (m, 1H), 1.85-1.78 (m, 1H), 1.82(s, 3H), 1.74-1.70 (m, 1H), 1.62-1.14 (m, 4H), 1.44 (s, 3H), 1.28-1.20 (m, 2H), 1.14 (d,  $J = 5.8$  Hz, 3H), 1.02-0.94 (m, 27H), 0.91 (d,  $J = 6.2$  Hz, 3H), 0.78 (d,  $J = 6.6$  Hz, 3H), 0.72-0.57 (m, 18H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  210.5, 171.1, 144.5, 140.8, 125.4, 114.8, 81.3, 80.7, 78.4, 66.0, 65.7, 63.0, 47.1, 45.9, 45.0, 35.2, 31.0, 29.7, 28.7, 28.2, 24.6, 21.0, 19.4, 14.9, 13.6, 7.1, 7.0, 6.9, 6.8, 6.6, 5.2, 4.8, 0.3; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{46}\text{H}_{94}\text{O}_8\text{Si}_4\text{Na}$  ( $\text{M}+\text{Na}$ ) 909.5924, found 909.5948.



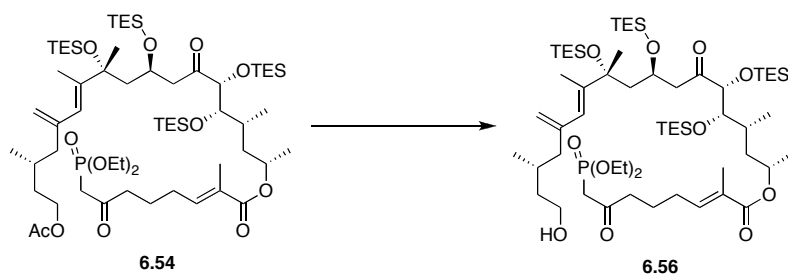
**TES ether 6.60:** To a stirred solution of aldol adduct **6.50** (440 mg, 0.496 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (20 mL) at rt was sequentially added DMAP (910 mg, 7.44 mmol) and TESCl (557 mg, 0.620 mL, 3.72 mmol). After 3 h, the reaction mixture was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 2% EtOAc / Hexanes, to give TES ether **6.60** (445 mg, 0.444 mmol, 90%) as a colorless oil:  $[\alpha]_D^{23} = -20.0$  (*c* 0.24, CHCl<sub>3</sub>); IR (neat) 2955, 2912, 2877, 1744, 1717, 1458, 1249, 1127, 1069, 1008, 840, 742 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  5.81 (s, 1H), 5.00 (s, 1H), 4.85 (s, 1H), 4.43-4.30 (m, 1H), 4.16-4.03 (m, 3H), 3.88-3.78 (m, 1H), 3.69-3.72 (m, 1H), 2.88 (dd, *J* = 17.4, 4.8 Hz, 1H), 2.73 (dd, *J* = 17.4, 7.2 Hz, 1H), 2.13 (dd, *J* = 13.8, 6.0 Hz, 1H), 2.04 (s, 3H), 1.82-1.94 (m, 3H), 1.79 (s, 3H), 1.64-1.75 (m, 3H), 1.46 (s, 3H), 1.39-1.51 (m, 2H), 1.20-1.28 (m, 1H), 1.14 (d, *J* = 6.0 Hz, 3H), 0.84-1.03 (m, 39H), 0.77 (d, *J* = 6.6 Hz, 3H), 0.56-0.71 (m, 24H), 0.11 (s, 9H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  208.4, 171.1, 144.3, 143.8, 124.3, 114.8, 81.1, 78.4, 77.3, 66.1 (2C), 62.9, 50.3, 48.7, 45.9, 45.2, 35.2, 30.7, 28.7, 26.9, 24.7, 21.0, 19.3, 14.8, 13.8, 7.2, 7.0, 6.9, 6.7, 5.4, 5.3, 5.0, 0.3; HRMS (ES<sup>+</sup>) calcd. for C<sub>52</sub>H<sub>108</sub>O<sub>8</sub>Si<sub>5</sub>Na (M+Na) 1023.6788, found 1023.6737.

**Alcohol 6.52:** To a stirred solution of TMS ether **6.60** (432 mg, 0.43 mmol) in THF / H<sub>2</sub>O (9 mL, 8:1) at -20°C was added HOAc (8 mL). After 5 h, the reaction was quenched with solid NaHCO<sub>3</sub>, filtered over Celite and extracted with ether (4 X 20 mL). The dried extract (MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 5-10-20% EtOAc / Hexanes, to give alcohol **6.52** (338 mg, 0.36 mmol, 85%) as a colorless oil:  $[\alpha]_D^{23} = -20.8$  (*c* 1.01, CHCl<sub>3</sub>); IR (neat) 3503, 2955, 2912, 2877, 1744, 1720, 1458, 1414, 1367, 1239, 1007, 741 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  5.85 (s, 1H), 5.02 (s, 1H), 4.86 (s, 1H), 4.50-4.40 (m, 1H), 4.18-4.03 (m, 3H), 3.85-3.77 (m, 1H), 3.67 (t, *J* = 5.0 Hz, 1H), 2.74-2.86 (m, 2H), 2.19 (br, OH), 2.14 (dd, *J* = 13.6, 6.3 Hz, 1H), 2.06 (s, 3H), 1.89-1.96 (m, 2H), 1.80 (s, 3H), 1.66-1.85 (m, 5H), 1.45 (s, 3H), 1.39-1.42 (m, 1H), 1.19 (d, *J* = 6.0 Hz, 3H), 1.10-1.22 (m, 1H), 0.95-1.05 (m, 36H), 0.90 (d, *J* = 6.4 Hz, 3H), 0.85 (d, *J* = 6.4 Hz, 3H), 0.59-0.72 (m, 24H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  208.4, 171.2, 144.3, 143.5, 124.4, 115.0, 82.1, 78.5, 77.3, 66.0, 65.9, 63.0, 50.0, 48.0, 45.9, 44.4, 35.2, 32.3, 28.7, 27.3, 24.2, 21.0, 19.3, 15.4, 15.0, 7.3, 7.1, 7.0, 6.9, 6.8, 5.3, 5.2, 4.9; HRMS (ES<sup>+</sup>) calcd. for C<sub>49</sub>H<sub>100</sub>O<sub>8</sub>Si<sub>4</sub>Na (M+Na) 951.6393, found 951.6418.



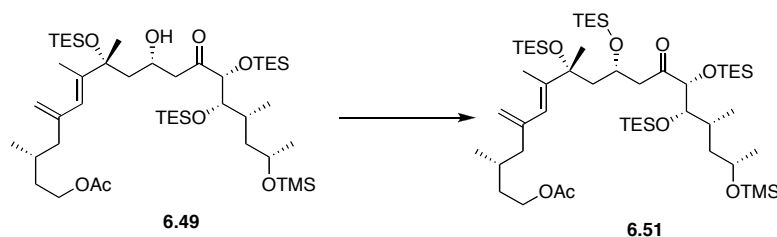
**Phosphonate 6.54:** To a stirred solution of acid **5.5** (837 mg, 2.73 mmol) in PhMe (6 mL) at rt was sequentially added Et<sub>3</sub>N (276 mg, 0.379 mL, 2.73 mmol) and 2, 4, 6-trichlorobenzoyl chloride (641 mg, 0.411 mL, 2.73 mmol). After 12 h, the resulted solution was concentrated *in vacuo*. DMAP (333 mg, 2.73 mmol) was added, followed by the addition of a solution of alcohol **6.52** (445 mg, 0.479 mmol) in PhMe (10.5 mL). After another 19 h, the reaction was quenched with sat. aq. NH<sub>4</sub>Cl (15 mL) and extracted with EtOAc (4 X 50 mL). The dried extract (MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 20-60% EtOAc / Hexanes, to give phosphonate **6.54** (450 mg, 0.100 mmol, 77%) as a colorless oil:  $[\alpha]_D^{23} = -20.3$  (*c* 1.23, CHCl<sub>3</sub>); IR (neat) 2955, 2913, 2877, 1740, 1716, 1458, 1368, 1243, 1056, 1019, 968, 742 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  6.69 (t, *J* = 6.4 Hz, 1H), 5.82 (s, 1H), 5.10-5.02 (m, 1H), 5.01 (s, 1H), 4.84 (s, 1H), 4.42-4.32 (m, 1H), 4.21-4.02 (m, 7H), 3.76-3.68 (m, 1H), 3.12 (d, *J* = 22.8 Hz, 2H), 2.85 (dd, *J* = 17.4, 4.1 Hz, 1H), 2.65-2.76 (m, 3H), 2.12-2.22 (m, 2H), 2.10-2.04 (m, 1H), 2.05 (s, 3H), 1.58-1.93 (m, 10H), 1.82 (s, 3H), 1.79 (s, 3H), 1.45 (s, 3H), 1.43-1.39 (m, 1H), 1.35 (t, *J* = 7.1 Hz, 6H), 1.24 (d, *J* = 5.9 Hz, 3H), 0.87-1.03 (m, 39H), 0.79 (d, *J* = 6.6 Hz, 3H), 0.55-0.68 (m, 24H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  208.3, 201.4, 171.1, 167.6, 144.3, 143.7,

140.5, 129.0, 124.4, 114.9, 81.1, 77.9, 77.34, 68.8, 66.2, 62.9, 62.6, 62.5, 50.3, 49.0, 45.9, 43.3, 41.6, 40.8, 35.2, 31.1, 28.7, 27.7, 26.9, 22.3, 21.0, 20.7, 19.3, 16.3, 16.2, 14.8, 14.3, 12.4, 7.2, 7.1, 7.0, 6.9, 6.7, 5.3, 5.2, 5.0; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{62}\text{H}_{121}\text{O}_{13}\text{Si}_4\text{PNa}$  ( $\text{M}+\text{Na}$ ) 1239.7520, found 1239.7563.



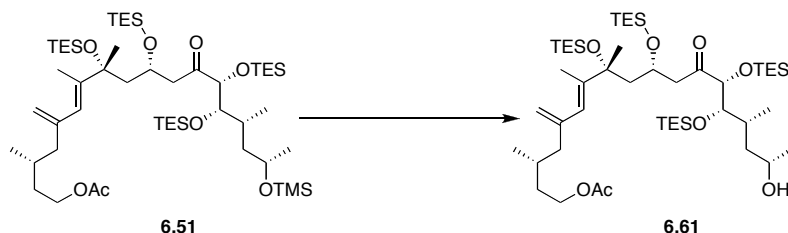
**Alcohol 6.56:** To a stirred solution of ester **6.54** (170 mg, 0.140 mmol) in MeOH (0.5 mL) at rt was added a saturated solution of  $\text{Ba}(\text{OH})_2 \cdot 8\text{H}_2\text{O}$  in MeOH (6.0 mL). After 20 min, the reaction mixture was purified directly by chromatography over silica gel, eluting with 20-60% EtOAc / Hexanes, to give alcohol **6.56** (150 mg, 0.127 mmol, 91%) as a colorless oil:  $[\alpha]_{\text{D}}^{23} = -20.3$  ( $c$  0.60,  $\text{CHCl}_3$ ); IR (neat) 3440, 2955, 2877, 1716, 1458, 1242, 1019, 969, 742  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):  $\delta$  6.69 (t,  $J = 6.2$  Hz, 1H), 5.82 (s, 1H), 5.10-5.03 (m, 1H), 5.00 (s, 1H), 4.83 (s, 1H), 4.42-4.31 (m, 1H), 4.20-4.08 (m, 5H), 3.63-3.74 (m, 3H), 3.12 (d,  $J = 22.8$  Hz, 2H), 2.78-2.72 (m, 2H), 2.66 (t,  $J = 7.1$  Hz, 2H), 2.04-2.19 (m, 3H), 1.89-1.61 (m, 10H), 1.81 (s, 3H), 1.78 (s, 3H), 1.44 (s, 3H), 1.43-1.38 (m, 1H), 1.35 (t,  $J = 7.1$  Hz, 6H), 1.24 (d,  $J = 5.9$  Hz, 3H), 1.03-0.92 (m, 36H), 0.87 (d,  $J = 6.4$  Hz, 3H), 0.79 (d,  $J = 6.6$  Hz, 3H), 0.55-0.71 (m, 24H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  208.5, 201.5, 167.7, 144.6, 143.5, 140.6, 129.0, 124.7,

114.7, 81.2, 78.0, 68.8, 66.1, 62.6, 62.5, 61.1, 50.3, 49.1, 46.1, 43.5, 43.4, 41.8, 40.9, 39.8, 31.1, 28.5, 27.8, 27.1, 22.3, 20.8, 19.5, 16.4, 16.3, 14.9, 14.2, 12.4, 7.3, 7.1, 7.0, 6.9, 6.8, 5.3, 5.2, 5.0; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{60}\text{H}_{19}\text{O}_{12}\text{Si}_4\text{PNa}$  ( $\text{M}+\text{Na}$ ) 1197.7414, found 1197.7423.



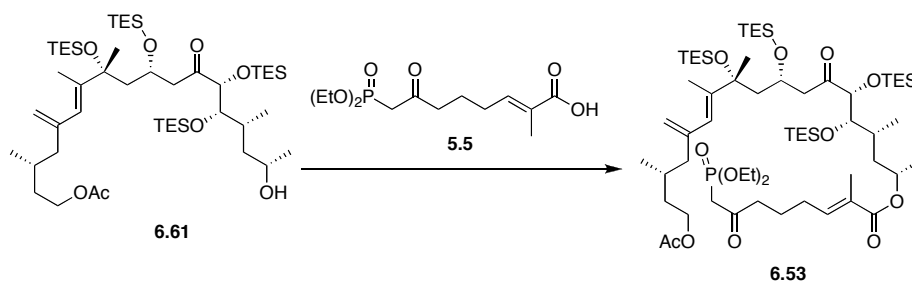
**TES ether 6.51:** To a stirred solution of aldol adduct **6.49** (295 mg, 0.332 mmol) in  $\text{CH}_2\text{Cl}_2$  (10 mL) at rt was sequentially added DMAP (608 mg, 4.98 mmol) and TESCl (375 mg, 0.418 mL, 2.49 mmol). After 3 h, the reaction mixture was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 2% EtOAc / Hexanes, to give TES ether **6.51** (290 mg, 0.289 mmol, 87%) as a colorless oil:  $[\alpha]_{\text{D}}^{23} = -31.4$  ( $c$  0.85,  $\text{CHCl}_3$ ); IR (neat) 2955, 2913, 2877, 1745, 1718, 1459, 1368, 1249, 1127, 1086, 1007, 841, 742  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  5.78 (s, 1H), 5.03 (s, 1H), 4.92 (s, 1H), 4.20-4.08 (m, 3H), 4.04 (d,  $J = 5.7$  Hz, 1H), 3.84-3.81 (m, 1H), 3.72-3.68 (m, 1H), 2.86 (dd,  $J = 18.0, 5.8$  Hz, 1H), 2.72 (dd,  $J = 17.4, 7.2$  Hz, 1H), 2.17 (dd,  $J = 13.8, 6.0$  Hz, 1H), 2.05 (s, 3H), 1.93-1.64 (m, 5H), 1.84 (s, 3H), 1.54-1.39 (m, 3H), 1.45 (s, 3H), 1.27-1.21 (m, 1H), 1.14 (d,  $J = 6.0$  Hz, 3H), 1.03-0.87 (m, 39H), 0.78 (d,  $J = 6.7$  Hz, 3H), 0.71-0.55 (m, 24H), 0.11 (s, 9H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  208.6, 171.1, 144.3, 143.0, 125.0, 115.1, 81.0, 78.3, 77.7, 66.1, 65.7, 63.0, 49.8, 48.5, 46.0, 44.9, 35.4, 31.0,

28.6, 27.9, 24.6, 21.0, 19.2, 14.5, 14.2, 7.2, 7.0, 6.9, 6.8, 5.2, 4.9, 0.4; HRMS (ES<sup>+</sup>) calcd. for C<sub>52</sub>H<sub>108</sub>O<sub>8</sub>Si<sub>5</sub>Na (M+Na) 1023.6788, found 1023.6785.



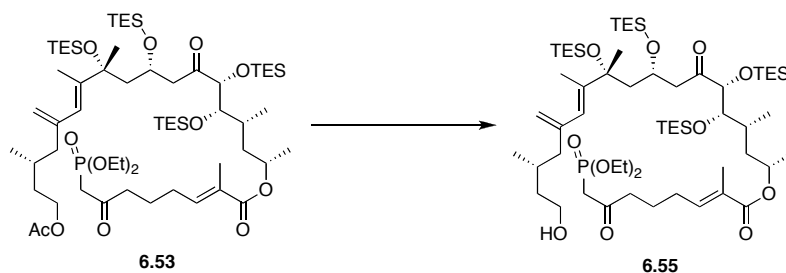
**Alcohol 6.61:** To a stirred solution of TMS ether **6.51** (290 mg, 0.289 mmol) in THF / H<sub>2</sub>O (6.52 mL, 8:1) at -20°C was added HOAc (4 X 1.45 mL) in 4 portions every 60 min. After 5 h, the reaction was quenched with solid NaHCO<sub>3</sub>, filtered over Celite and extracted with ether (4 X 15 mL). The dried extract (MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 5-10% EtOAc / Hexanes, to give alcohol **6.61** (220 mg, 0.237 mmol, 82%) as a colorless oil:  $[\alpha]_D^{23} = -31.6$  (*c* 1.01, CHCl<sub>3</sub>); IR (neat) 3510, 2956, 2912, 2877, 1744, 1720, 1458, 1414, 1368, 1239, 1062, 1006, 741 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 5.78 (s, 1H), 5.02 (s, 1H), 4.90 (s, 1H), 4.22-4.08 (m, 4H), 3.81-3.77 (m, 1H), 3.69-3.66 (m, 1H), 2.94 (dd, *J* = 18.3, 6.1 Hz, 1H), 2.78 (dd, *J* = 18.3, 5.7 Hz, 1H), 2.15 (dd, *J* = 13.1, 5.7 Hz, 1H), 2.04 (s, 3H), 1.93-1.60 (m, 6H), 1.83 (s, 3H), 1.45-1.34 (m, 2H), 1.44 (s, 3H), 1.30-1.20 (m, 1H), 1.17 (d, *J* = 6.0 Hz, 3H), 1.04-0.87 (m, 39H), 0.82 (d, *J* = 6.0 Hz, 3H), 0.72-0.56 (m, 24H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 208.8, 171.2, 144.4, 142.6, 125.2, 115.1, 81.4, 78.1, 77.7, 66.2, 65.4, 63.1, 49.7, 49.0, 45.9, 44.2, 35.3, 32.5, 28.6, 28.0, 24.3, 21.0,

19.2, 15.5, 14.6, 7.2, 7.0, 6.8, 5.3, 5.2, 4.8; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{49}\text{H}_{100}\text{O}_8\text{Si}_4\text{Na}$  ( $\text{M}+\text{Na}$ ) 951.6393, found 951.6398.



**Phosphonate 6.53:** To a stirred solution of acid **5.5** (450 mg, 1.47 mmol) in PhMe (3.2 mL) at rt was sequentially added  $\text{Et}_3\text{N}$  (149 mg, 0.204 mL, 1.47 mmol) and 2, 4, 6-trichlorobenzoyl chloride (346 mg, 0.222 mL, 1.47 mmol). After 12 h, the resulted solution was concentrated *in vacuo*. DMAP (180 mg, 1.47 mmol) was added, followed by the addition of a solution of alcohol **6.61** (240 mg, 0.258 mmol) in PhMe (5.7mL). After another 19 h, the reaction was quenched with sat. aq.  $\text{NH}_4\text{Cl}$  (8 mL) and extracted with EtOAc (4 X 50 mL). The dried extract ( $\text{MgSO}_4$ ) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 20-60% EtOAc / Hexanes, to give phosphonate **6.53** (235 mg, 0.193 mmol, 78%) as a colorless oil:  $[\alpha]_{\text{D}}^{23} = -23.6$  ( $c$  0.83,  $\text{CHCl}_3$ ); IR (neat) 2955, 2912, 2877, 1734, 1716, 1458, 1369, 1241, 1056, 1019, 970, 741  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  6.69 (t,  $J$  = 6.3 Hz, 1H), 5.79 (s, 1H), 5.10-5.00 (m, 1H), 5.03 (s, 1H), 4.92 (s, 1H), 4.22-4.08 (m, 8H), 3.72 (dd,  $J$  = 5.1, 3.6 Hz, 1H), 3.13 (d,  $J$  = 27.8 Hz, 2H), 2.90 (dd,  $J$  = 18.0, 5.9 Hz, 1H), 2.77 (dd,  $J$  = 17.8, 6.0

Hz, 1H), 2.68 (t,  $J = 7.2$  Hz, 2H), 2.18-2.07 (m, 3H), 2.06 (s, 3H), 1.94-1.68 (m, 9H), 1.842 (s, 3H), 1.82 (s, 3H), 1.46 (s, 3H), 1.43-1.39 (m, 2H), 1.36 (t,  $J = 7.0$  Hz, 6H), 1.25 (d,  $J = 6.0$  Hz, 3H), 1.03-0.92 (m, 36H), 0.90 (d,  $J = 6.4$  Hz, 3H), 0.80 (d,  $J = 6.7$  Hz, 3H), 0.70-0.55 (m, 24H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  208.4, 201.3, 171.1, 167.6, 144.3, 142.7, 140.5, 127.9, 125.1, 115.1, 80.9, 77.8, 77.6, 68.8, 65.6, 63.0, 62.9, 62.8, 48.7, 49.0, 45.9, 43.4, 41.7, 40.6, 35.3, 31.4, 28.6, 28.0, 27.7, 22.3, 21.0, 20.7, 19.2, 16.3, 16.2, 14.7, 14.5, 12.4, 7.2, 7.0, 6.9, 6.8, 5.3, 5.2, 4.9; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{62}\text{H}_{121}\text{O}_{13}\text{Si}_4\text{PNa}$  ( $\text{M}+\text{Na}$ ) 1239.7520, found 1239.7458.



**Alcohol 6.55:** To a stirred solution of ester **6.53** (230 mg, 0.189 mmol) in MeOH (10 mL) at rt was added  $\text{Ba}(\text{OH})_2 \cdot 8\text{H}_2\text{O}$  (66.4 mg, 0.189 mmol). After 1 h, the reaction mixture was purified directly by chromatography over silica gel, eluting with 20-60% EtOAc / Hexanes, to give alcohol **6.55** (204 mg, 0.168 mmol, 89%) as a colorless oil:  $[\alpha]_{\text{D}}^{23} = -27.0$  ( $c$  0.80,  $\text{CHCl}_3$ ); IR (neat) 3434, 2955, 2877, 1716, 1458, 1376, 1242, 1019, 742  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):  $\delta$  6.68 (t,  $J = 6.8$  Hz, 1H), 5.78 (s, 1H), 5.11-5.00 (m, 1H), 5.00 (s, 1H), 4.86 (s, 1H),



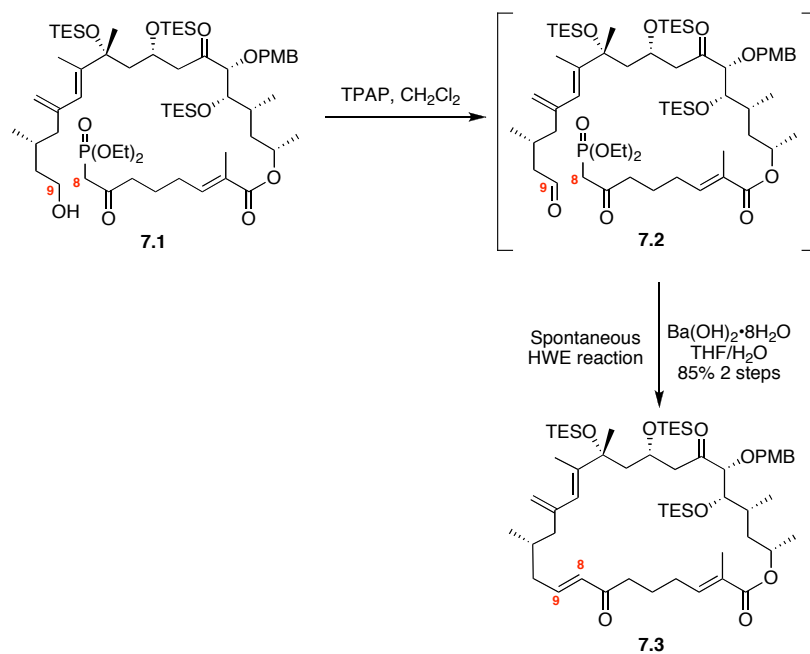
4.19-4.05 (m, 6H), 3.73-3.66 (m, 3H), 3.07 (d,  $J = 27.8$  Hz, 2H), 2.93 (dd,  $J = 18.0$ , 6.1 Hz, 1H), 2.78 (dd,  $J = 18.3$ , 5.7 Hz, 1H), 2.65 (t,  $J = 7.1$  Hz, 2H), 2.22-2.11 (m, 3H), 1.92-1.59 (m, 8H), 1.81 (s, 6H), 1.43 (s, 3H), 1.42-1.39 (m, 3H), 1.33 (t,  $J = 7.0$  Hz, 6H), 1.23 (d,  $J = 6.3$  Hz, 3H), 1.02-0.85 (m, 39H), 0.78 (d,  $J = 6.5$  Hz, 3H), 0.69-0.52 (m, 24H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  209.0, 201.4, 167.6, 144.8, 142.3, 140.5, 129.0, 125.5, 114.9, 80.8, 77.8, 77.7, 68.8, 65.6, 62.6, 62.5, 61.0, 49.5, 49.0, 46.2, 43.3, 41.6, 40.6, 40.0, 31.4, 28.3, 28.1, 27.7, 22.3, 20.7, 19.3, 16.3, 16.2, 14.7 (2C), 12.4, 7.2, 7.0, 6.9, 6.8, 5.2, 5.1, 4.9; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{60}\text{H}_{119}\text{O}_{12}\text{Si}_4\text{PNa}(\text{M}+\text{Na})$  1197.7414, found 1197.7422.

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1. (a) Zhang, W.; Carter, R. G. *Org. Lett.* **2005**, 7, 4209. (b) Zhang, W. Ph.D. Dissertation, Oregon State University, 2006.
  2. Preparation of LDA Solution: To a solution of diisopropylamine (101.9 mg, 0.14 mL, 1.0 mmol) in THF (0.46 mL) at  $-78^\circ\text{C}$  was added *n*-BuLi (0.4 mL, 1.0 mmol, 2.5 M in THF). After 5 min, the white slurry was warmed to  $-10^\circ\text{C}$  and stirred for an additional 15 min.
  3. Zhang, W.; Carter, R. G.; Yokochi, A. F. T. *J. Org. Chem.* **2004**, 69, 2569.

## CHAPTER 7. COMPLETION OF THE SYNTHESIS

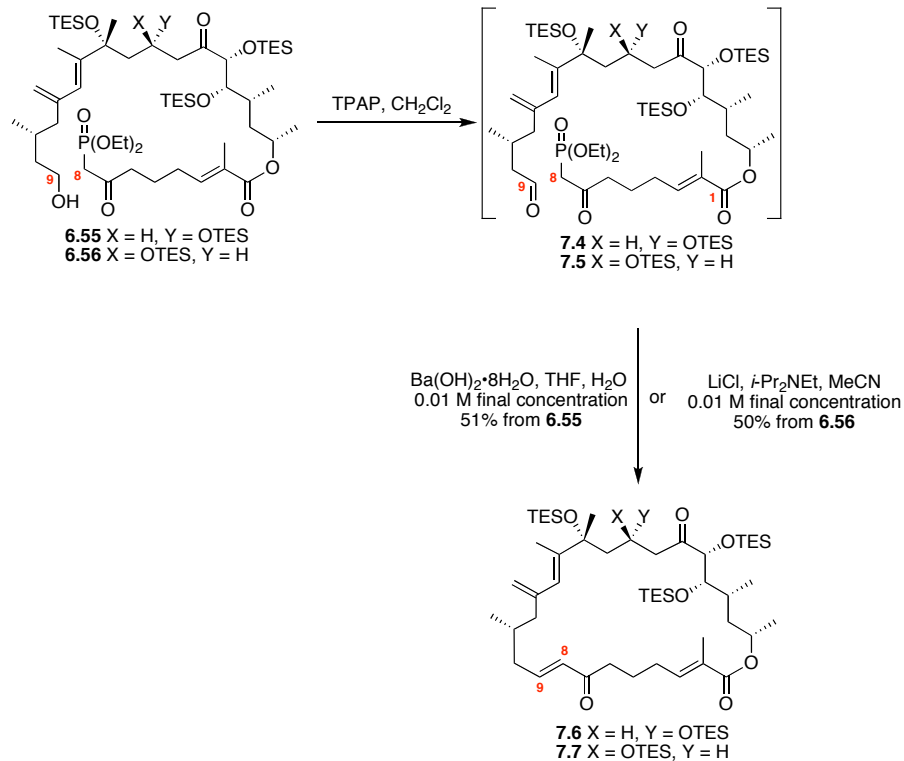
### 7.1 Macrocyclization

Once the synthesis of the phosphonate alcohol was accomplished, our sights were focused on the key macrocyclization. In our 1<sup>st</sup> generation synthesis of amphidinolide B,<sup>1</sup> we had developed the first successful macrocyclization of this natural product via a spontaneous intramolecular Horner-Wadsworth-Emmons olefination. The cyclization was driven to completion by the addition of  $\text{Ba}(\text{OH})_2 \cdot 8\text{H}_2\text{O}$  to give macrocycle **7.3** in good yield.

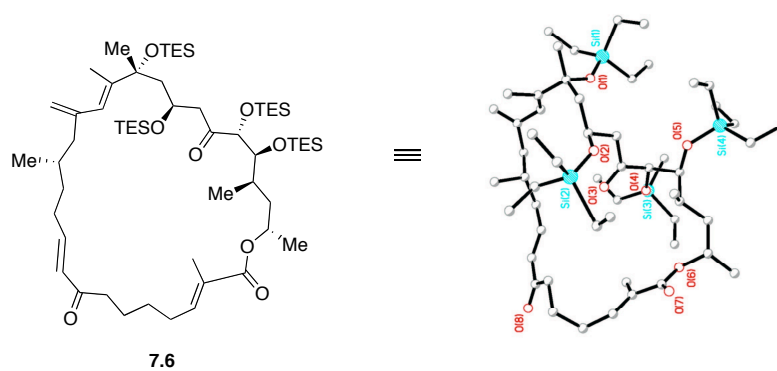


**Scheme 7.1.** Our Developed Strategy for the Macrocyclization

Equipped with the knowledge gained from our previous research, we applied the similar conditions on phosphonate alcohol **6.56**. Gratifyingly, significant amounts of the macrocycle **7.7** formed during the TPAP oxidation, appeared to undergo spontaneous intramolecular Horner-Wadsworth-Emmons olefination to provide the desired macrocycle. The conversion could be driven to completion by the addition of LiCl and Hunig's base.<sup>2</sup> A similar sequence was followed for construction of the 18*S* macrocycle **7.6**. In this case, Ba(OH)<sub>2</sub> proved more effective for driving the macrocyclization to completion. Additionally, we were pleased to observe that macrocycle **7.6** crystallized upon standing - allowing us to confirm the stereochemistry in the 26-membered macrocycle (Figure 7.1).



**Scheme 7.2.** Synthesis of Macrocycle **7.6** and **7.7**

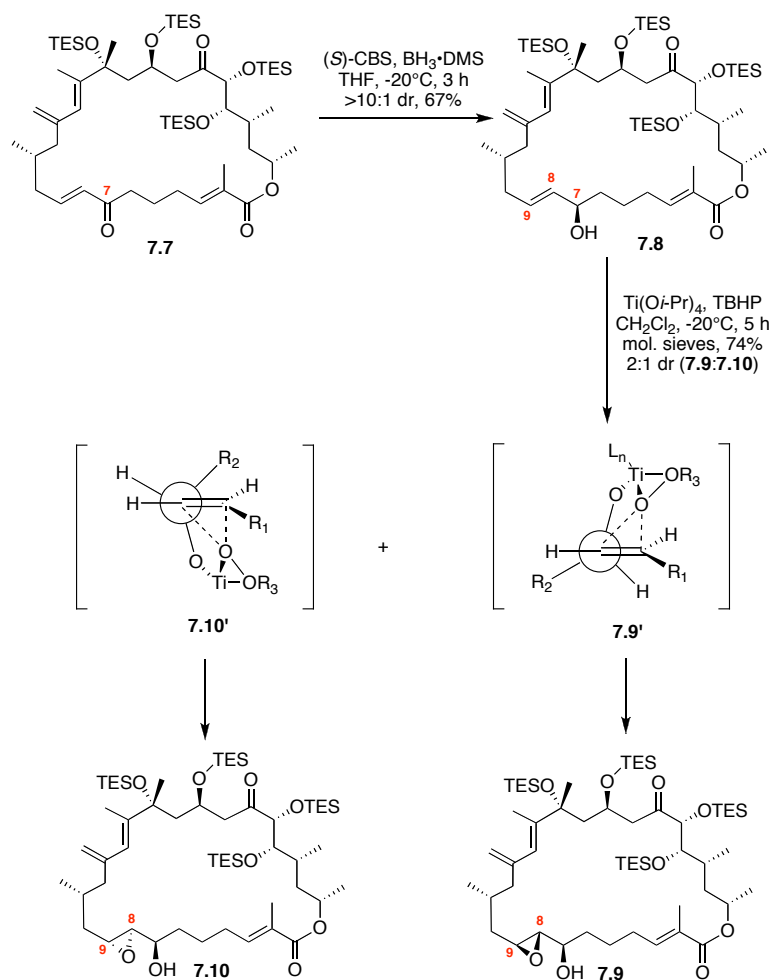


**Figure 7.1.** ORTEP Representation of Macrocycle **7.6**

## 7.2 Epoxidation of C<sub>8,9</sub> Alkene

With an efficient route into macrocycles **7.6** and **7.7**, the final challenges that remained were the incorporation of the C<sub>6</sub>-C<sub>9</sub> allylic epoxide moiety and deprotection of silyl groups. We performed our initial explorations on the more readily available 18*R* macrolactone **7.7** (Scheme 7.3). Regio- and stereoselective reduction of the C<sub>7</sub> carbonyl functionality could be accomplished with the (*S*)-CBS reagent.<sup>3</sup> The possible reduction at the C<sub>20</sub> ketone was not observed, presumably due to the increased steric congestion caused by the C<sub>21</sub> stereocenter. We had next intended to epoxidize the alkene using Sharpless conditions;<sup>4</sup> however, the presumed steric congestion of the C<sub>7</sub> alcohol thwarted this approach. Walsh and co-workers have recently shown that the *threo* (*syn*) epoxy alcohol can be obtained from a Ti(*Oi*-Pr)<sub>4</sub> / TBHP system.<sup>5</sup> As proposed by Adam and co-workers,<sup>6</sup> the binding of the allylic alkoxide to the titanium peroxy complex favors a dihedral angle of 70-90°. In Walsh's work, this dihedral angle led to a modest (~2:1) preference for the *syn* diastereomer in the epoxidation of a chiral *E*-disubstituted allylic alcohol.<sup>5</sup> In contrast, a 40-50° dihedral angle for VO(acac)<sub>2</sub> / TBHP system resulted in a moderate diastereoselectivity (~1.8:1) favoring the *anti* epoxy alcohol.<sup>5</sup> We were gratified to find that a similar reactivity profile appeared to take place with our system. The epoxidation led to the formation of both the *syn* (from transition-state **7.9'**) and *anti* (from transition-state **7.10'**) diastereomers - favoring the *syn* stereochemistry (2:1 dr). It is worth noting that the relative stereochemical

assignments are based on literature precedent.<sup>5</sup> We cannot at this time rigorously establish the relative stereochemistries of these two epoxy alcohols

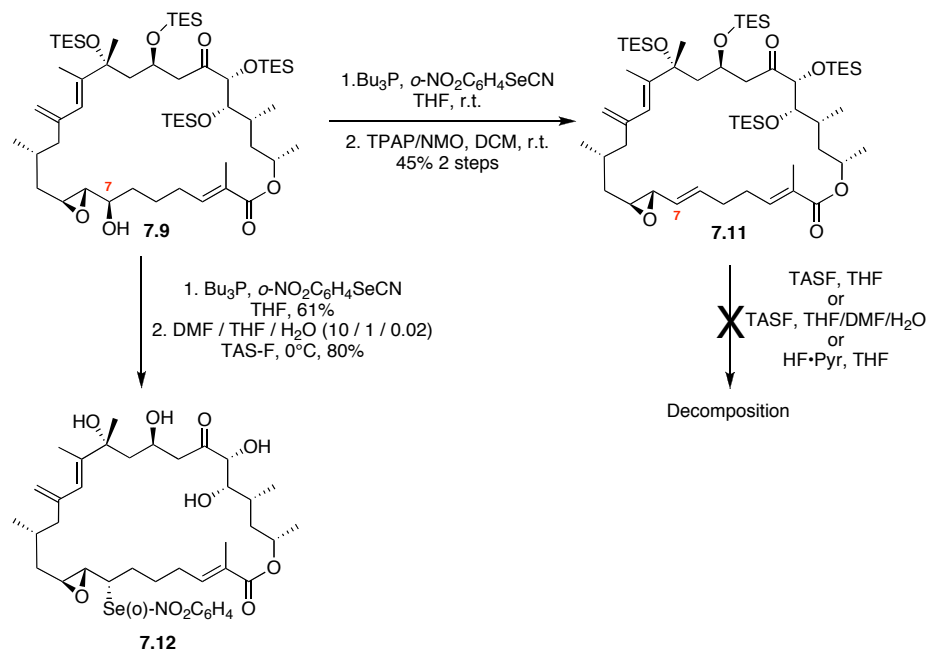


**Scheme 7.3.** Synthesis of Epoxy Alcohol **7.9** and **7.10**

### 7.3 Formation of C<sub>6,7</sub> Alkene and the Attempts to Remove TES Groups

As the *syn* diastereomer **7.9** contained the stereochemistry proposed for amphidinolide B<sub>2</sub>, we initially proceeded forward with that diastereomer (Scheme 7.4). Selenide incorporation using a large excess of *o*-nitrophenylselenium nitrile

and  $\text{PBu}_3$  (30 equivalent)<sup>7</sup> and subsequent elimination under our recently developed TPAP / NMO conditions<sup>8</sup> yielded the fully functionalized macrocycle **7.11**. Unfortunately, all attempts to remove the silyl protecting groups under fluoride or acidic conditions led to decomposition. We were surprised by these unexpected results since Fürstner and co-workers reported a successful desilylation using TAS-F<sup>9</sup> on a similar system in their recent synthesis of amphidinolide G and H.<sup>10</sup> Suspecting that the allylic epoxide might be the culpable functionality, we next explored global deprotection on the epoxy selenide. We were quite pleased to find that treatment of the epoxy selenide with TAS-F cleanly removed all silyl protecting groups to provide the polyol **7.12**.

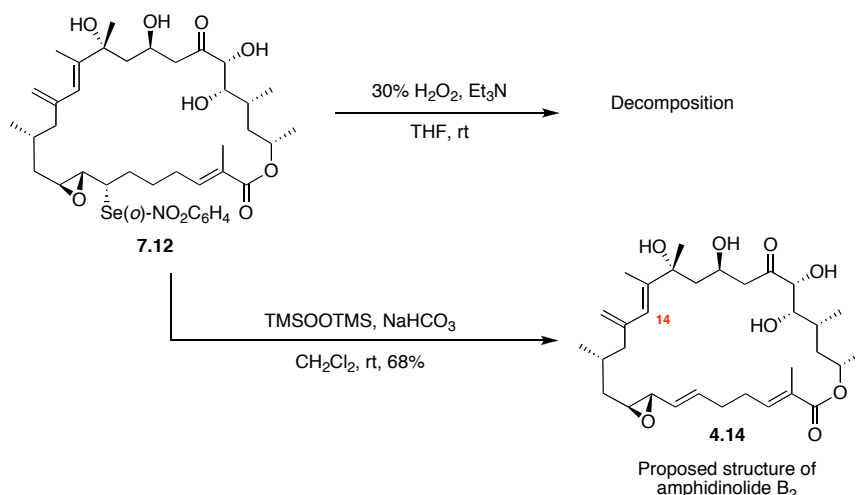


**Scheme 7.4.** Global Deprotection of TES Groups.

#### 7.4 Completion of the Proposed Structure of Amphidinolide B<sub>2</sub>

With the polyol in hand, the only challenge that remained was the oxidation and elimination of the selenide. The standard (H<sub>2</sub>O<sub>2</sub>) conditions<sup>11</sup> led to the decomposition of compound **7.12**. This issue was not completely surprising as we have previously encountered this problem in our azaspiracid work<sup>12</sup> as well as in an earlier generation approach to amphidinolide B.<sup>1</sup> We have attributed this deleterious reactivity to the  $\alpha$ -hydroxy ketone moiety or the C<sub>21,22</sub> diol structure. A H<sub>2</sub>O<sub>2</sub>-induced Baeyer-Villiger oxidation of the  $\alpha$ -hydroxy ketone or the oxidative cleavage of 1,2-diol to carboxylic acids might lead to the decomposition.<sup>13</sup> Our previously employed TPAP / NMO conditions are not compatible with the polyol functionality of **7.12**. A logical solution to this problem would be an alternative reagent that would not affect the  $\alpha$ -hydroxy ketone and 1,2-diol functionality. Bistrimethylsilylperoxide has been used as a replacement of H<sub>2</sub>O<sub>2</sub> in the metal catalyzed epoxidation of alkenes,<sup>14</sup> also has been employed to oxidize phosphonates to phosphates;<sup>15</sup> however, no precedents for the oxidation of a selenide have been reported. We were gratified to find that bistrimethylsilylperoxide (TMSOOTMS) cleanly facilitated the desired transformation to reveal compound **4.14**, the proposed structure of amphidinolide B<sub>2</sub>. Surprisingly, this synthetic product (**4.14**) *did not match* with the spectra data provided for the natural product amphidinolide B<sub>2</sub>.<sup>16</sup>

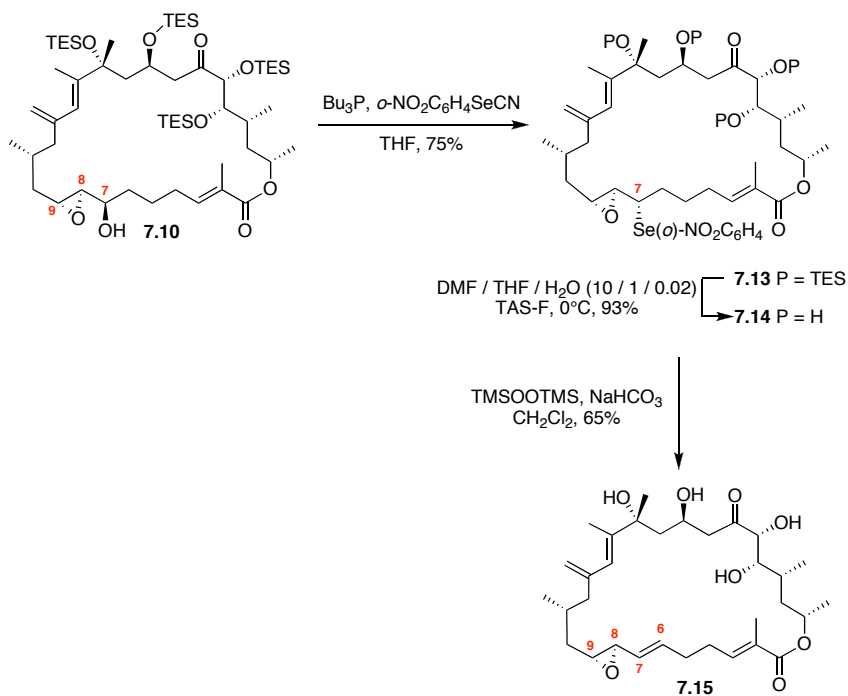




**Scheme 7.7.** Synthesis of The Proposed Structure of Amphidinolide B<sub>2</sub>.

### 7.5 Synthesis of C<sub>8,9</sub> Epoxide Diastereomer of Amphidinolide B<sub>2</sub>

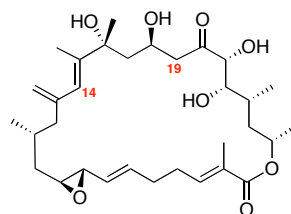
We followed a similar sequence on *anti*-epoxy alcohol **7.10** to afford C<sub>8,9</sub> epoxide diastereomer of amphidinolide B<sub>2</sub> (Scheme 7.5). Interestingly, the selenation had better yield (75% vs. 61%) and significantly shorter reaction time (30 min vs. 4 hours) compared to that of the *syn*-epoxy alcohol **7.9**. This observation was in agreement with our stereochemical assignments on epoxy alcohol **7.9** and **7.10**. In the formation of compound **7.13**, the nucleophilic attack from the selenide would not be hindered by the epoxide ring, while this effect would appear on substrate **7.9** due to the *syn*-epoxy alcohol relationship. Unfortunately, compound **7.15** also did not correlate with the reported data for amphidinolide B<sub>2</sub>.<sup>16</sup>



**Scheme 7.5.** Synthesis of C<sub>8,9</sub> Epoxide Diastereomer of Amphidinolide B<sub>2</sub>

## 7.6 Proposed Structure of Amphidinolide B<sub>2</sub>

Comparison of the  $^1\text{H}$  NMR data is shown in Table 7.1. The most significant differences are in the chemical shifts and coupling constants of H<sub>14</sub> and H<sub>19</sub>. In both cases, the  $^1\text{H}$  NMR shift for the H<sub>14</sub> alkene was shifted significantly downfield and the  $^1\text{H}$  NMR shift for H<sub>19b</sub> moved upfield as compared to the natural product data.

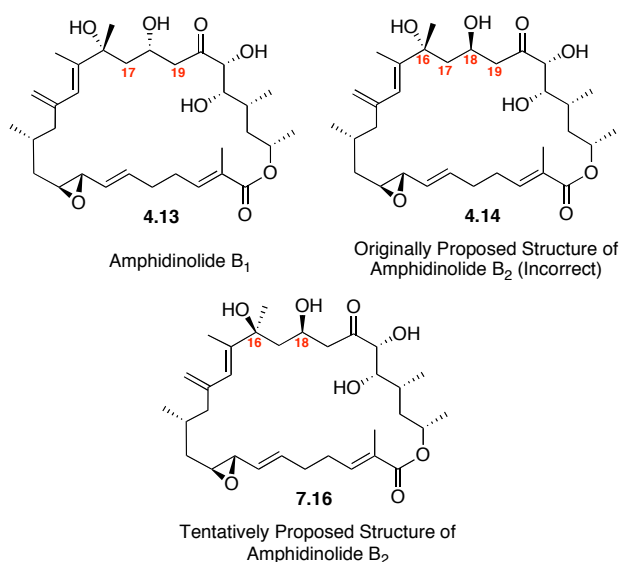


**4.14**  
Originally proposed structure of  
amphidinolide B<sub>2</sub>

Position	Natural amphidinolide B <sub>2</sub>	Synthesized amphidinolide B <sub>2</sub>	7.15
H <sub>14</sub>	5.93 ppm, br, s	6.06 ppm, s	6.08 ppm, s
H <sub>19a</sub>	3.09 ppm, dd <i>J</i> = 2.3, 8.8 Hz	3.05 ppm, m	2.90 ppm, dd <i>J</i> = 9.9, 17.1 Hz
H <sub>19b</sub>	2.63 ppm, dd <i>J</i> = 8.6, 17.7 Hz	2.48 ppm, dd <i>J</i> = 8.0, 17.0 Hz	2.45 ppm, m

**Table 7.1.** Comparison of the <sup>1</sup>H NMR Data

Careful inspection of the isolation paper revealed that the stereochemical analysis of amphidinolide B<sub>2</sub> was based primarily on the differences in the <sup>1</sup>H NMR in the C<sub>17</sub>-C<sub>19</sub> region of the natural product as compared to amphidinolide B<sub>1</sub> (**4.13**). It is important to note that Shimizu and Clardy<sup>16</sup> obtained X-ray crystallographic structure of natural product **4.13**. It is clear from our work that the structural differences between amphidinolide B<sub>1</sub> and B<sub>2</sub> are more complicated than initially expected. On the basis of this information, we have concluded that the proposed structure of amphidinolide B<sub>2</sub> is incorrect. Based on our tentative <sup>1</sup>H NMR data analysis, we suspect that the culprit stereochemistry is in fact the C<sub>16</sub> tertiary alcohol. We speculate that a common *syn* relationship is present between C<sub>16</sub> and C<sub>18</sub> in amphidinolide B<sub>2</sub> (Figure 7.2).

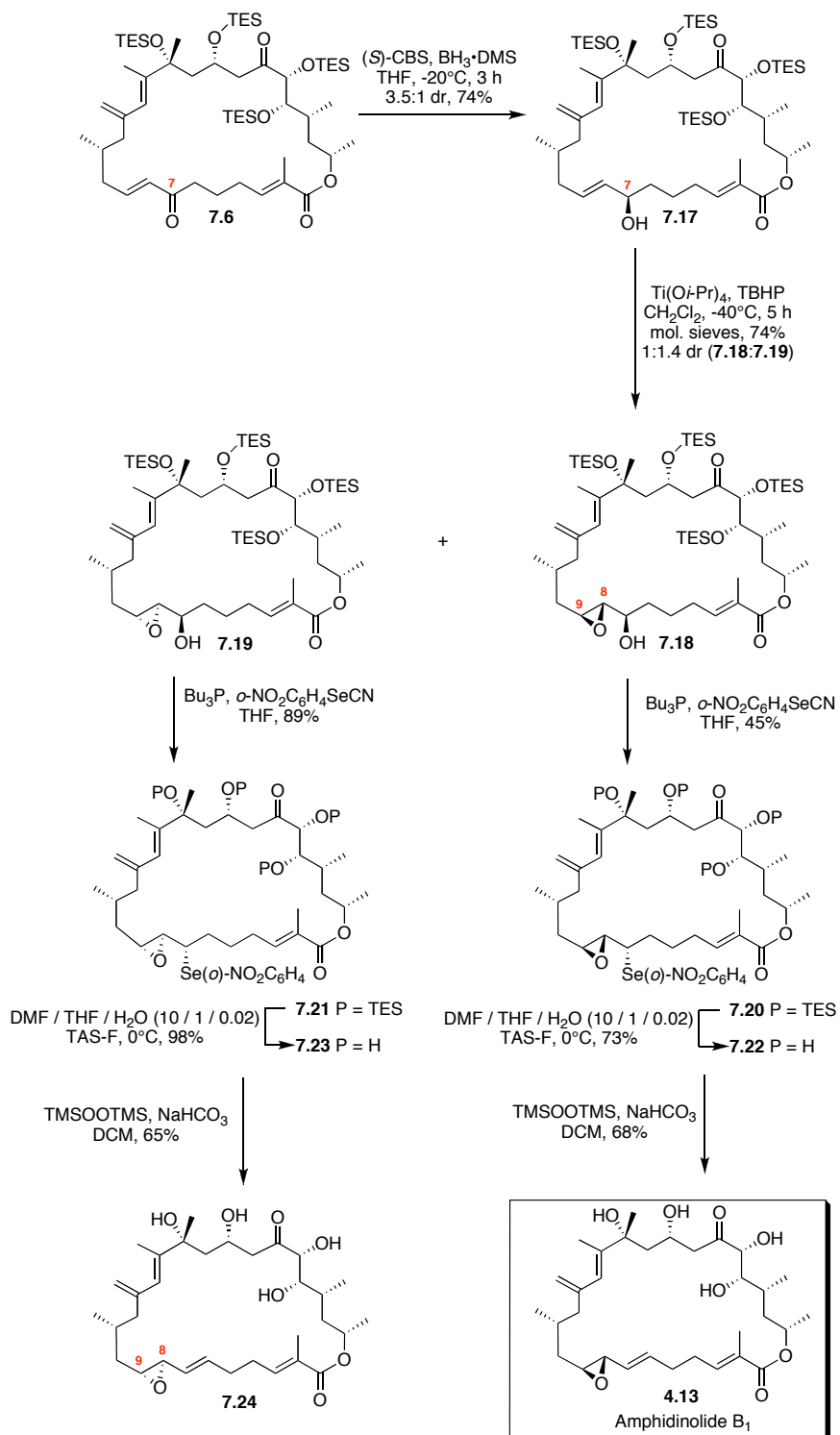


**Figure 7.2.** Tentatively Proposed Structure of Amphidinolide B<sub>2</sub>

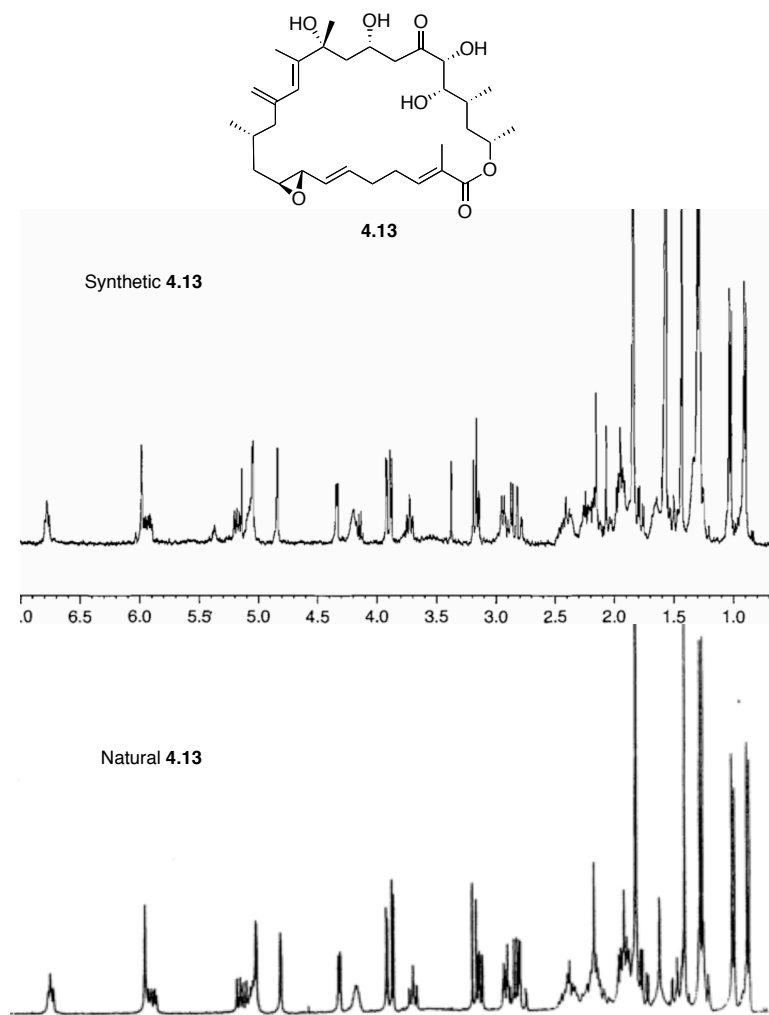
### 7.7 Completion of the Synthesis of Amphidinolide B<sub>1</sub>

Next, we shifted our focus to the total synthesis of amphidinolide B<sub>1</sub> (**4.13**) (Scheme 7.6). We applied an analogous strategy for the synthesis of **4.13** as was described for the 18*R* series. It appears that a slight reversal in selectivity in the epoxidation occurs with the 18*S* stereochemistry - now with a modest preference for the undesired C<sub>8,9</sub> epoxide, probably due to the geometry change of the macrocycle caused by the 18*S* stereocenter. This is supported spectroscopically by the downfield shift for C<sub>8</sub> & C<sub>9</sub> proton (5.52 & 5.65 ppm, respectively) in the <sup>1</sup>H NMR of the 18*S* allylic alcohol **7.17** compared to that (5.43 & 5.55 ppm, respectively) of its C<sub>18</sub> epimer **7.8**. Again, assignment of the relative

stereochemistries were based on literature precedent.<sup>5</sup> Fortunately, these diastereomers are chromatographically separable. Conversion of both epoxides to the selenides, followed by TAS-F deprotection yielded the penultimate intermediates. Finally, we were grateful to find that tandem selenide oxidation / elimination using our bis-TMS peroxide conditions yielded the natural product amphidinolide B<sub>1</sub> (**4.13**) and its C<sub>8,9</sub> epoxide diastereomer **7.27**. The synthesized material **4.13** matched with the spectra data reported by Kobayashi and co-workers for amphidinolide B<sub>1</sub> (Figure 7.3).<sup>17</sup>



**Scheme 7.6.** Synthesis of Amphidinolide B<sub>1</sub> and its C<sub>8,9</sub> Epoxide Isomer



**Figure 7.3.** Comparison of the <sup>1</sup>H NMR Data For the Synthetic and Natural Amphidinolide B<sub>1</sub> (4.13)

## 7.8 Conclusion

In summary, we have successfully cyclized the 26-membered macrocycle via an intramolecular Horner-Wadsworth-Emmons olefination and removed the TES protecting groups on the selenide moiety with TAS-F. We also developed the

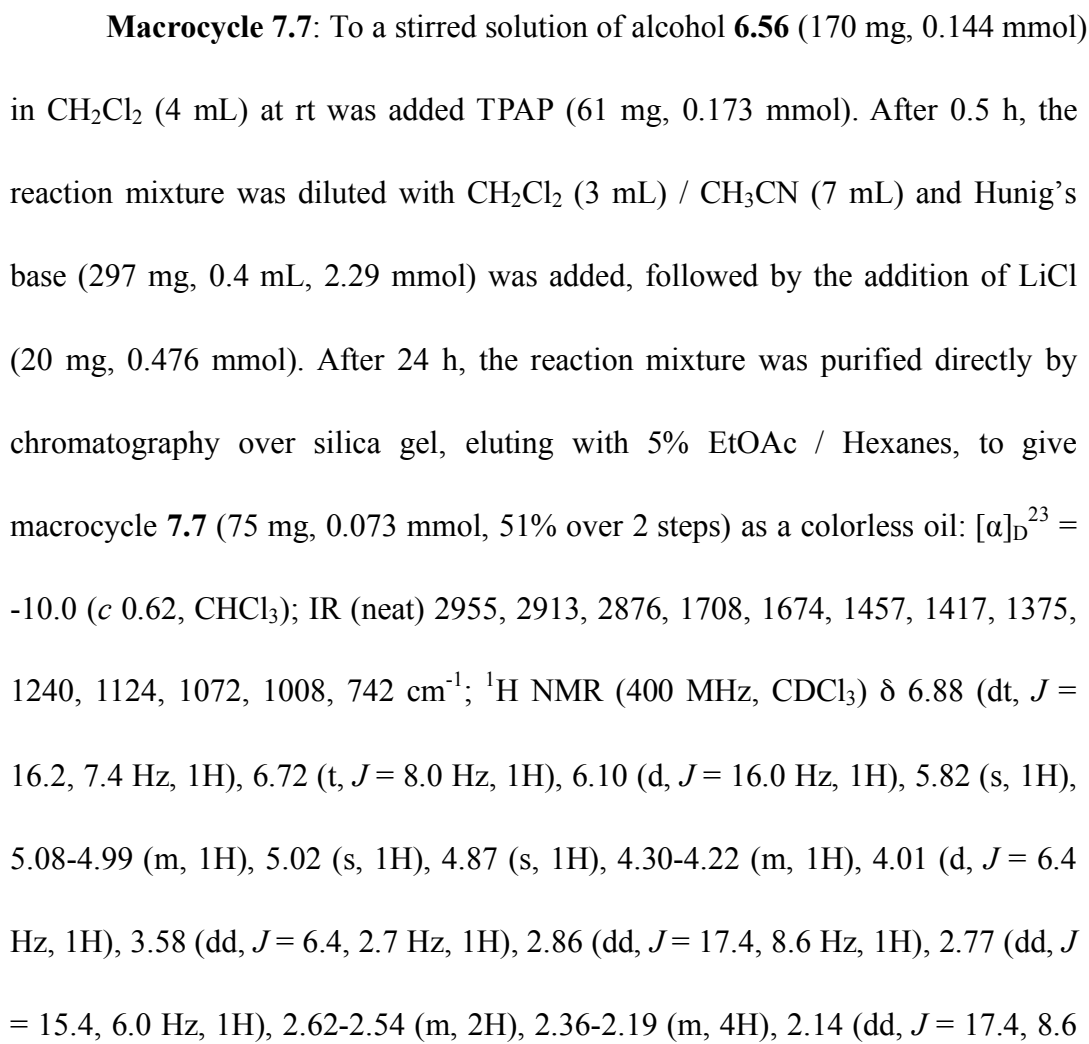
mild conditions for the oxidation and elimination of selenide using TMSOOTMS. The total syntheses of amphidinolide B<sub>1</sub> and the proposed structure of amphidinolide B<sub>2</sub> were finally accomplished with a longest linear sequence of 29 steps. The originally proposed structure of amphidinolide B<sub>2</sub> was found to be incorrect based on our careful analysis of the structural data.

## 7.9 References

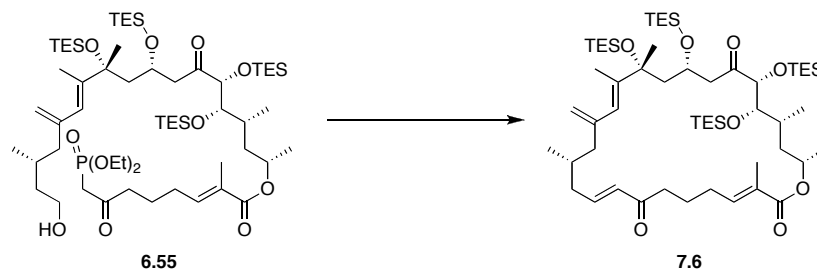
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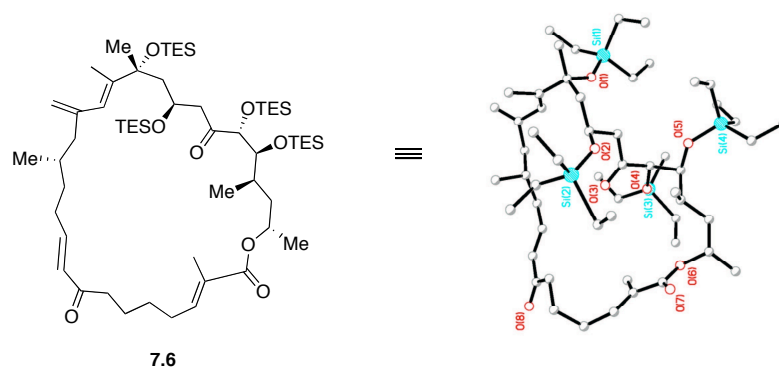


Hz, 1H), 2.05-1.76 (m, 7H), 1.80 (s, 3H), 1.79 (s, 3H), 1.70-1.60 (m, 1H), 1.57-1.51 (m, 1H), 1.41 (s, 3H), 1.25 (d,  $J = 6.2$  Hz, 3H), 1.05-0.93 (m, 36H), 0.84 (d,  $J = 6.6$  Hz, 3H), 0.76 (d,  $J = 6.6$  Hz, 3H), 0.73-0.58 (m, 24H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  208.1, 201.0, 167.6, 147.3, 147.1, 144.0, 142.8, 140.8, 132.4, 129.3, 124.7, 115.1, 80.6, 77.4 (2C), 68.5, 65.1, 50.6, 49.0, 46.1, 41.7, 40.2, 37.2, 31.3, 30.8, 27.8, 27.4, 23.1, 21.0, 19.7, 15.3, 12.8, 12.5, 7.3, 7.1, 7.0, 6.9, 5.3, 5.2, 4.9; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{56}\text{H}_{106}\text{O}_8\text{Si}_4\text{Na}$  ( $\text{M}+\text{Na}$ ) 1041.6863, found 1041.6824.

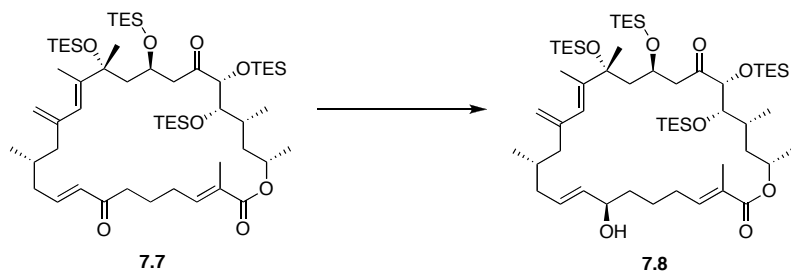


**Macrocycle 7.6:** To a stirred solution of alcohol **6.55** (125 mg, 0.106 mmol) in  $\text{CH}_2\text{Cl}_2$  (4 mL) at rt was added TPAP (45 mg, 0.127 mmol). After 0.5 h, the reaction mixture was diluted with THF (6.5 mL) /  $\text{H}_2\text{O}$  (16  $\mu\text{L}$ ) and  $\text{Ba}(\text{OH})_2 \cdot 8\text{H}_2\text{O}$  (3 x 74 mg, 0.636 mmol) was added in 3 portions every 30 min. After another 2 h, the reaction mixture was purified directly by chromatography over silica gel, eluting with 5% EtOAc / Hexanes, to give macrocycle **7.6** (54 mg,

0.053 mmol, 50% over 2 steps) as colorless crystals:  $[\alpha]_D^{23} = -27.0$  ( $c$  0.40,  $\text{CHCl}_3$ ); IR (neat) 2955, 2925, 2876, 1727, 1708, 1675, 1458, 1417, 1260, 1127, 1064, 1009, 741  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.15-7.03 (m, 1H), 6.73 (t,  $J = 6.9$  Hz, 1H), 6.19 (d,  $J = 16.3$  Hz, 1H), 5.78 (s, 1H), 5.00 (s, 1H), 5.05-4.97 (m, 1H), 4.82 (s, 1H), 4.18-4.09 (m, 1H), 4.11 (d,  $J = 5.1$  Hz, 1H), 3.62-3.58 (m, 1H), 3.00-2.93 (m, 2H), 2.85-2.70 (m, 1H), 2.62-2.50 (m, 1H), 2.37-2.21 (m, 3H), 2.15-2.00 (m, 2H), 1.91-1.63 (m, 7H), 1.80 (s, 6H), 1.45-1.35 (m, 2H), 1.41 (s, 3H), 1.25 (d,  $J = 6.1$  Hz, 3H), 1.08-0.83 (m, 39H), 0.75-0.47 (m, 27H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  206.2, 201.0, 167.6, 147.2, 144.7, 141.8, 140.6, 132.4, 129.2, 125.8, 115.4, 80.5, 79.6, 77.9, 68.3, 64.9, 49.7, 48.6, 46.7, 43.2, 41.1, 37.5, 31.0, 29.2, 28.9, 27.8, 22.6, 21.0, 18.8, 15.6, 13.3, 12.5, 7.3, 7.1, 7.0, 5.2, 4.8; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{56}\text{H}_{106}\text{O}_8\text{Si}_4\text{Na}$  ( $\text{M}+\text{Na}$ ) 1041.6863, found 1041.6812.

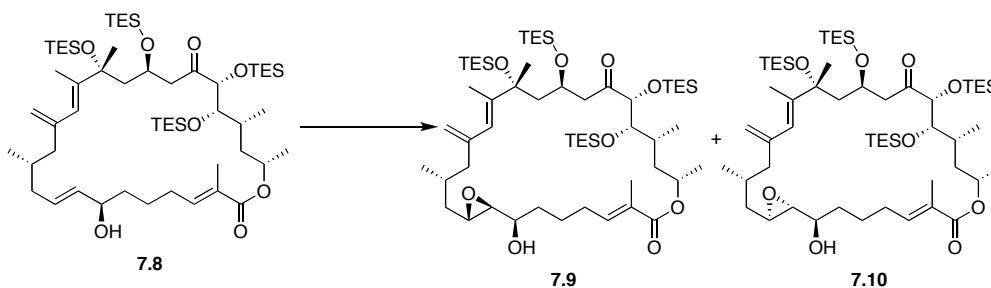


**Figure 1.** ORTEP Representation of macrocycle **7.6**



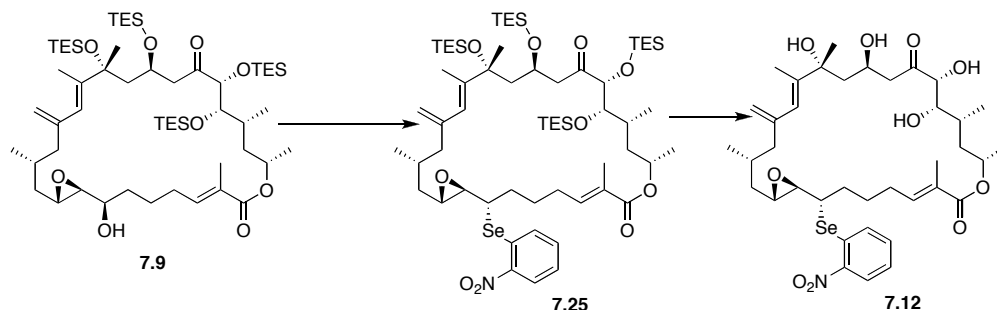
**Allylic alcohol 7.8:** To a stirred solution of macrocycle **7.7** (107 mg, 0.105 mmol) in  $\text{CH}_2\text{Cl}_2$  (5.4 mL) at  $-20^\circ\text{C}$  was sequentially added (*S*)-CBS (0.42 mL, 0.42 mmol, 1 M in PhMe) and  $\text{BH}_3\cdot\text{DMS}$  (0.84 mL, 0.84 mmol, 1 M in THF). After 45 min, the reaction was quenched with MeOH (0.3 mL), diluted with aq.  $\text{NaHCO}_3$  (5 mL) and extracted with  $\text{Et}_2\text{O}$  (3 x 8 mL). The dried extract ( $\text{MgSO}_4$ ) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 3-6% EtOAc / Hexanes, to give allylic alcohol **7.8** (72 mg, 0.0704 mmol, 67%) as a colorless oil:  $[\alpha]_{\text{D}}^{23} = -16.3$  ( $c$  0.30,  $\text{CHCl}_3$ ); IR (neat) 3431, 2954, 2913, 2876, 1708, 1674, 1458, 1414, 1376, 1241, 1128, 1073, 1009, 742  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  6.80 (t,  $J = 8.0$  Hz, 1H), 5.89 (s, 1H), 5.63-5.52 (m, 1H), 5.45 (dd,  $J = 15.4, 7.5$  Hz, 1H), 5.10-5.00 (m, 1H), 4.98 (s, 1H), 4.82 (s, 1H), 4.32-4.20 (m, 1H), 4.19-4.09 (m, 1H), 4.04 (d,  $J = 6.2$  Hz, 1H), 3.57 (dd,  $J = 6.2, 2.5$  Hz, 1H), 2.87 (dd,  $J = 17.0, 9.0$  Hz, 1H), 2.49 (d,  $J = 16.8$  Hz, 1H), 2.28-2.20 (m, 2H), 2.13-1.98 (m, 3H), 1.90-1.61 (m, 9H), 1.86 (s, 3H), 1.75 (s, 3H), 1.55-1.46 (m, 2H), 1.39 (s, 3H), 1.26 (d,  $J = 6.0$  Hz, 3H), 1.06-0.94 (m, 36H), 0.82 (d,  $J = 6.4$  Hz, 3H), 0.73-0.60 (m, 27H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  208.5,

167.8, 144.6, 142.0, 141.3, 134.6, 130.8, 128.2, 125.4, 114.4, 80.6, 78.0, 72.9, 68.4, 65.2, 50.3, 49.2, 45.4, 41.8, 39.7, 36.8, 31.3, 30.4, 28.8, 28.2, 24.0, 20.9, 19.6, 15.4, 12.5, 12.4, 7.3, 7.1, 7.0, 5.3, 5.2, 4.9; HRMS (ES<sup>+</sup>) calcd. for C<sub>56</sub>H<sub>108</sub>O<sub>8</sub>Si<sub>4</sub>Na (M+Na) 1043.7019, found 1043.7052



**Epoxide 7.9 & 7.10:** To a stirred solution of allylic alcohol **7.8** (70 mg, 0.0685 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (6 mL) at -20°C was sequentially added 4Å MS (50 mg), TBHP (37 µL, 0.206 mmol, 5.5 M in decane) and Ti(O-*i*Pr)<sub>4</sub> (23.3 mg, 24 µL, 0.082 mmol). After 5 h, the reaction was quenched with aq. NaHCO<sub>3</sub> (3 mL) and extracted with Et<sub>2</sub>O (3 x 7 mL). The dried extract (MgSO<sub>4</sub>) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 6-10% EtOAc / Hexanes, to give epoxide **7.9** (35 mg, 0.0342 mmol, 50%) and epoxide **7.10** (17 mg, 0.0166 mmol, 24%) as colorless oils. **7.9**: [α]<sub>D</sub><sup>23</sup> = -29.0 (c 0.42, CHCl<sub>3</sub>); IR (neat) 3431, 2954, 2923, 2876, 1708, 1647, 1458, 1414, 1377, 1242, 1128, 1073, 1009, 742 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 6.78 (t, *J* = 8.0 Hz, 1H), 5.88 (s, 1H), 5.05-4.98 (m, 1H), 5.01 (s, 1H), 4.86 (s, 1H), 4.30-4.20 (m, 1H), 4.05 (d, *J* = 6.1 Hz, 1H), 3.59 (dd, *J* = 6.1, 2.0 Hz, 1H), 3.52-3.43 (m, 1H), 3.22-3.15 (m, 1H),

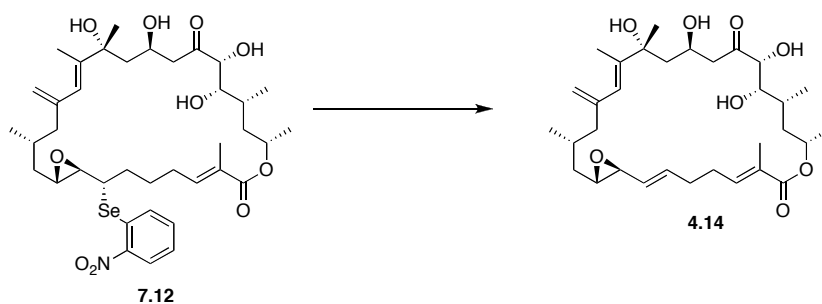
2.91 (dd,  $J=16.3, 9.1$  Hz, 1H), 2.70 (dd,  $J=6.0, 2.0$  Hz, 1H), 2.44-2.25 (m, 3H), 2.18 (dd,  $J=13.0, 5.6$  Hz, 1H), 2.10-1.63 (m, 12H), 1.87 (s, 3H), 1.78 (s, 3H), 1.41 (s, 3H), 1.26 (d,  $J=6.0$  Hz, 3H), 1.15-1.09 (m, 1H), 1.07-0.89 (m, 36H), 0.80 (d,  $J=6.4$  Hz, 3H), 0.76-0.59 (m, 27H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  208.4, 167.7, 144.0, 142.1, 141.4, 128.6, 125.0, 114.9, 80.8, 78.2, 77.5, 71.8, 68.3, 65.6, 62.8, 56.4, 50.6, 49.1, 46.6, 42.1, 38.9, 33.1, 30.1, 29.4, 28.8, 28.3, 23.7, 21.0, 19.8, 15.5, 12.5, 12.4, 7.3, 7.1, 7.0, 5.3, 5.2, 5.0; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{56}\text{H}_{108}\text{O}_9\text{Si}_4\text{Na}$  ( $\text{M}+\text{Na}$ ) 1059.6968, found 1059.7009. **7.10**:  $[\alpha]_{\text{D}} = -26.2$  ( $c$  0.60,  $\text{CHCl}_3$ ); IR (neat) 3482, 2954, 2923, 2876, 1708, 1458, 1414, 1377, 1240, 1128, 1008, 742  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  6.79 (t,  $J=7.7$  Hz, 1H), 5.87 (s, 1H), 5.05-4.98 (m, 1H), 4.99 (s, 1H), 4.84 (s, 1H), 4.30-4.20 (m, 1H), 4.09 (d,  $J=5.9$  Hz, 1H), 3.75-3.69 (m, 1H), 3.57 (dd,  $J=5.6, 3.2$  Hz, 1H), 2.91-2.78 (m, 3H), 2.68-2.60 (m, 2H), 2.38-2.20 (m, 3H), 2.07 (dd,  $J=12.9, 6.2$  Hz, 1H), 1.93-1.62 (m, 10H), 1.85 (s, 3H), 1.76 (s, 3H), 1.50-1.44 (m, 2H), 1.40 (s, 3H), 1.25 (d,  $J=6.0$  Hz, 3H), 1.05-0.93 (m, 39H), 0.74-0.59 (m, 27H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  208.6, 167.8, 144.1, 142.5, 141.5, 128.4, 124.9, 114.9, 81.0, 78.2, 77.4, 69.0, 68.4, 65.5, 60.7, 54.9, 50.3, 49.2, 46.3, 41.7, 38.4, 33.3, 30.7, 30.3, 29.0, 27.9, 23.7, 21.0, 20.0, 15.3, 13.2, 12.4, 7.3, 7.1, 7.0, 6.9, 5.3, 5.2, 4.9; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{56}\text{H}_{108}\text{O}_9\text{Si}_4\text{Na}$  ( $\text{M}+\text{Na}$ ) 1059.6968, found 1059.7009.



**Selenide 7.25:** To a stirred solution of epoxide **7.9** (42 mg, 0.0405 mmol) in THF (2 mL) at rt was sequentially added *o*-NO<sub>2</sub>C<sub>6</sub>H<sub>4</sub>SeCN (184 mg, 0.809 mmol) and PBu<sub>3</sub> (164 mg, 202  $\mu$ L, 0.809 mmol). After 5 h, the reaction mixture was concentrated *in vacuo* purified by chromatography over silica gel, eluting with Hexanes then with 4% EtOAc / Hexanes, to give crude selenide **7.25** (30 mg) as yellow oils which was used directly in next step without further purification.

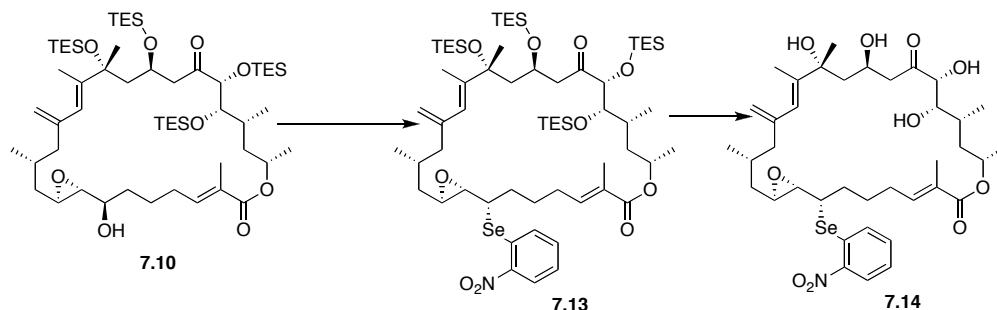
**Polyol 7.12:** To a stirred solution of selenide **7.25** (30 mg) in THF /DMF / H<sub>2</sub>O (10:1:0.02, 1.8 mL / 180  $\mu$ L / 3.6  $\mu$ L) at 0°C was added TAS-F (33.7 mg, 0.123 mmol). The reaction mixture was then warmed up to rt. After 2 h, the reaction mixture was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 10-65% EtOAc / Hexanes, to give polyol **7.12** (15 mg, 0.0196 mmol, 48% over 2 steps) as a yellow solid:  $[\alpha]_D^{23} = -20.1$  (*c* 0.12, CHCl<sub>3</sub>); IR (neat) 3447, 2925, 2854, 1701, 1520, 1456, 1334, 1273, 759, 732 cm<sup>-1</sup>; <sup>1</sup>H





**Proposed structure of Amphidinolide B<sub>2</sub> (4.14):** To a stirred solution of selenide **7.12** (6.0 mg, 0.00784 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (1.2 mL) at rt was sequentially

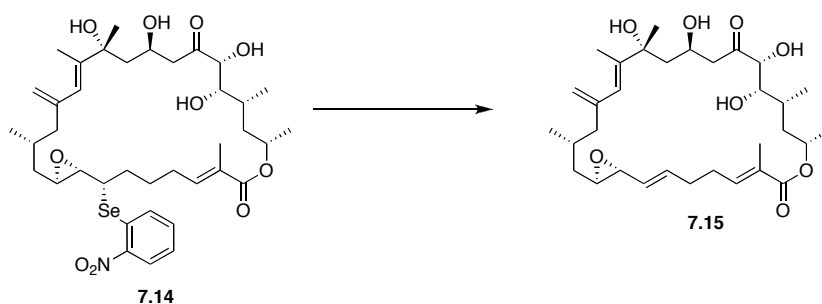
added NaHCO<sub>3</sub> (60 mg, 0.714 mmol) and TMSOOTMS (41.7 mg, 50  $\mu$ L, 0.233 mmol). After 1.5 h, the yellow color vanished and the reaction mixture was purified by preparative TLC, eluting with 60% EtOAc / Hexanes, to give allylic epoxide **4.14** (3.0 mg, 0.00533 mmol, 68%):  $[\alpha]_D^{23} = -52.3$  (*c* 0.21, CHCl<sub>3</sub>); IR (neat) 3446, 2923, 2853, 1701, 1457, 1273, 1120 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.74 (t, *J* = 6.4 Hz, 1H), 6.06 (s, 1H), 5.92 (ddd, *J* = 15.0, 8.9, 4.4 Hz, 1H), 5.20 (dd, *J* = 15.5, 8.8 Hz, 1H), 5.11-5.07 (m, 1H), 5.07 (s, 1H), 4.86 (s, 1H), 4.28 (d, *J* = 5.4, 1H), 4.14 (s, OH), 4.14-4.09 (m, 1H), 3.73 (d, *J* = 5.6 Hz, OH), 3.69 (t, *J* = 9.5 Hz, 1H), 3.50 (d, *J* = 6.8 Hz, 1H), 3.23 (dd, *J* = 8.7, 2.2 Hz, 1H), 3.08-3.03 (m, 2H), 2.95 (d, *J* = 9.2 Hz, OH), 2.53-2.45 (m, 1H), 2.45-2.36 (m, 1H), 2.28 (dd, *J* = 13.2, 1.8 Hz, 1H), 2.17-2.12 (m, 3H), 1.97-1.93 (m, 4H), 1.85 (s, 3H), 1.82-1.79 (m, 1H), 1.80 (s, 3H), 1.78-1.75 (m, 1H), 1.64-1.60 (m, 1H), 1.34 (s, 3H), 1.31 (d, *J* = 6.1 Hz, 3H), 1.17-1.12 (m, 1H), 1.03 (d, *J* = 6.7 Hz, 3H), 0.98 (d, *J* = 5.6 Hz, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  212.53, 167.68, 144.65, 141.59, 139.52, 136.29, 128.41, 128.34, 124.88, 114.59, 78.14, 75.58, 69.28, 68.23, 61.45, 59.5, 47.14, 46.41, 44.14, 39.98, 39.36, 33.25, 31.04, 29.32, 28.27, 26.69, 21.19, 17.52, 15.91, 15.17, 12.60; HRMS (ES<sup>+</sup>) calcd. for C<sub>32</sub>H<sub>50</sub>O<sub>8</sub>Na (M+Na) 585.3403, found 585.3390.



**Selenide 7.13:** To a stirred solution of epoxide **7.10** (12 mg, 0.0116 mmol) in THF (0.7 mL) at rt was sequentially added *o*-NO<sub>2</sub>C<sub>6</sub>H<sub>4</sub>SeCN (53 mg, 0.232 mmol) and PBu<sub>3</sub> (47 mg, 58  $\mu$ L, 0.232 mmol). After 0.5 h, the reaction mixture was concentrated *in vacuo* purified by chromatography over silica gel, eluting with Hexanes then with 4% EtOAc / Hexanes, to give crude selenide **7.13** (10.6 mg) as a yellow oil which was used directly in next step without further purification.

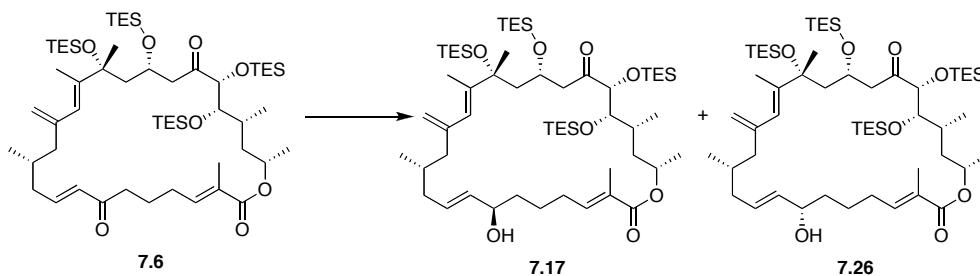
**Polyol 7.14:** To a stirred solution of selenide **7.13** (10.6 mg) in THF / DMF / H<sub>2</sub>O (10:1:0.02, 1.0 mL / 100  $\mu$ L / 2.0  $\mu$ L) at 0°C was added TAS-F (12 mg, 0.0434 mmol). The reaction mixture was then warmed up to rt. After 2 h, the reaction mixture was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 10-65% EtOAc / Hexanes, to give polyol **7.14** (6.2 mg, 0.00811 mmol, 70% over 2 steps) as a yellow oil:  $[\alpha]_D = -47.0$  (*c* 0.30, CHCl<sub>3</sub>); IR (neat) 3446, 2925, 2854, 1701, 1515, 1456, 1332, 1271, 757, 731 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.16 (d, *J* = 8.1 Hz, 1H), 7.70 (d, *J* = 8.0 Hz, 1H), 7.55 (t, *J* = 7.4 Hz, 1H), 7.42 (t, *J* = 7.8 Hz, 1H), 6.85 (t, *J* = 6.3 Hz, 1H), 6.02 (s, 1H),

5.11-5.05 (m, 1H), 5.04 (s, 1H), 4.81 (s, 1H), 4.38 (d,  $J = 3.6$  Hz, 1H), 4.32-4.25 (m, 1H), 4.17-4.12 (m, 1H), 3.89 (d,  $J = 4.6$  Hz, 1H), 3.72-3.65 (m, 1H), 3.62-3.55 (m, 1H), 3.10-3.05 (m, 2H), 2.84 (dd,  $J = 14.8, 9.2$  Hz, 1H), 2.52 (dd,  $J = 14.8, 2.5$  Hz, 1H), 2.38 (d,  $J = 9.5$  Hz, 1H), 2.40-2.20 (m, 3H), 1.97-1.74 (m, 8H), 1.83 (s, 3H), 1.76 (s, 3H), 1.70-1.60 (m, 3H), 1.36 (s, 3H), 1.33 (d,  $J = 6.1$  Hz, 3H), 1.06 (d,  $J = 6.7$  Hz, 3H), 0.87 (d,  $J = 6.2$  Hz, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  212.0, 167.7, 147.7, 144.6, 141.3, 140.9, 133.7, 132.3, 130.1, 128.6, 126.5, 126.1, 125.6, 115.1, 78.0, 77.0, 75.5, 69.1, 67.7, 60.6, 58.4, 45.7, 45.6, 43.7, 43.1, 40.4, 39.2, 33.8, 30.3, 29.5, 28.9, 28.0, 27.0, 21.2, 19.7, 16.3, 15.2, 12.5; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{38}\text{H}_{55}\text{NO}_{10}\text{NaSe}(\text{M}+\text{Na})$  788.2889, found 788.2897.



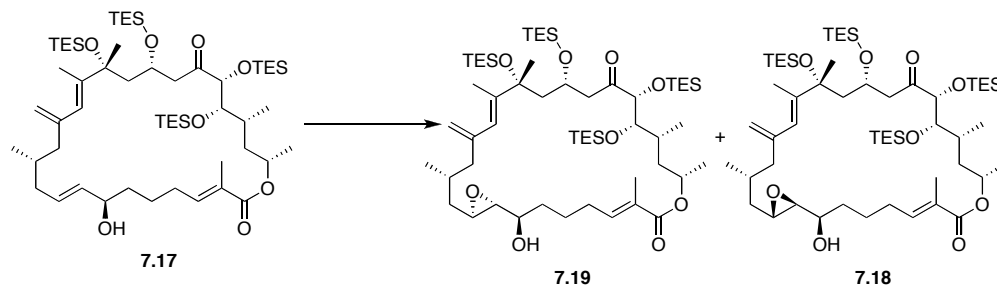
**Allylic epoxide 7.15:** To a stirred solution of selenide **7.14** (2.5 mg, 0.00327 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (0.5 mL) at rt was sequentially added NaHCO<sub>3</sub> (20 mg, 0.238 mmol) and TMSO-OTMS (19.1 mg, 23 μL, 0.107 mmol). After 1.5 h, the yellow color vanished and the reaction mixture was purified by preparative TLC,

eluting with 60% EtOAc / Hexanes, to give allylic epoxide **7.15** (1.2 mg, 0.00213 mmol, 65%):  $[\alpha]_D = -27.5$  ( $c$  0.12,  $\text{CHCl}_3$ ); IR (neat) 3443, 2924, 2852, 1703, 1457, 1379, 1272, 1118  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  6.71-6.63 (m, 1H), 6.08 (s, 1H), 5.88-5.78 (m, 1H), 5.26 (dd,  $J = 15.1, 8.4$  Hz, 1H), 5.12-5.05 (m, 1H), 5.05 (s, 1H), 4.84 (s, 1H), 4.29 (dd,  $J = 5.7, 1.5$  Hz, 1H), 4.30-4.20 (m, 1H), 3.71 (t,  $J = 7.9$  Hz, 1H), 3.66 (d,  $J = 5.7$  Hz, 1H), 3.19 (d,  $J = 9.6$  Hz, 1H), 3.12 (dd,  $J = 8.6, 2.2$  Hz, 1H), 3.01-2.87 (m, 2H), 2.46 (dd,  $J = 14.4, 2.1$  Hz, 1H), 2.36-2.20 (m, 5H), 2.00-1.74 (m, 6H), 1.83 (s, 3H), 1.77 (s, 3H), 1.60-1.55 (m, 1H), 1.35 (s, 3H), 1.33-1.28 (m, 1H), 1.30 (d,  $J = 6.1$  Hz, 3H), 1.18-1.11 (m, 1H), 1.03 (d,  $J = 6.6$  Hz, 3H), 0.88 (d,  $J = 6.5$  Hz, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  212.90, 167.45, 144.94, 141.56, 140.29, 136.40, 128.71, 128.60, 125.65, 114.95, 78.20, 76.56, 75.86, 68.49, 68.18, 60.18, 60.01, 45.84, 45.13, 43.77, 39.43, 39.29, 33.62, 30.96, 30.26, 28.54, 27.02, 21.22, 20.02, 15.82, 15.22, 12.74; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{32}\text{H}_{50}\text{O}_8\text{Na}$  ( $\text{M}+\text{Na}$ ) 585.3403, found 585.3394.



**Allylic alcohol 7.17:** To a stirred solution of macrocycle **7.6** (50 mg, 0.049 mmol) in  $\text{CH}_2\text{Cl}_2$  (2.3 mL) at  $-30^\circ\text{C}$  was sequentially added (*S*)-CBS (0.196 mL, 0.196 mmol, 1 M in PhMe) and  $\text{BH}_3\cdot\text{DMS}$  (0.3934 mL, 0.393 mmol, 1 M in THF). After 45 min, the reaction was quenched with MeOH (0.3 mL), diluted with aq.  $\text{NaHCO}_3$  (3 mL) and extracted with  $\text{Et}_2\text{O}$  (4 x 6 mL). The dried extract ( $\text{MgSO}_4$ ) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 3-6% EtOAc / Hexanes, to give allylic alcohol **7.17** (29 mg, 0.028 mmol, 58%) and **7.26** (8 mg, 0.0078 mmol, 16%) as colorless oils. **7.17**:  $[\alpha]_{\text{D}}^{23} = -23.6$  (*c* 0.25,  $\text{CHCl}_3$ ); IR (neat) 3503, 2954, 2911, 2876, 1707, 1458, 1376, 1240, 1128, 1007,  $742\text{ cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  6.81 (t,  $J = 6.9$  Hz, 1H), 5.76 (s, 1H), 5.70-5.62 (m, 1H), 5.53 (dd,  $J = 13.2, 6.3$  Hz, 1H), 5.00-4.95 (m, 1H), 4.98 (s, 1H), 4.84 (s, 1H), 4.15-4.04 (m, 3H), 3.58 (dd,  $J = 5.7, 2.4$  Hz, 1H), 2.93 (dd,  $J = 18.4, 6.2$  Hz, 1H), 2.78 (dd,  $J = 18.4, 6.0$  Hz, 1H), 2.28-2.22 (m, 2H), 2.18-2.10 (m, 2H), 2.08-1.99 (m, 1H), 1.89-1.57 (m, 9H), 1.84 (s, 3H), 1.81 (s, 3H), 1.48-1.40 (m, 2H), 1.45 (s, 3H), 1.26 (d,  $J = 6.1$  Hz, 3H), 1.07-0.91 (m, 36H), 0.88 (d,  $J = 6.6$  Hz, 3H), 0.74-0.53 (m, 27H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  207.8, 167.8, 145.5,

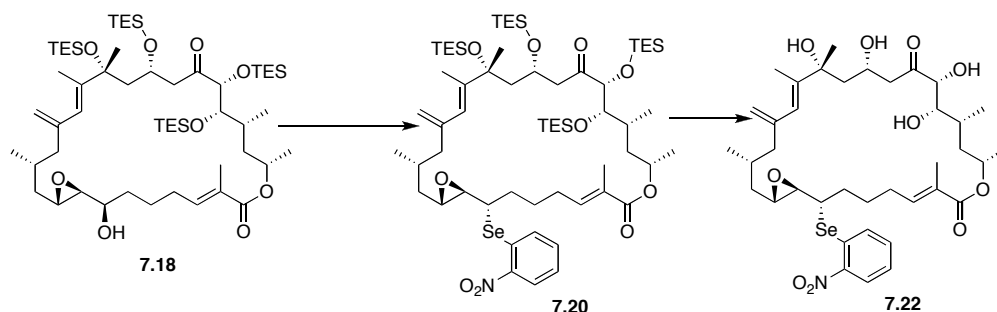
142.5 (2C), 134.9, 129.7, 127.9, 125.7, 114.9, 80.6, 79.2, 77.9, 71.9, 68.2, 65.3, 49.6, 49.2, 45.6, 42.2, 39.4, 37.0, 31.8, 29.8, 29.0, 28.1, 23.6, 21.0, 19.9, 14.7, 12.7, 12.4, 7.2, 7.1, 7.0, 6.9, 6.8, 5.3, 5.2, 4.7; HRMS (ES<sup>+</sup>) calcd. for C<sub>56</sub>H<sub>108</sub>O<sub>8</sub>Si<sub>4</sub>Na (M+Na) 1043.7019, found 1043.7072. **7.26**: [ $\alpha$ ]<sub>D</sub> = -29.1 (*c* 0.80, CHCl<sub>3</sub>); IR (neat) 3481, 2954, 2876, 1707, 1458, 1241, 1130, 1008, 742 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.74 (t, *J* = 7.4 Hz, 1H), 5.77 (s, 1H), 5.75-5.70 (m, 1H), 5.55 (dd, *J* = 15.5, 7.0 Hz, 1H), 5.00-4.96 (m, 1H), 4.99 (s, 1H), 4.83 (s, 1H), 4.22-4.15 (m, 1H), 4.11-4.06 (m, 1H), 4.07 (d, *J* = 5.8 Hz, 1H), 3.57 (dd, *J* = 5.6, 2.3 Hz, 1H), 2.96 (dd, *J* = 18.4, 7.4 Hz, 1H), 2.78 (dd, *J* = 18.3, 4.2 Hz, 1H), 2.23-2.08 (m, 4H), 1.95-1.52 (m, 11H), 1.834 (s, 3H), 1.80 (s, 3H), 1.48-1.40 (m, 1H), 1.43 (s, 3H), 1.25 (d, *J* = 6.1 Hz, 3H), 1.06-0.87 (m, 39H), 0.73-0.55 (m, 27H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  207.4, 167.9, 145.3, 142.2, 141.7, 134.2, 130.9, 128.3, 125.9, 114.9, 80.9, 79.0, 77.7, 73.2, 68.4, 65.1, 49.2, 48.8, 45.5, 42.1, 39.8, 36.8, 31.7, 30.1, 28.5, 28.3, 24.5, 21.0, 19.4, 15.0, 13.1, 12.4, 7.3, 7.1, 7.0, 6.9, 6.8, 5.2, 5.1, 4.8; HRMS (ES<sup>+</sup>) calcd. for C<sub>56</sub>H<sub>108</sub>O<sub>8</sub>Si<sub>4</sub>Na (M+Na) 1043.7019, found 1043.6984.



**Epoxide 7.18 & 7.19:** To a stirred solution of allylic alcohol **7.17** (29 mg, 0.0284 mmol) in  $\text{CH}_2\text{Cl}_2$  (2.2 mL) at  $-40^\circ\text{C}$  was sequentially added 4Å MS (20 mg), TBHP (15.5  $\mu\text{L}$ , 0.0852 mmol, 5.5 M in decane) and  $\text{Ti}(\text{O}-i\text{Pr})_4$  (16.1 mg, 16.6  $\mu\text{L}$ , 0.0567 mmol). After 5 h, the reaction was quenched with aq.  $\text{NaHCO}_3$  (3 mL) and extracted with  $\text{Et}_2\text{O}$  (4 x 4 mL). The dried extract ( $\text{MgSO}_4$ ) was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 6-10%  $\text{EtOAc}$  / Hexanes, to give epoxide **7.19** (13.6 mg, 0.0131 mmol, 46%) and epoxide **7.18** (9.1 mg, 0.00867 mmol, 31%) as colorless oils. **7.19**:  $[\alpha]_{\text{D}}^{23} = -32.3$  (*c* 0.73,  $\text{CHCl}_3$ ); IR (neat) 3482, 2954, 2911, 2876, 1706, 1458, 1380, 1239, 1131, 1073, 1009, 742  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  6.83 (t,  $J = 7.0$  Hz, 1H), 5.76 (s, 1H), 5.00-4.92 (m, 1H), 4.98 (s, 1H), 4.83 (s, 1H), 4.20-4.10 (m, 1H), 4.10 (d,  $J = 5.5$  Hz, 1H), 3.58 (dd,  $J = 5.4, 1.7$  Hz, 1H), 3.60-3.50 (m, 1H), 3.12-3.05 (m, 1H), 2.98-2.92 (m, 2H), 2.90-2.82 (m, 1H), 2.77 (dd,  $J = 5.0, 2.0$  Hz, 1H), 2.40-2.30 (m, 2H), 2.17 (dd,  $J = 12.7, 4.3$  Hz, 1H), 2.12-2.00 (m, 1H), 1.91-1.61 (m, 8H), 1.82 (s, 3H), 1.79 (s, 3H), 1.50-1.40 (m, 3H), 1.42 (s, 3H), 1.30-1.20 (m, 1H), 1.26 (d,  $J = 6.0$  Hz, 3H), 1.08-0.87 (m, 39H), 0.75-0.50 (m, 27H);  $^{13}\text{C}$  NMR

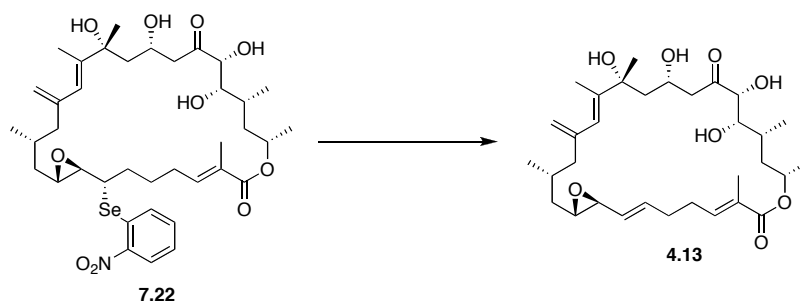


(75 MHz, CDCl<sub>3</sub>)  $\delta$  208.1, 167.8, 144.7, 141.8, 141.7, 128.2, 125.9, 115.0, 80.5, 79.4, 77.9, 70.4, 68.2, 65.1, 60.8, 55.8, 49.8, 48.9, 45.9, 42.9, 39.3, 33.6, 30.2, 29.2, 28.6, 28.5, 24.0, 21.1, 19.5, 15.1, 12.9, 12.4, 7.3, 7.1, 6.9, 5.2, 4.7; HRMS (ES<sup>+</sup>) calcd. for C<sub>56</sub>H<sub>108</sub>O<sub>9</sub>Si<sub>4</sub>Na (M+Na) 1059.6968, found 1059.7001. **7.18**:  $[\alpha]_D^{23} = -25.7$  (*c* 0.42, CHCl<sub>3</sub>); IR (neat) 3482, 2954, 2876, 1708, 1458, 1378, 1240, 1130, 1008, 742 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  6.76 (t, *J* = 6.1 Hz, 1H), 5.83 (s, 1H), 5.05-4.90 (m, 1H), 5.01 (s, 1H), 4.86 (s, 1H), 4.15-4.10 (m, 1H), 4.08 (d, *J* = 5.8 Hz, 1H), 3.58 (dd, *J* = 5.5, 2.6 Hz, 1H), 3.59-3.49 (m, 1H), 3.11-3.03 (m, 1H), 2.95-2.91 (m, 2H), 2.77 (dd, *J* = 5.5, 2.0 Hz, 1H), 2.35-2.20 (m, 3H), 2.02-1.59 (m, 10H), 1.83 (s, 3H), 1.81 (s, 3H), 1.44 (s, 3H), 1.50-1.40 (m, 3H), 1.25 (d, *J* = 6.0 Hz, 3H), 1.06-0.89 (m, 39H), 0.74-0.52 (m, 27H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  207.7, 167.8, 144.8, 142.3, 141.5, 128.4, 125.4, 115.3, 80.8, 78.9, 77.8, 71.5, 68.4, 65.3, 61.8, 55.5, 49.2 (2C), 45.4, 41.7, 38.8, 33.3, 30.4, 29.6, 28.5, 28.3, 23.9, 21.0, 20.0, 15.0, 13.2, 12.5, 7.2, 7.1, 7.0, 6.9, 5.2, 4.8; HRMS (ES<sup>+</sup>) calcd. for C<sub>56</sub>H<sub>108</sub>O<sub>9</sub>Si<sub>4</sub>Na (M+Na) 1059.6968, found 1059.6982.



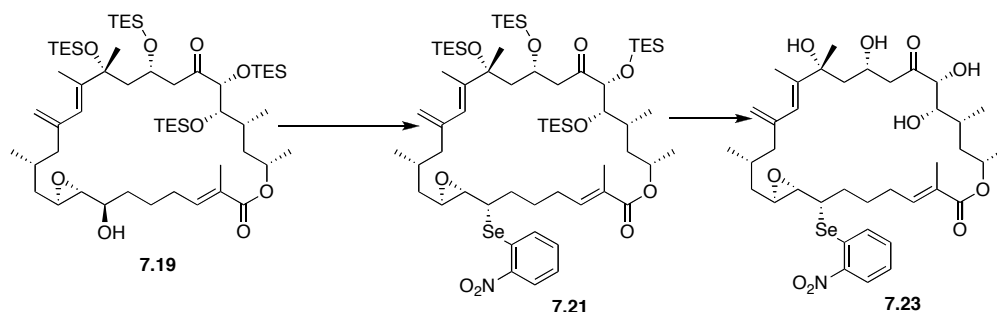
**Selenide 7.20:** To a stirred solution of epoxide **7.18** (8.5 mg, 0.00819 mmol) in THF (0.5 mL) at rt was sequentially added *o*-NO<sub>2</sub>C<sub>6</sub>H<sub>4</sub>SeCN (37 mg, 0.164 mmol) and PBu<sub>3</sub> (33.2 mg, 41 μL, 0.164 mmol). After 5 h, the reaction mixture was concentrated *in vacuo* purified by chromatography over silica gel, eluting with Hexanes then with 4% EtOAc / Hexanes, to give crude selenide **7.20** (4.5 mg) as a yellow oil which was used directly in next step without further purification.

**Polyol 7.22:** To a stirred solution of selenide **7.20** (4.5 mg) in THF / DMF / H<sub>2</sub>O (10:1:0.02, 0.50 mL / 50 μL / 1 μL) at 0°C was added TAS-F (5 mg, 0.0180 mmol). The reaction mixture was then warmed up to rt. After 2 h, the reaction mixture was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 10-65% EtOAc / Hexanes, to give polyol **7.22** (2.0 mg, 0.00261 mmol, 32% over 2 steps) as a yellow oil:  $[\alpha]_D^{23} = -41.7$  (*c* 0.12, CHCl<sub>3</sub>); IR (neat) 3446, 2924, 2854, 1701, 1519, 1457, 1378, 1334, 1121, 759, 732 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.17 (d, *J* = 8.0 Hz, 1H), 7.70 (d, *J* = 8.0 Hz, 1H), 7.55 (t, *J* = 7.4



**Amphidinolide B<sub>1</sub> (4.13):** To a stirred solution of selenide **7.22** (2.0 mg, 0.00261 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (0.4 mL) at rt was sequentially added NaHCO<sub>3</sub> (20 mg, 0.238 mmol) and TMSO-OTMS (16.6 mg, 20 μL, 0.0929 mmol). After 1.5 h, the yellow color vanished and the reaction mixture was purified by preparative TLC, eluting with 60% EtOAc / Hexanes, to give amphidinilide B<sub>1</sub> (**4.13**) (1.0 mg,

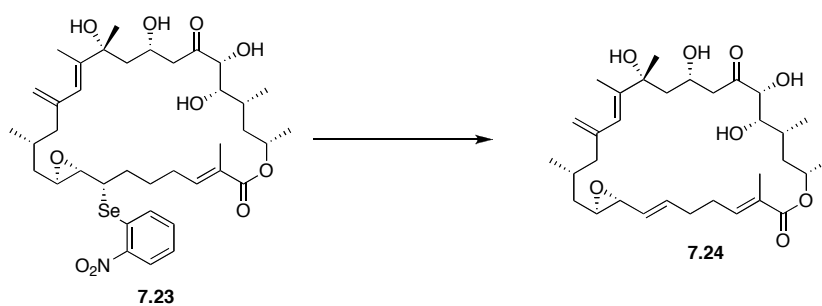
0.00178 mmol, 68%):  $[\alpha]_D^{23} = -63.7$  ( $c$  0.08,  $\text{CHCl}_3$ ), Literature Value:<sup>1</sup>  $-62.5$  ( $c$  0.39,  $\text{CHCl}_3$ );  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  6.78 (t,  $J = 7.8$  Hz, 1H), 5.99 (s, 1H), 5.93 (ddd,  $J = 15.2, 8.5, 4.8$  Hz, 1H), 5.18 (dd,  $J = 15.8, 8.6$  Hz, 1H), 5.06 (m, 1H), 5.05 (s, 1H), 4.84 (s, 1H), 4.34 (dd,  $J = 4.8, 1.4$  Hz, 1H), 4.20 (m, 1H), 3.92 (d,  $J = 3.3$  Hz, 1H), 3.88 (d,  $J = 5.0$  Hz, 1H), 3.73 (ddd,  $J = 10.3, 8.8, 1.5$  Hz, 1H), 3.19 (d,  $J = 10.0$  Hz, 1H), 3.16 (dd,  $J = 8.3, 2.0$  Hz, 1H), 2.94 (ddd,  $J = 8.9, 2.6, 2.2$  Hz, 1H), 2.86 (d,  $J = 7.3$  Hz, 1H), 2.80 (dd,  $J = 15.9, 3.2$  Hz, 1H), 2.43 (m, 1H), 2.38 (m, 1H), 2.25 (m, 1H), 2.20 (m, 1H), 2.17 (m, 1H), 2.16 (s, 1H), 1.98-1.91 (m, 4H), 1.80 (s, 6H), 1.76 (dd,  $J = 14.5, 5.2$  Hz, 1H), 1.64 (m, 1H), 1.49 (ddd,  $J = 13.6, 10.9, 3.0$  Hz, 1H), 1.44 (s, 3H), 1.31 (m, 1H), 1.30 (d,  $J = 6.1$  Hz, 3H), 1.26 (m, 1H), 1.03 (d,  $J = 6.6$  Hz, 3H), 0.90 (d,  $J = 6.4$  Hz, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  212.51, 167.77, 144.47, 143.17, 140.02, 135.52, 128.58, 128.45, 124.40, 114.92, 77.86, 76.07, 75.69, 68.44, 66.70, 60.19, 46.99, 45.98, 45.35, 39.52, 39.39, 33.31, 30.95, 29.35, 28.44, 26.88, 21.07, 18.27, 15.71, 15.15, 12.50; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{32}\text{H}_{50}\text{O}_8\text{Na}$  ( $\text{M}+\text{Na}$ ) 585.3403, found 585.3411.



**Selenide 7.21:** To a stirred solution of epoxide **7.19** (14.5 mg, 0.0139 mmol) in THF (0.8 mL) at rt was sequentially added *o*-NO<sub>2</sub>C<sub>6</sub>H<sub>4</sub>SeCN (63 mg, 0.279 mmol) and PBu<sub>3</sub> (56.7 mg, 70  $\mu$ L, 0.279 mmol). After 1 h, the reaction mixture was concentrated *in vacuo* purified by chromatography over silica gel, eluting with Hexanes then with 4% EtOAc / Hexanes, to give crude selenide **7.21** (15.2 mg) as a yellow oil which was used directly in next step without further purification.

**Polyol 7.23:** To a stirred solution of selenide **7.21** (15.2 mg, 0.0122 mmol) in THF / DMF / H<sub>2</sub>O (10:1:0.02, 1.6 mL / 0.16 mL / 3.2  $\mu$ L) at 0°C was added TAS-F (16.8 mg, 0.0610 mmol). The reaction mixture was then warmed up to rt. After 2 h, the reaction mixture was concentrated *in vacuo* and purified by chromatography over silica gel, eluting with 10-65% EtOAc / Hexanes, to give polyol **7.23** (9.1 mg, 0.0119 mmol, 86% over 2 steps) as yellow oils:  $[\alpha]_D^{23} = -51.8$  (*c* 0.44, CHCl<sub>3</sub>); IR (neat) 3447, 2926, 2855, 1701, 1514, 1456, 1332, 1271, 1037, 902, 756, 731 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.26 (d, *J* = 8.3 Hz, 1H), 7.73

(d,  $J = 8.0$  Hz, 1H), 7.55 (t,  $J = 7.3$  Hz, 1H), 7.36 (t,  $J = 7.9$  Hz, 1H), 6.81 (t,  $J = 7.0$  Hz, 1H), 5.97 (s, 1H), 5.10-5.03 (m, 1H), 5.03 (s, 1H), 4.83 (s, 1H), 4.47 (s, OH), 4.40-4.32 (m, 1H), 4.14 (dd,  $J = 14.2, 7.2$  Hz, 1H), 3.88-3.78 (m, 1H), 3.42-3.35 (m, 1H), 3.05-2.97 (m, 1H & OH), 2.85 (d,  $J = 4.8$  Hz, 2H), 2.50-2.38 (br, 1H), 2.30-2.20 (m, 2H), 2.07 (s, OH), 2.11-2.02 (m, 1H), 1.96-1.65 (m, 11H), 1.83 (s, 6H), 1.44 (s, 3H), 1.40-1.32 (m, 2H), 1.33 (d,  $J = 5.8$  Hz, 3H), 1.08 (d,  $J = 6.3$  Hz, 3H), 0.90 (d,  $J = 4.4$  Hz, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  211.6, 167.9, 147.6, 144.0, 143.5, 141.2, 133.6, 132.3, 130.6, 128.7, 126.4, 126.0, 124.0, 115.5, 78.0, 76.1, 75.0, 69.5, 66.3, 60.7, 58.7, 46.1, 45.8, 44.8, 43.8, 40.5, 39.5, 33.5, 30.4 (2C), 28.2, 27.7, 27.1, 20.8, 19.6, 16.5, 15.2, 12.5; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{38}\text{H}_{55}\text{NO}_{10}\text{NaSe}$  ( $\text{M}+\text{Na}$ ) 788.2889, found 788.2934.



**Allylic epoxide 7.24:** To a stirred solution of selenide **7.23** (2.7 mg, 0.00353 mmol) in  $\text{CH}_2\text{Cl}_2$  (0.5 mL) at rt was sequentially added  $\text{NaHCO}_3$  (30 mg, 0.357 mmol) and TMSO-OTMS (22.5 mg, 27  $\mu\text{L}$ , 0.126 mmol). After 1.5 h, the

yellow color vanished and the reaction mixture was purified by preparative TLC, eluting with 60% EtOAc / Hexanes, to give allylic epoxide **7.24** (1.3 mg, 0.00231 mmol, 65%):  $[\alpha]_{\text{D}}^{23} = +10.0$  ( $c$  0.13,  $\text{CHCl}_3$ ); IR (neat) 3422, 2923, 2853, 1701, 1457, 1377, 1261, 1103;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  6.72 (t,  $J = 4.9$  Hz, 1H), 6.04 (s, 1H), 5.88-5.80 (m, 1H), 5.28 (dd,  $J = 15.4, 8.4$  Hz, 1H), 5.10-5.05 (m, 1H), 5.06 (s, 1H), 4.84 (s, 1H), 4.35 (dd,  $J = 4.2, 1.5$  Hz, 1H), 4.30-4.20 (m, 1H), 4.13 (s, 1H), 3.79 (t,  $J = 9.8$  Hz, 1H), 3.77 (d,  $J = 4.6$  Hz, 1H), 3.40 (d,  $J = 10.2$  Hz, 1H), 3.09 (dd,  $J = 8.5, 2.1$  Hz, 1H), 2.98-2.93 (m, 2H), 2.76 (dd,  $J = 15.4, 2.4$  Hz, 1H), 2.38-2.23 (m, 5H), 2.35 (s, 1H), 1.98 (m, 1H), 1.95 (m, 1H), 1.88 (m, 1H), 1.85 (m, 1H), 1.84 (s, 6H), 1.78 (dd,  $J = 14.6, 4.7$  Hz, 1H), 1.70 (m, 1H), 1.64 (m, 1H), 1.32 (m, 1H), 1.30 (d,  $J = 6.1$  Hz, 3H), 1.13 (m, 1H), 1.03 (d,  $J = 6.6$  Hz, 3H), 0.83 (d,  $J = 6.4$  Hz, 3H)  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  212.64, 167.52, 144.58, 143.16, 140.79, 135.90, 128.87, 128.68, 124.34, 114.91, 78.27, 76.07, 75.46, 68.31, 66.79, 60.29, 59.18, 46.45, 46.10, 45.60, 39.55, 33.25, 31.24, 30.45, 29.71, 28.55, 27.08, 21.04, 19.90, 15.74, 15.41, 12.69; HRMS ( $\text{ES}^+$ ) calcd. for  $\text{C}_{32}\text{H}_{50}\text{O}_8\text{Na}$  ( $\text{M}+\text{Na}$ ) 585.3403, found 585.3409.

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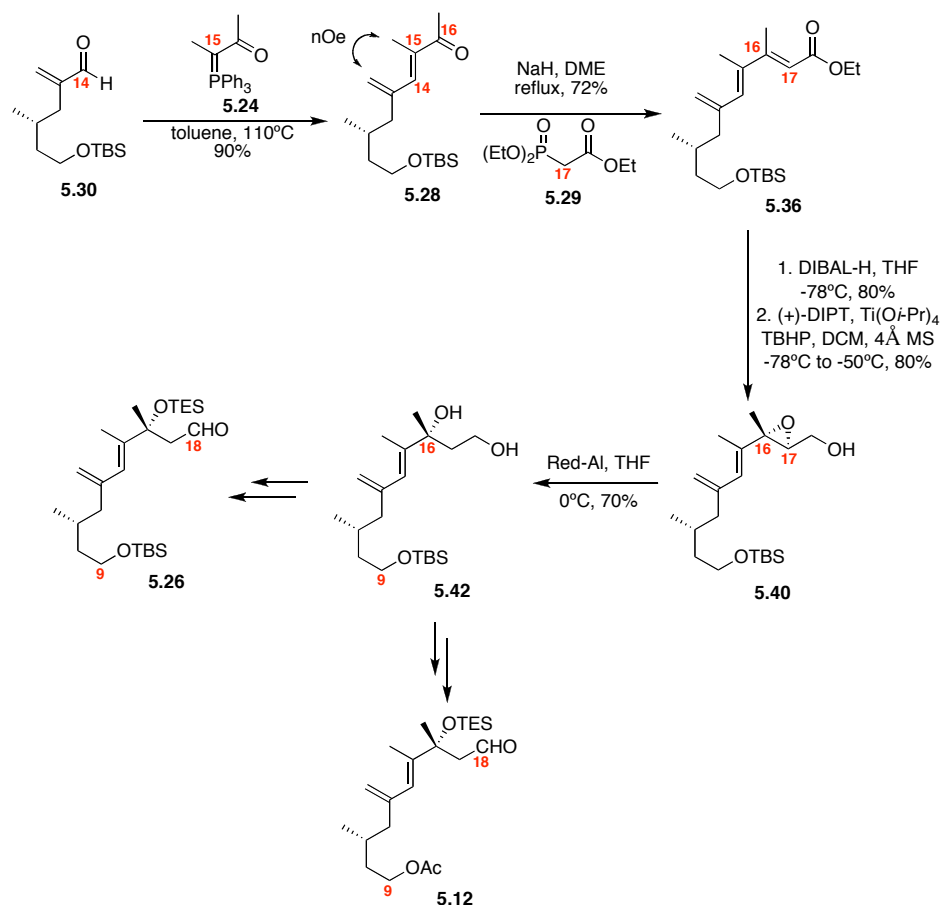
1. Bauer, I.; Maranda, L.; Shimizu, Y.; Peterson, R. W.; Cornell, L.; Steiner, J. R.; Clardy, J. *J. Am. Chem. Soc.* **1994**, *116*, 2657-58.

## CHAPTER 8. CONCLUSION AND FUTURE WORK

### 8.1 General Conclusion

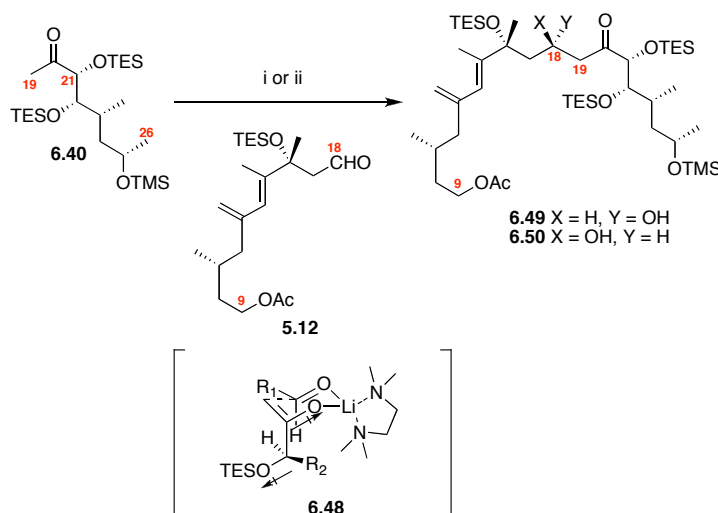
During our endeavors toward amphidinolide B<sub>1</sub> and B<sub>2</sub>, we developed several important protocols. Our metal catalyst-free strategy yielded the unusual highly substituted C<sub>13</sub>-C<sub>15</sub> diene efficiently (Scheme 8.1). Utilizing a Wittig reaction between aldehyde **5.30** and ylide **5.24**, we could synthesize this difficult C<sub>13</sub>-C<sub>15</sub> diene moiety in good yield and excellent *E/Z* selectivity. Two other highlights of our approach are a HWE reaction to build C<sub>16</sub>-C<sub>17</sub> alkene and a Sharpless epoxidation / regioselective epoxide opening sequence to yield C<sub>16</sub> tertiary alcohol.





**Scheme 8.1.** Synthesis of Diene Subunits

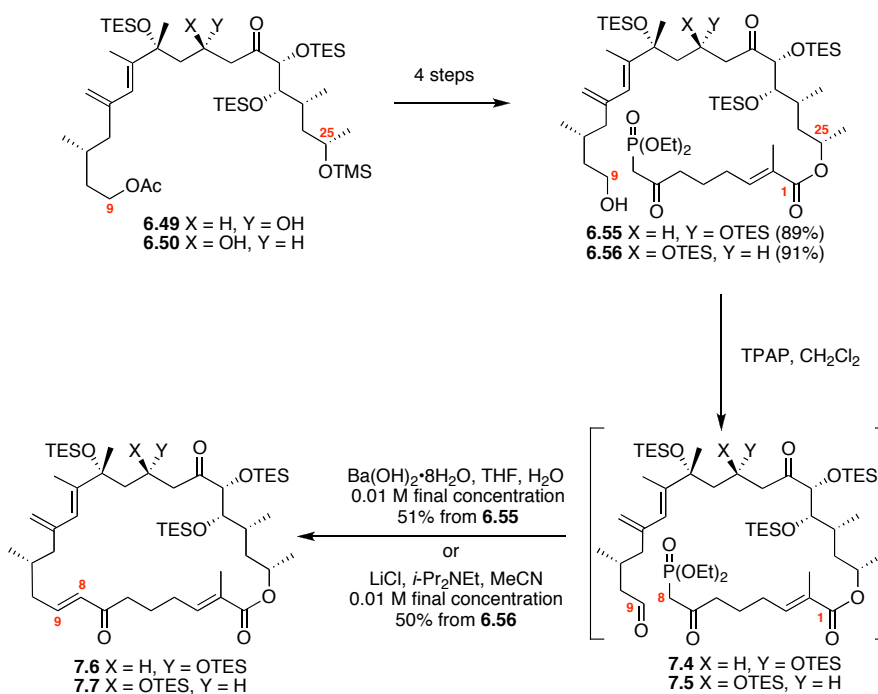
Using C<sub>21</sub> TES-protected methyl ketone **6.40**, the non-chelation-controlled aldol reaction led to 18*R* isomer **6.50** in 1:8 dr (**6.49**:**6.50**) at -100°C (Scheme 8.2). Alternatively, the 18*S* stereoisomer **6.49** was generated in 1.2:1 dr (**6.49**:**6.50**) at -40°C. While we are still exploring the nature of the diastereoselectivity, one possible explanation could be that a transition state **6.48**, which minimizes the dipoles of the C<sub>21</sub> C–O σ bond and the enolate, determines the stereochemical outcome of the reaction.



(i) LDA, TMEDA, THF,  $-100^{\circ}\text{C}$  then add **5.12**, 65% (1:8 dr, **6.49**:**6.50**); (ii) LDA, TMEDA, THF,  $-40^{\circ}\text{C}$  then add **5.12**, 66% (1.2:1 dr, **6.49**:**6.50**)

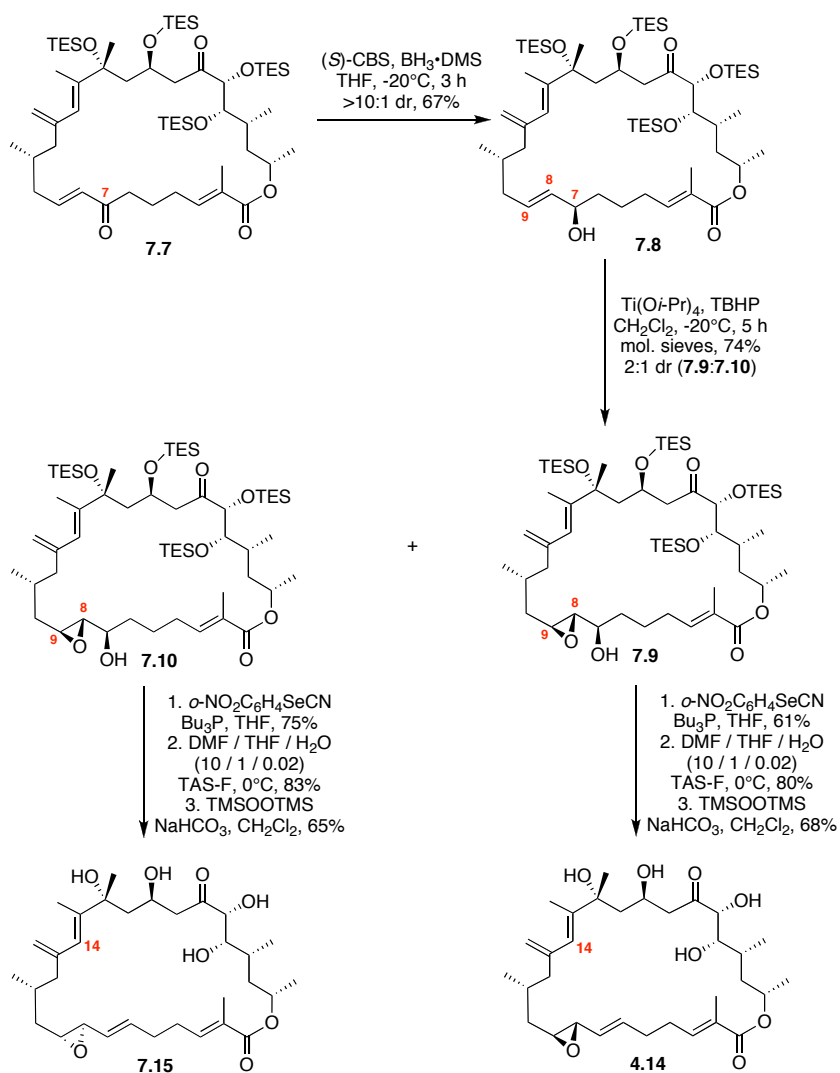
**Scheme 8.2.** Aldol Coupling between Methyl Ketone **6.40** and Aldehyde **5.12**

Another highlight of our work is the macrocyclization of the 26-membered lactone ring (Scheme 8.3) Further chemical elaboration of aldol adducts **6.49/6.50** gave rise to phosphonate alcohol **6.55/6.56**. When phosphonate **6.55/6.56** was exposed to TPAP /  $\text{CH}_2\text{Cl}_2$ , significant amounts of the macrocycle **7.6/7.7** formed via a spontaneous intramolecular Horner-Wadsworth–Emmons olefination. The conversion could be driven to completion by the addition of LiCl and Hunig's base or  $\text{Ba}(\text{OH})_2 \cdot 8\text{H}_2\text{O}$ , respectively.



**Scheme 8.3.** Macrocyclization of 26-Membered Ring

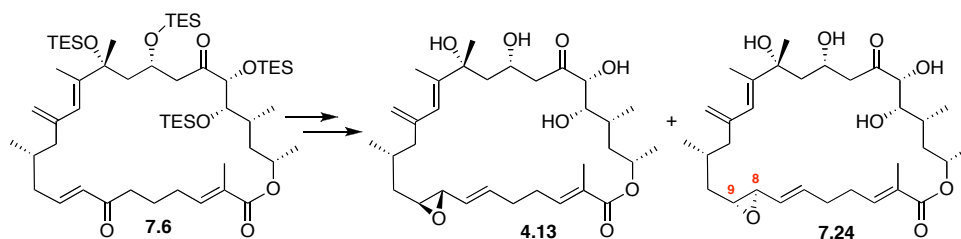
The key steps of the incorporation of the allylic epoxide moiety include a region- and stereoselective reduction of the C<sub>7</sub> carbonyl functionality with the (*S*)-CBS reagent, a Ti(*Oi*-Pr)<sub>4</sub> / TBHP-mediated epoxidation, and a TMSOOTMS induced oxidation and *in situ* elimination of a selenide (Scheme 8.4). The proposed structure of amphidinolide B<sub>2</sub> (**4.14**) and its C<sub>8,9</sub> epoxide diastereomer **7.15** were finally synthesized with a longest linear of 29 steps. To our surprise, these synthesized compounds **4.14** and **7.15** *did not match* with the spectra data provided for amphidinolide B<sub>2</sub>. In both cases, the <sup>1</sup>H NMR shift for the H<sub>14</sub> alkene was shifted significantly downfield as compared to the natural product data.



**Scheme 8.4.** Synthesis of Proposed Structure of Amphidinolide B<sub>2</sub> and its C8,9 Epoxide Diastereomer

Careful inspection of the isolation paper revealed that the stereochemical analysis of amphidinolide B<sub>2</sub> was based primarily on the differences in the <sup>1</sup>H NMR in the C<sub>17</sub>–C<sub>19</sub> region of the natural product as compared to amphidinolide

B<sub>1</sub> (**4.13**). It is important to note that Shimizu and Clardy obtained X-ray crystallographic structure of natural product **4.13**. It is clear from our work that the structural differences between amphidinolide B<sub>1</sub> and B<sub>2</sub> are more complicated than initially expected. On the basis of this information, we have concluded that the proposed structure of amphidinolide B<sub>2</sub> is incorrect. We applied an analogous strategy for the synthesis of **4.13** as was described for the 18*R* series (Scheme 8.5). The synthesized material **4.13** matched with the spectra data reported by Kobayashi and co-workers for amphidinolide B<sub>1</sub>.

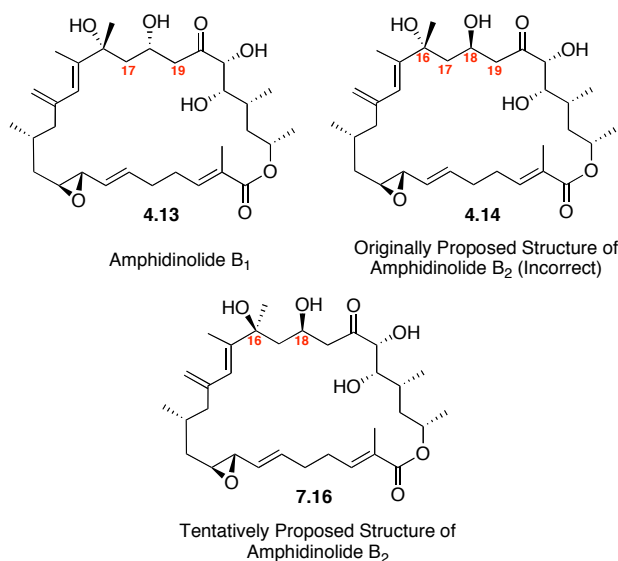


**Scheme 8.5.** Synthesis of Amphidinolide B<sub>1</sub> and its C<sub>8,9</sub> Epoxide Diastereomer

## 8.2 Proposed Future Work

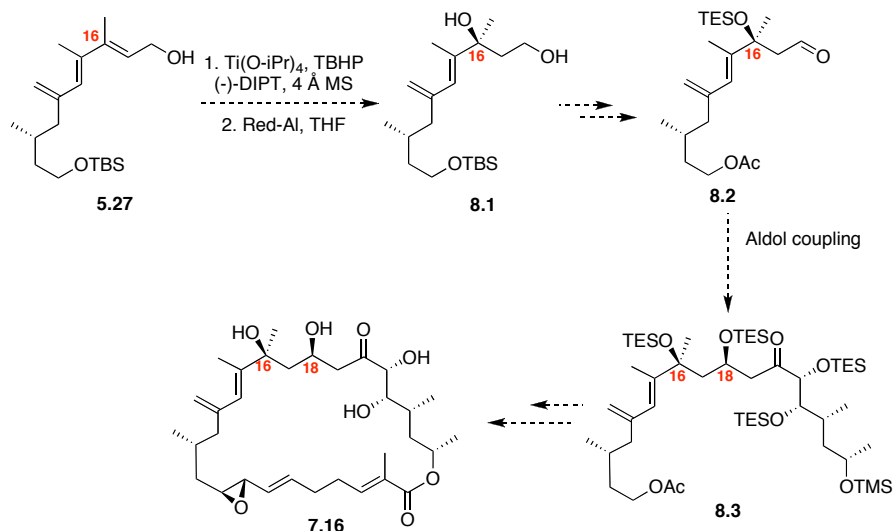
We have developed a synthetic route for amphidinolide B and its analogs. First syntheses of amphidinolide B<sub>1</sub> and the proposed structure of amphidinolide B<sub>2</sub> have been accomplished based on this strategy; however the originally proposed structure of amphidinolide B<sub>2</sub> was found to be incorrect. Consequently, our next target would be the correction of the proposed structure of amphidinolide B<sub>2</sub>. We intend to do extensive 2D NMR on compound **4.14** to firmly assign each

H and C for the compound. Based on our tentative assignments of the data we have already collected, we suspect that the culprit stereochemistry is in fact the C<sub>16</sub> tertiary alcohol. We speculate that a common *syn* relationship is present between C<sub>16</sub> and C<sub>18</sub> in amphidinolide B<sub>2</sub> (Figure 8.1).



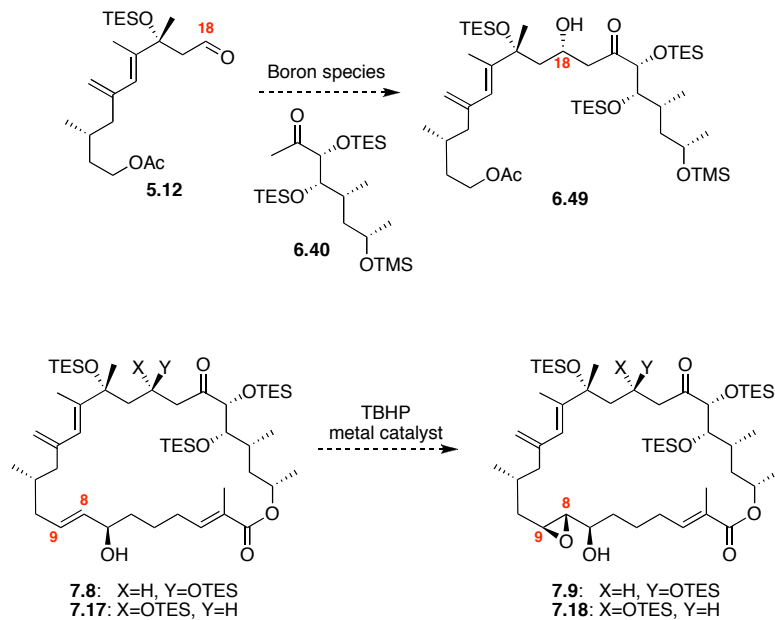
**Figure 8.1.** Tentatively Proposed Structure of Amphidinolide B<sub>2</sub>

The first compound we would like to synthesize will be epimeric at C<sub>16</sub>-compound **7.16** (Scheme 8.6). The epimer stereochemistry can be readily available from the Sharpless epoxidation and the following transformation could be accomplished using analogous strategy for the synthesis of the proposed structure of amphidinolide B<sub>2</sub>.



**Scheme 8.6.** Proposed Synthesis of Compound 7.16

Besides the correction of the proposed structure of amphidinolide B<sub>2</sub>, we also intend to investigate other options to improve the diastereoselectivity of several steps including the aldol coupling to afford 18*S* isomer and the epoxidation to install C<sub>8</sub>-C<sub>9</sub> epoxide (Scheme 8.7). Boron enolate based asymmetric aldol reactions<sup>1</sup> would be a potential option for the aldol coupling. We have already shown that  $\text{Ti}(\text{O}-i\text{Pr})_4$  can be used to access both diastereomers of the C<sub>8</sub>-C<sub>9</sub> epoxide. We would like to explore other transition metal oxidants (e.g.  $\text{VO}(\text{acac})_2$ ) to see if an improved diastereoselectivity can be obtained.



**Scheme 8.7.** Proposed Optimization of Current Work

### 8.3 References

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**APPENDIX: X-RAY CRYSTALLOGRAPHIC DATA**

**X-ray Crystal Structure Determination.** X-ray diffraction intensity data were collected with a Bruker Smart Apex CCD diffractometer using MoK $\alpha$  – radiation (0.71073 Å). Crystallographic data and some details of data collections and refinements for the investigated structures are given in Tables A1-A16. The structures were solved using direct methods, completed by subsequent difference Fourier syntheses, and refined by full matrix least-squares procedures on  $F^2$ . The non-hydrogen atoms in all structures were refined with anisotropic thermal parameters. Highly disordered solvent molecules were treated by SQUEEZE (Van der Sluis, P. & Spek, A. L. (1990) *Acta Cryst. Sect. A*, A46, 194-201). All software and scattering factor sources are contained in the SHELXTL (5.10) program package (G. Sheldrick, Bruker XRD, Madison, WI).

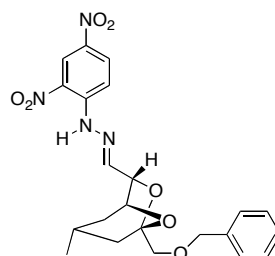
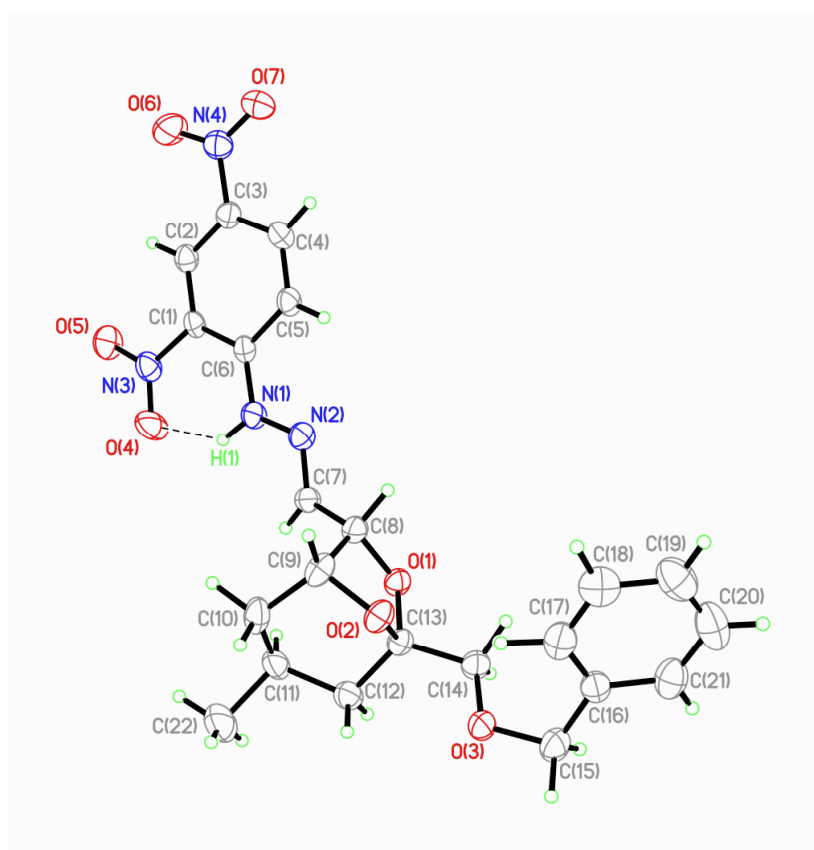
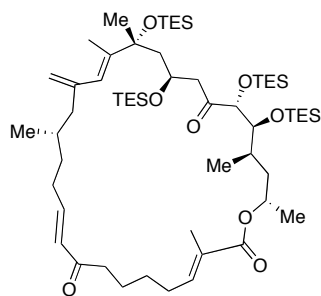
**2,4-Dinitrohydrazone 2.20:****2.20**



Table 1. Crystal data and structure refinement for rc4.

Identification code	rc4	
Empirical formula	C <sub>22</sub> H <sub>24</sub> N <sub>4</sub> O <sub>7</sub>	
Formula weight	456.45	
Temperature	293(2) K	
Wavelength	0.71073 Å	
Crystal system	Monoclinic	
Space group	C2	
Unit cell dimensions	a = 24.757(4) Å	a = 90°.
	b = 6.6425(10) Å	b = 131.966(2)°.
	c = 18.281(3) Å	g = 90°.
Volume	2235.3(6) Å <sup>3</sup>	
Z	4	
Density (calculated)	1.356 Mg/m <sup>3</sup>	
Absorption coefficient	0.103 mm <sup>-1</sup>	
F(000)	960	
Crystal size	0.48 x 0.05 x 0.02 mm <sup>3</sup>	
Theta range for data collection	1.50 to 25.00°.	
Index ranges	-29 ≤ h ≤ 29, -7 ≤ k ≤ 7, -21 ≤ l ≤ 21	
Reflections collected	8138	

Independent reflections	3905 [R(int) = 0.0334]
Completeness to theta = 25.00°	100.0 %
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	1.000 and 0.837
Refinement method	Full-matrix least-squares on F <sup>2</sup>
Data / restraints / parameters	3905 / 1 / 394
Goodness-of-fit on F <sup>2</sup>	0.986
Final R indices [I>2sigma(I)]	R1 = 0.0477, wR2 = 0.0561
R indices (all data)	R1 = 0.0965, wR2 = 0.0675
Absolute structure parameter	-0.5(11)
Largest diff. peak and hole	0.122 and -0.118 e.Å <sup>-3</sup>

**Macrocycle 7.6:**

7.6

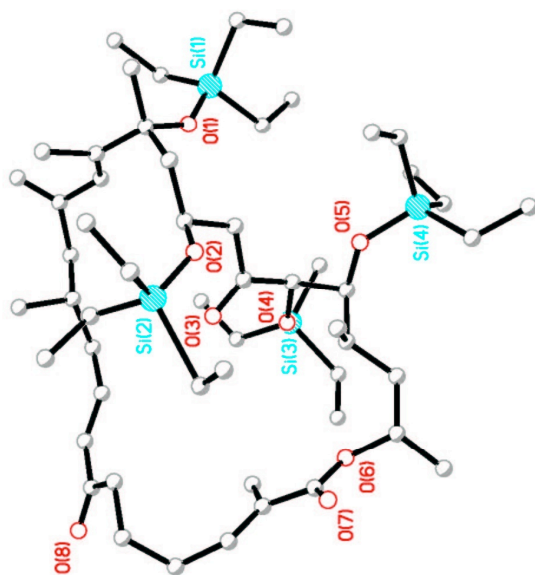
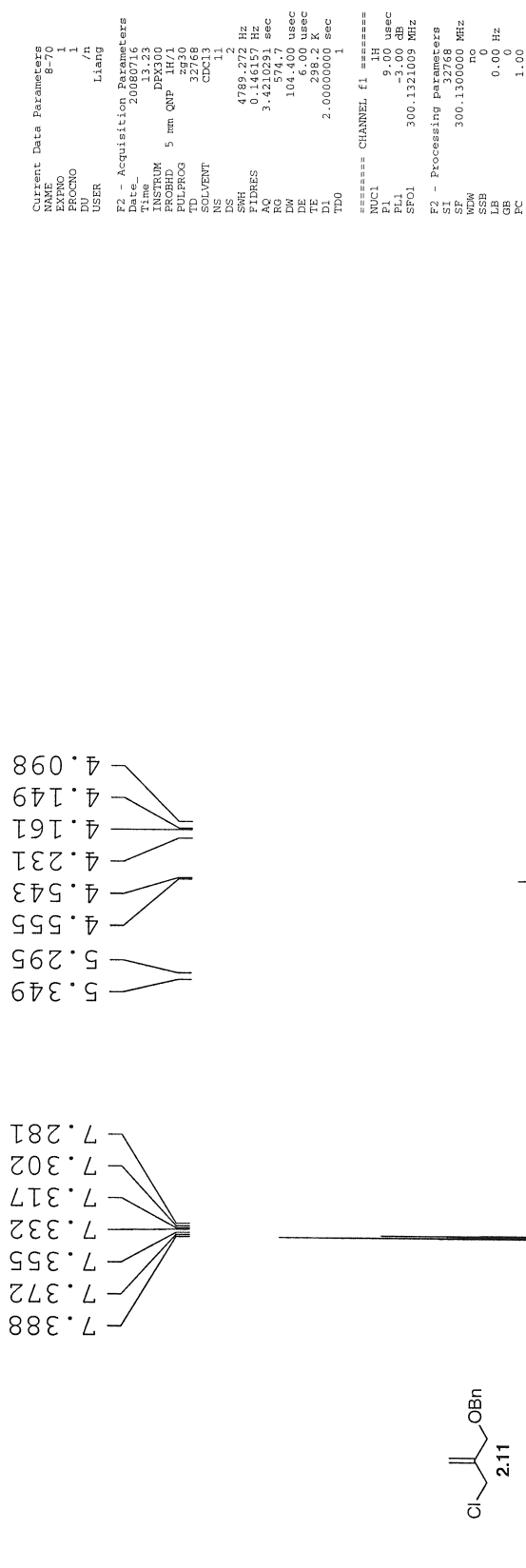


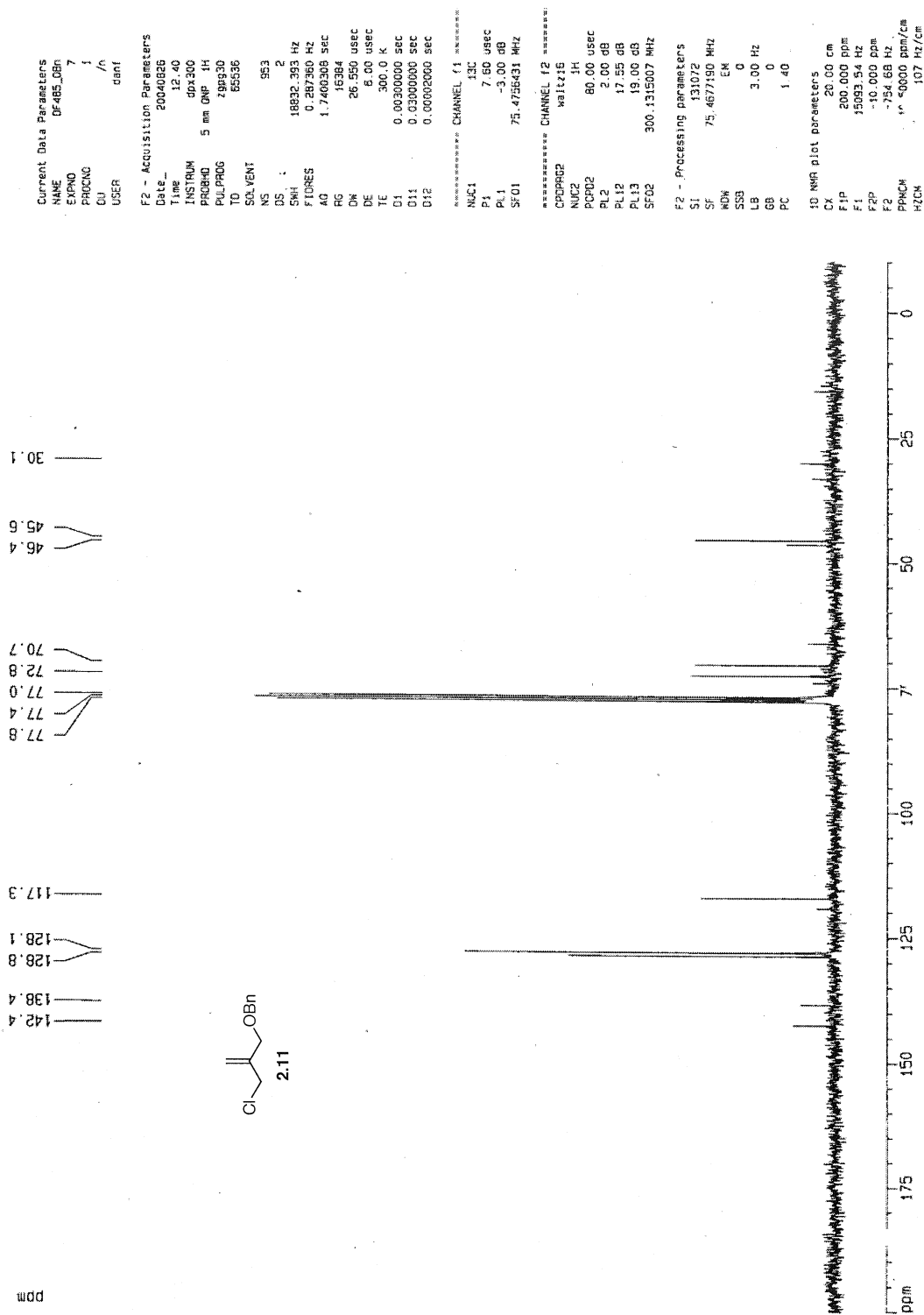
Table 1. Crystal data and structure refinement for rcr34.

Identification code	rcr34
Empirical formula	C <sub>56</sub> H <sub>106</sub> O <sub>8</sub> Si <sub>4</sub>
Formula weight	1019.77
Temperature	173(2) K
Wavelength	0.71073 Å
Crystal system	Monoclinic
Space group	P2(1)
Unit cell dimensions	a = 11.5018(14) Å      a = 90°. b = 22.813(3) Å      b = 107.376(2)°. c = 12.7063(15) Å      c = 90°.
Volume	3181.9(7) Å <sup>3</sup>
Z	2
Density (calculated)	1.064 Mg/m <sup>3</sup>
Absorption coefficient	0.139 mm <sup>-1</sup>
F(000)	1124
Crystal size	0.38 x 0.36 x 0.09 mm <sup>3</sup>
Theta range for data collection	1.68 to 25.00°.
Index ranges	-13 ≤ h ≤ 13, -27 ≤ k ≤ 27, -15 ≤ l ≤ 15
Reflections collected	30468

Independent reflections	11188 [R(int) = 0.0335]
Completeness to theta = 25.00°	100.0 %
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	0.9876 and 0.9491
Refinement method	Full-matrix least-squares on F <sup>2</sup>
Data / restraints / parameters	11188 / 13 / 809
Goodness-of-fit on F <sup>2</sup>	1.043
Final R indices [I>2sigma(I)]	R1 = 0.0600, wR2 = 0.1442
R indices (all data)	R1 = 0.0769, wR2 = 0.1563
Absolute structure parameter	0.00(12)
Largest diff. peak and hole	0.420 and -0.367 e.Å <sup>-3</sup>

**APPENDIX: NMR DATA**







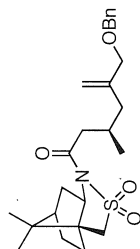
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PROCNO 1  
DU /n  
USER danf

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PROBHD 5 mm BBO Z-G  
PULPROG zg30  
TD 32768  
SOLVENT CDCl3  
NS 32  
DS 2  
SWH 5995.204 Hz  
FIDRES 0.182559 Hz  
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RG 1149.4  
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DE 6.00 usec  
TE 300.0 K  
D1 2.0000000 sec

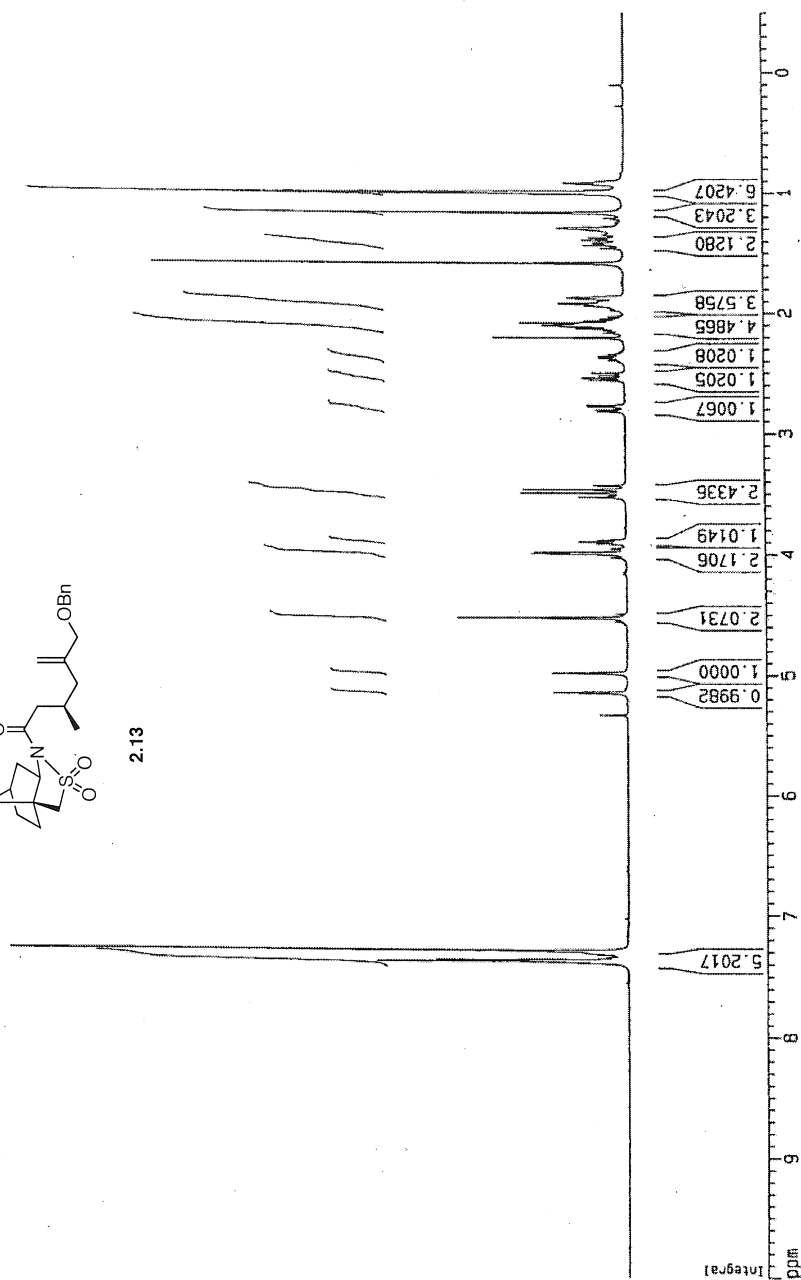
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SF01 400.0126001 MHz

F2 - Processing parameters  
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SF 400.0100000 MHz  
WDW EM  
SSB 0  
LB 0.70 Hz  
GB 0  
PC 1.00

1D NMR plot parameters  
CX 21.00 cm  
FIP 10.000 ppm  
F1 4000.10 Hz  
F2P -0.500 ppm  
F2 -200.01 Hz  
PPMCH 0.50000 ppm/c  
HZCM 200.00500 Hz/cm



2.13



Current Data Parameters  
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 PROCNO 1  
 DU /h  
 USER denf

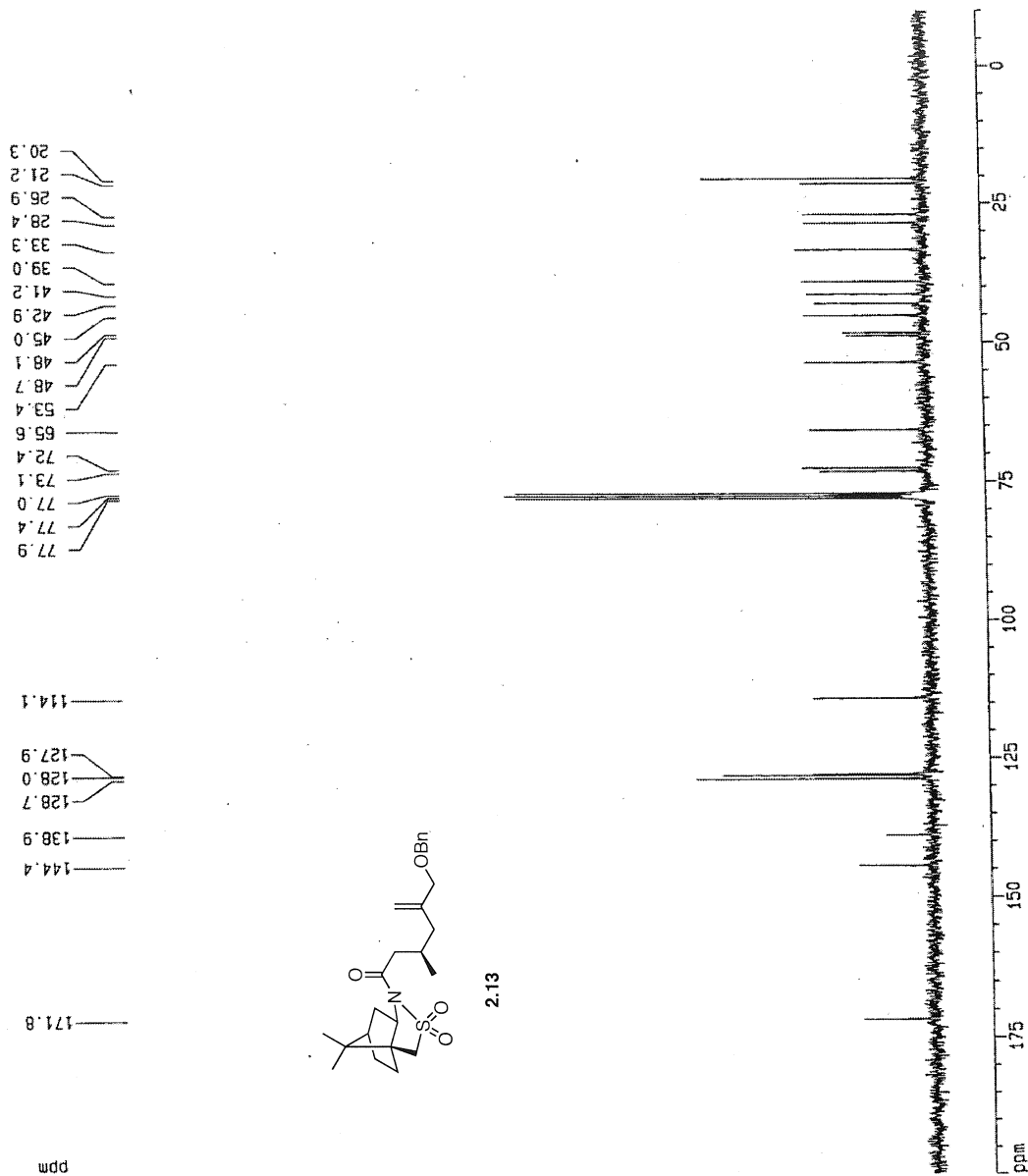
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 FIDRES 0.287350 Hz  
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 DE 6.00 usec  
 TE 300.0 K  
 D1 0.60300000 sec  
 D11 0.63000000 sec  
 D12 0.60002000 sec

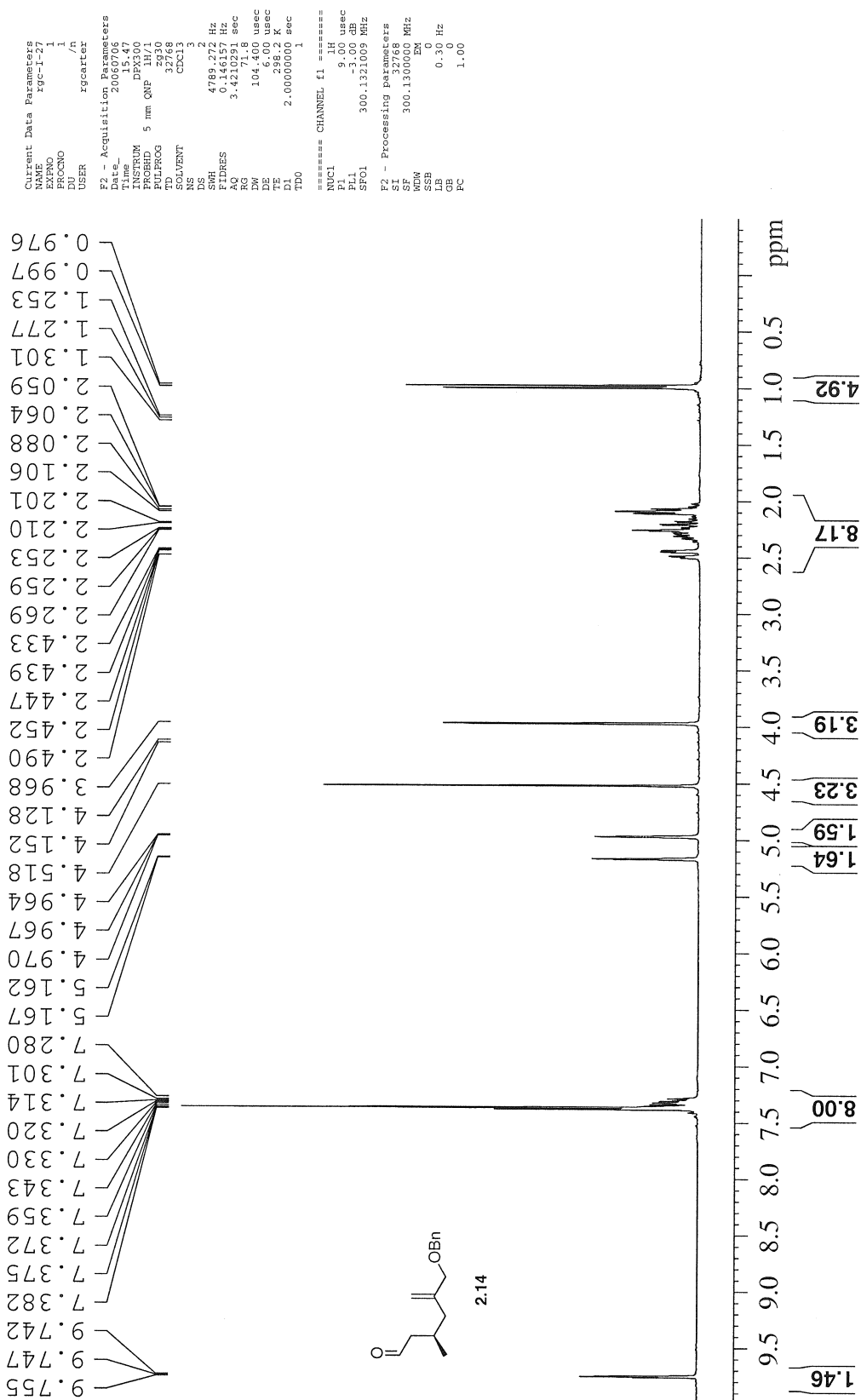
===== CHANNEL f1 =====  
 NUC1 13C  
 P1 7.60 usec  
 PL1 -3.00 dB  
 SF01 75.4756431 MHz

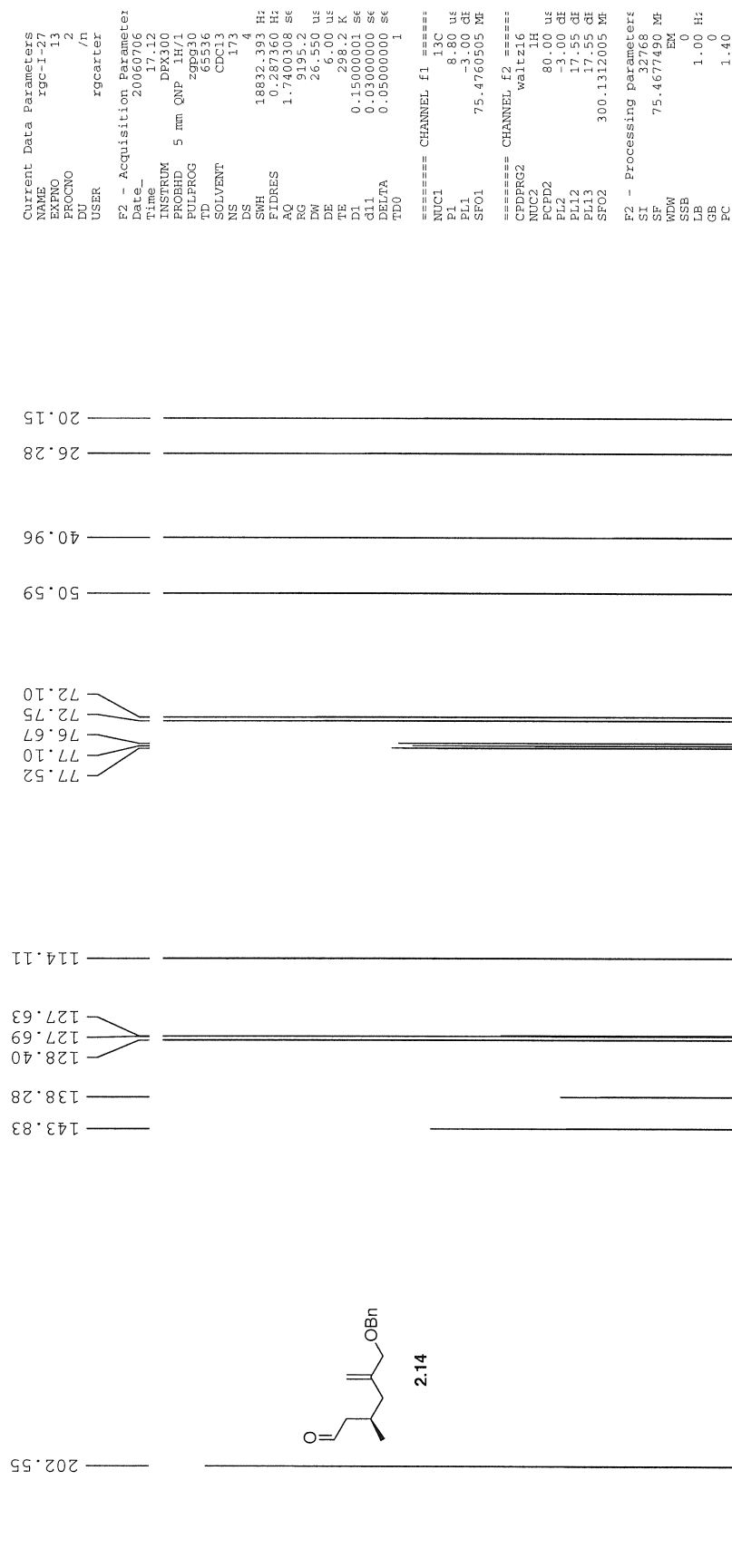
===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 80.00 usec  
 PL2 2.00 dB  
 PL12 17.55 dB  
 PL13 19.00 dB  
 SF02 300.1315007 MHz

F2 - Processing parameters  
 SI 131072  
 SF 75.4577150 MHz  
 MDW EM  
 SSB 0  
 LB 3.00 Hz  
 GB 0  
 PC 1.40

1D NMR plot parameters  
 CX 30.00 cm  
 F1P 200.000 ppm  
 F1 15093.54 Hz  
 F2P -10.000 ppm  
 F2 -754.88 Hz  
 PPMH 10.50000 ppm/cm  
 HZCM 792.41107 Hz/cm







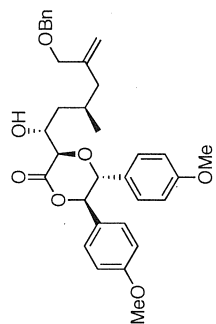
Current Data Parameters  
 NAME DF496\_A1001  
 EXPNO 14  
 PROCNO 1  
 DU /n  
 USER danf

F2 - Acquisition Parameters  
 Date\_ 20040707  
 Time 17.24  
 INSTRUM spect  
 PROBHD 5 mm BBO Z-G  
 PULPROG zgpg30  
 TO 32768  
 SOLVENT CDCl3  
 NS 32  
 DS 2  
 SWH 5995.204 Hz  
 FIDRES 0.162959 Hz  
 AQ 2.7329011 sec  
 RG 512  
 DM 63.400 usec  
 DE 6.00 usec  
 TE 300.0 K  
 D1 2.0000000 sec

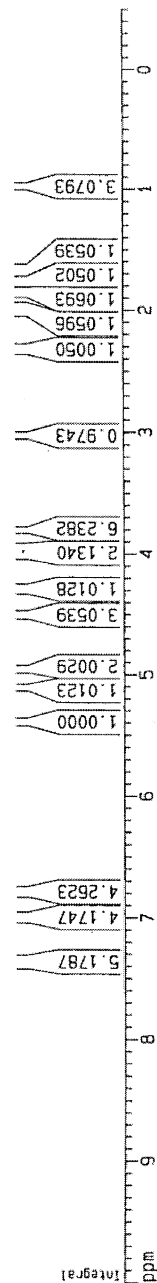
===== CHANNEL f1 =====  
 NUC1 1H  
 P1 11.30 usec  
 PL1 0.00 dB  
 SF01 400.0126001 MHz

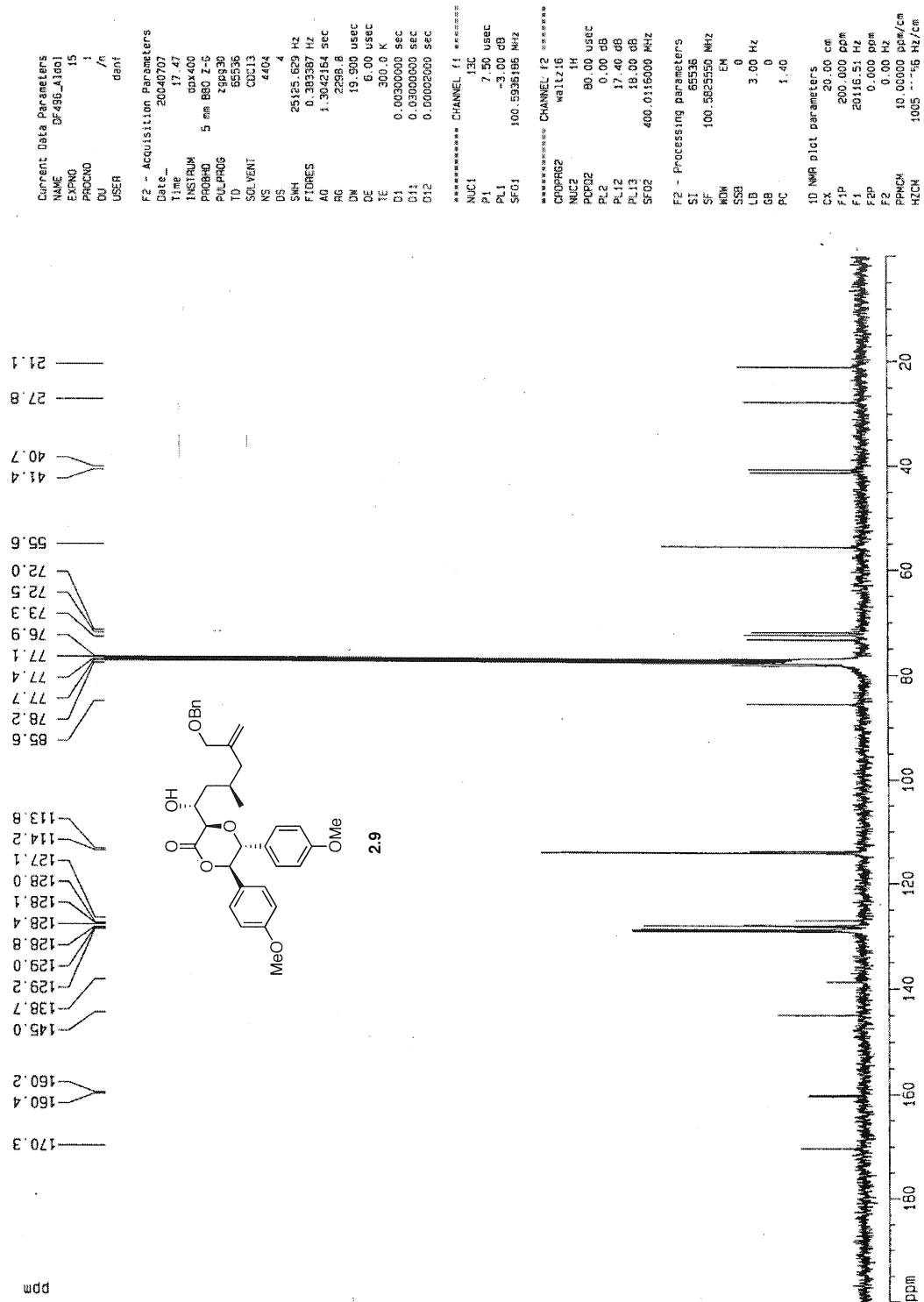
F2 - Processing parameters  
 SI 65536  
 SF 400.0100000 MHz  
 NQW EN  
 SSB 0  
 LB 0.70 Hz  
 GB 0  
 PC 1.00

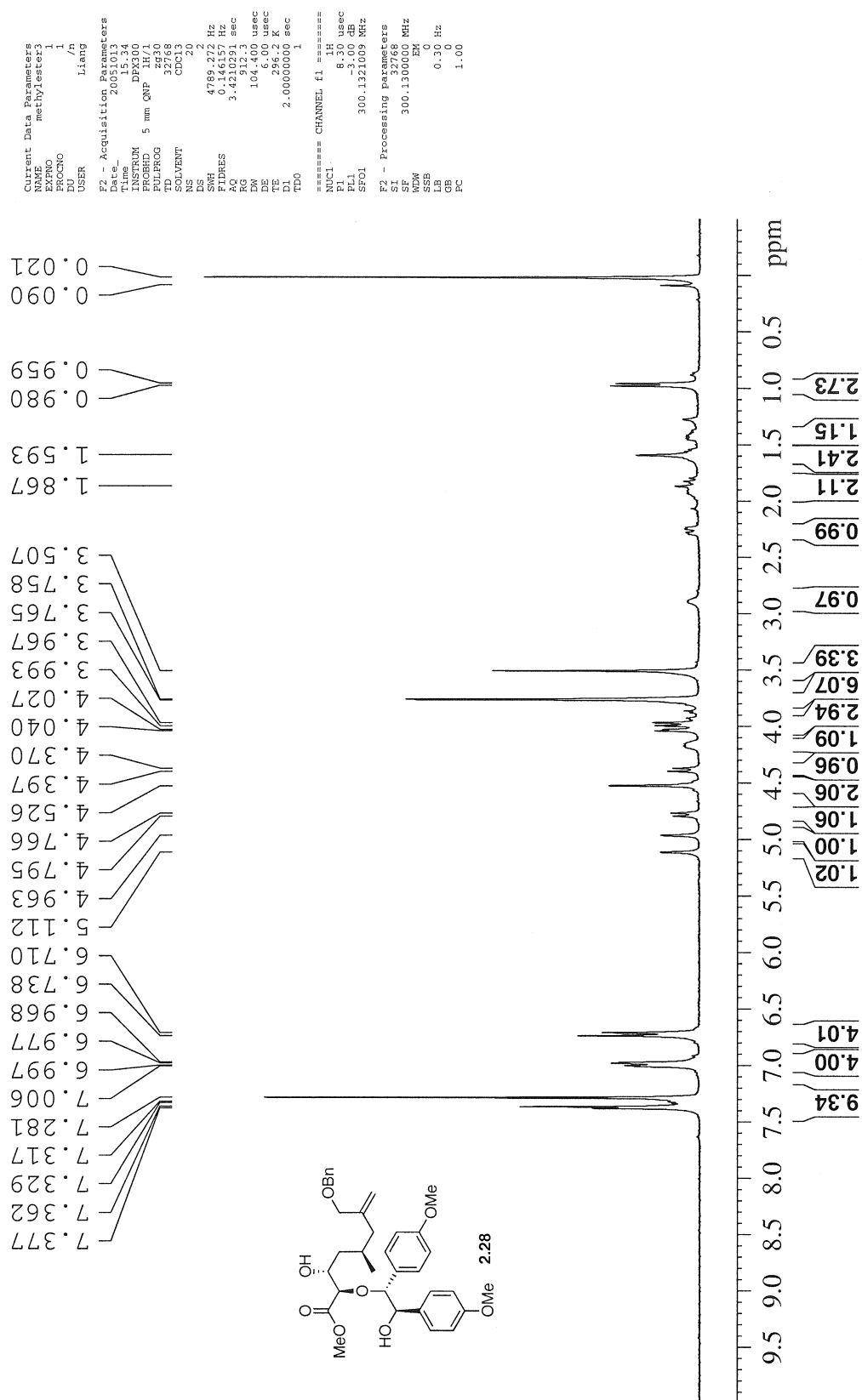
1D NMR plot parameters  
 CX 21.00 cm  
 F1P 10.000 ppm  
 F1 4000.10 Hz  
 F2P -0.500 ppm  
 F2 -200.01 Hz  
 PPMCK 0.50000 ppm/c  
 HZCM 200.00500 Hz/cm

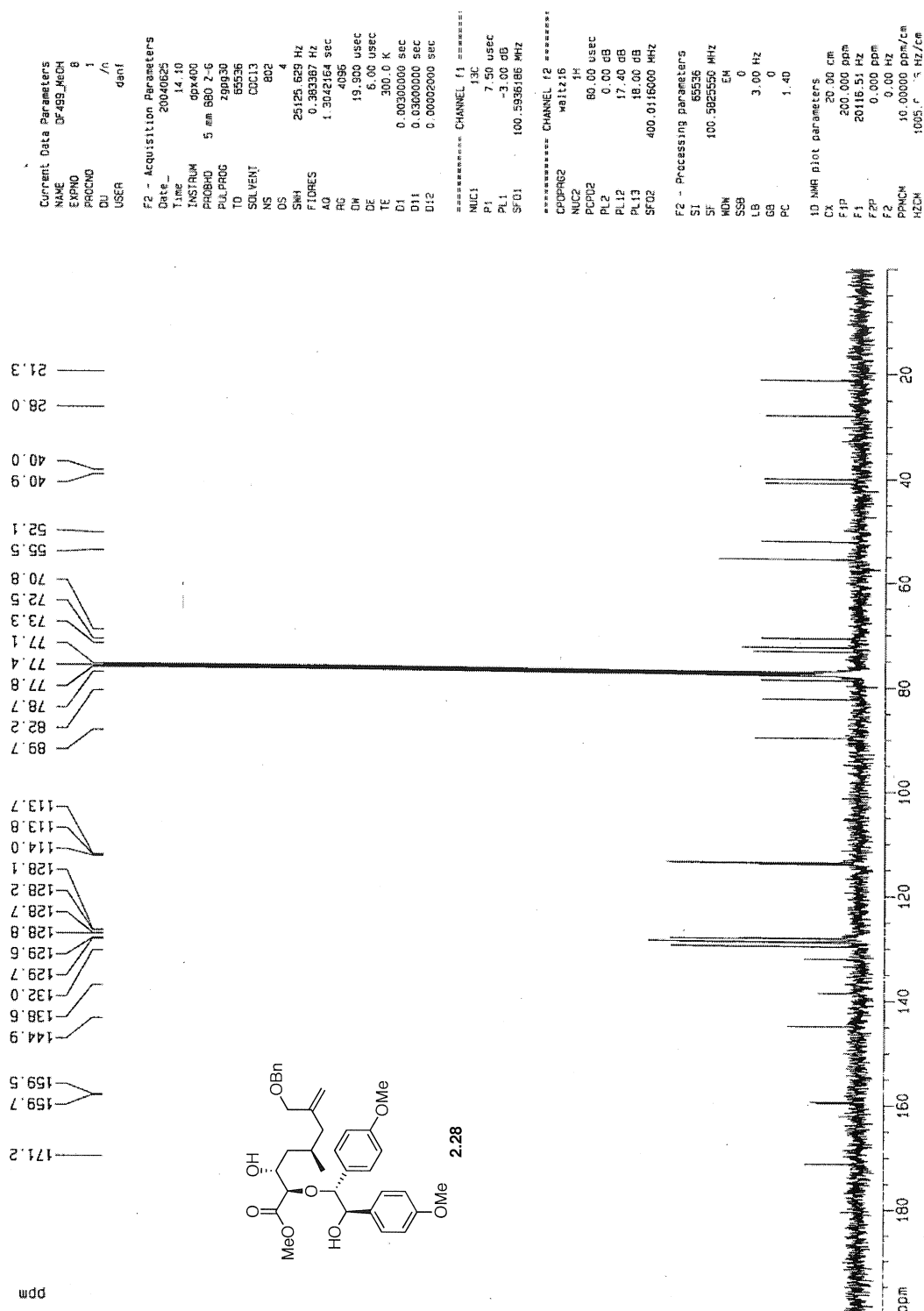


2.9

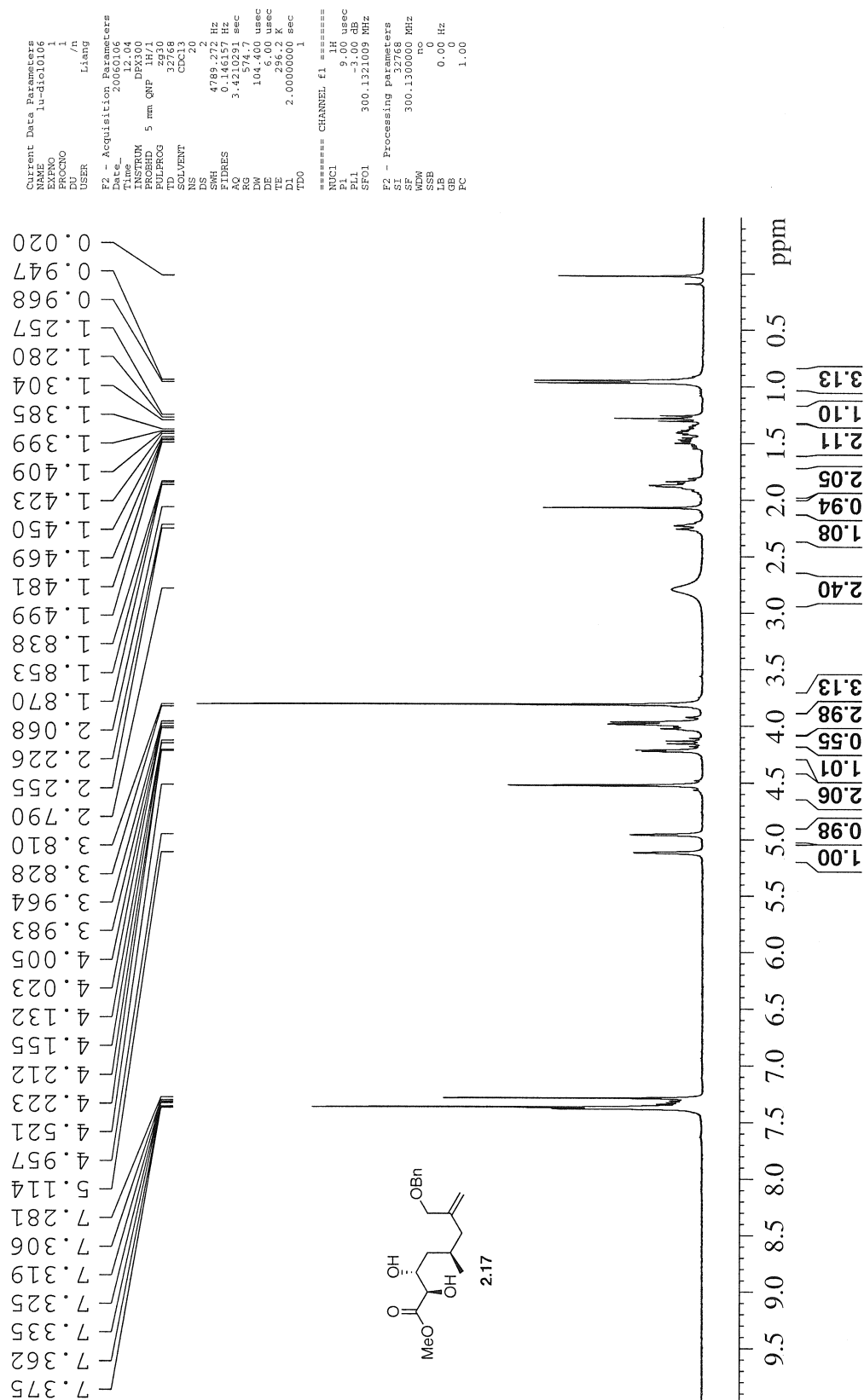












Current Data Parameters  
 NAME: 2F500\_CAN  
 EXPNO: 7  
 PROCNO: 1  
 DU: /n  
 USER: dant

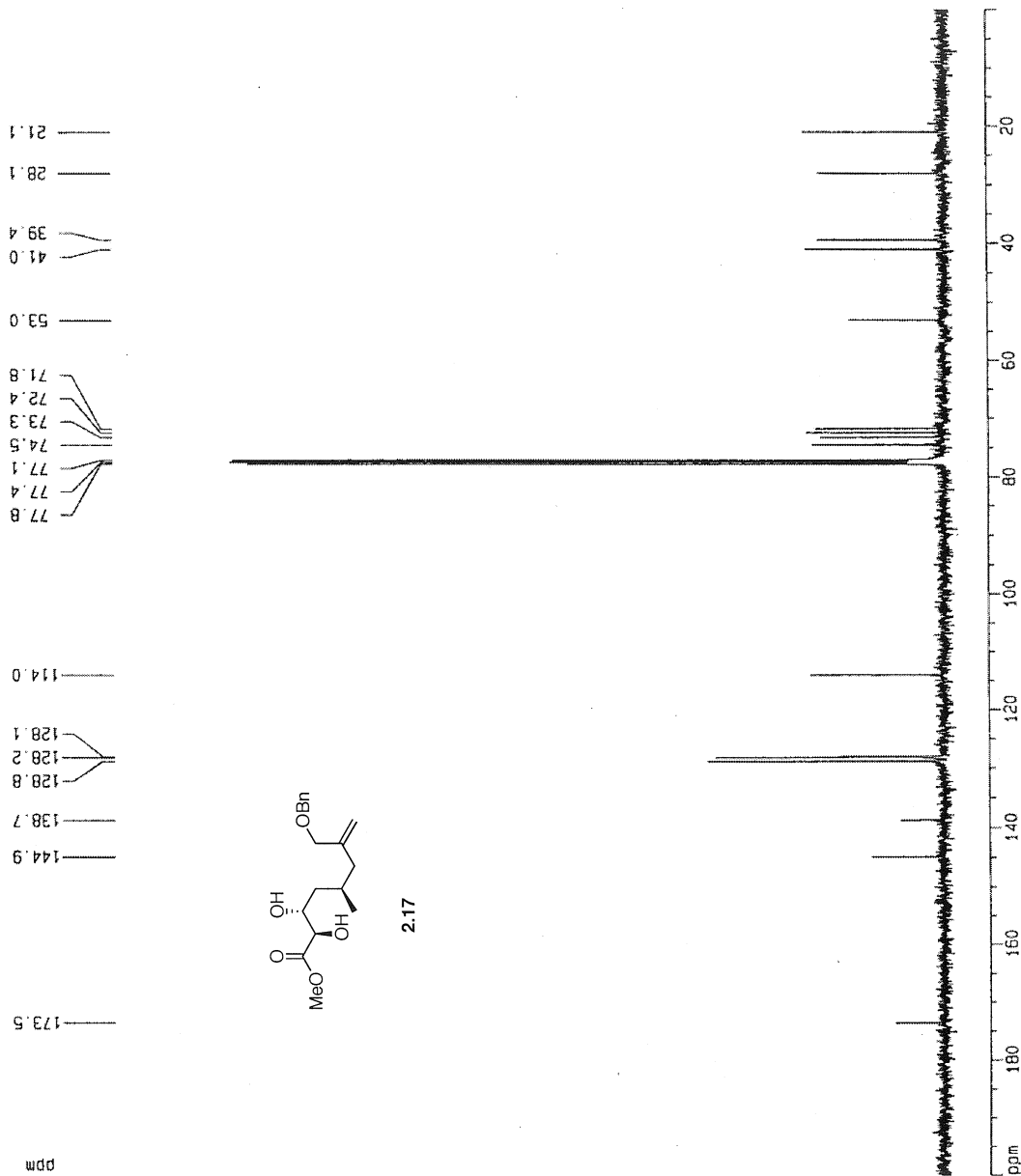
F2 - Acquisition Parameters  
 Date\_: 20040423  
 Time: 16.02  
 INSTRUM: dd400  
 PROBO: 5 mm BBO 2-G  
 PULPROG: zgpg30  
 TD: 65536  
 SOLVENT: CDCl3  
 NS: 1024  
 DS: 4  
 SWH: 25125.629 Hz  
 FIDRES: 0.383387 Hz  
 AQ: 1.3042164 sec  
 RG: 1024  
 DW: 19.900 usec  
 DE: 6.00 usec  
 TE: 300.0 K  
 D1: 0.00300000 sec  
 D11: 0.03000000 sec  
 D12: 0.00020000 sec

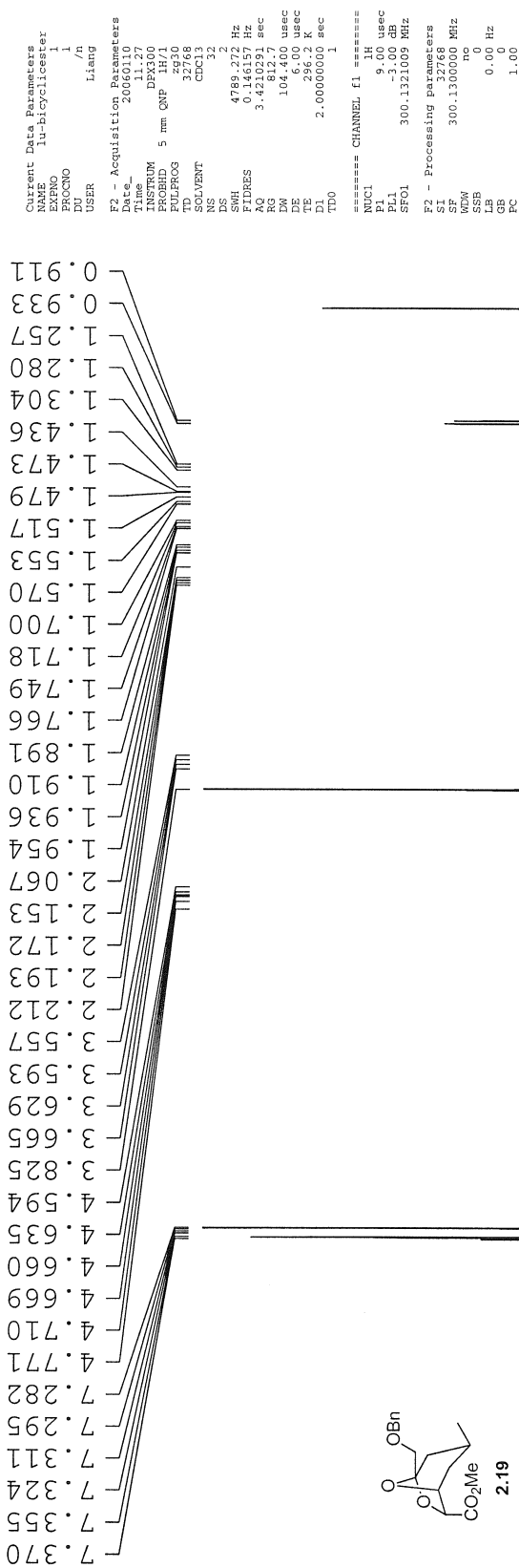
===== CHANNEL f1 =====  
 NUC1: 13C  
 P1: 7.50 usec  
 PL1: -3.00 dB  
 SF01: 100.5936186 MHz

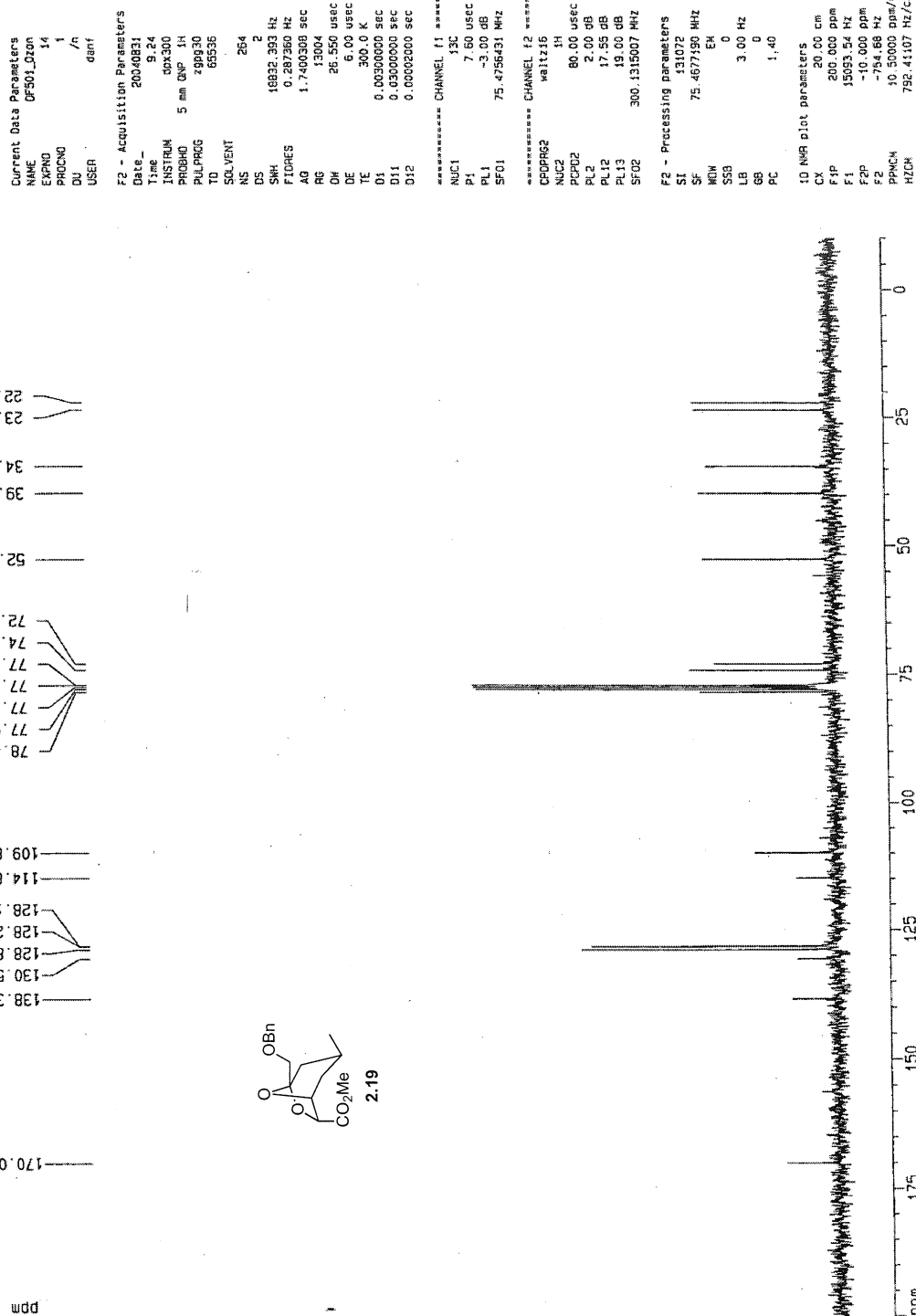
===== CHANNEL f2 =====  
 CPDPRG2: waltz16  
 NUC2: 1H  
 PCPD2: 80.00 usec  
 PL2: 0.00 dB  
 PL12: 17.40 dB  
 PL13: 18.00 dB  
 SF02: 400.0115000 MHz

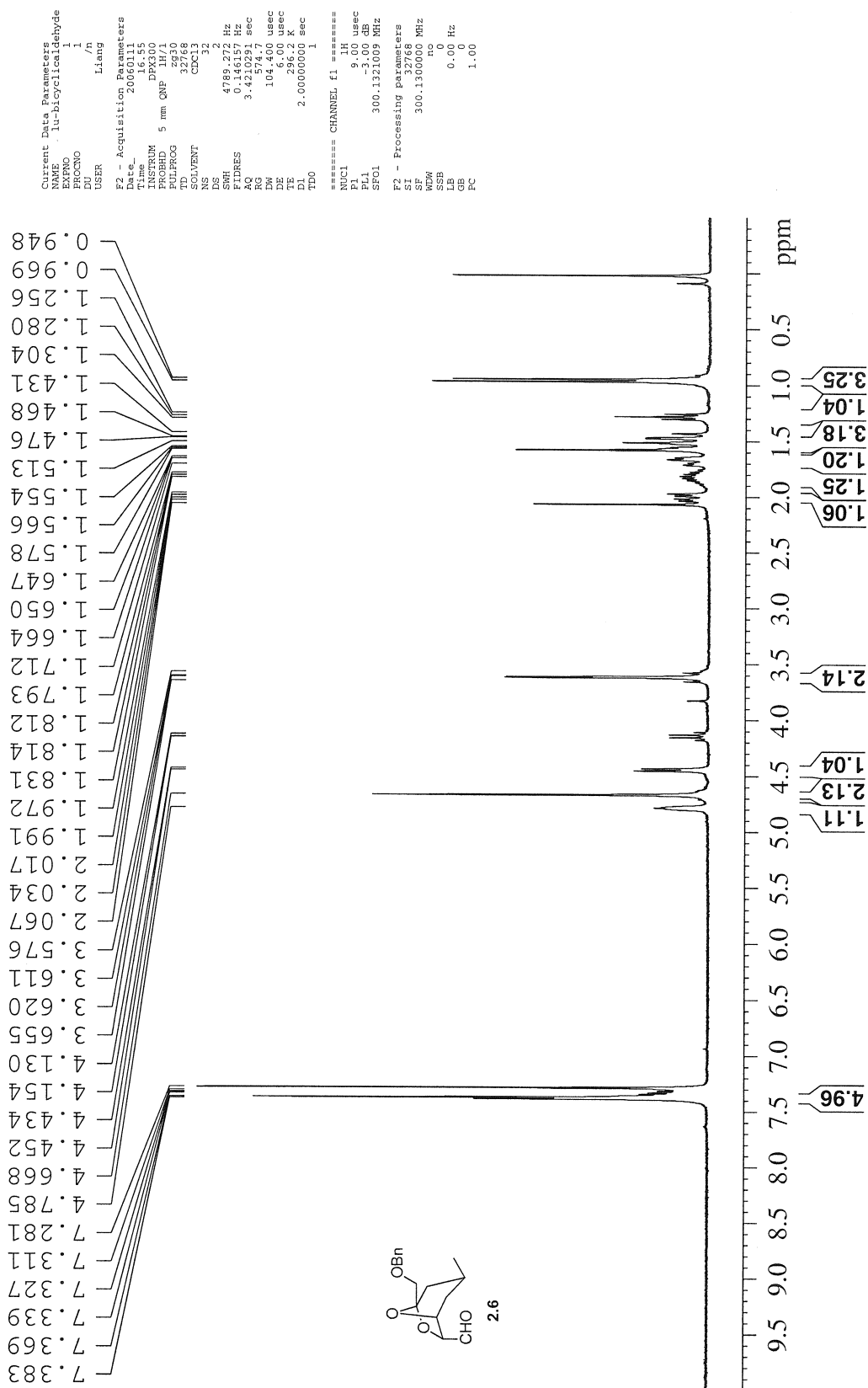
F2 - Processing parameters  
 SI: 65536  
 SF: 100.5825550 MHz  
 WDW: EM  
 SSB: 0  
 LB: 3.00 Hz  
 GB: 0  
 PC: 1.40

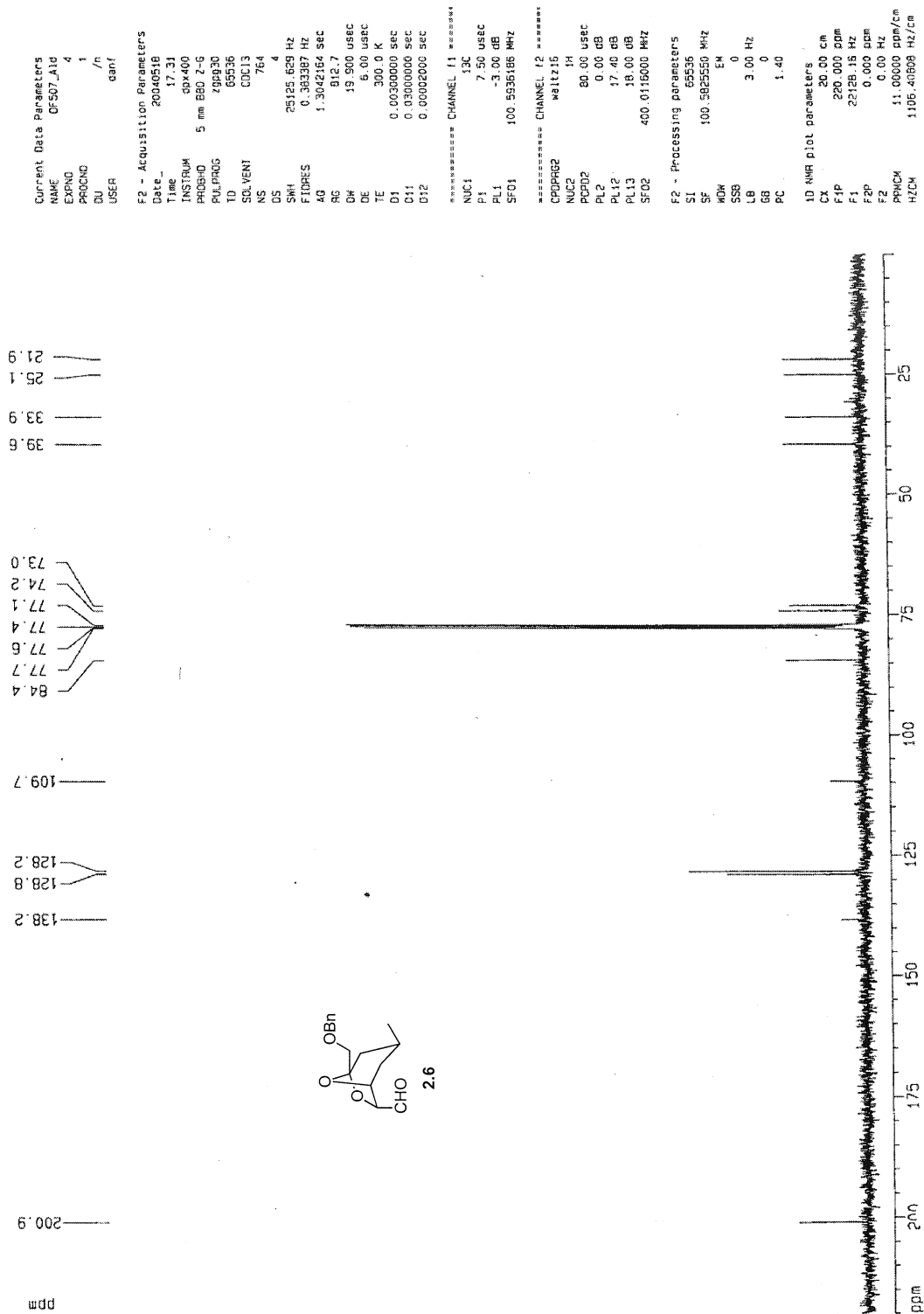
10 NMR plot parameters  
 CX: 20.00 cm  
 FIP: 200.000 ppm  
 F1: 20116.51 Hz  
 F2: 0.000 ppm  
 F3: 0.00 Hz  
 PPMH: 10.0000 ppm/cm  
 HZCM: 1005.82556 Hz/cm









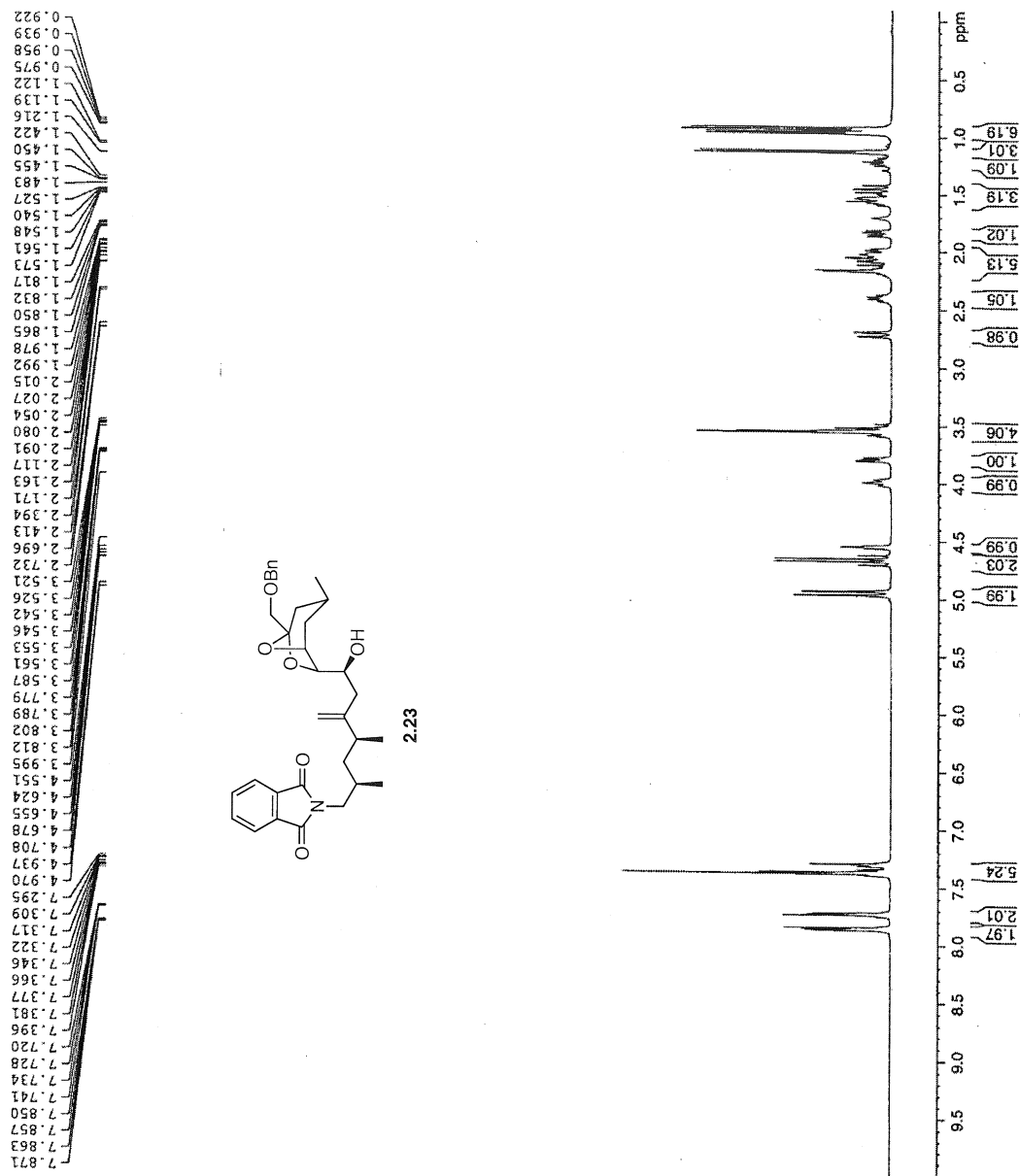


Current Data Parameters  
 NAME XZ-XI-72  
 EXNO 1  
 PROCNO 2  
 EXPNO 1  
 USER xzhou

F2 - Acquisition Parameters  
 Date\_ Time 20080819 19:47  
 INSTRN DFX400  
 PROBD 5 mm BBO BB-1H  
 PULPROG zgpg30  
 TD 32768  
 SOLVENT CDCl3  
 NS 32  
 DS 4  
 SWH 6410.252 Hz  
 FIDRES 0.195625 Hz  
 AQ 2.5559840 sec  
 RG 143.7  
 SH 76.000 usec  
 DE 6.000 usec  
 TE 298.2 K  
 D1 2.0000000 sec  
 TDO 1

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 14.70 usec  
 PL 0.00 dB  
 STOL 400.0128001 MHz

F2 - Processing parameters  
 SI 32768  
 SF 400.0100000 MHz  
 MDW 0  
 SSB 0  
 LB 0.70 Hz  
 GB 0  
 PC 1.00

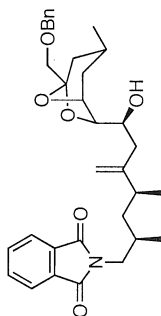
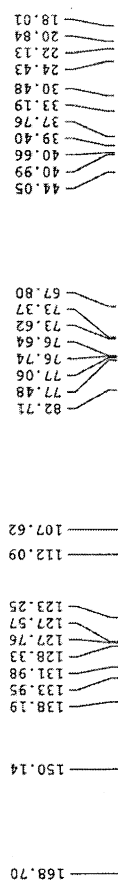


C13

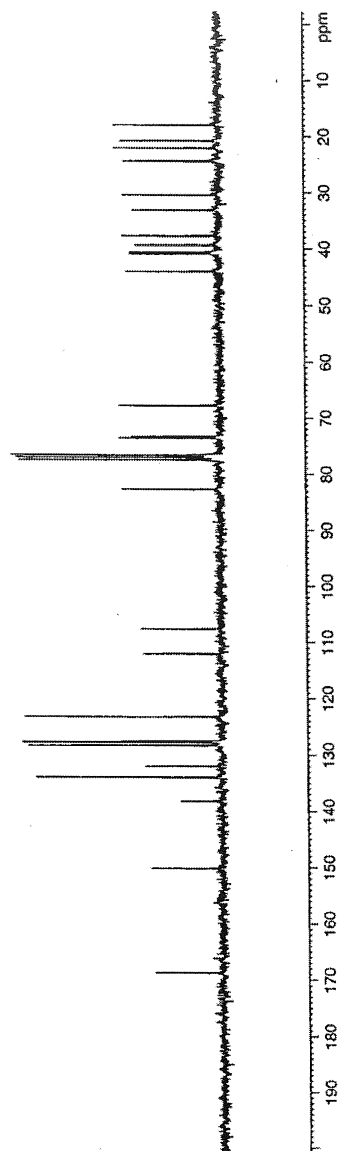
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Current Data Parameters
NAME      XZ-XI-89
EXPNO     8
PROCNO    1
INSTRUM    5
PULPROG    zgpg30
SOLVENT    CDCl3
NS         284
DS         4
SMH        16832.393 Hz
FIDRES     0.287360 Hz
AQ         1.148155 sec
RG         919.2
DM         26.550 usec
DE         6.00 usec
TE         300.2 K
D1         0.15000000 sec
d11        0.03000000 sec
DELTA     0.05000000 sec
TD        1
===== CHANNEL f1 =====
NUC1       13C
P1         8.80 usec
PL         0.00 dB
SFO1       75.4760505 MHz
===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
PCPD2      80.00 usec
P22        19.00 dB
P23        17.50 dB
P24        17.50 dB
P25        17.55 dB
SFO2       300.1312005 MHz
===== Processing parameters =====
SI         32768
SF         75.4677490 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
EC         1.40

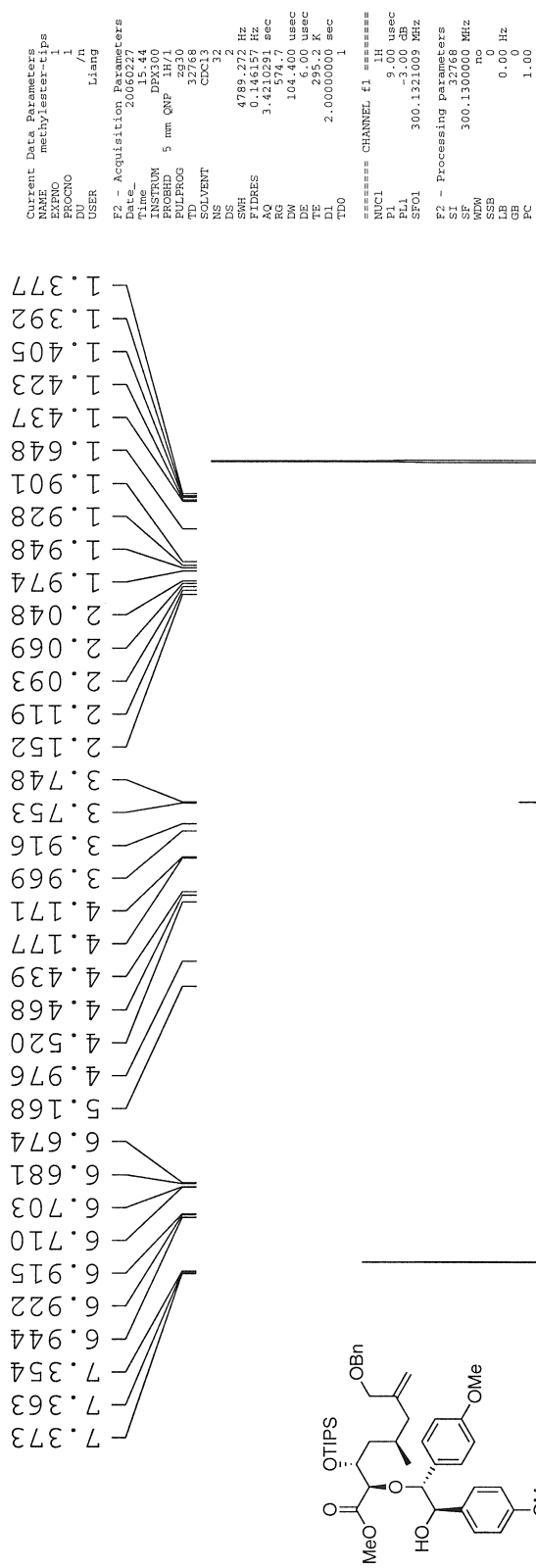
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2.23







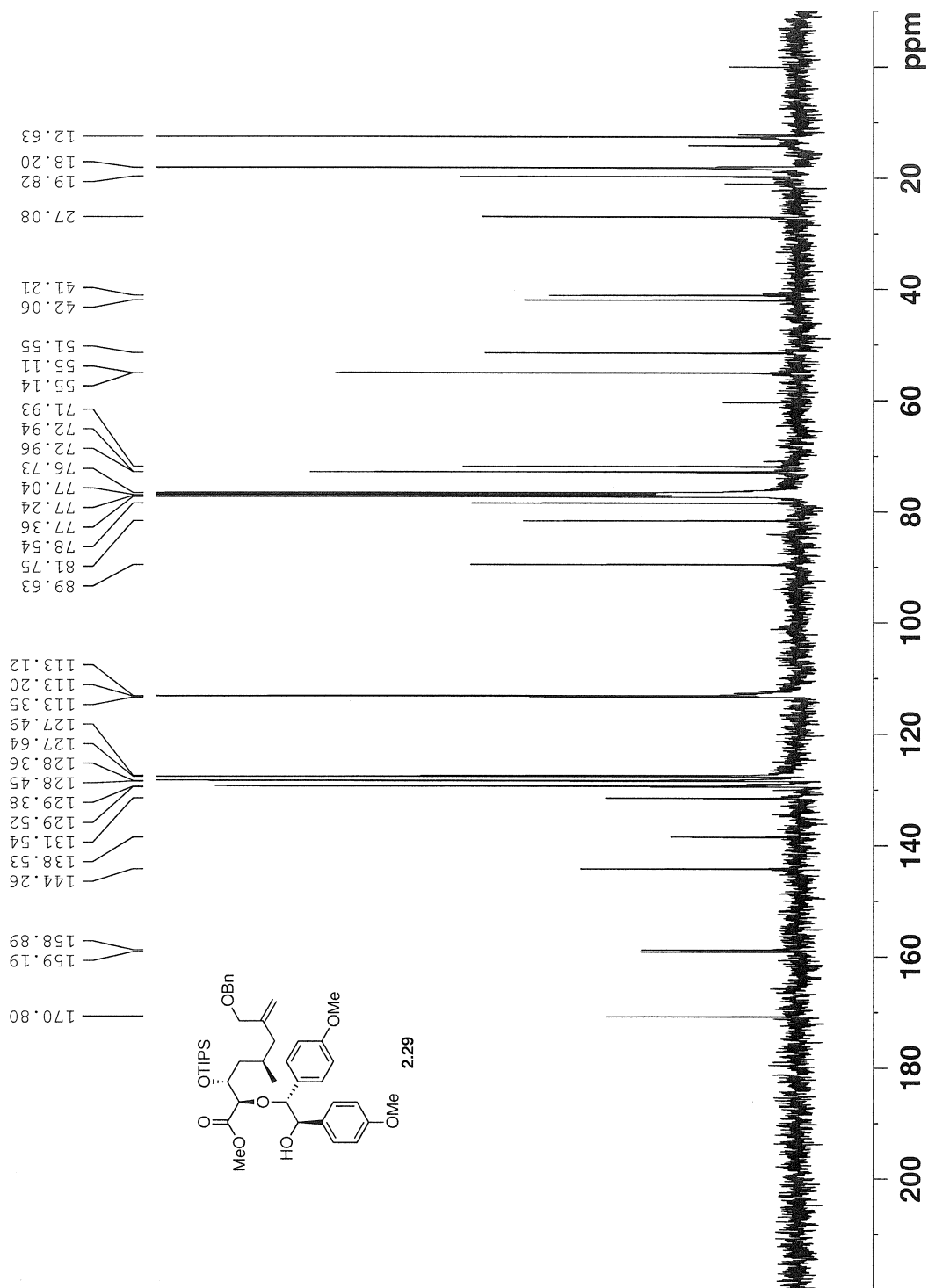
Current Data Parameters  
NAME LUL-033106-1  
EXPNO 2  
PROCNO 1  
DU /n  
USER liang

F2 - Acquisition Parameters  
Date\_ 20060331  
Time 15.07  
INSTRUM DFX400  
PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
TD 65536  
SOLVENT CDCl3  
NS 1504  
DS 4  
SWH 25125.629 Hz  
FIDRES 0.383387 Hz  
AQ 1.3042164 sec  
RG 7298.2  
DW 19.900 usec  
DE 6.00 usec  
TE 298.2 K  
D1 0.15000001 sec  
d11 0.03000000 sec  
DELTA 0.05000000 sec  
TD0 1

===== CHANNEL f1 =====  
NUC1 13C  
P1 7.80 usec  
PL1 -3.00 dB  
SFO1 100.5936591 MHz

===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 135.00 usec  
PL2 17.40 dB  
PL12 17.40 dB  
PL13 17.40 dB  
SFO2 400.0116000 MHz

F2 - Processing parameters  
SI 32768  
SF 100.5825950 MHz  
WDW EM  
SSB 0  
LB 3.00 Hz  
GB 0  
FC 1.40

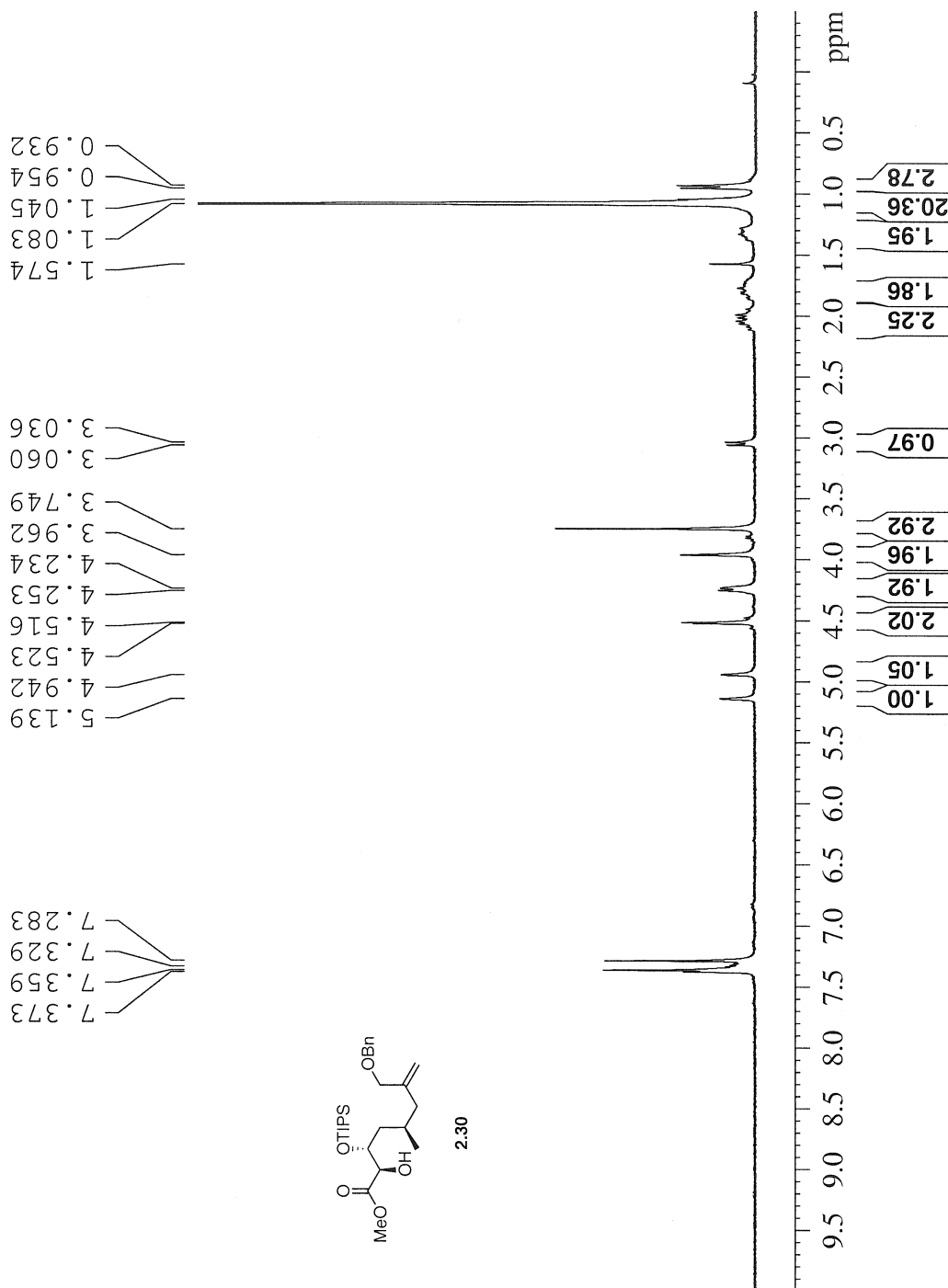


Current Data Parameters  
 NAME E11-030206-5  
 EXPNO 1  
 PROCNO 1  
 DU /n  
 USER Liang

F2 - Acquisition Parameters  
 Date\_ 20060302  
 Time 16:57  
 INSTRUM spect  
 PROBD 5 mm QNP 1H/1  
 PULPROG zg30  
 TD 32768  
 SOLVENT CDCl3  
 NS 14  
 DS 2  
 SWH 4789.272 Hz  
 FIDRES 0.146157 Hz  
 AQ 3.420291 sec  
 RG 327.68  
 DW 104.400 usec  
 DE 6.00 usec  
 TE 295.2 K  
 D1 2.0000000 sec  
 TDO 1

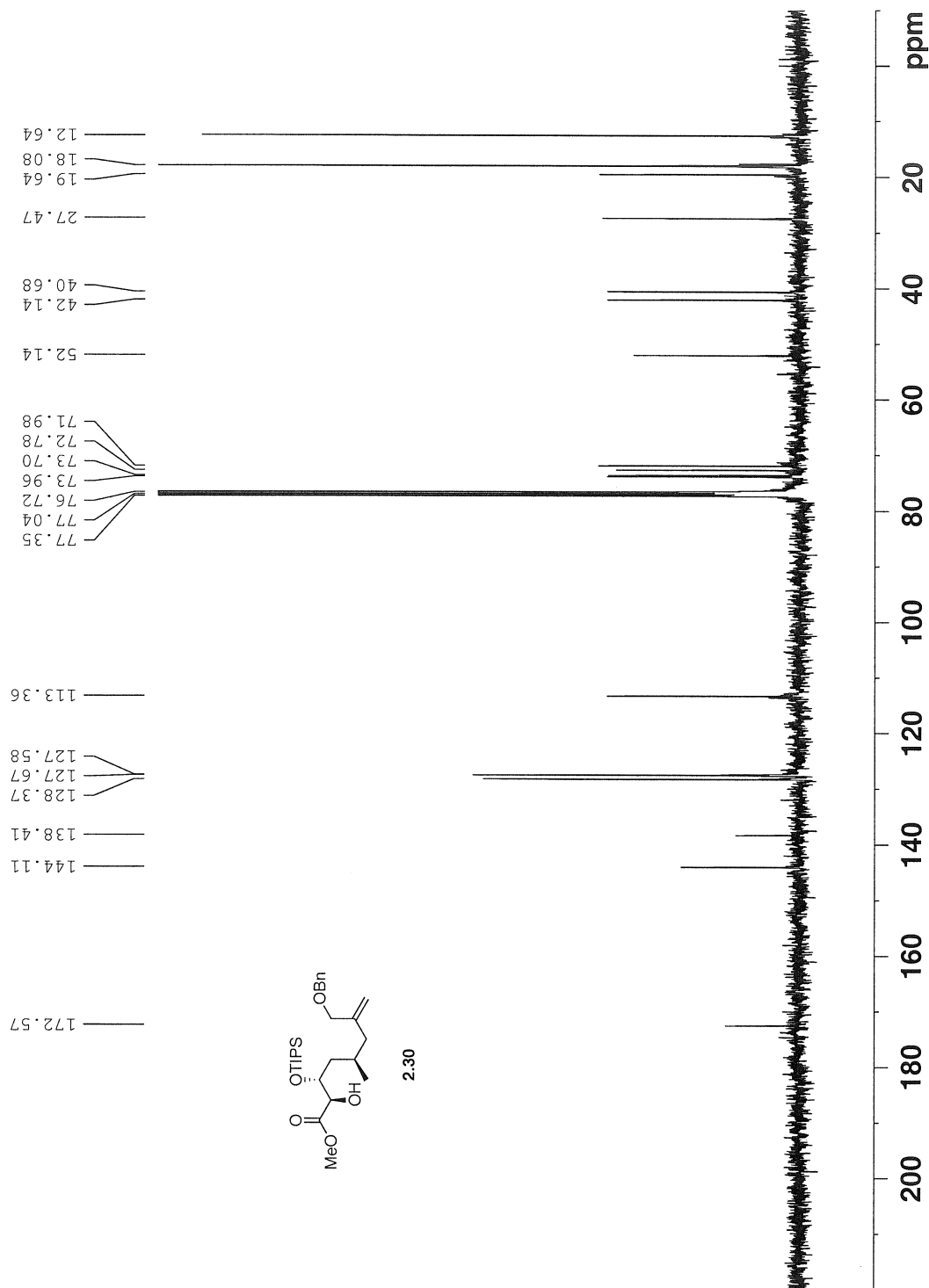
===== CHANNEL f1 =====  
 NUC1 1H  
 P1 9.00 usec  
 PL1 -1.00 dB  
 SFO1 300.1321009 MHz

F2 - Processing parameters  
 SI 32768  
 SF 300.1300000 MHz  
 WDW no  
 SSB 0  
 LB 0.00 Hz  
 GB 0  
 PC 1.00



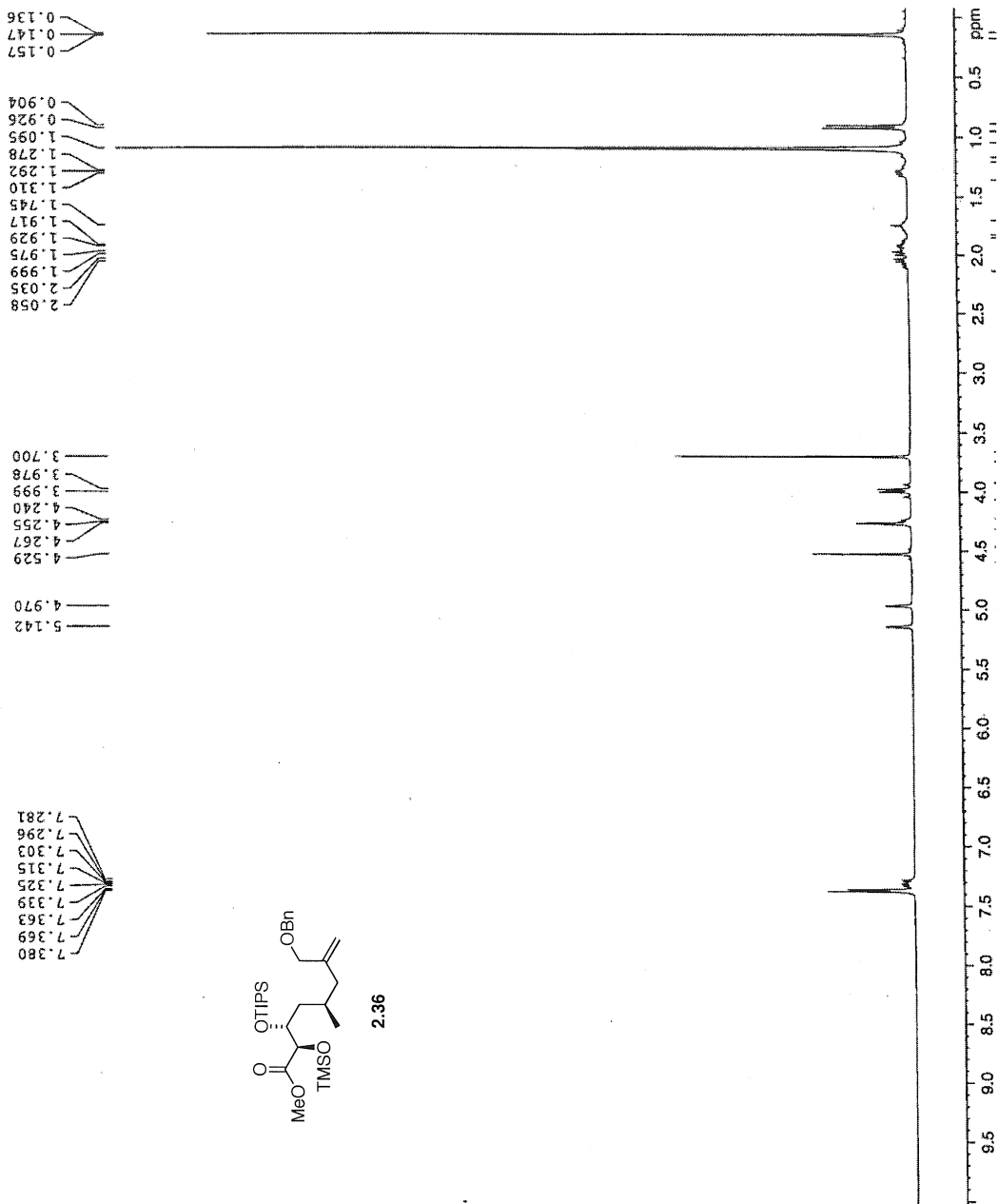


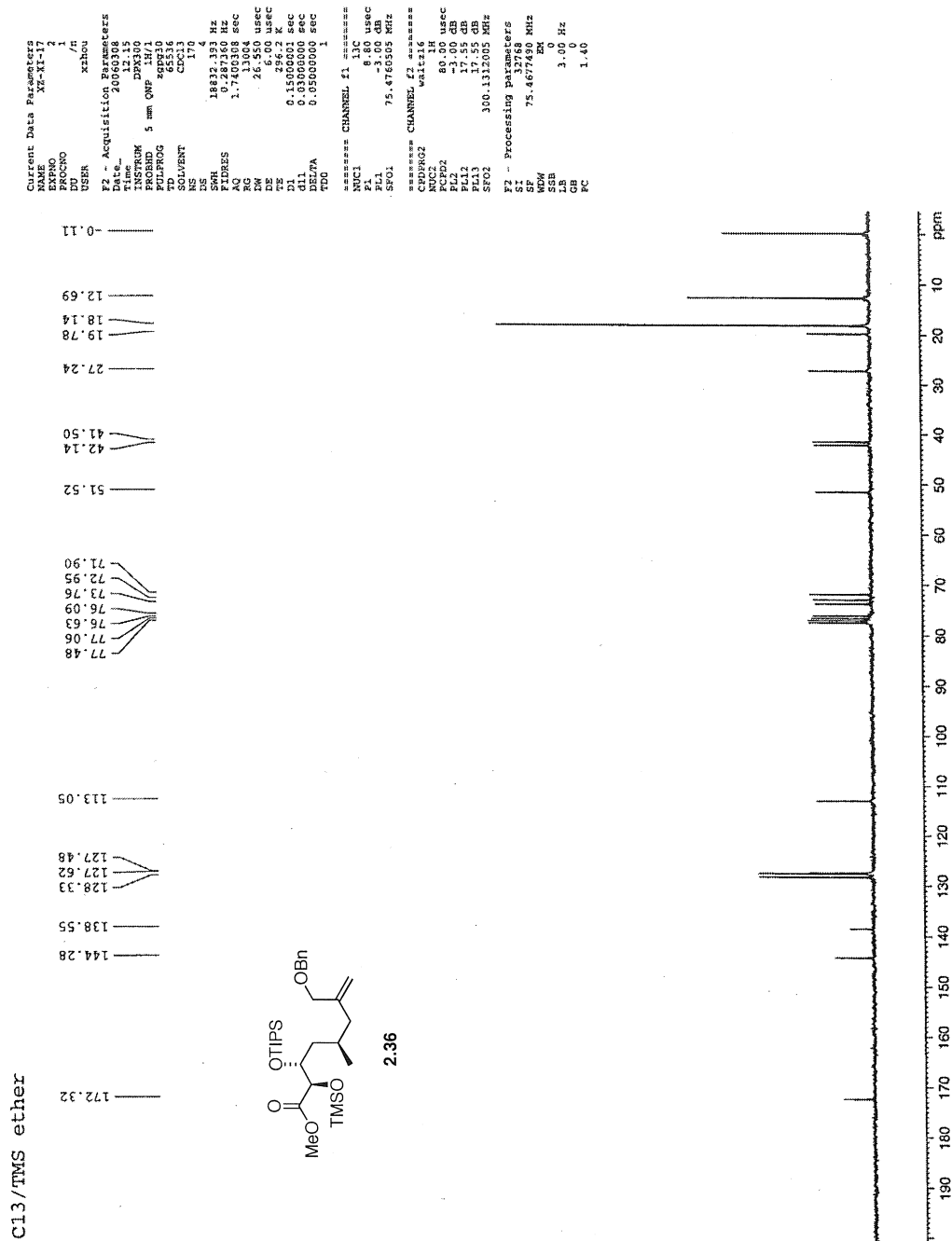
2.30



Current Data Parameters  
 NAME LUL-27-exp040606  
 EXPNO 2  
 PROCNO 1  
 DU /n  
 USER Liang  
 F2 - Acquisition Parameters  
 Date\_ 20060407  
 Time 10.31  
 INSTRUM DEX400  
 PROBHD 5 mm BBO BB-1H  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT  
 NS 1780  
 DS 4  
 SWH 25125.629 Hz  
 FIDRES 0.383387 Hz  
 AQ 1.3042164 s  
 RG 4597.6  
 DW 19.900 us  
 DE 6.00 us  
 TE 298.2 K  
 D1 0.15000001 s  
 d11 0.03000000 s  
 DELTA 0.05000000 s  
 TD0 1  
 ===== CHANNEL f1 =====  
 NUC1 <sup>13</sup>C  
 P1 7.80 us  
 PL1 -2.00 dB  
 SFO1 100.5936591 MHz  
 ===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 <sup>1</sup>H  
 PCPD2 135.00 us  
 PL2 17.40 dB  
 PL12 17.40 dB  
 PL13 17.40 dB  
 SFO2 400.0116000 MHz  
 F2 - Processing parameters  
 SI 32768  
 SF 100.5825950 MHz  
 WDW EM  
 SSB 0  
 LB 3.00 Hz  
 GB 0  
 PC 1.40

Current Data Parameters  
 NAME XZ-XI-17  
 EXPNO 1  
 F2 - Acquisition Parameters  
 F2 200.136 MHz  
 INSTRUM DEK300  
 PROBHD 5 mm QNP 1H/1  
 PULPROG zgpg30  
 TD 32768  
 SFO1 300.136000 MHz  
 NS 32  
 DS 2  
 SWH 4789.272 Hz  
 FIDRES 0.14617 Hz  
 AQ 3.421453 sec  
 RG 327.68  
 DQ 194.400 usec  
 DE 6.00 usec  
 TE 295.2 K  
 TF 2.0000000 sec  
 TPO 1  
 ===== CHANNEL f1 =====  
 NUC1 1H  
 P1 9.00 usec  
 PL -3.00 dB  
 SFO1 300.1361003 MHz  
 F2 - Processing parameters  
 SI 32768  
 SF 300.136000 MHz  
 WSW 0  
 SSB 0  
 LB 0.30 Hz  
 GR 0  
 PC 1.00



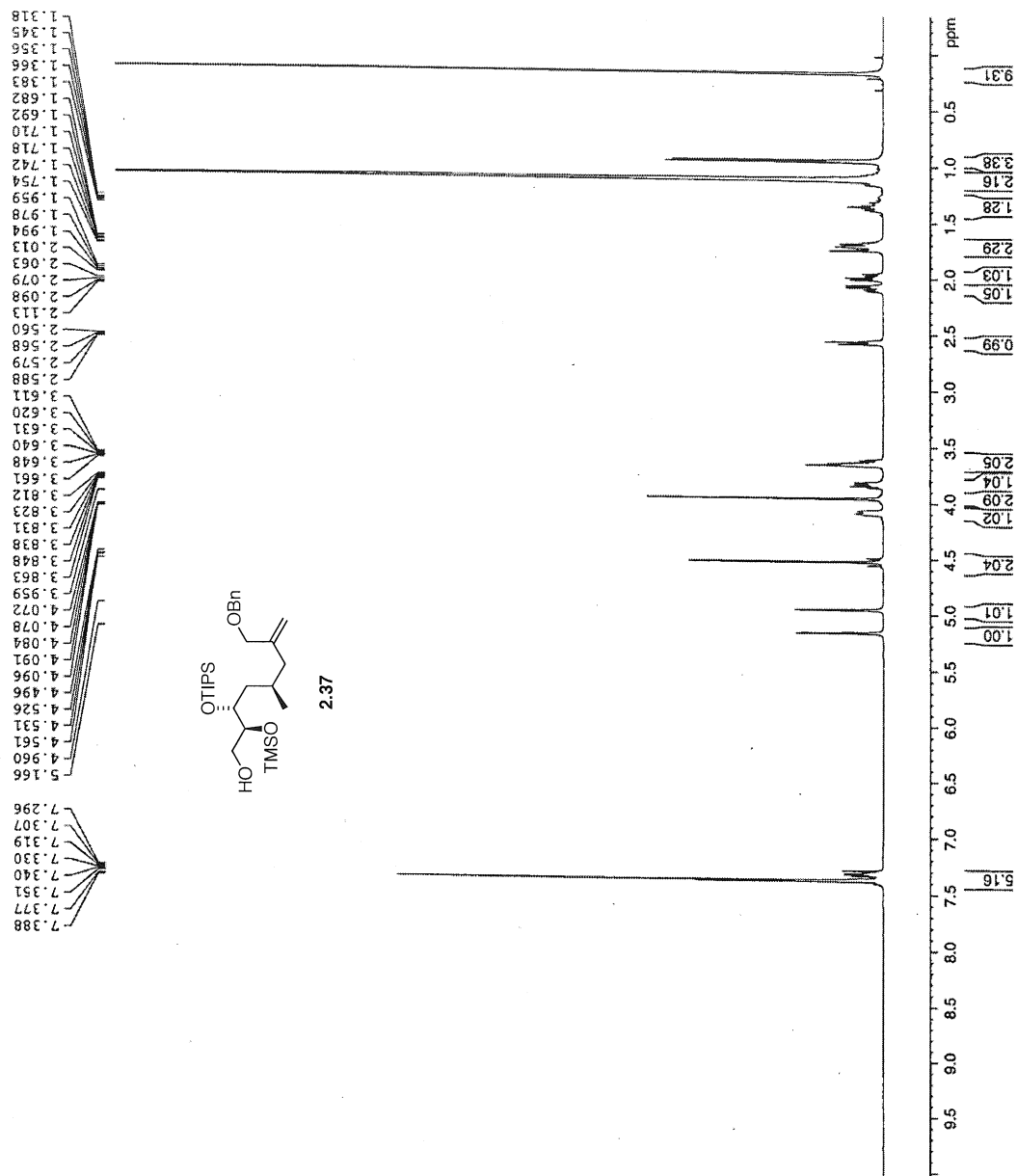


Current Data Parameters  
 NAME XZ-XI-92  
 PROGNO 1  
 PROCNO 1  
 DU /m  
 USER xzhou

F2 - Acquisition Parameters  
 Date\_ 20060726  
 Time 17.18  
 INSTRUM WPA400  
 PULPROG zgpg30  
 TD 32768  
 SOLVENT CDCl3  
 NS 32  
 DS 2  
 SWH 6410.256 Hz  
 FIDRES 0.195625 Hz  
 AQ 2.555949 sec  
 EQ 78.000 usec  
 DE 6.00 usec  
 TE 298.2 K  
 D1 2.0000000 sec  
 TDO 1

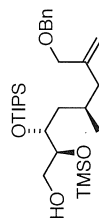
===== CHANNEL f1 =====  
 NUC1 1H  
 PL1 14.70 usec  
 PL1 0.00 dB  
 SFO1 400.0128001 MHz

F2 - Processing Parameters  
 SI 32768  
 SF 400.0100000 MHz  
 EX 0  
 LS 0  
 GB 0.70 Hz  
 PC 1.00

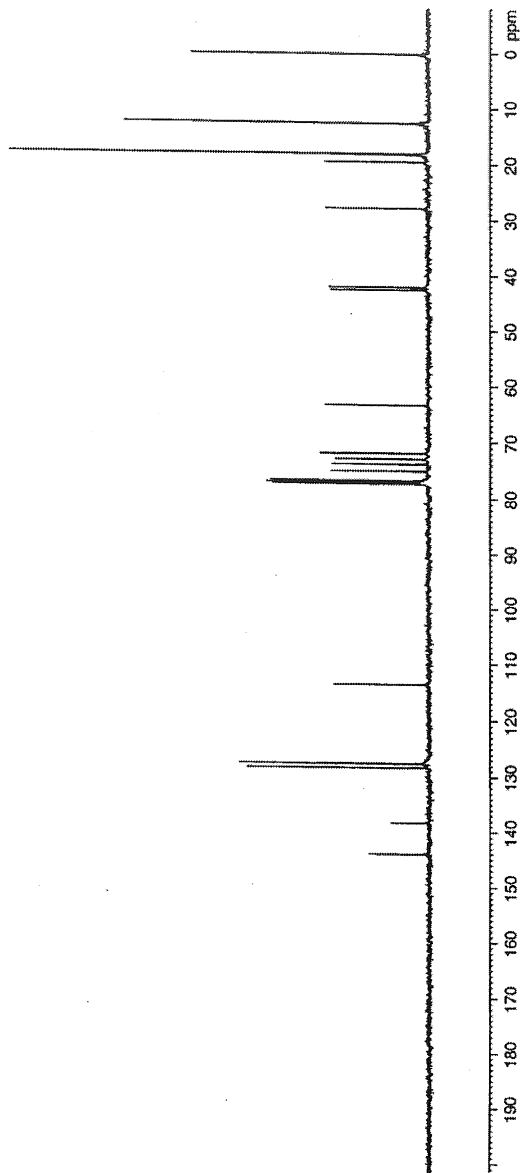


Current Data Parameters  
 NAME XZ-XI-92  
 EXPNO 1  
 PROCNO 1  
 F2 - Acquisition Parameters  
 Date\_ 20060726  
 Time 17.24  
 INSTRUM DFX400  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT DMSO  
 NS 352  
 DS 4  
 SWH 25125.629 Hz  
 FIDRES 0.383387 Hz  
 AQ 1.3042164 sec  
 RG 327.680  
 DE 6.00 usec  
 TE 298.2 K  
 DELTA 0.4500000 sec  
 CH1 0.4300000 sec  
 DELTA 0.0500000 sec  
 TDO 1  
 ===== CHANNEL f1 =====  
 NUC1 13C  
 P1 7.80 usec  
 PL1 -3.00 dB  
 SFO1 100.5936591 MHz  
 ===== CHANNEL f2 =====  
 CPOPG2 waitz16  
 NUC2 1H  
 P2 135.00 usec  
 PL2 17.40 dB  
 PL12 17.40 dB  
 PL13 17.40 dB  
 SFO2 400.0116000 MHz  
 F2 - Processing parameters  
 SI 32768  
 SF 100.5853500 MHz  
 WDW EM  
 SSB 0  
 LB 3.00 Hz  
 GB 0  
 PC 1.40

144.03  
 138.34  
 128.40  
 127.68  
 127.60  
 113.66  
 77.38  
 77.06  
 76.74  
 75.11  
 73.89  
 72.99  
 71.98  
 63.41  
 42.66  
 42.18  
 27.88  
 19.59  
 18.22  
 12.68  
 0.27



2.37



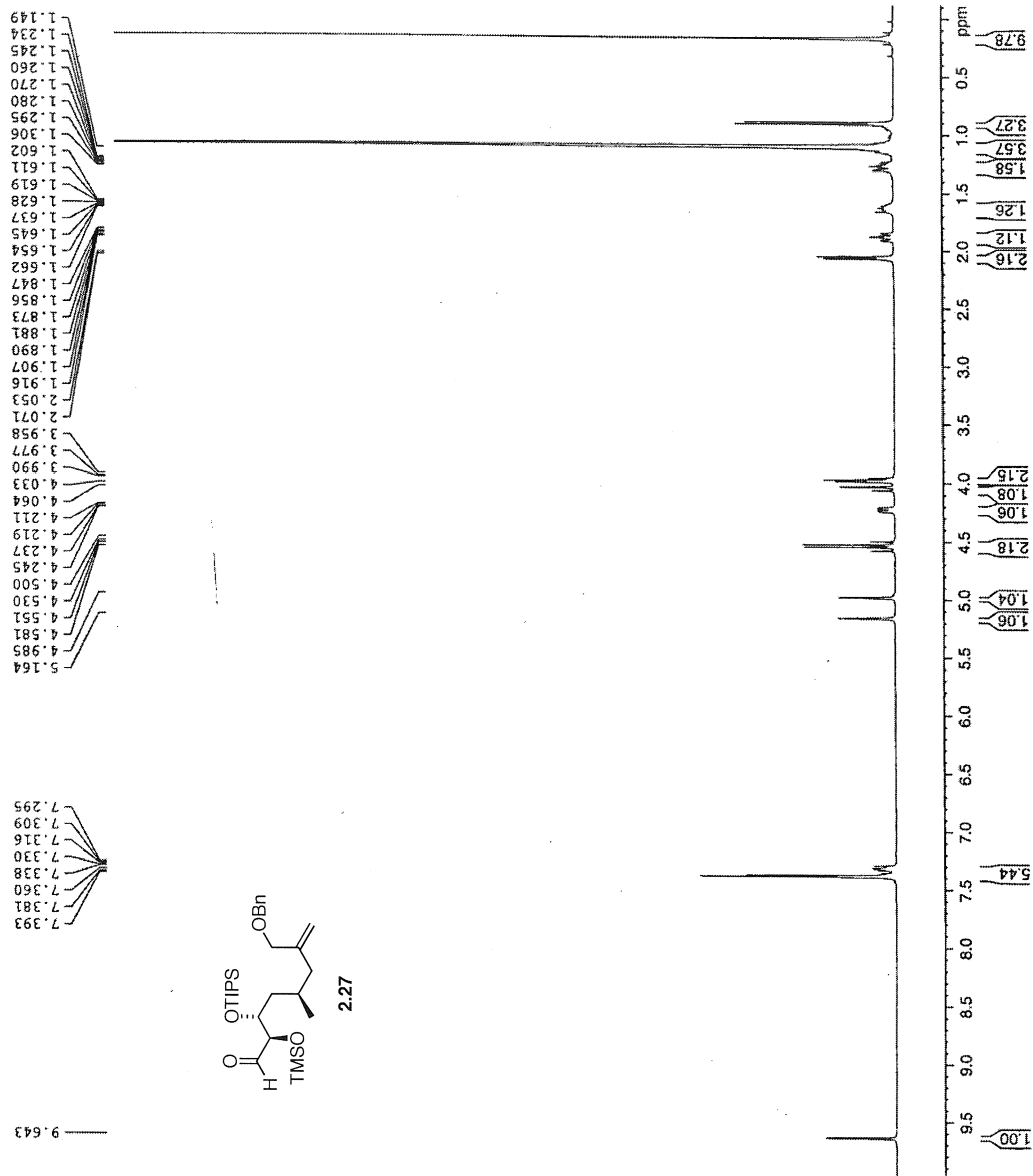


Current Data Parameters  
 NAME XZ-XI-36  
 EXPNO 1  
 PROCNO 1  
 DU /m  
 USER xzhou

F2 - Acquisition Parameters  
 Date\_ 20060330  
 Time 16.41  
 PROBNM 5 mm BBO BB-1H  
 PULPROG zg30  
 TD 32768  
 CHAN1 1  
 NS 2  
 DS 2  
 SWH 6410.256 Hz  
 FIDRES 0.121824 Hz  
 AQ 2.555550 sec  
 RG 50.8  
 FM 78.000 usec  
 SFO 400.146000 MHz  
 PC 258.2 usec  
 D1 2.0000000 sec  
 TD0 1

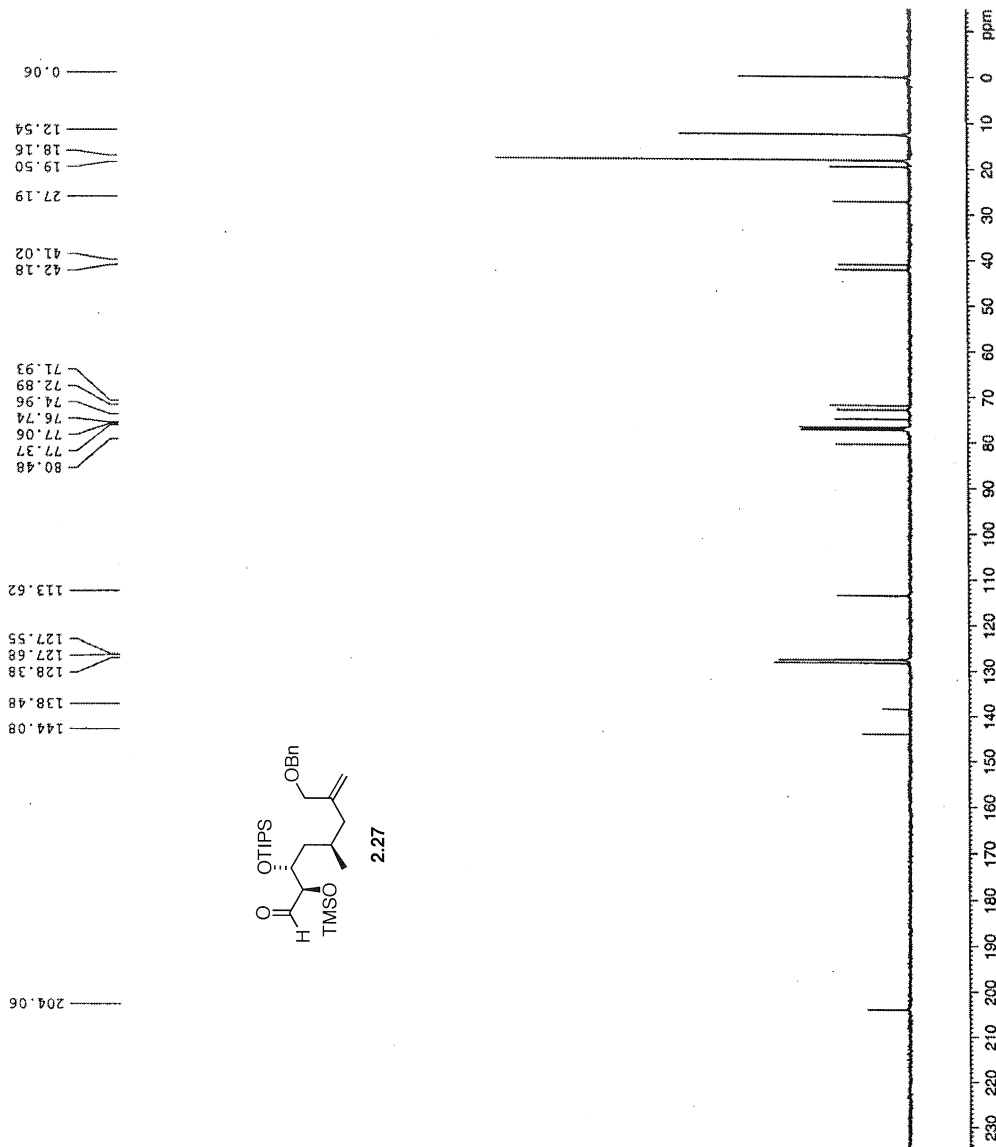
===== CHANNEL f1 =====  
 NUC1 1H  
 P1 14.70 usec  
 PL1 0.00 dB  
 SFO1 400.0146001 MHz

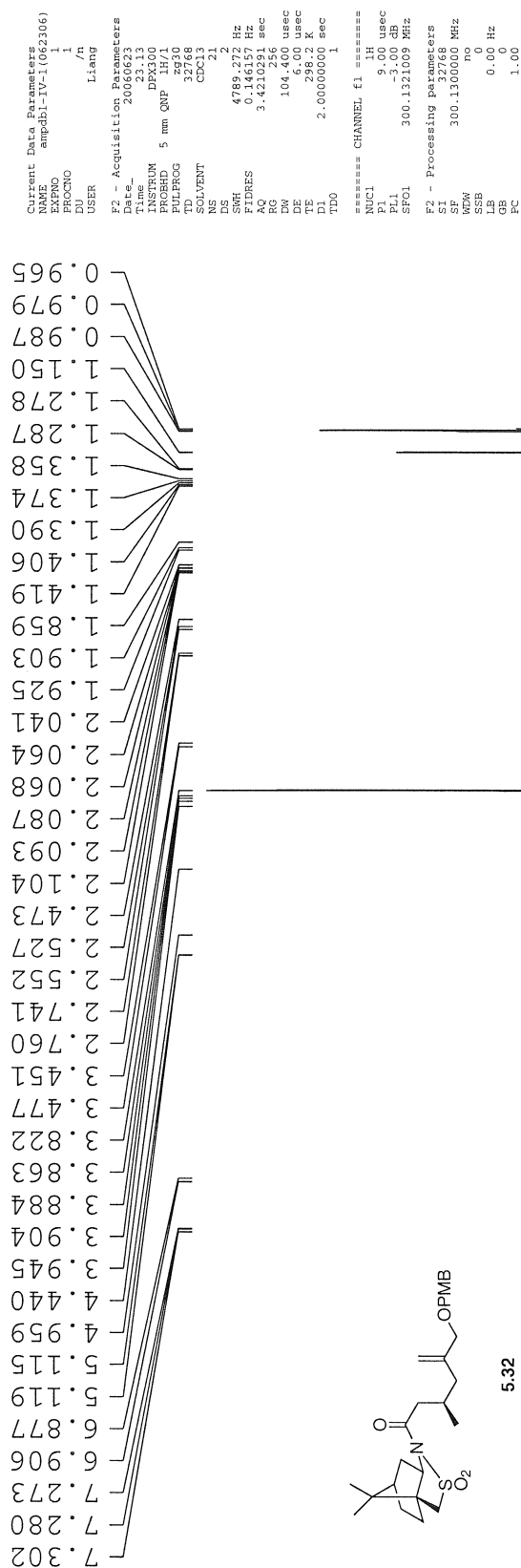
F2 - Processing Parameters  
 SI 32768  
 SF 400.0146001 MHz  
 WDW EM  
 SSF 0  
 LB 0.70 Hz  
 GB 0  
 PC 1.00

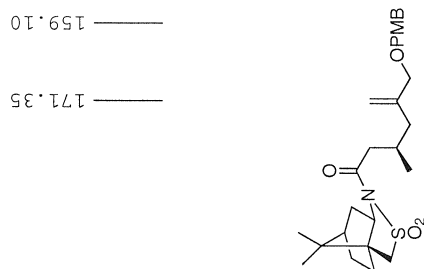


## C13/aldehyde

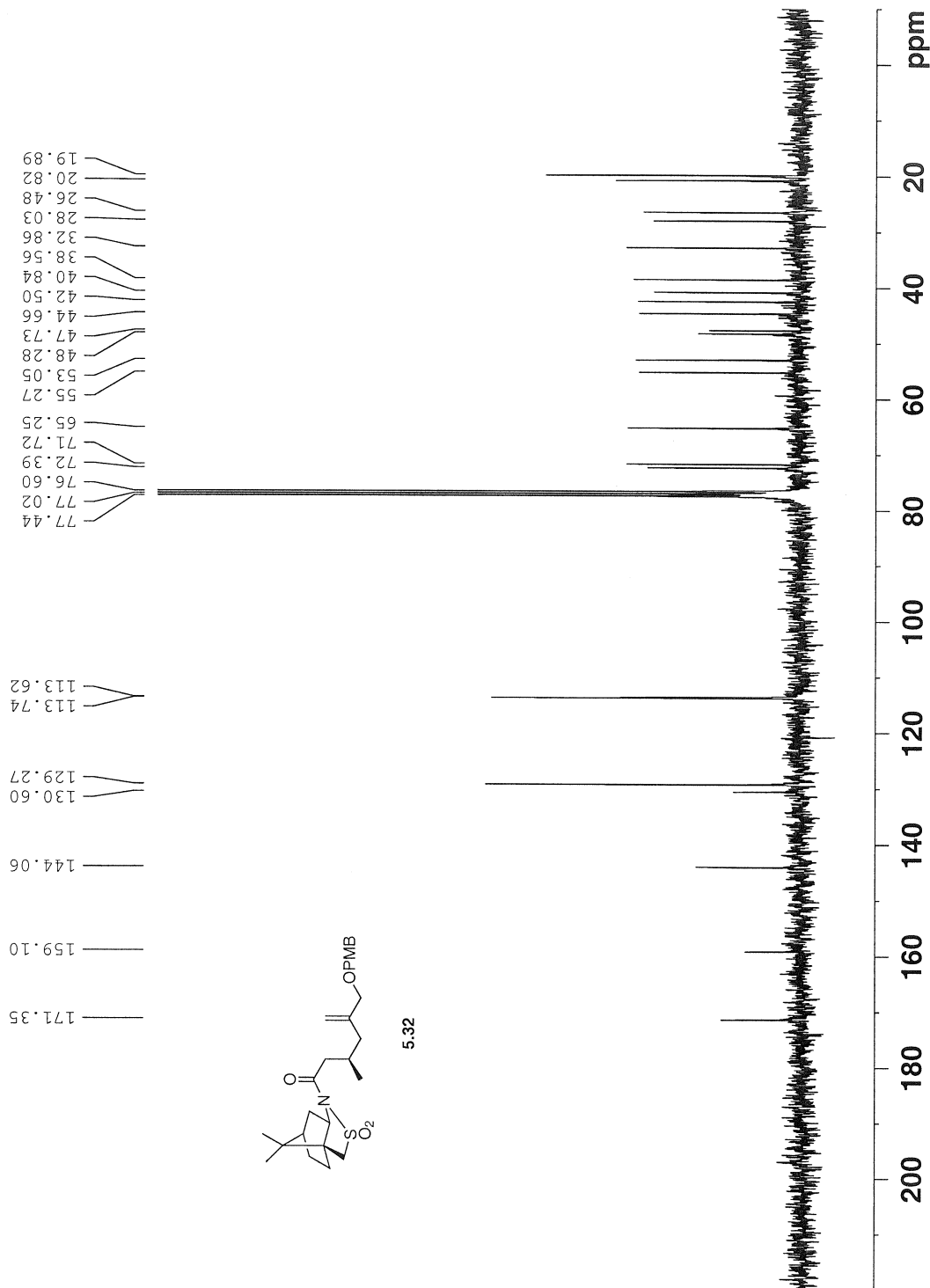
Current Data Parameters  
 NAME XZ-XI-16  
 EXPNO 2  
 PROCNO 1  
 USER xzhou  
 F2 - Acquisition Parameters  
 File 2060330  
 Time 16.46  
 INSTRUM 5 mm BBO BB-1H  
 PROBD 5 mm BBO BB-1H  
 TD 65536  
 SOLVENT 250  
 NS 250  
 SWH 25125.679 Hz  
 FIDRES 0.381387 Hz  
 AQ 1.364194 sec  
 RG 327.5  
 IN 19.900 usec  
 DE 6.00 usec  
 TE 298.2 K  
 FL 0.1500000 sec  
 d11 0.0300000 sec  
 DELTA 0.0500000 sec  
 TD0 1  
 ===== CHANNEL f1 =====  
 NUC1 13C  
 P1 7.80 usec  
 PL 0.00 dB  
 SFO1 100.593551 MHz  
 ===== CHANNEL f2 =====  
 NUC2 1H  
 P2 135.00 usec  
 PL2 17.40 dB  
 SFO2 400.011600 MHz  
 F2 - Processing Parameters  
 SI 327.5  
 SF 100.5825950 MHz  
 WDW EM  
 SSB 0  
 GB 0  
 PC 1.40



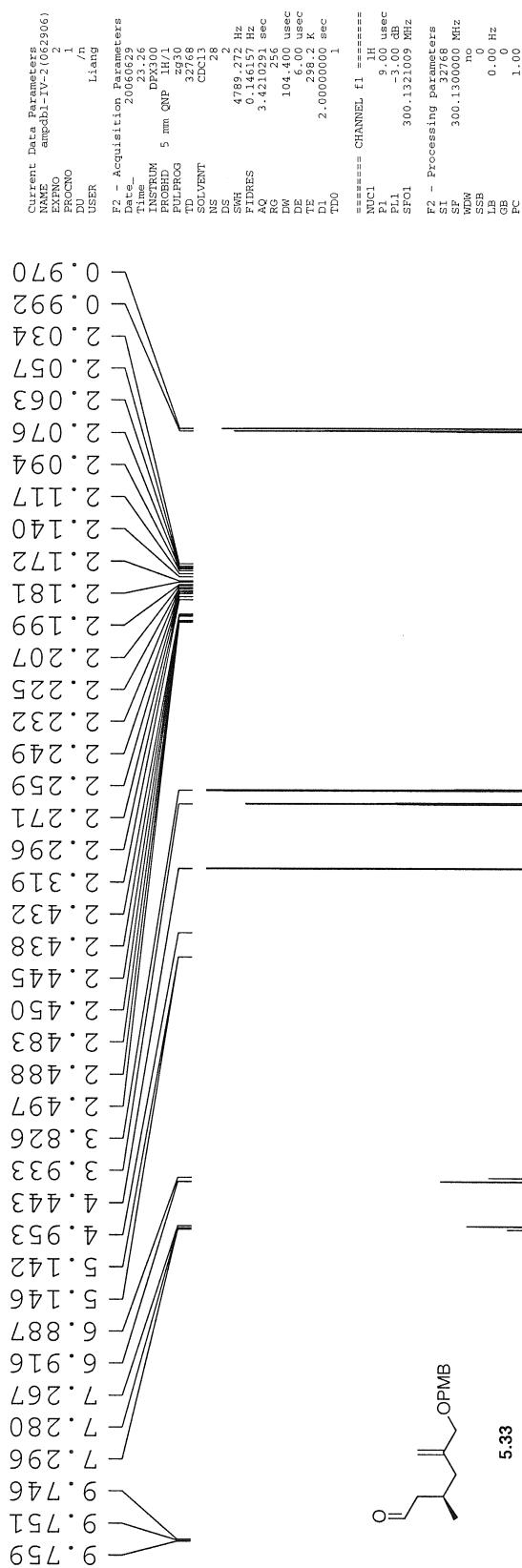


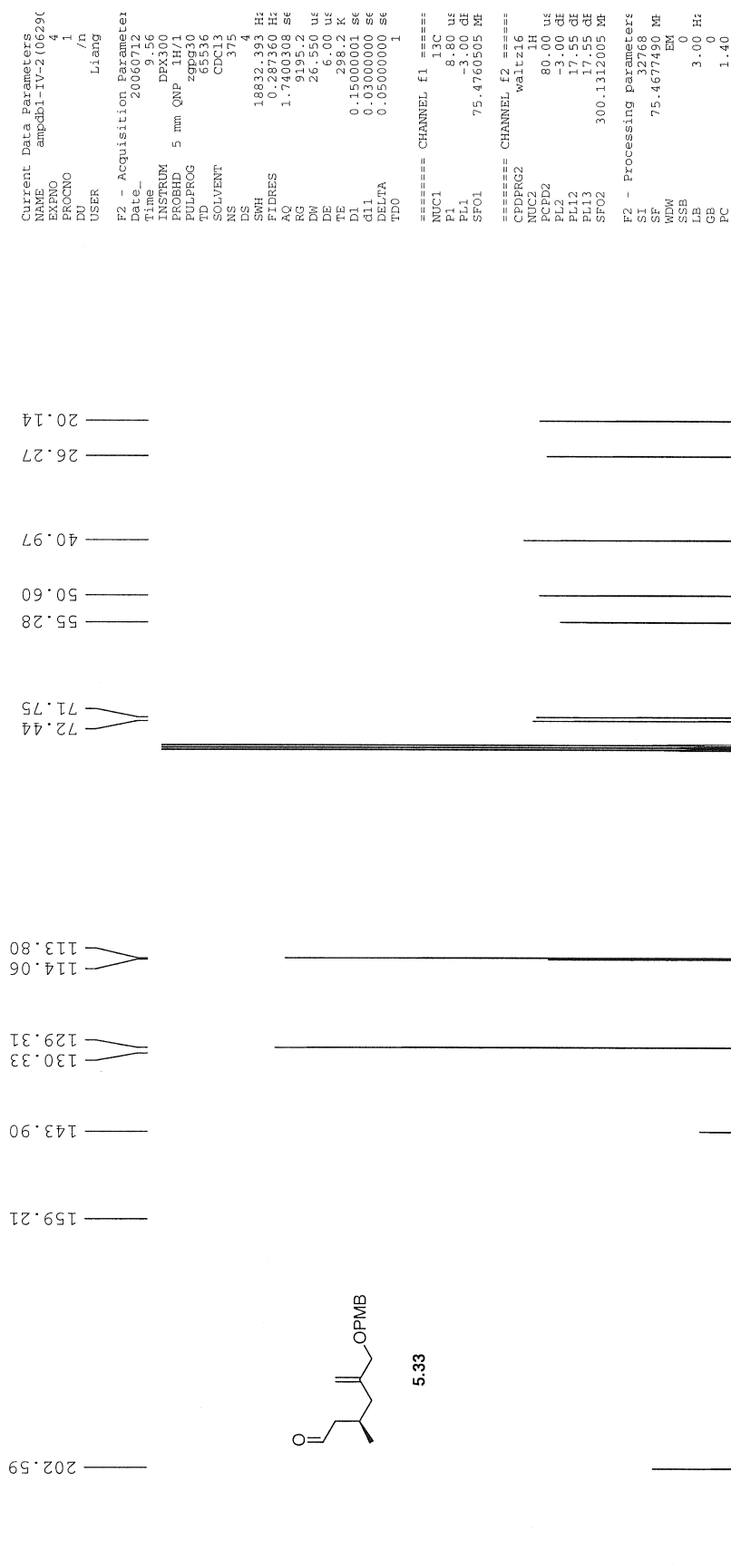


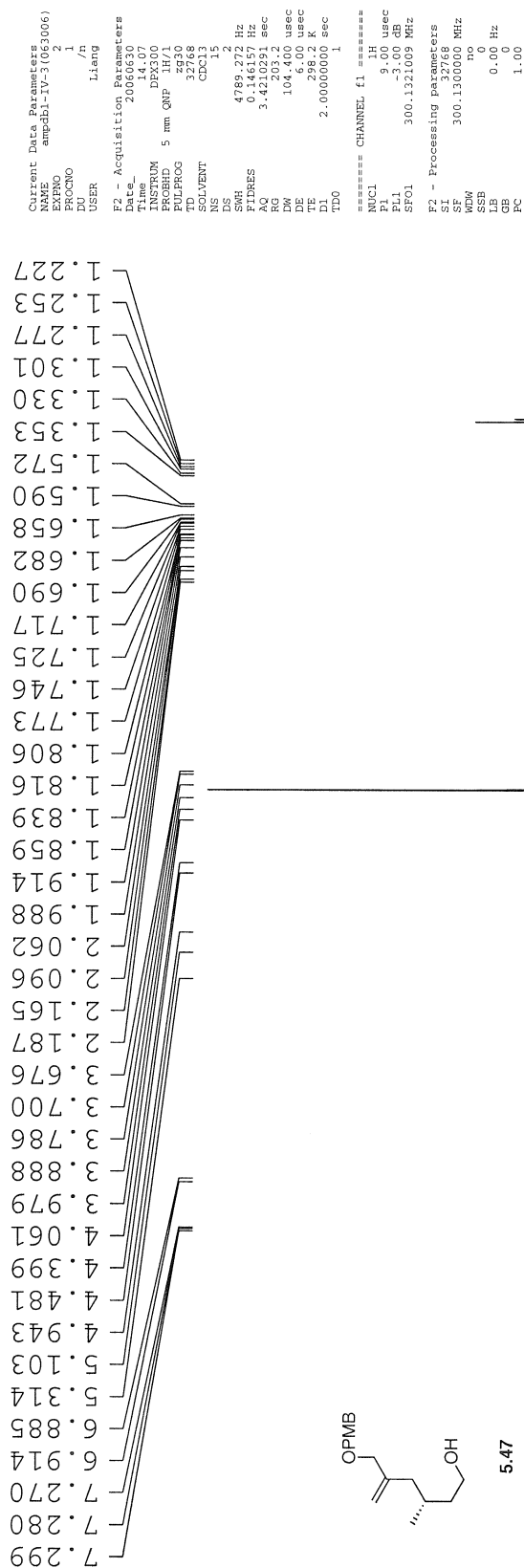
5.32



Current Data Parameters  
 NAME ampdh1-IV-110623(  
 EXPNO 2  
 PROCNO 1  
 DU /n  
 USER Liang  
 F2 - Acquisition Parameters  
 Date\_ 20060623  
 Time 23.38  
 INSTRUM DEX300  
 PROBHD 5 mm QNP 1H/1  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 694  
 DS 4  
 SWH 18832.393 Hz  
 FIDRES 0.287360 Hz  
 AQ 1.7400308 s  
 RG 9195.2  
 DW 26.550 us  
 DE 6.00 us  
 TE 298.2 K  
 D1 0.15000001 s  
 d11 0.03000000 s  
 DELTA 0.05000000 s  
 TDO 1  
 ===== CHANNEL f1 =====  
 NUC1 13C  
 P1 8.80 us  
 PL1 -3.00 dB  
 SFO1 75.4760505 MHz  
 ===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 80.00 us  
 PL2 -3.00 dB  
 PL12 17.55 dB  
 PL13 17.55 dB  
 SFO2 300.1312005 MHz  
 F2 - Processing parameters  
 SI 32768  
 SF 75.4677490 MHz  
 WDW EM  
 SSB 0  
 LB 3.00 Hz  
 GB 0  
 FC 1.40







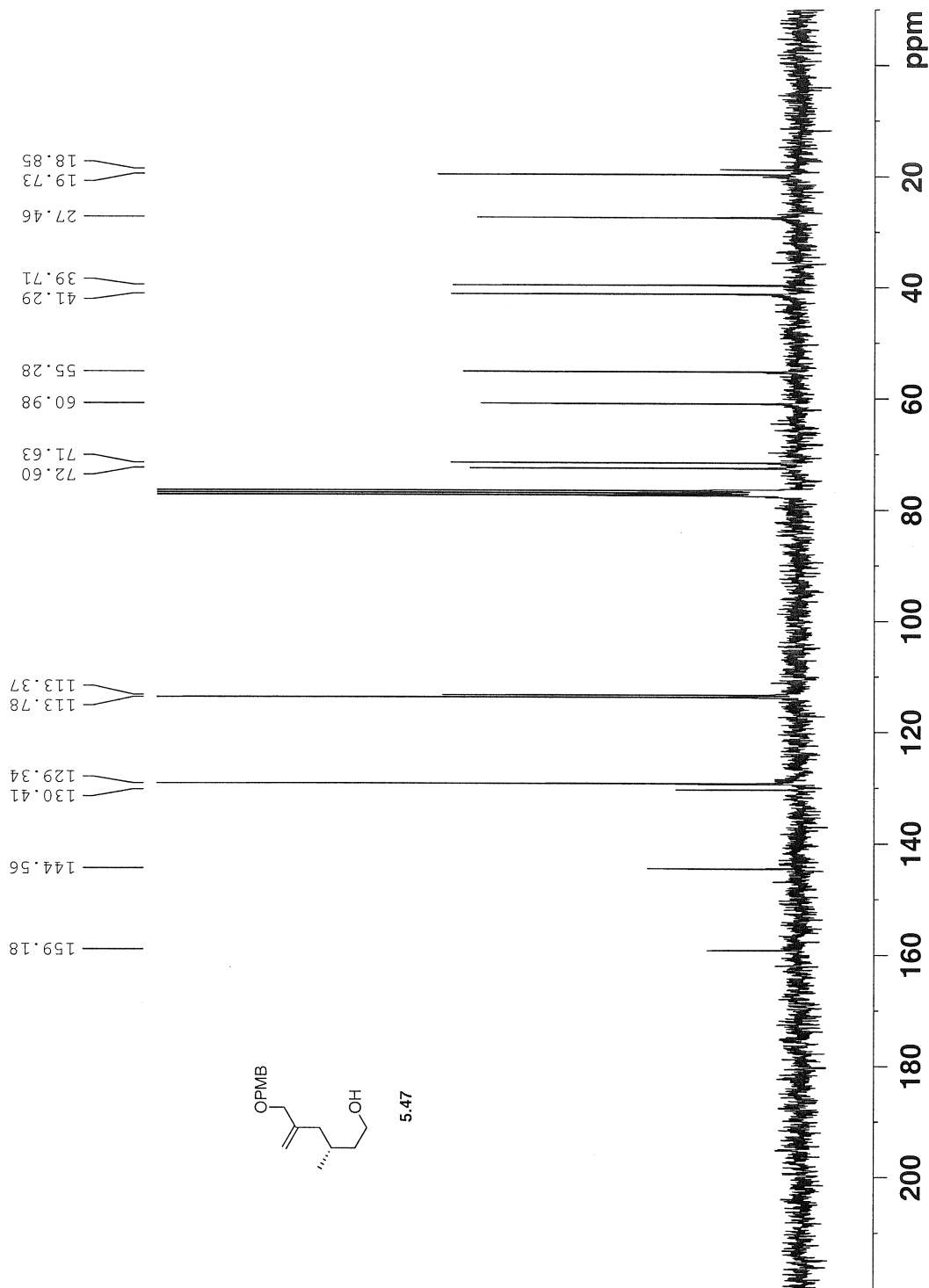
Current Data Parameters  
NAME ampdb1-IV-310630(  
EXPNO 1  
PROCNO 1  
DU /n  
USER Liang

F2 - Acquisition Parameters  
Date\_ 20060630  
Time 23.00  
INSTRUM DFX300  
PROBHD 5 mm QNP 1H/1  
PULPROG zgpg30  
TD 65536  
SOLVENT CDCl3  
NS 604  
DS 4  
SWH 18832.393 Hz  
FIDRES 0.287360 Hz  
AQ 1.7400308 sec  
RG 11585.2  
DW 26.550 usec  
DE 6.00 usec  
TE 298.2 K  
D1 0.15000001 sec  
d11 0.03000000 sec  
DELTA 0.05000000 sec  
TD0 1

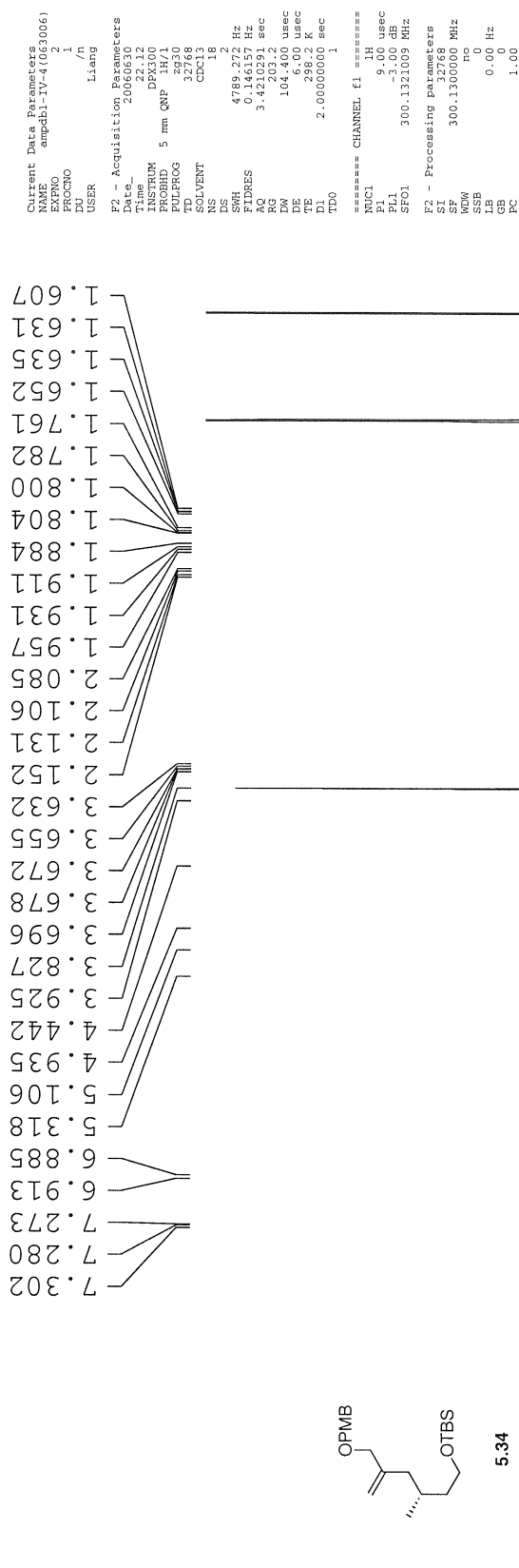
===== CHANNEL f1 =====  
NUC1 13C  
P1 8.80 usec  
PL1 -2.50 dB  
SFO1 75.4760505 MHz

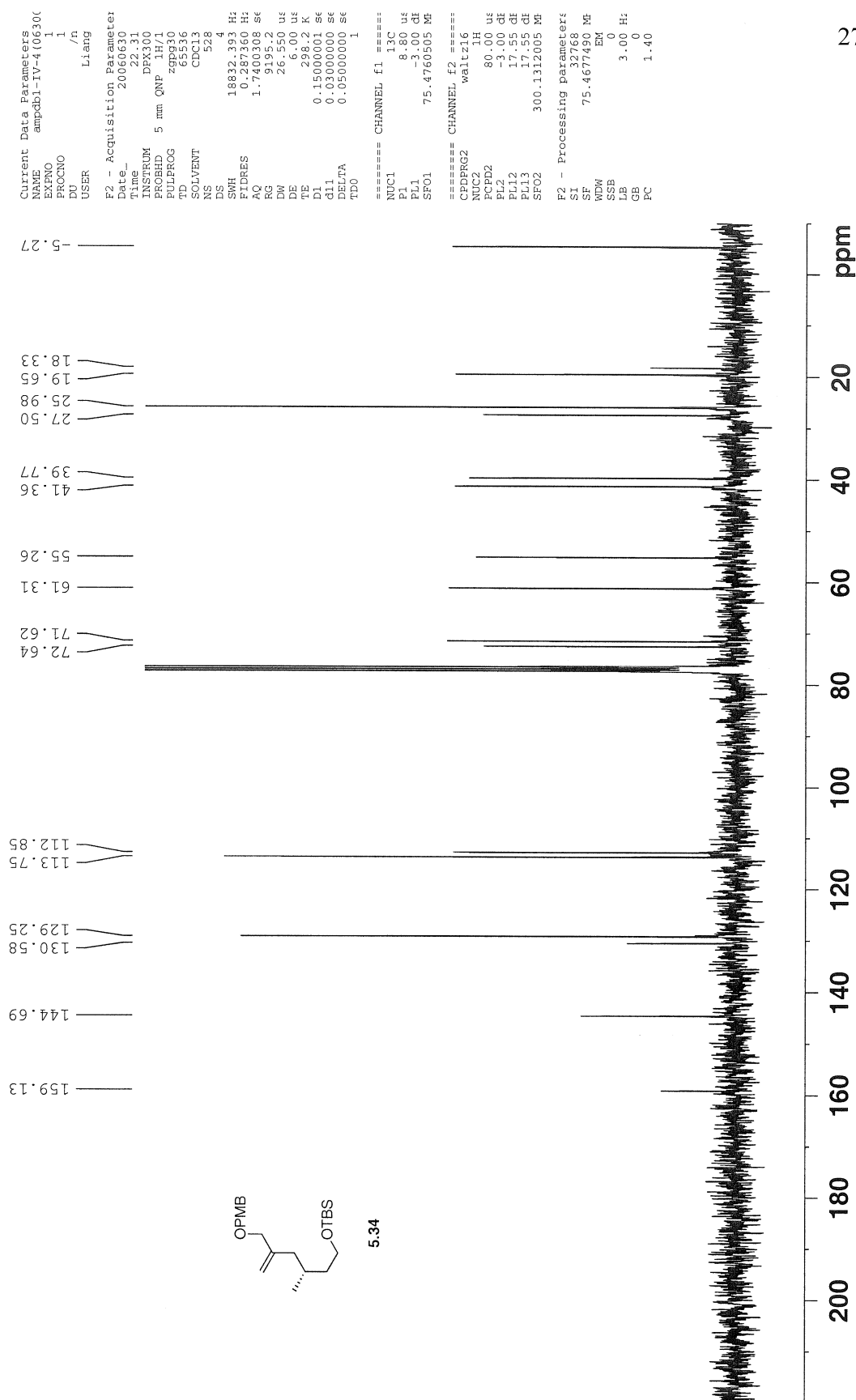
===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 80.00 usec  
PL2 -3.00 dB  
PL12 17.55 dB  
PL13 17.55 dB  
SFO2 300.1312005 MHz

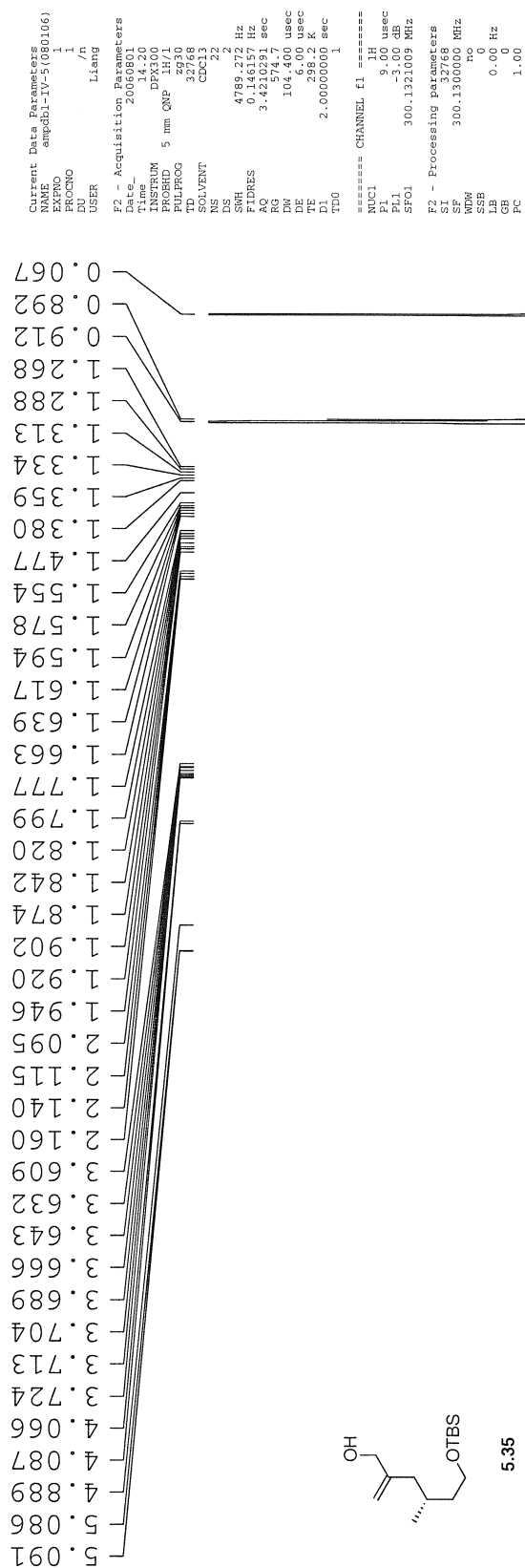
F2 - Processing parameters:  
SI 32768  
SF 75.4677490 MHz  
WDW EM  
SSB 0  
LB 0  
GB 0  
PC 1.40

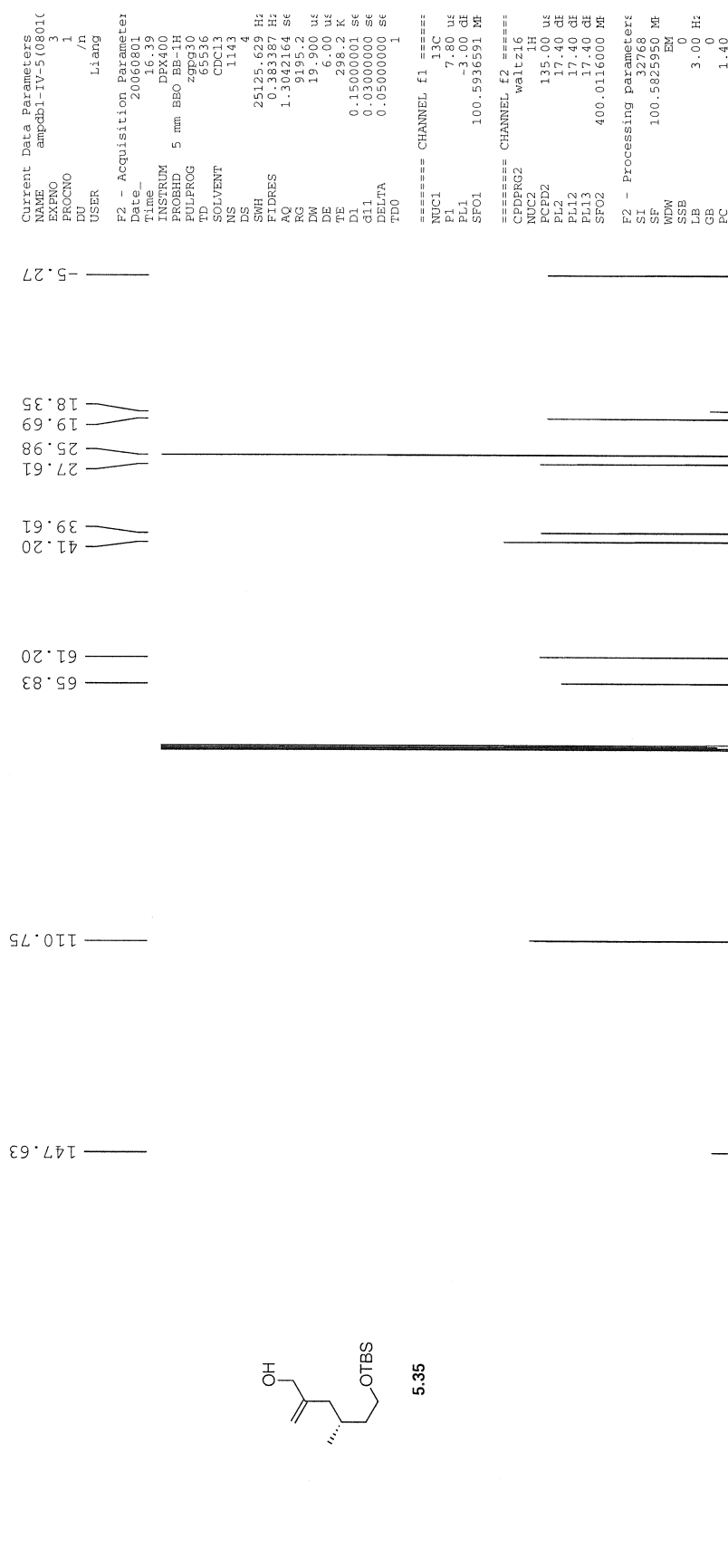


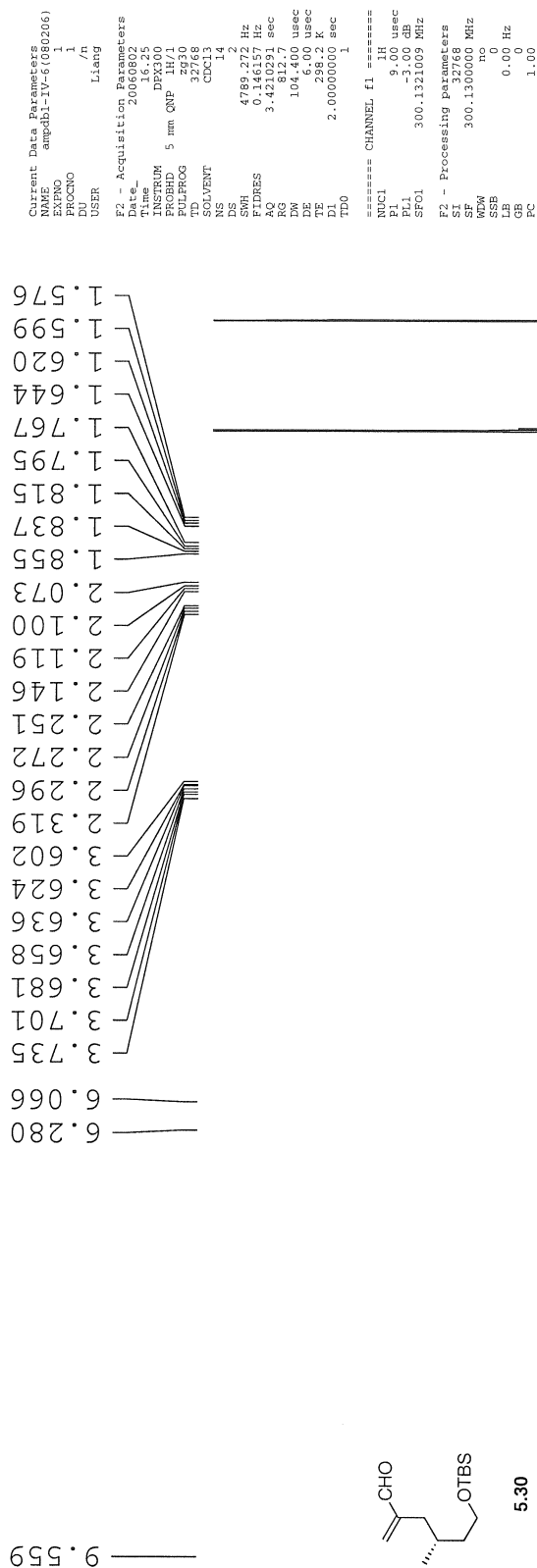


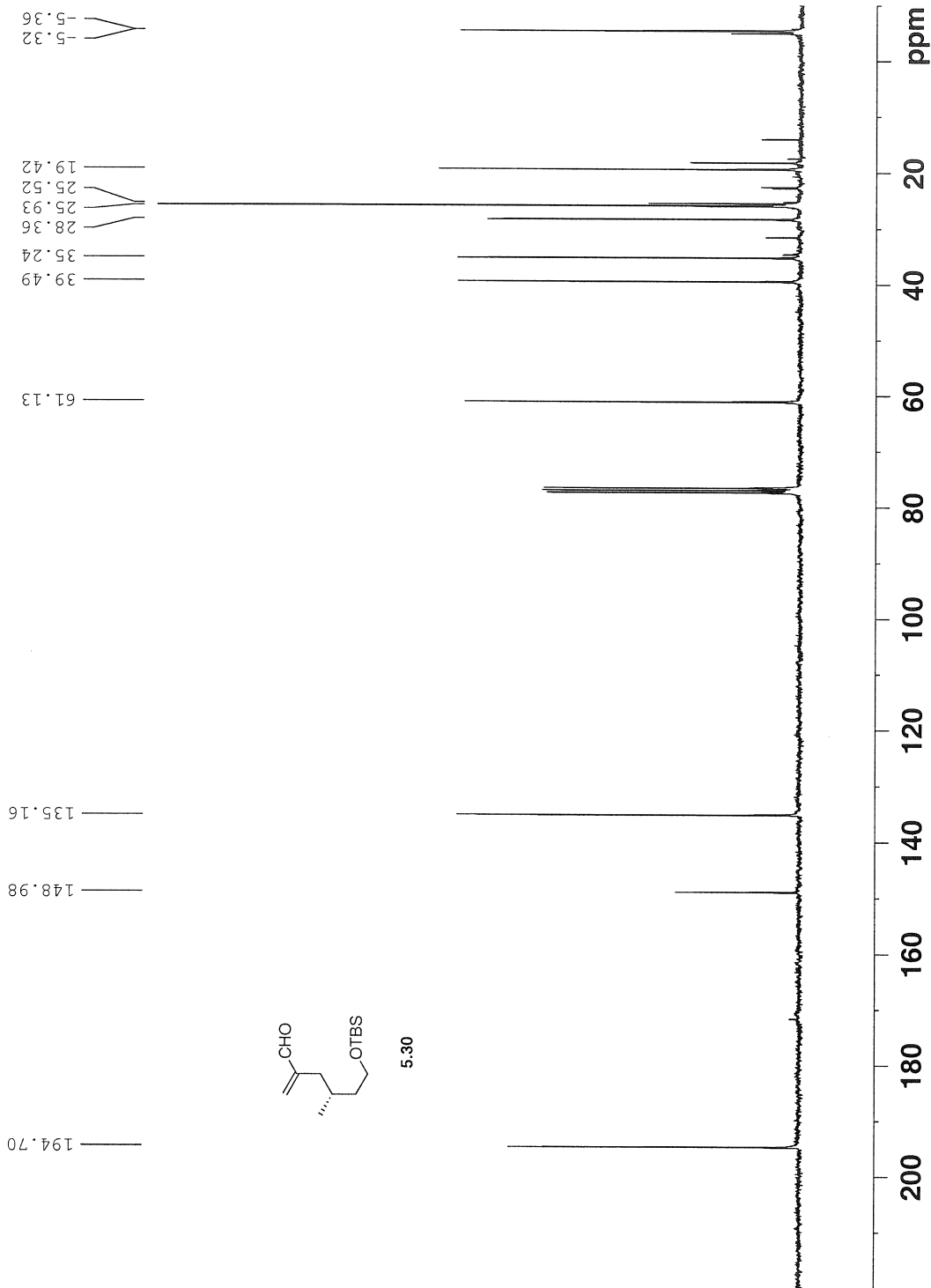




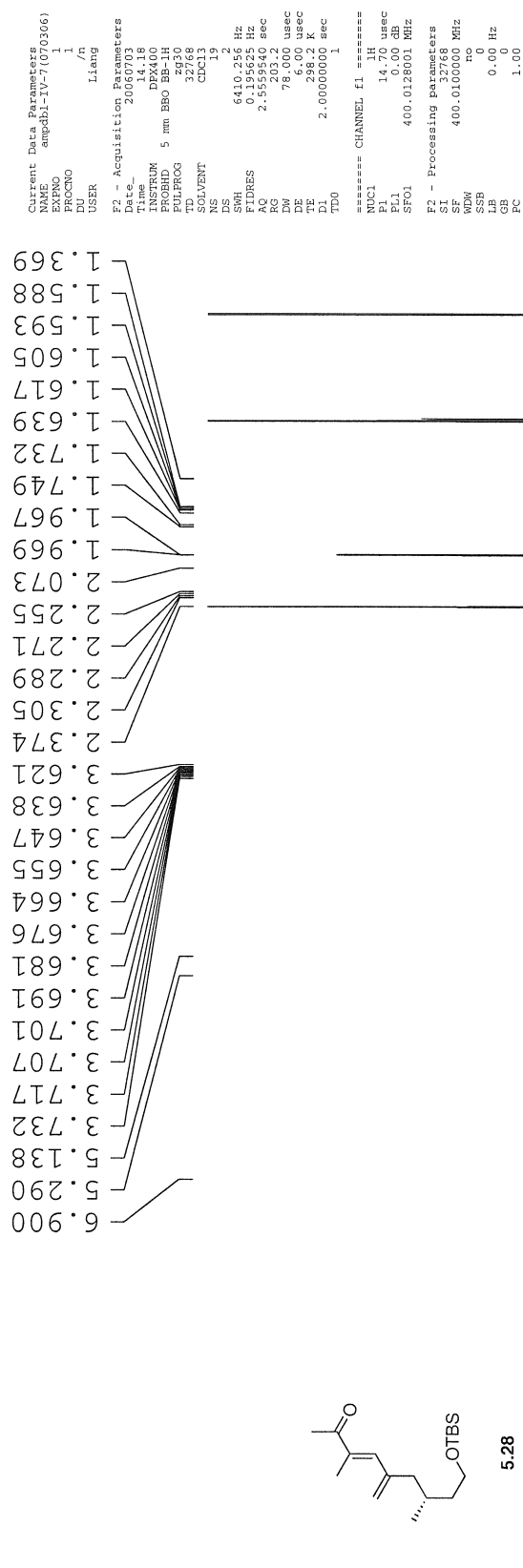








Current Data Parameters  
 NAME: amdb1-1V-DMP-topaprot(07)  
 PROCNO: 1  
 DO: /n  
 USER: Liang  
 F2 - Acquisition Parameters  
 Date\_: 20060702  
 Time: 14:16  
 INSTRUM: DFX100  
 PROBRD: 5 mm QNP 1H/1  
 PULPROG: zgpg30  
 TD: 65536  
 SOLVENT: CDCl3  
 NS: 1850  
 DS: 4  
 SWH: 18832.393 Hz  
 FIDRES: 0.287360 Hz  
 AQ: 1.7400208 sec  
 RG: 261.250  
 EN: 26.550 usec  
 DE: 6.00 usec  
 TE: 300.2 K  
 D1: 0.15000000 sec  
 d11: 0.03000000 sec  
 DELTA: 0.05000000 sec  
 TDO: 1  
 ===== CHANNEL f1 =====  
 NUC1: 13C  
 P1: 8.00 usec  
 PL1: -3.00 dB  
 SF01: 75.4760505 MHz  
 ===== CHANNEL f2 =====  
 CPDPRG2: waltz16  
 NUC2: 1H  
 P2: 80.00 usec  
 PL2: -3.00 dB  
 PL12: 17.55 dB  
 PL13: 17.55 dB  
 SS04: 300.1312605 MHz  
 F2 - Processing parameters  
 SI: 32768  
 SF: 75.4677491 MHz  
 WDW: EM  
 SSB: 0  
 LB: 3.00 Hz  
 GB: 0  
 PC: 1.40



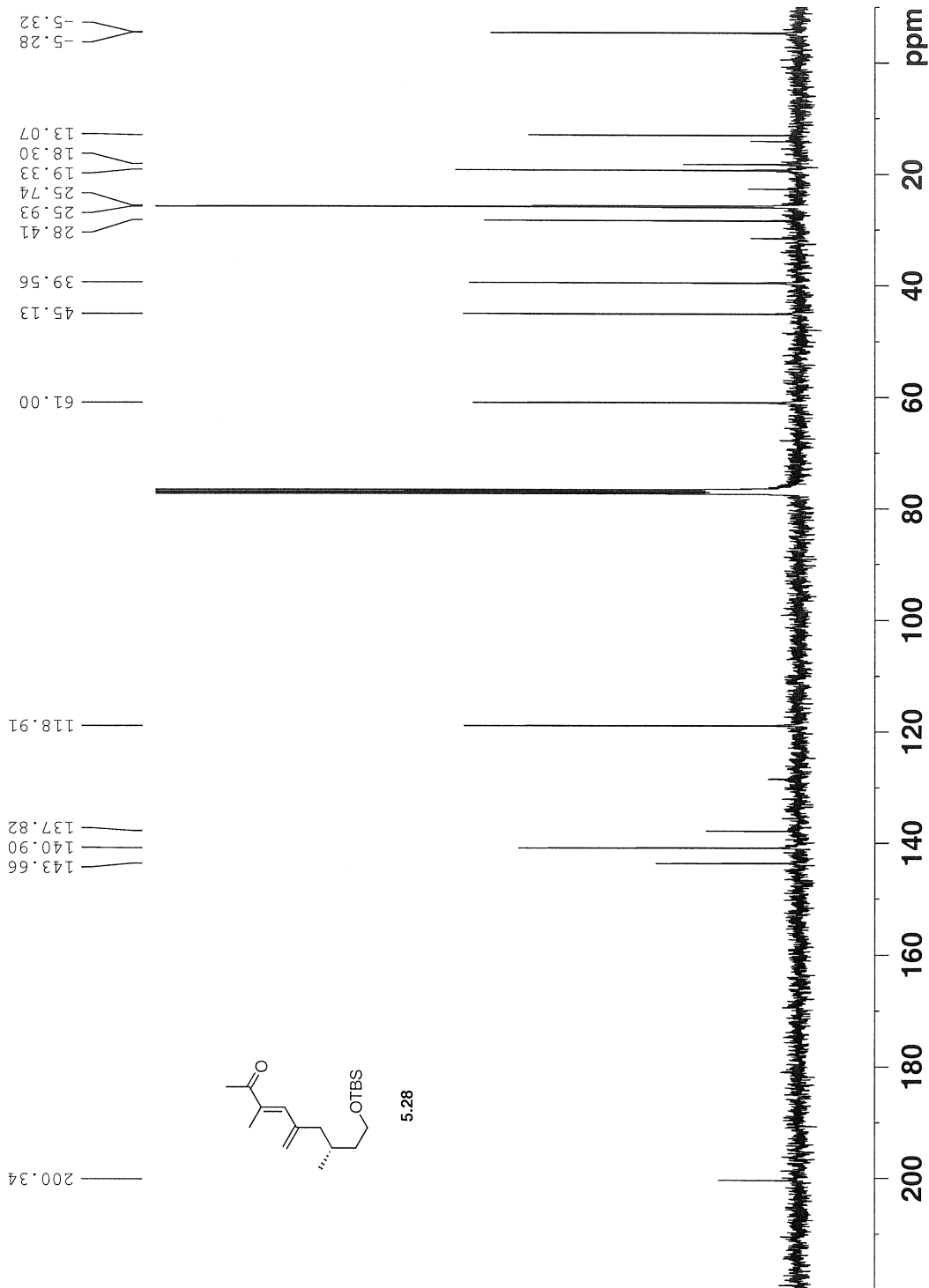
Current Data Parameters  
NAME ampd1-IV-7(0703)  
EXPNO 2  
PROCNO 1  
DU /n  
USER Liang

F2 - Acquisition Parameters  
Date\_ 20060703  
Time 17.50  
INSTRUM DFX400  
PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
TD 65536  
SOLVENT CDCl3  
NS 2004  
DS 4  
SWH 25125.629 Hz  
FIDRES 0.383387 Hz  
AQ 1.3042164 s  
RG 4597.6  
DW 19.900 us  
DE 6.00 us  
TE 298.2 K  
D1 0.15000001 s  
d11 0.03000000 s  
DELTA 0.05000000 s  
TD0 1

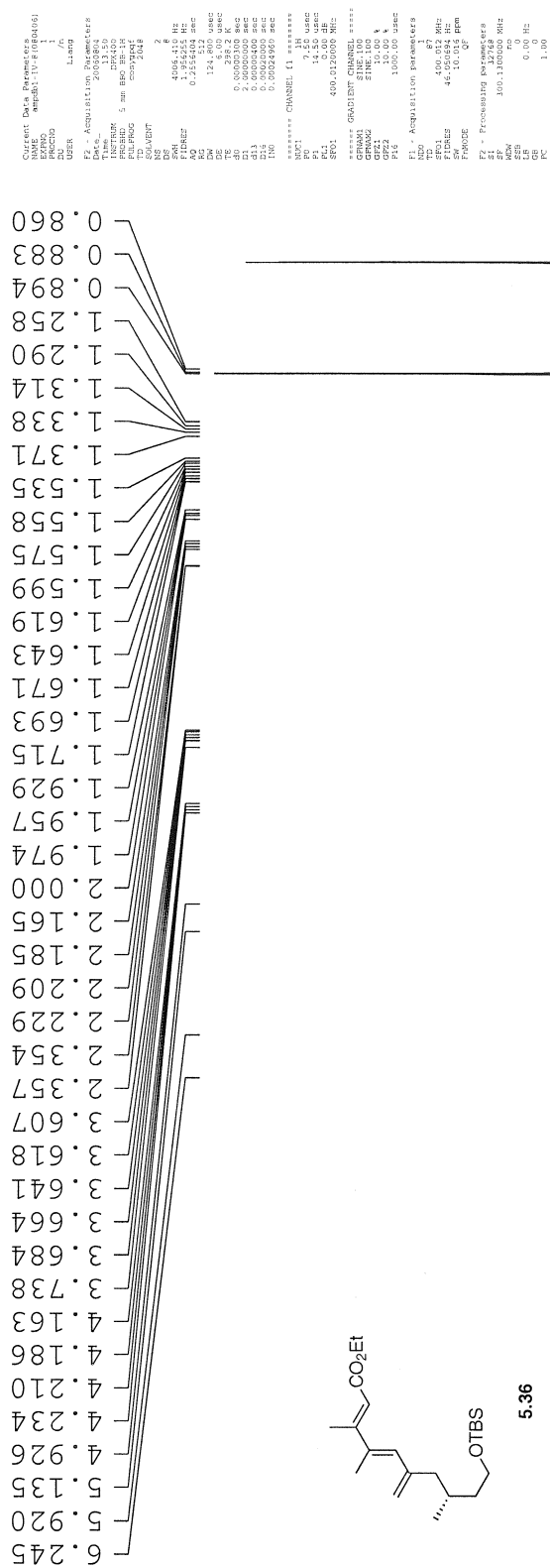
===== CHANNEL f1 =====  
NUC1 13C  
P1 7.80 us  
PL1 -3.80 dB  
SFO1 100.5936591 MHz

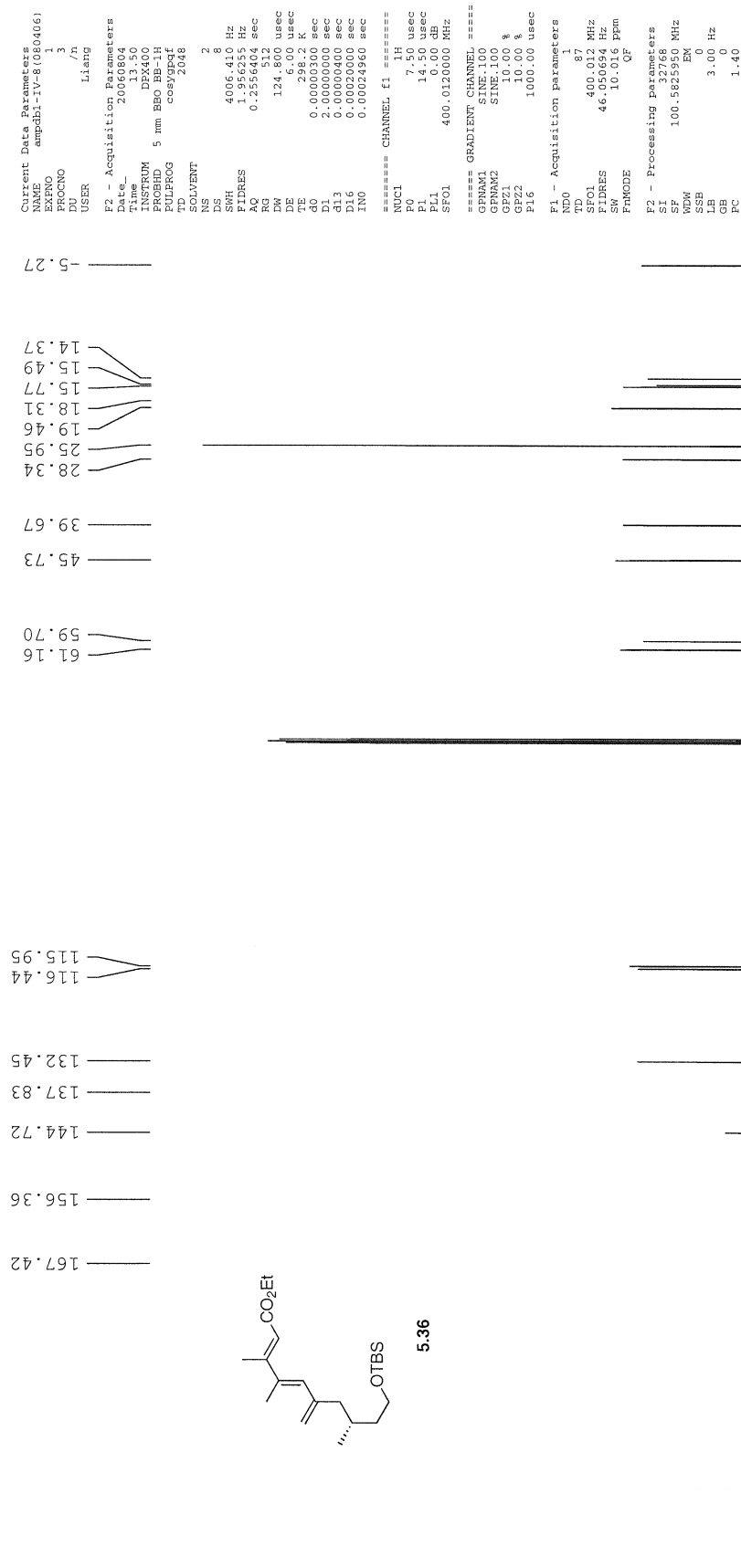
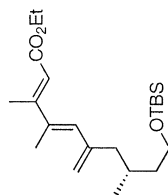
===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 135.00 us  
PL2 17.40 dB  
PL12 17.40 dB  
PL13 17.40 dB  
SFO2 400.0116000 MHz

F2 - Processing parameters  
SI 32768  
SF 100.5825950 MHz  
WDW EM  
SSB 0  
LB 3.00 Hz  
GB 0  
PC 1.40

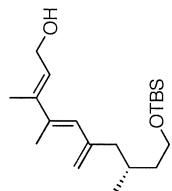




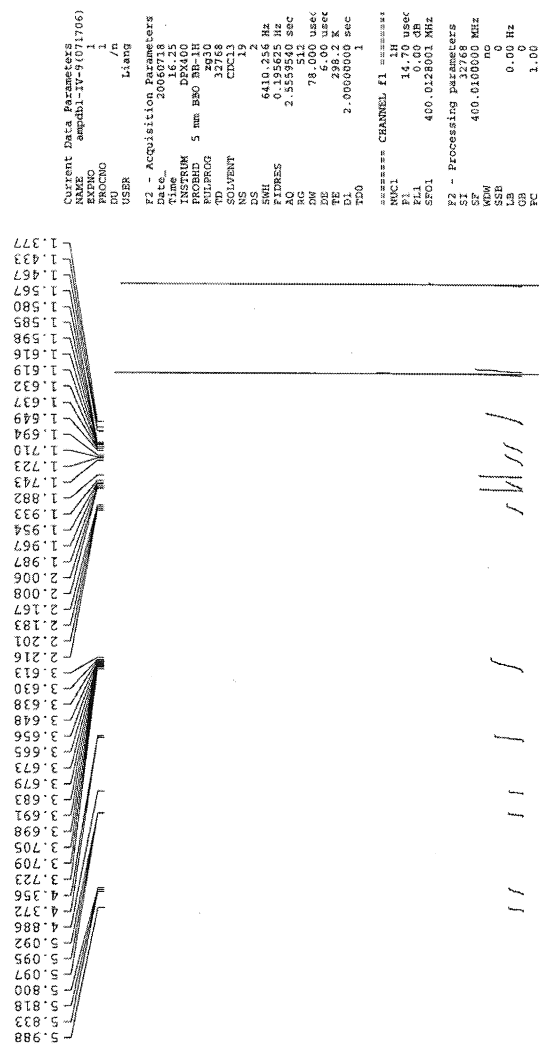


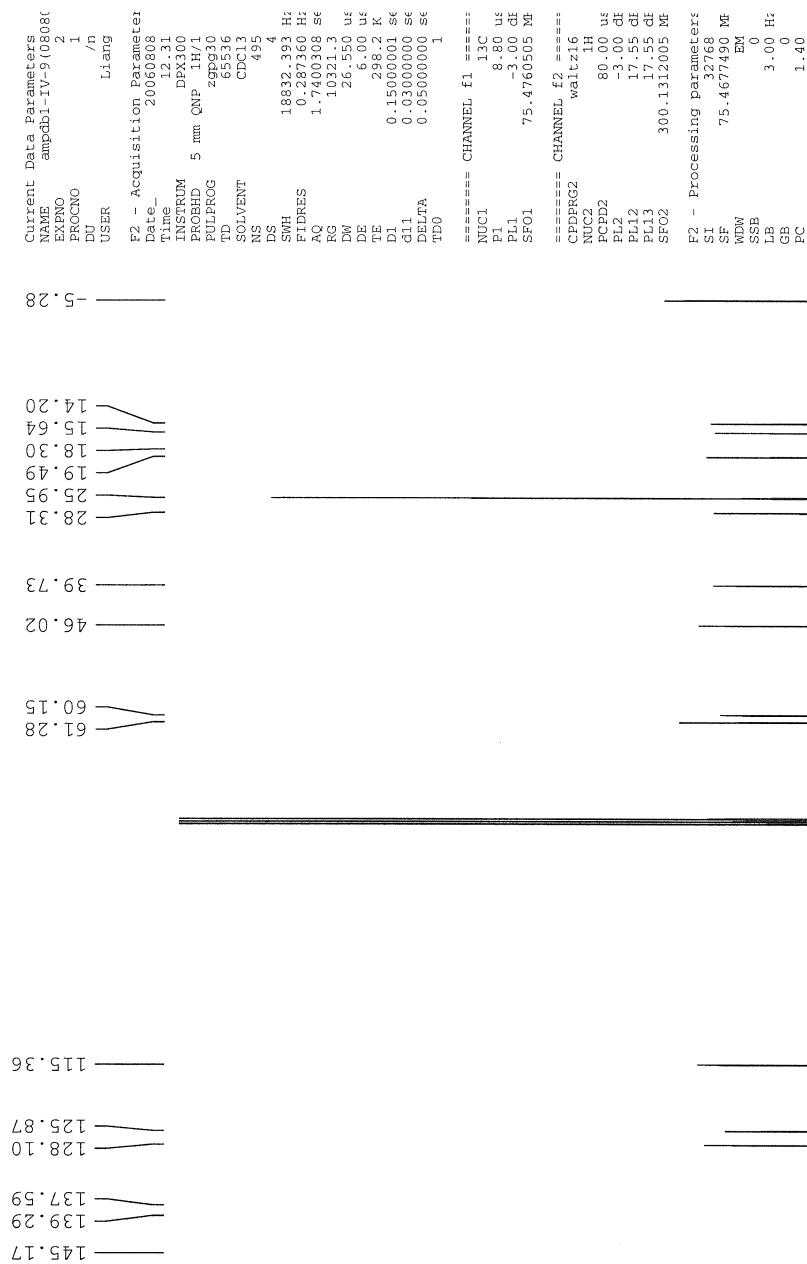


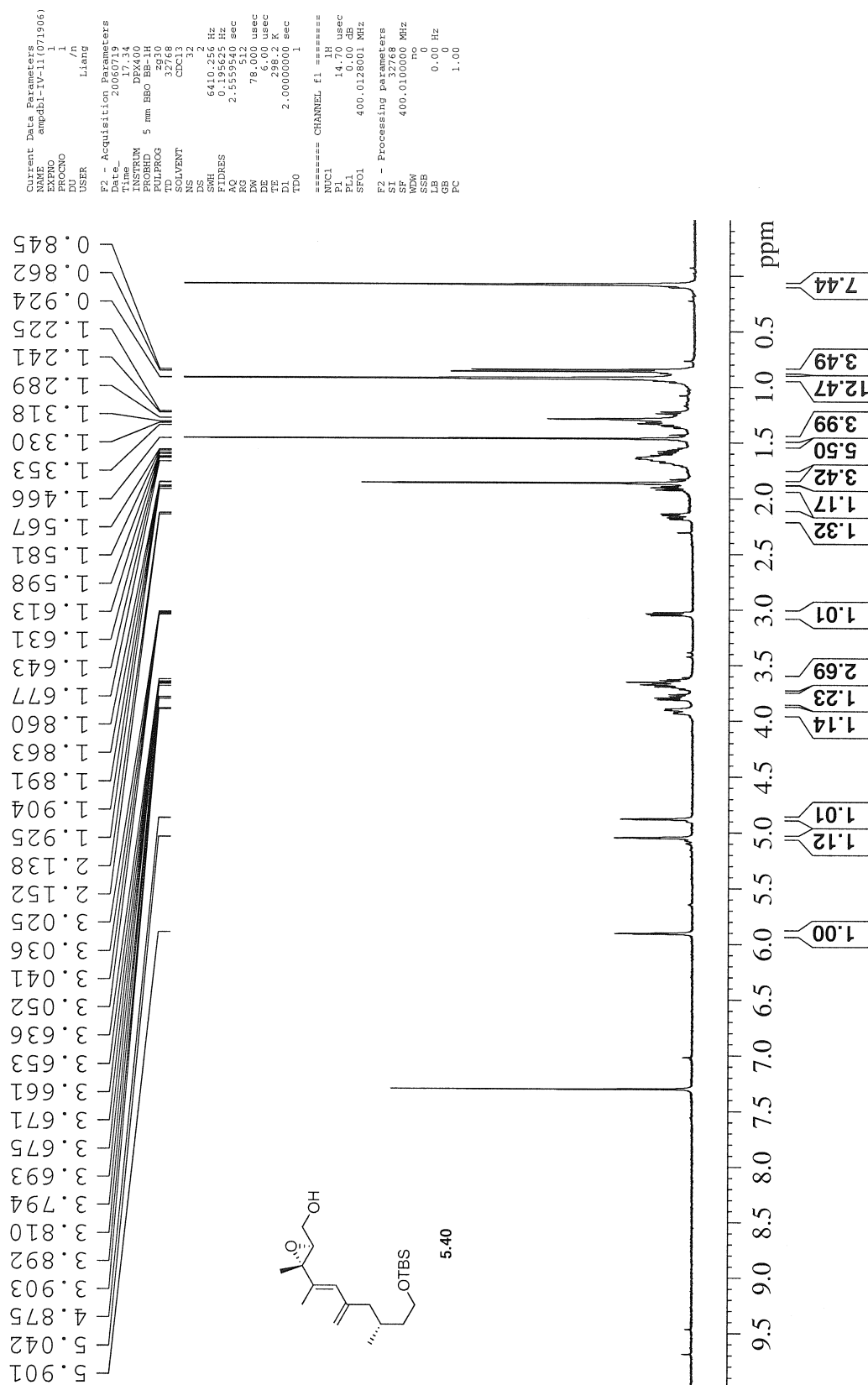
alcohol



5.27







```

Current Data Parameters
NAME      ampbdl-iv-11-2 (071
EXPNO     2
PROCNO    1
DU         /n
USER      Liang

F2 - Acquisition Parameters
Date_     20060719
Time      17.50
INSTRUM   DEX300
PROBHD    5 mm QNP 1H/1
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         1074
DS         4
SWH        18832.393 Hz
FIDRES     0.287360 Hz
AQ         1.7400308 sec
RG         9195.2
DW         26.550 use
DE         6.00 use
TE         298.2 K
T1         0.1500001 sec
T1RHO     0.3300000 sec
DELTA     0.05000000 sec
TD0        1

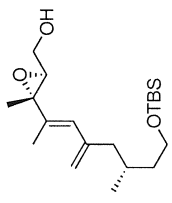
===== CHANNEL f1 =====
NUC1       13C
P1         8.80 use
PL1        -3.00 dB
SFO1       75.4760505 MHz

===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
PCPD2      80.00 use
PL2        -3.00 dB
PL12       17.55 dB
PL13       17.55 dB
SFO2       300.1312005 MHz

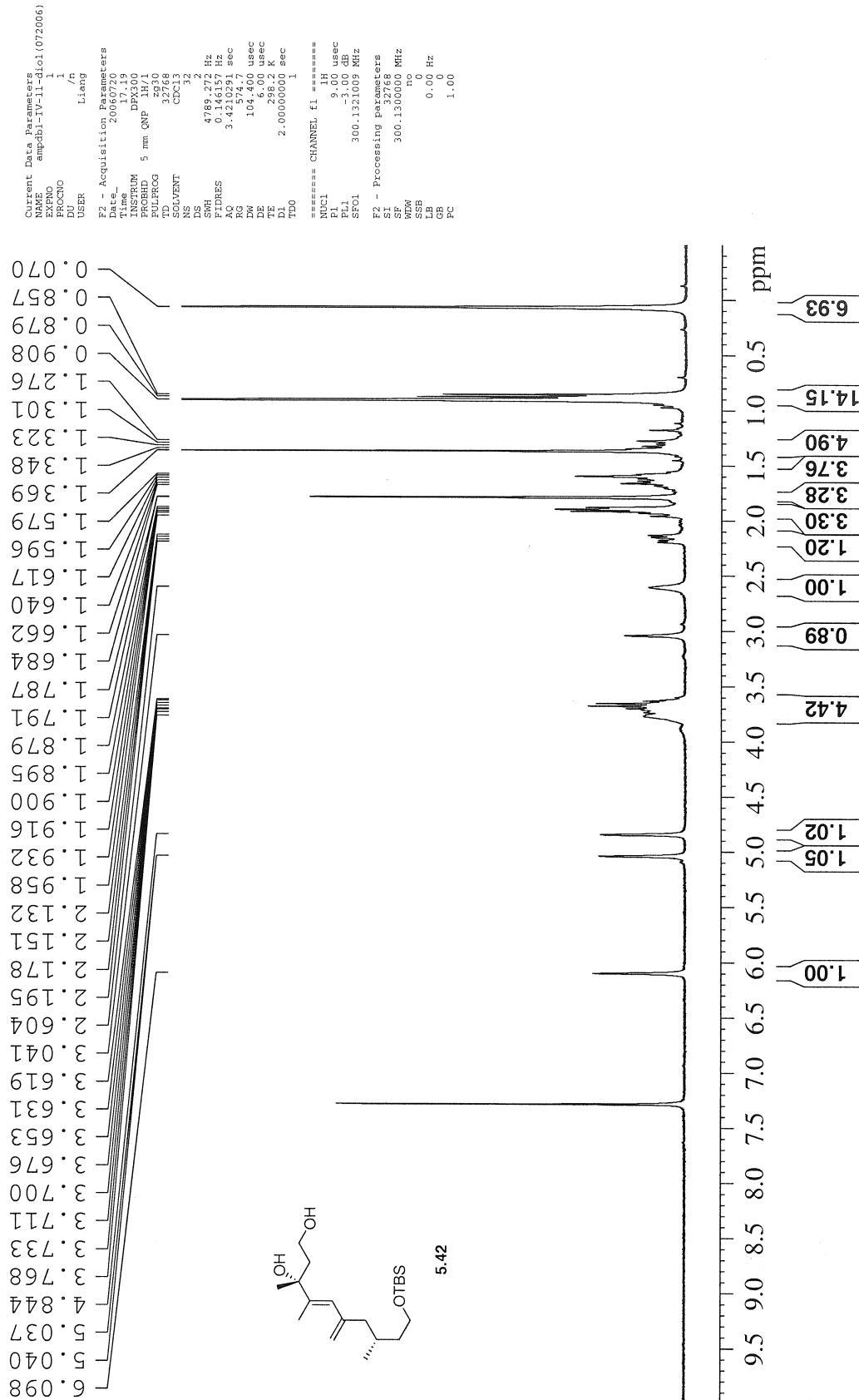
F2 - Processing parameters
SI         32768
SF         75.4677490 MHz
WDW        EM
SSB        0
LB         3.00 Hz
GB         0
PC         1.40

```

144.00  
 137.53  
 126.17  
 115.19  
 63.67  
 61.29  
 45.59  
 39.77  
 28.23  
 25.98  
 19.34  
 18.32  
 16.75  
 14.78  
 -5.27



5.40



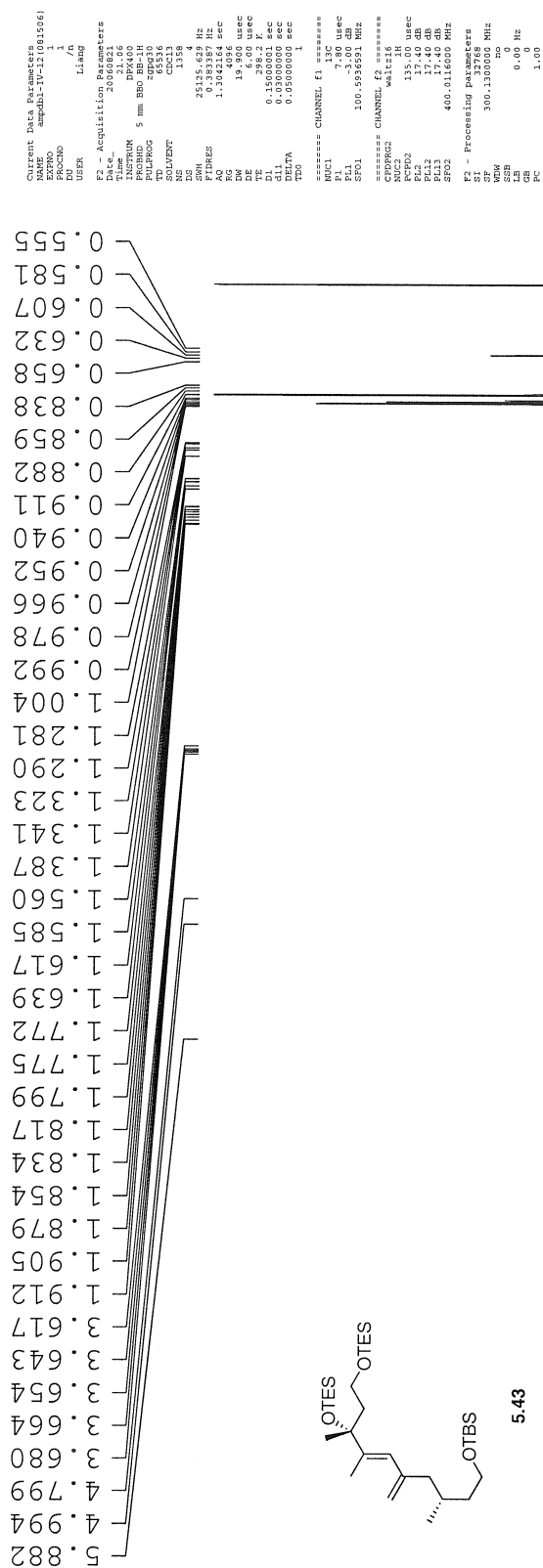


```

Current Data Parameters
=====
NAME      ampbelt-IV-11-doi(072
EXPNO     4
PROCNO    1
PROBHD     5 mm BBO BH-1H
PULPROG    zgpg30
TD         65536
F2 - Acquisition Parameters
=====
Date_      20060720
Time       17.40
INSTRUM    DP4400
PROBHD     5 mm BBO BH-1H
PULPROG    zgpg30
TD         65536
SOLVENT    NS
NS          1943
DS          4
SWH         25125.629 Hz
FIDRES      0.38387 Hz
AQ          1.339764 sec
RG          157.900 usec
WDW          EM
SSB          0
GB          0
DE         288.2 K
TE          0.1500000 sec
D11         0.03000001 sec
DELTA       0.05000000 sec
TD0         1
===== CHANNEL f1 =====
NUC1        13C
P1          7.80 usec
PL1         -3.00 dB
SFO1        100.5936591 MHz
===== CHANNEL f2 =====
WALTZ16     walz16
NUC2        1H
P2          135.00 usec
PL2         1.40 dB
SFO2        400.0116000 MHz
===== F2 - Processing Parameters =====
SI          32768
SF          100.5825950 MHz
WDW          EM
SSB          0
GB          0
PC          1.40
=====

```





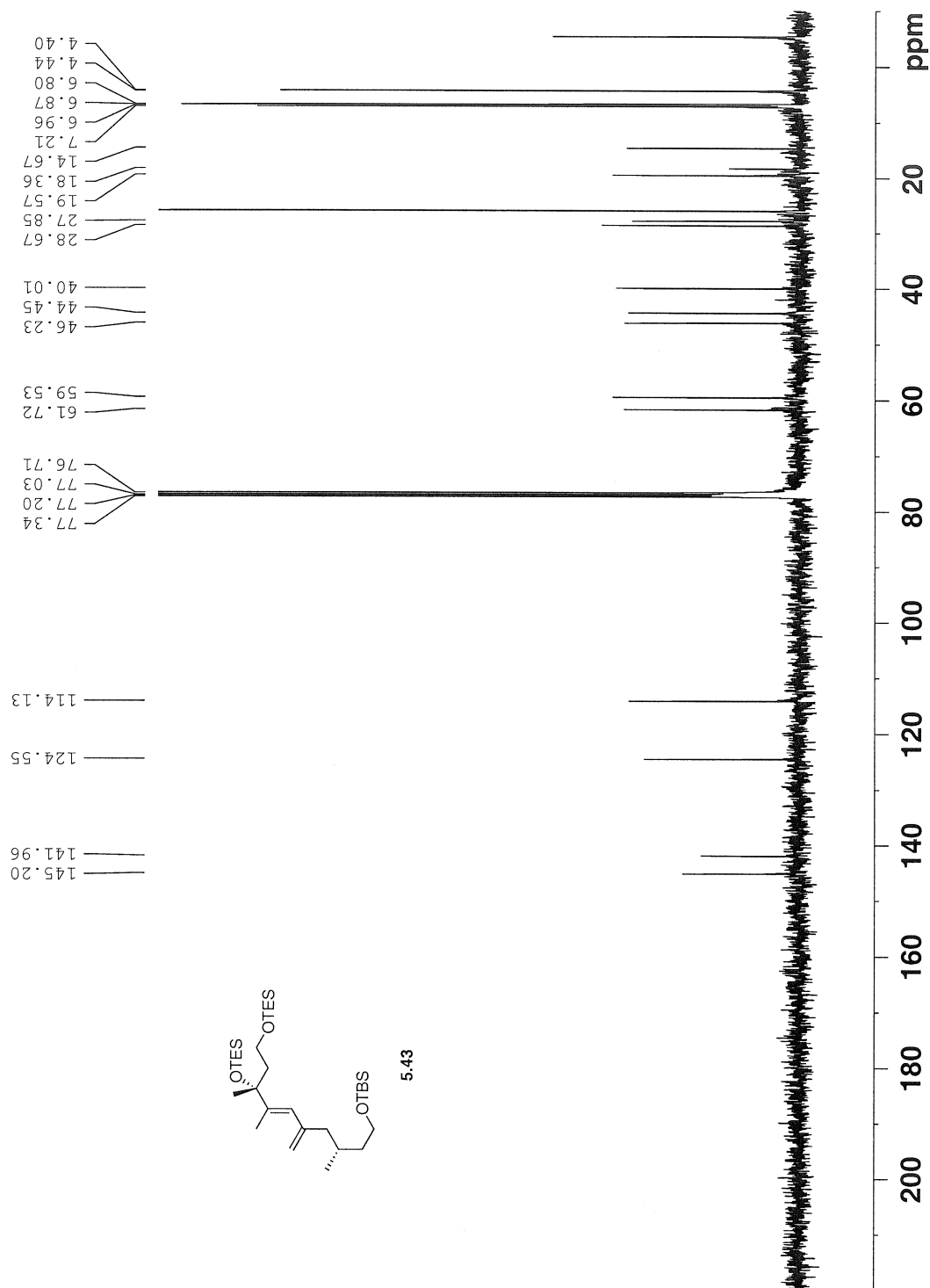
Current Data Parameters  
 NAME ampdb1-1V-121081  
 EXPNO 1  
 PROCNO 3  
 DU /n  
 USER Liang

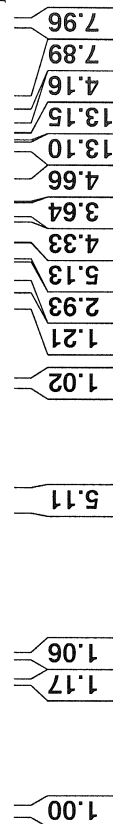
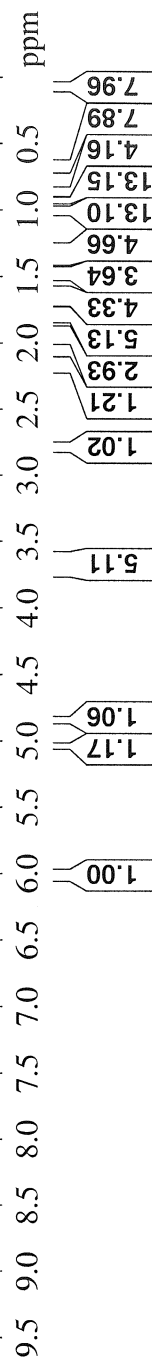
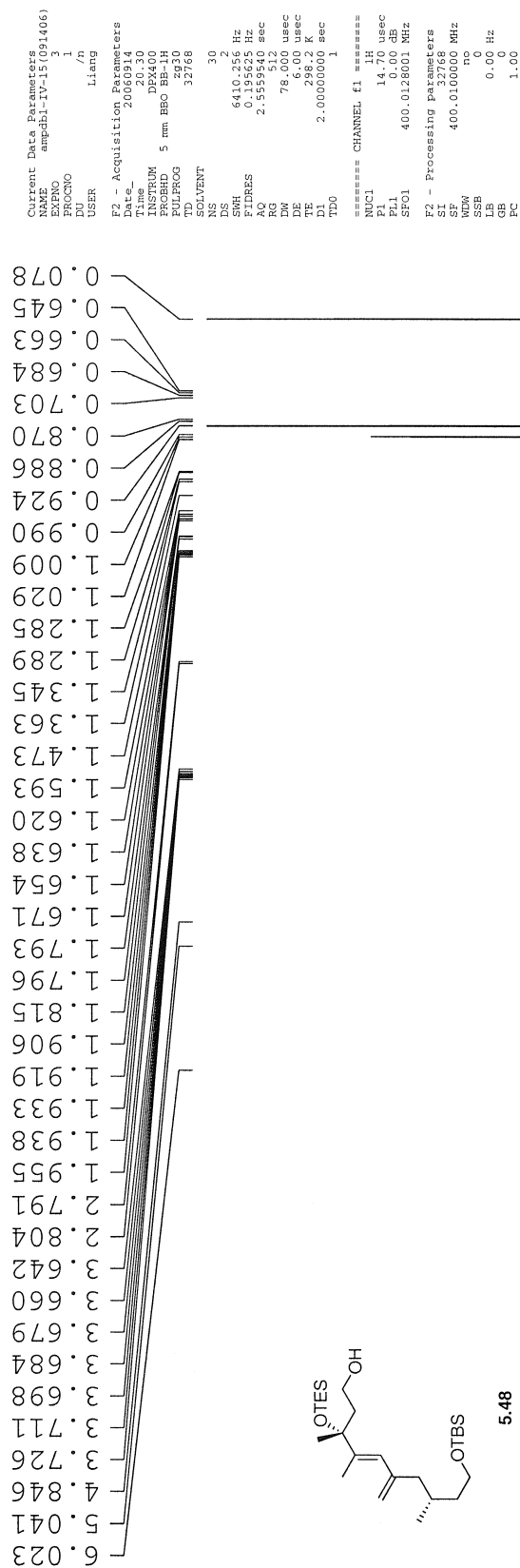
F2 - Acquisition Parameters  
 Date\_ 20060821  
 Time 21.06  
 INSTRUM DPX400  
 PROBHD 5 mm BBO BB-1H  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 1358  
 SSB 4  
 SWH 25125.628 Hz  
 FIDRES 0.383387 Hz  
 AQ 1.3042164 s  
 RG 4096  
 DW 19.900 us  
 DE 6.00 us  
 TE 298.2 K  
 D1 0.15000001 s  
 d11 0.03000000 s  
 DELTA 0.05000000 s  
 TD0 1

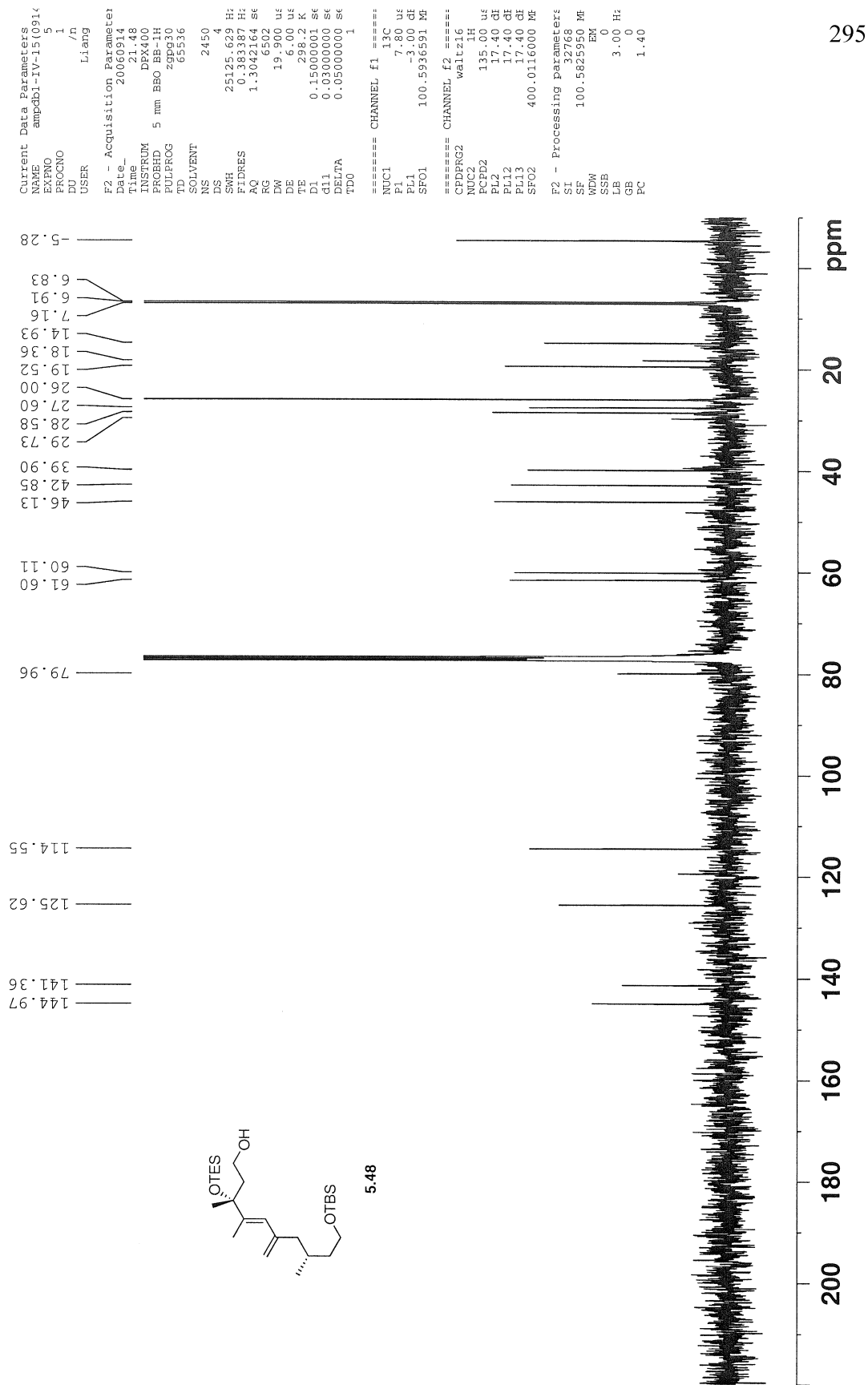
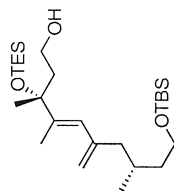
===== CHANNEL f1 =====  
 NUC1 13C  
 PL 7.80 us  
 PL1 3.00 us  
 SFO1 100.5936591 MHz

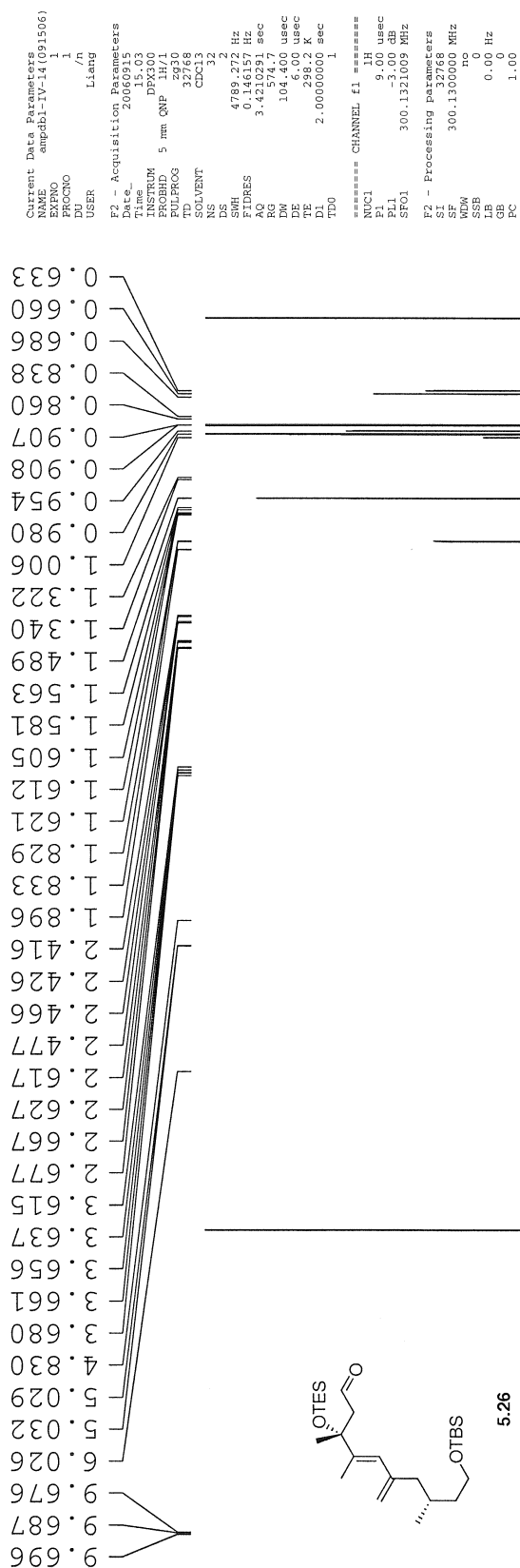
===== CHANNEL f2 =====  
 CDEPRG2 waltz16  
 NUC2 1H  
 PCPD2 135.00 us  
 PL2 17.40 us  
 PL12 17.40 us  
 PL13 17.40 us  
 SFO2 400.0116000 MHz

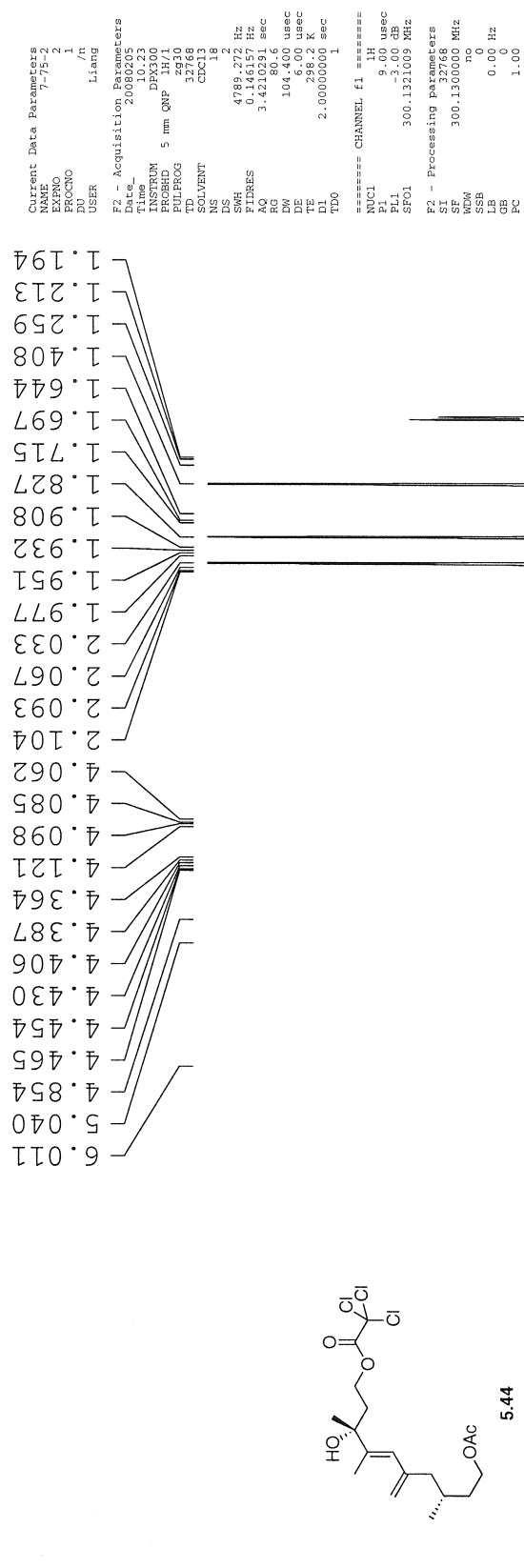
F2 - Processing parameters  
 SI 32768  
 SF 100.5825950 MHz  
 WDW EM  
 SSB 0  
 LB 3.00 Hz  
 GB 0  
 PC 1.40

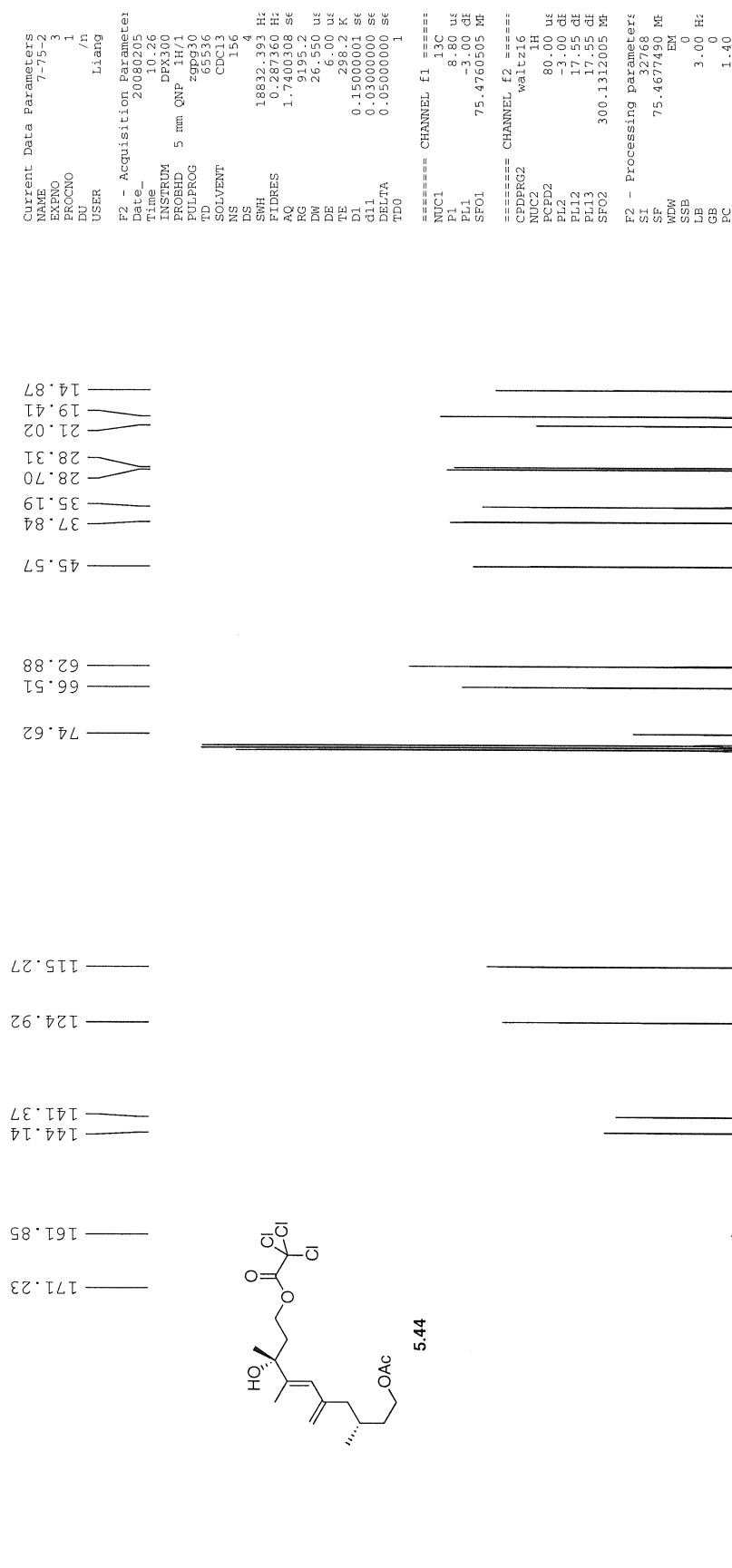
















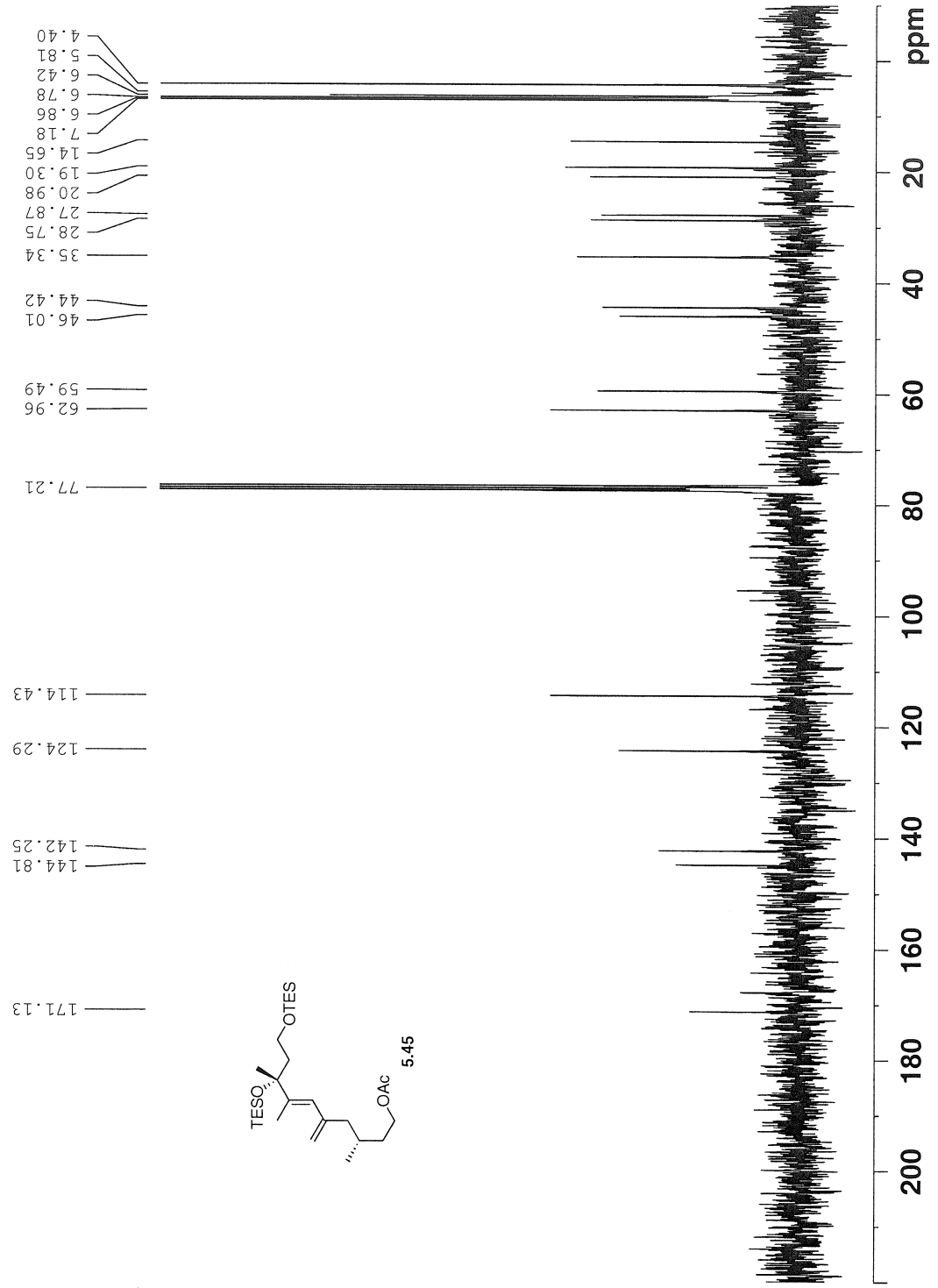
Current Data Parameters  
NAME 7-76-2  
EXPNO 2  
PROCNO 1  
DU /n  
USER Liang

F2 - Acquisition Parameters  
Date\_ 20080214  
Time 15.30  
INSTRUM DEX300  
PROBHD 5 mm QNP 1H/1  
PULPROG zgpg30  
TD 65536  
SOLVENT CDC13  
NS 189  
DS 4  
SWH 18832.393 Hz  
FIDRES 0.287360 Hz  
AQ 1.7400308 s  
RG 9195.2  
DM 26.550 us  
DE 6.00 us  
TE 298.2 K  
D1 0.15000001 s  
d11 0.03000000 s  
DELTA 0.05000000 s  
TD0 1

===== CHANNEL f1 =====  
NUC1 13C  
P1 8.80 us  
PL1 -3.00 dB  
SFO1 75.4760505 MHz

===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 80.00 us  
PL2 -3.00 dB  
PL12 17.55 dB  
PL13 17.55 dB  
SFO2 300.1312005 MHz

F2 - Processing parameters  
SI 32768  
SF 75.4677490 MHz  
WDW EM  
SSB 0  
LB 3.00 Hz  
GB 0  
FC 1.40



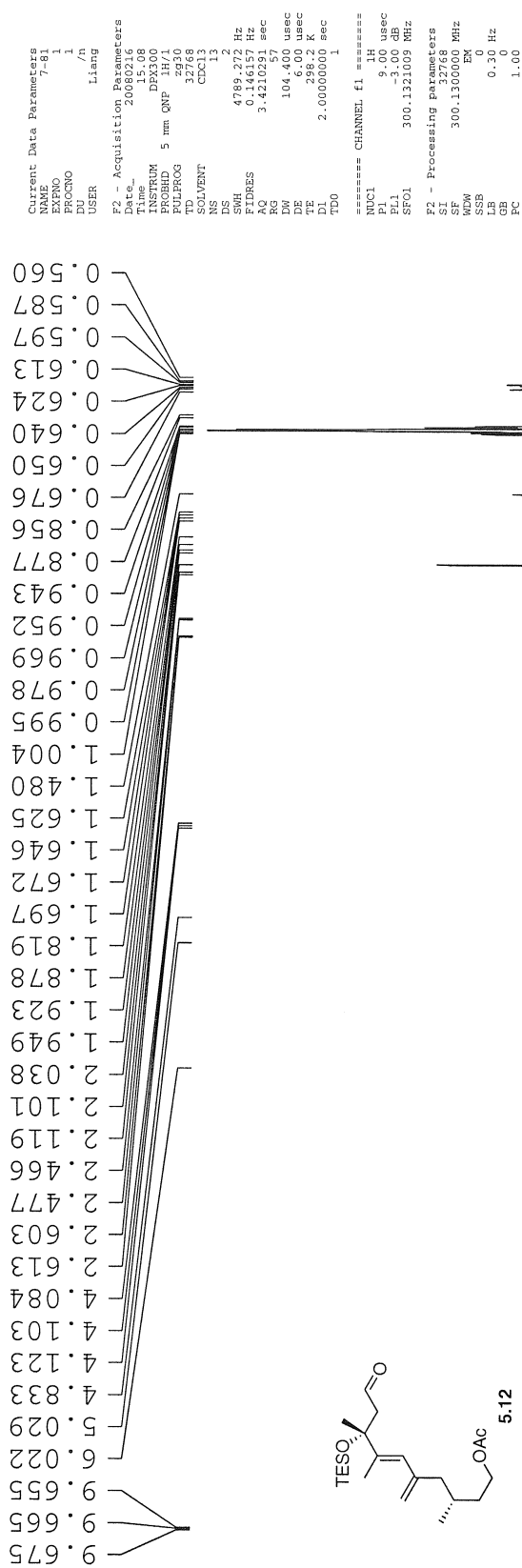
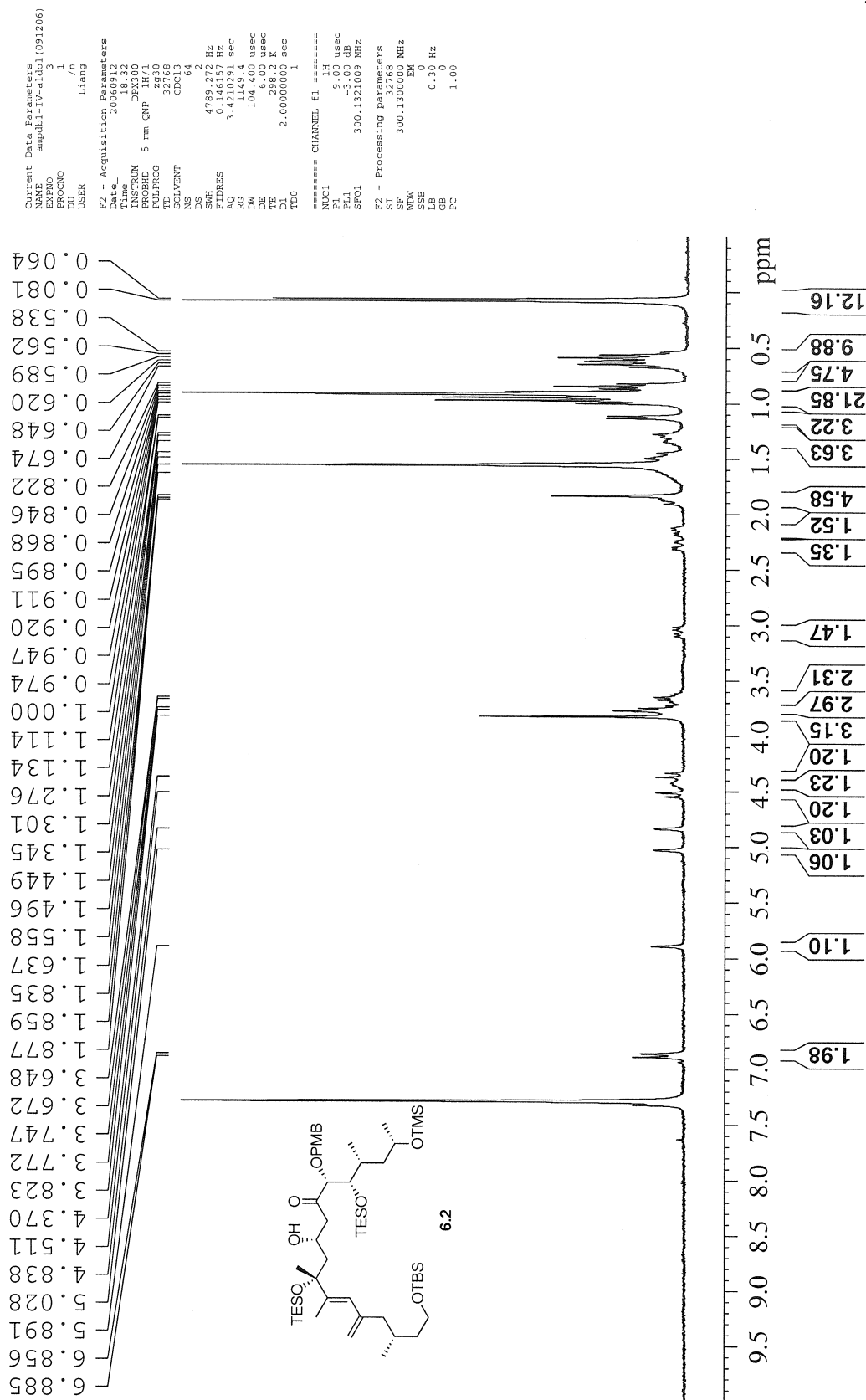
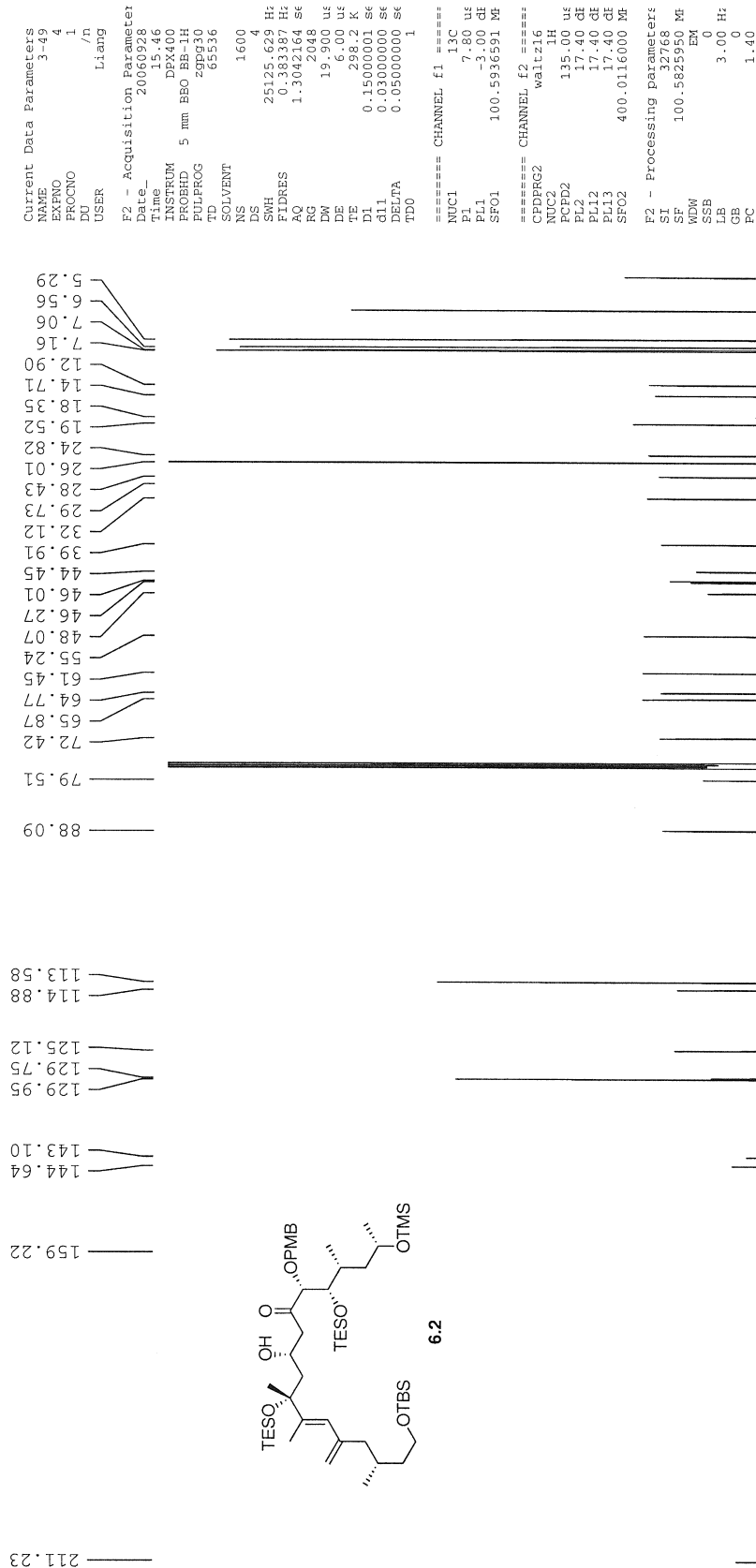


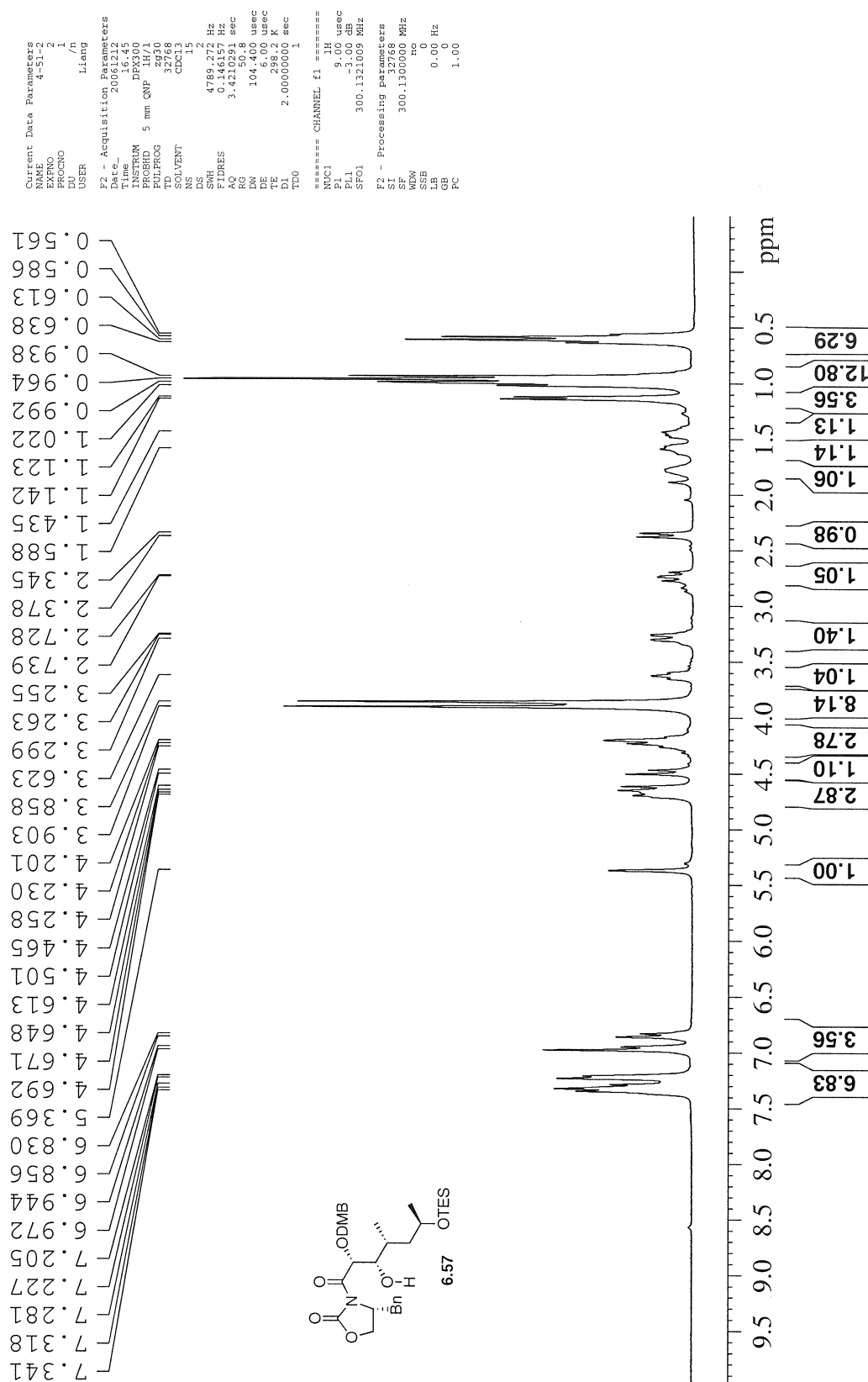
Figure 1 is a line graph illustrating the evolution of the average number of nodes per cluster over time (x-axis, 0 to 100) for various values of  $\alpha$  (y-axis, 0 to 202.96). The graph shows that as  $\alpha$  increases, the average number of nodes per cluster increases and the clusters become more stable over time.

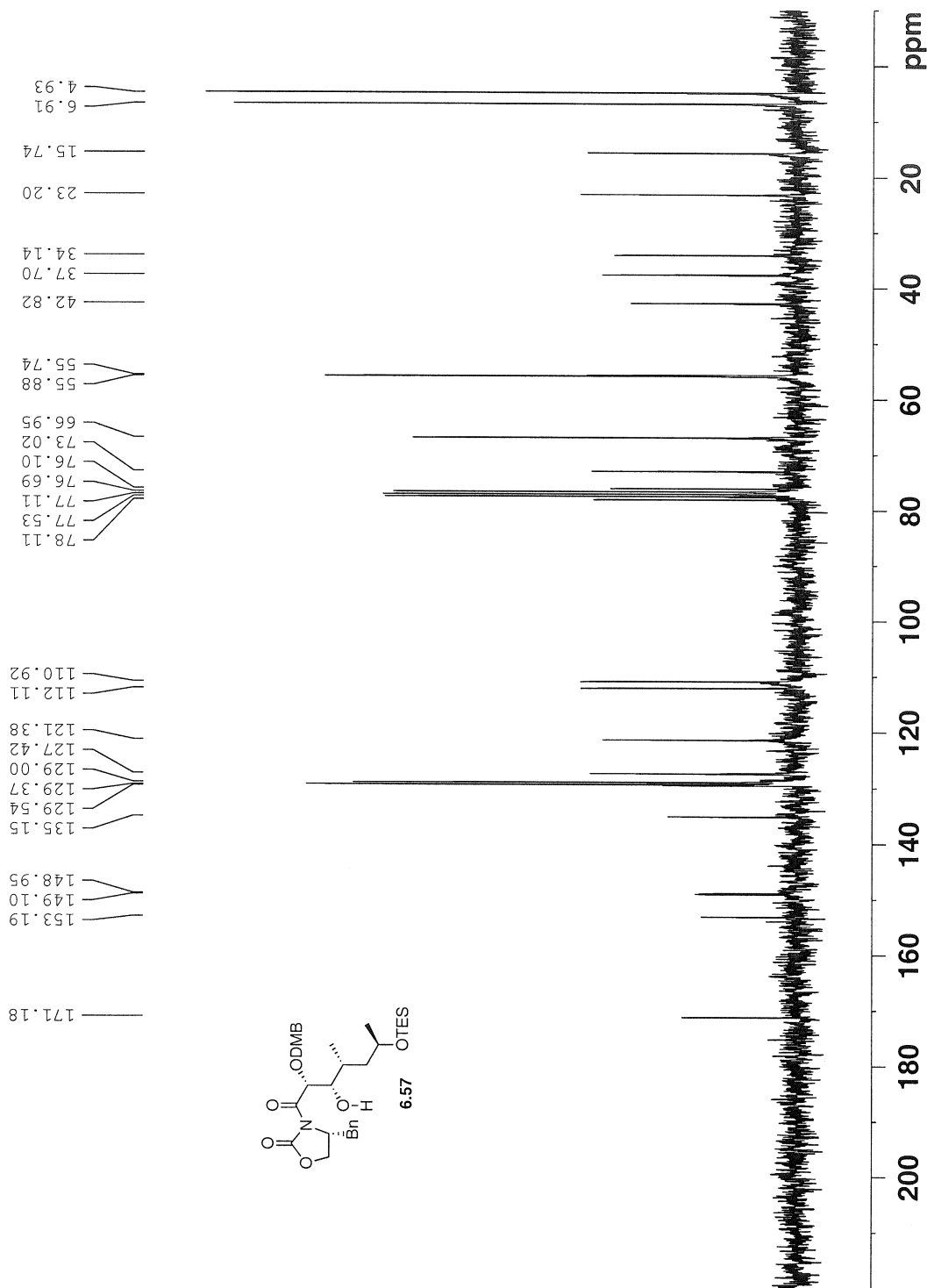
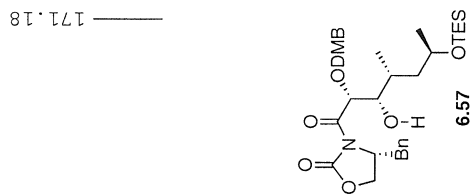
$\alpha$	Approximate Average Number of Nodes per Cluster at Time 100
5.80	~5.8
6.57	~6.6
6.74	~6.7
7.08	~7.1
14.67	~14.7
19.23	~19.2
20.94	~20.9
27.86	~27.9
28.65	~28.7
35.29	~35.3
45.75	~45.8
54.02	~54.0
62.84	~62.8
115.14	~115.1
125.74	~125.7
141.22	~141.2
144.26	~144.3
171.18	~171.2
202.96	~203.0











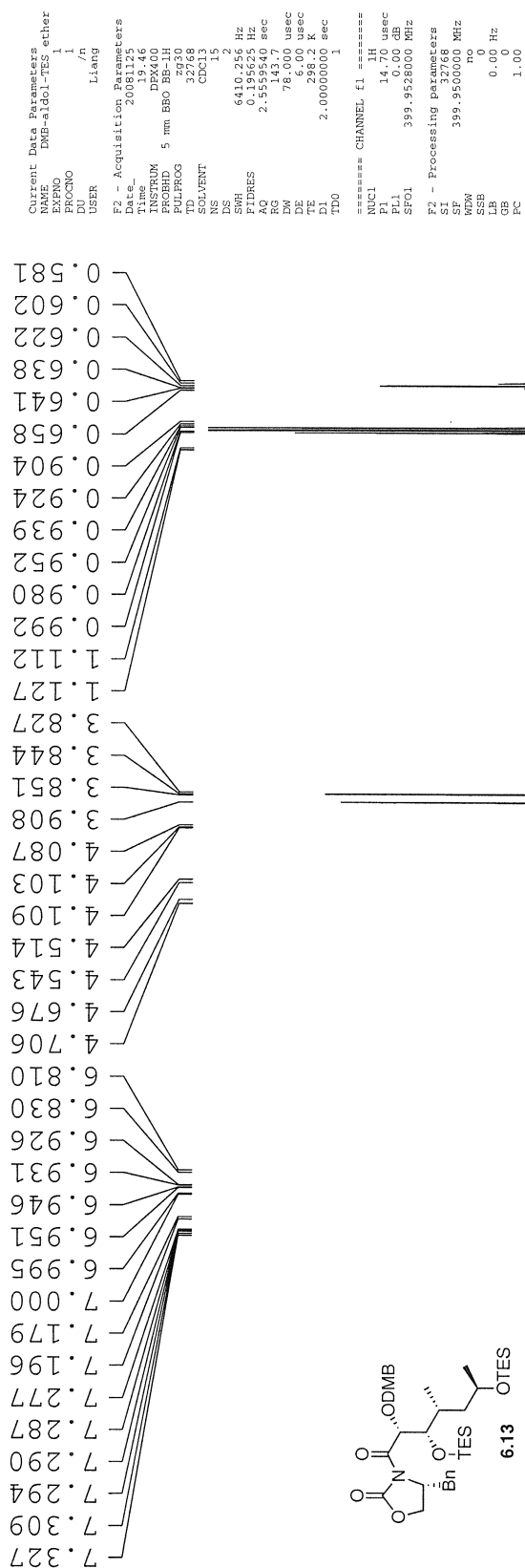
Current Data Parameters  
 NAME 4-51-2  
 EXPNO 4  
 PROCNO 1  
 DU /n  
 USER Liang

F2 - Acquisition Parameters  
 Date\_ 20061212  
 Time 17.03  
 INSTRUM DPX300  
 PROBHD 5 mm QNP 1H/1  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 58  
 DS 4  
 SWH 18832.393 Hz  
 FIDRES 0.287360 Hz  
 AQ 1.7400308 sec  
 RG 16384  
 DW 26.550 us  
 DE 6.00 us  
 TE 298.2 K  
 D1 0.15000001 sec  
 d11 0.03000000 sec  
 DELTA 0.05000000 sec  
 TDO 1

===== CHANNEL f1 =====  
 NUC1 13C  
 P1 8.80 us  
 PL1 -3.00 dB  
 SFO1 75.476505 MHz

===== CHANNEL f2 =====  
 NUC2 1H  
 P2 80.00 us  
 PL2 -3.00 dB  
 PL12 17.55 dB  
 PL13 17.55 dB  
 SFO2 300.1312005 MHz

F2 - Processing parameters  
 SI 32768  
 SF 75.4677450 MHz  
 WDW EM  
 SSB 0  
 LB 3.00 Hz  
 GB 0  
 PC 1.40





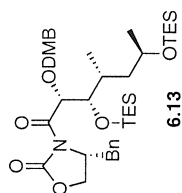
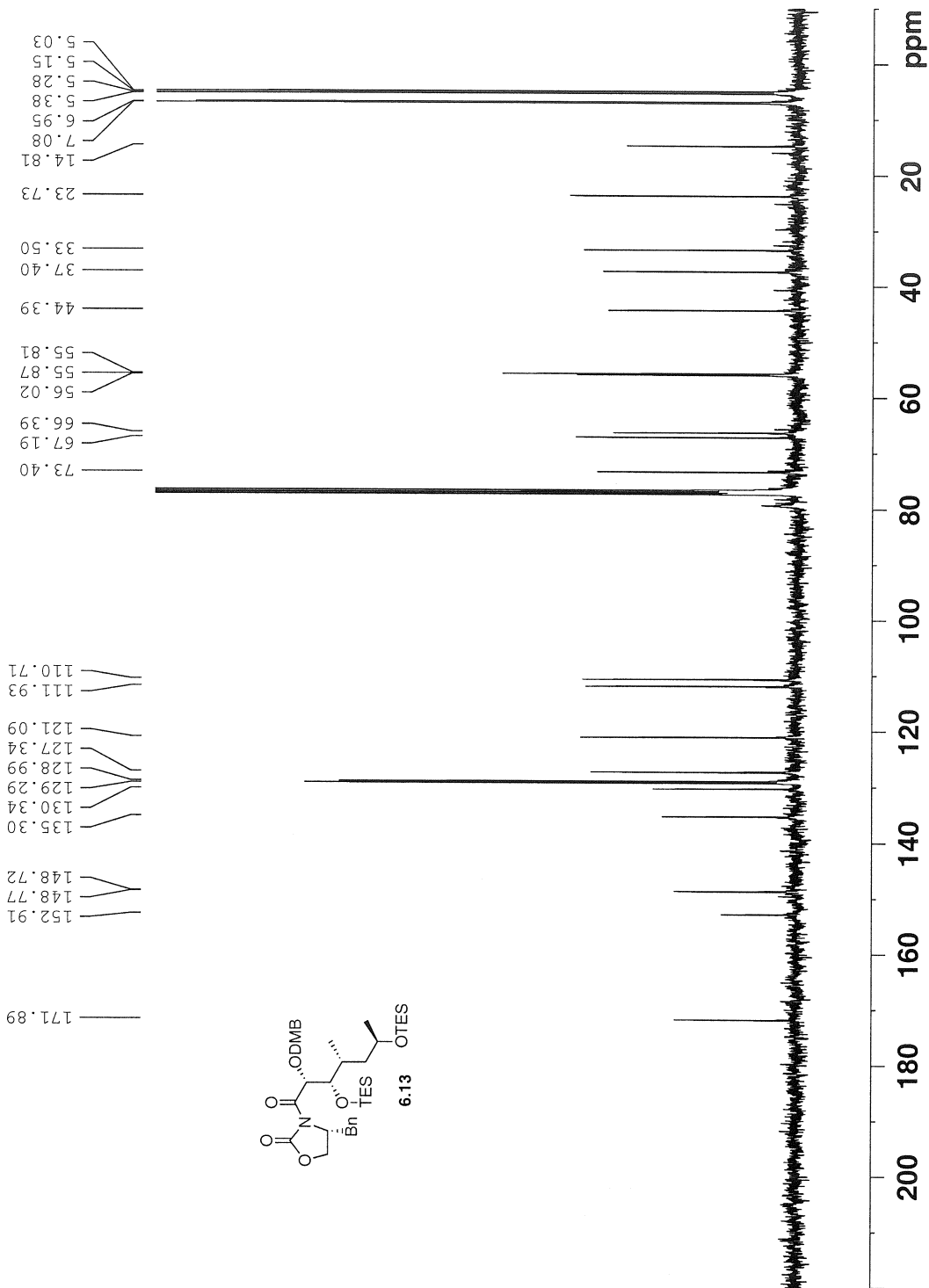
Current Data Parameters  
NAME DMB-aldol-TES etl  
EXPNO 3  
PROCNO 1  
DU /n  
USER Liang

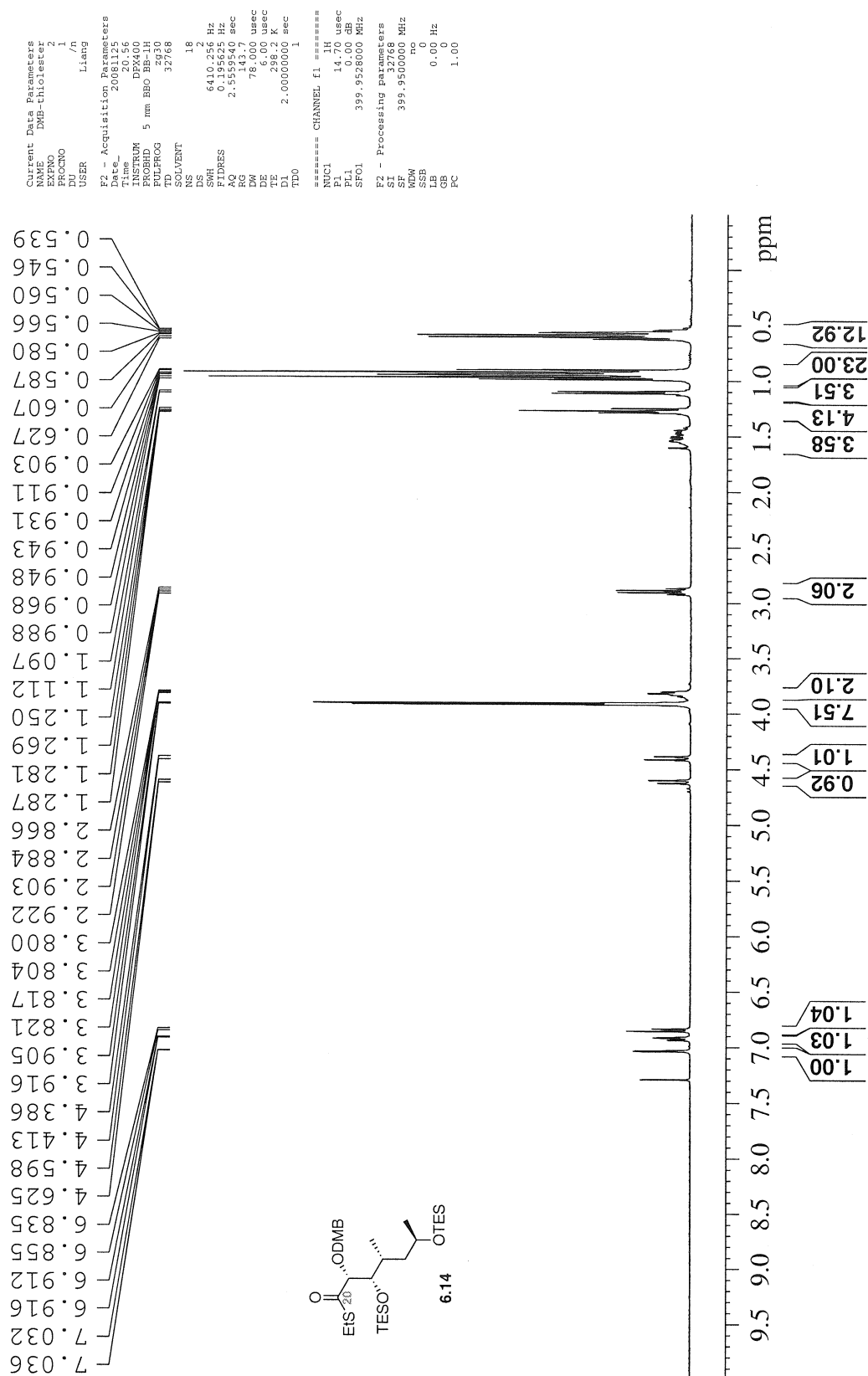
F2 - Acquisition Parameters  
Date\_ 20081125  
Time 20.41  
INSTRUM DFX400  
PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
TD 65536  
SOLVENT NS  
NS 1641  
DS 4  
SWH 25125.629 Hz  
FIDRES 0.383387 Hz  
AQ 1.3042164 s  
RG 16384  
DW 19.900 us  
DE 6.00 us  
TE 298.2 K  
D1 0.15000001 s  
d11 0.03000000 s  
DELTA 0.05000000 s  
TD0 1

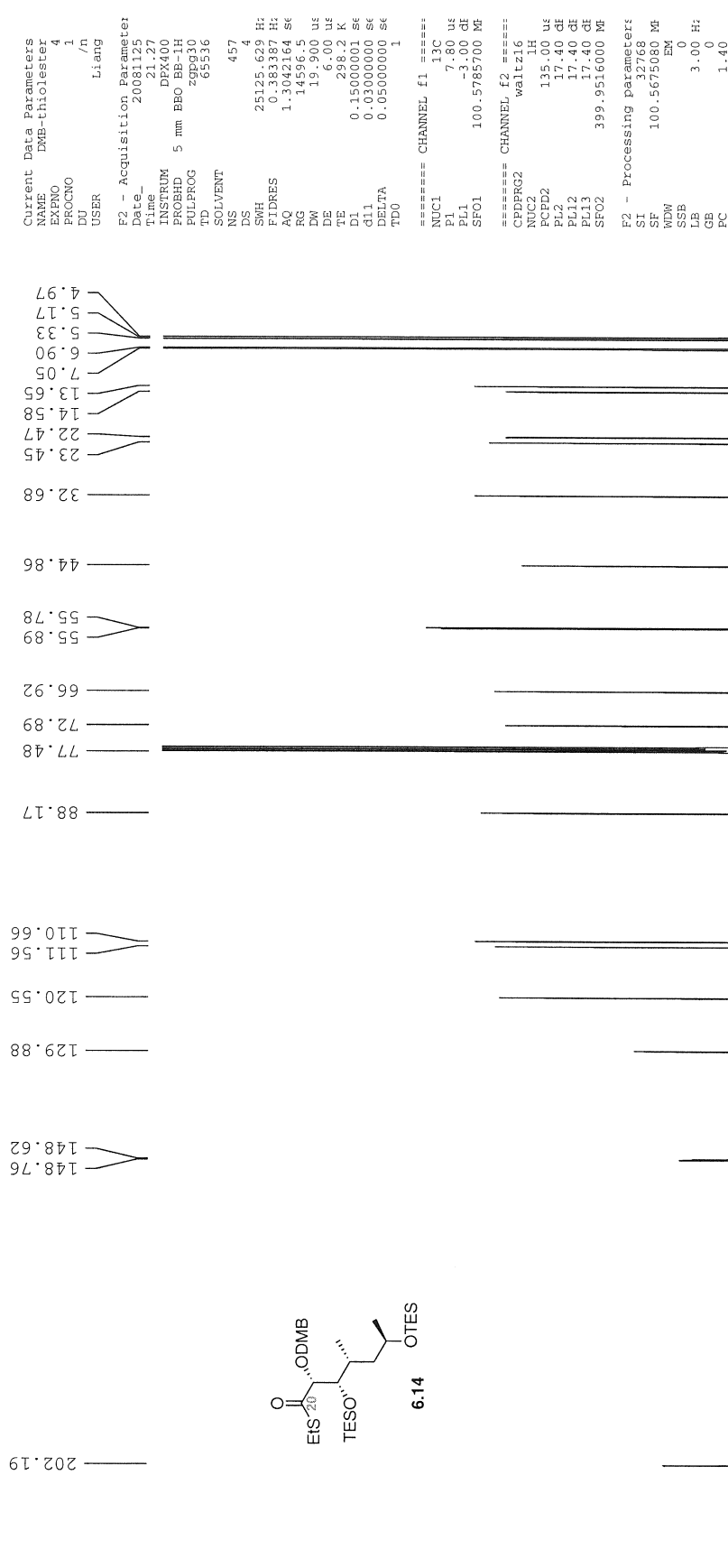
===== CHANNEL f1 =====  
NUC1 13C  
PL1 7.80 us  
PL12 -3.00 dB  
SFO1 100.5785700 MHz

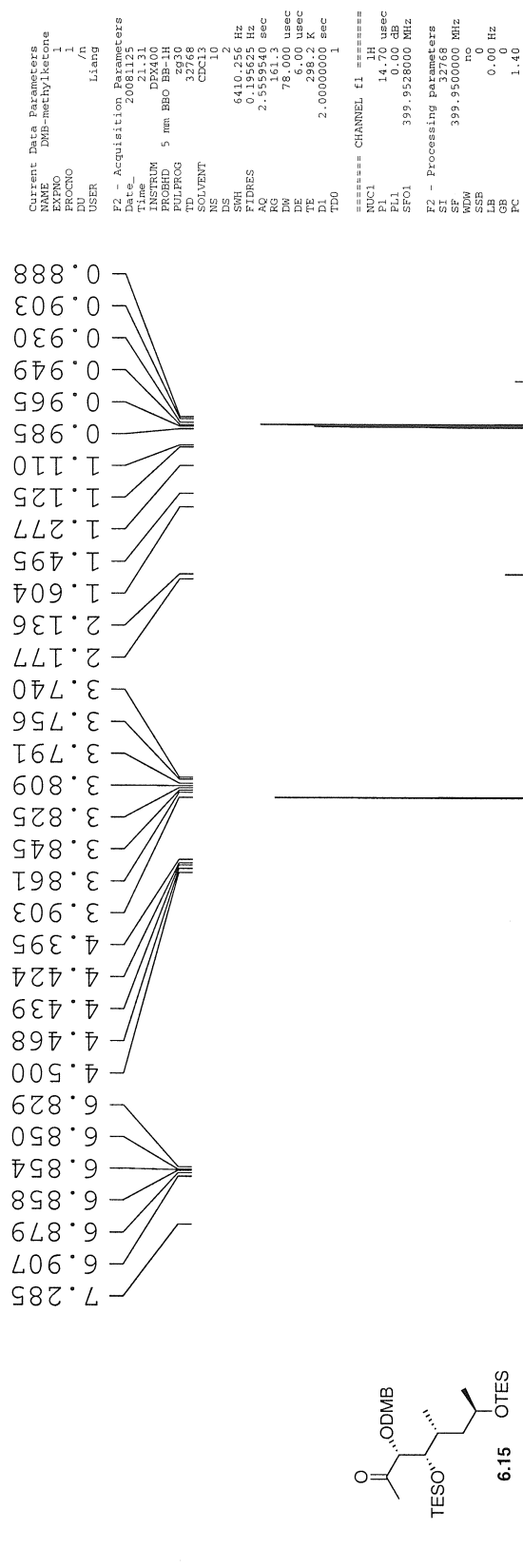
===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 135.00 us  
PL2 17.40 dB  
PL12 17.40 dB  
PL13 17.40 dB  
SFO2 399.9516000 MHz

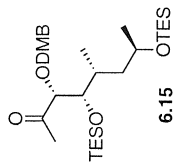
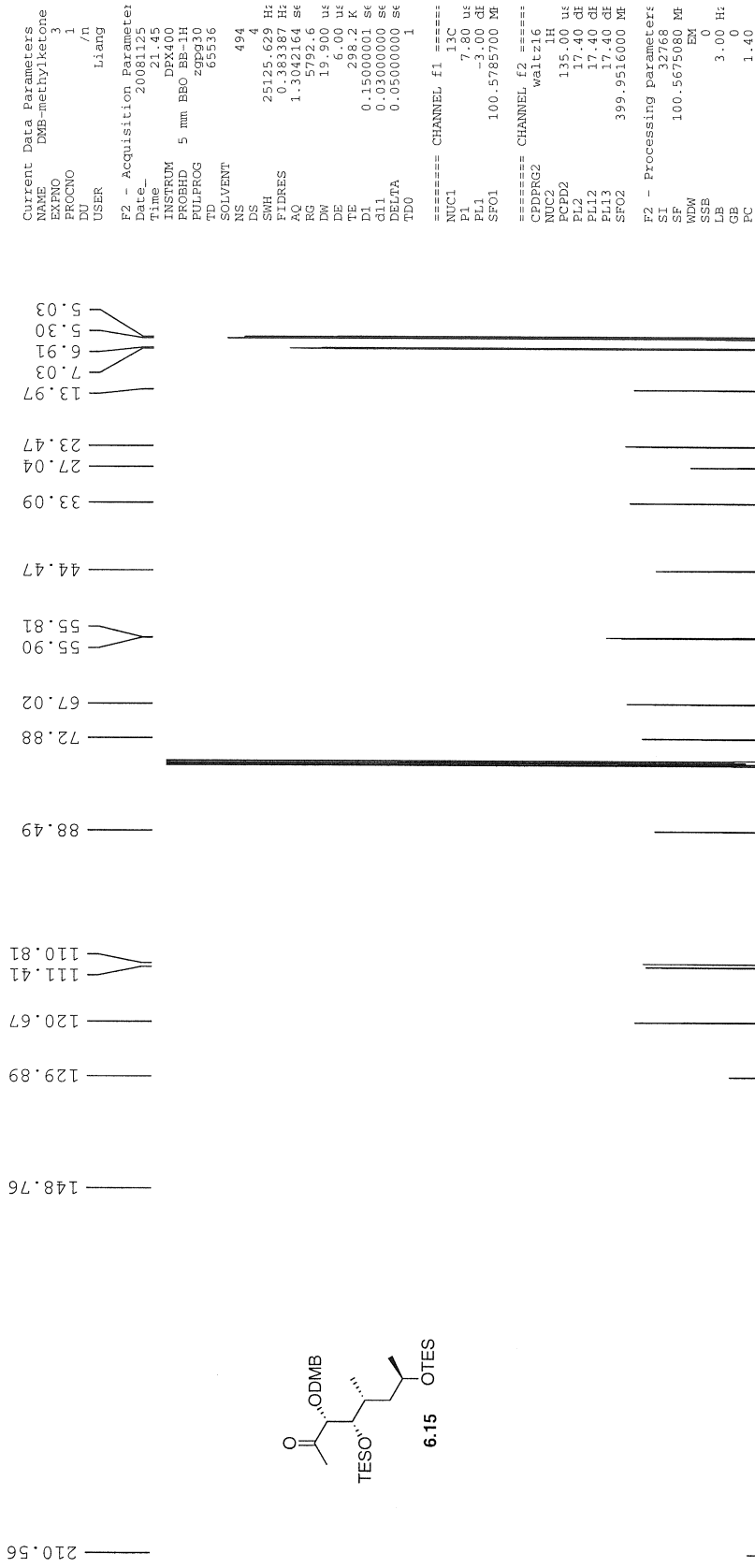
F2 - Processing parameters  
SI 32768  
SF 100.5675080 MHz  
WDW EM  
SSB 0  
GB 0  
PC 3.00 Hz  
1.40

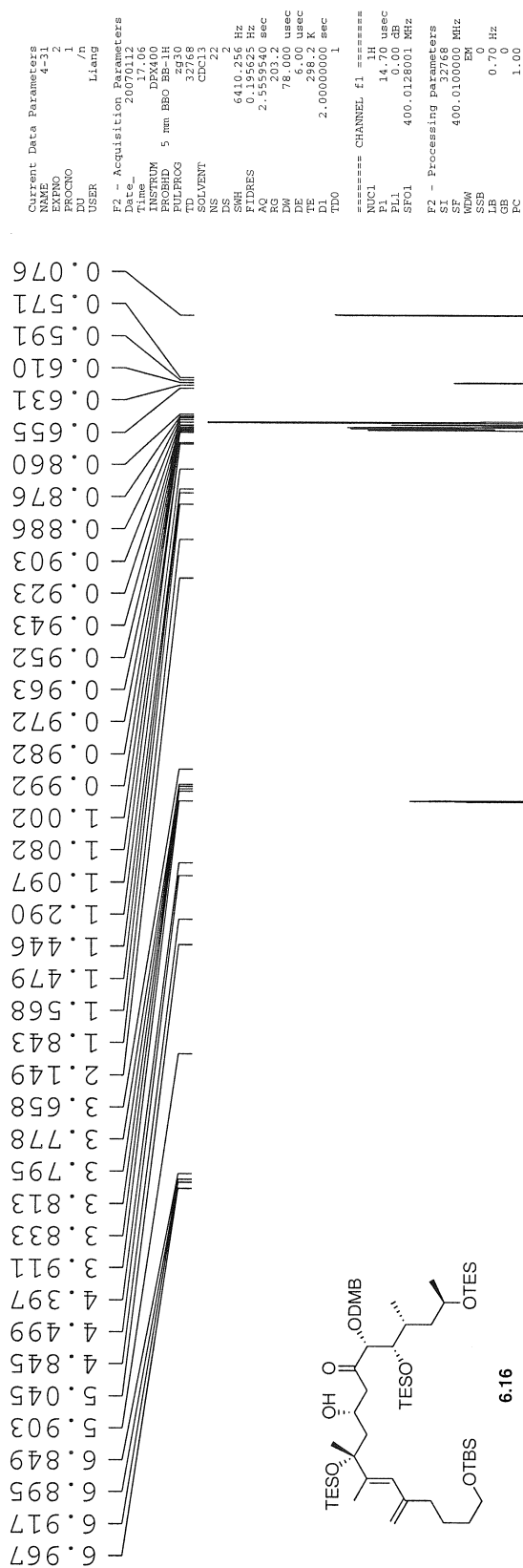




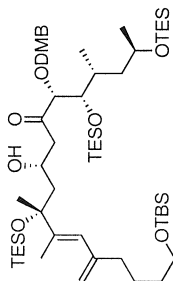




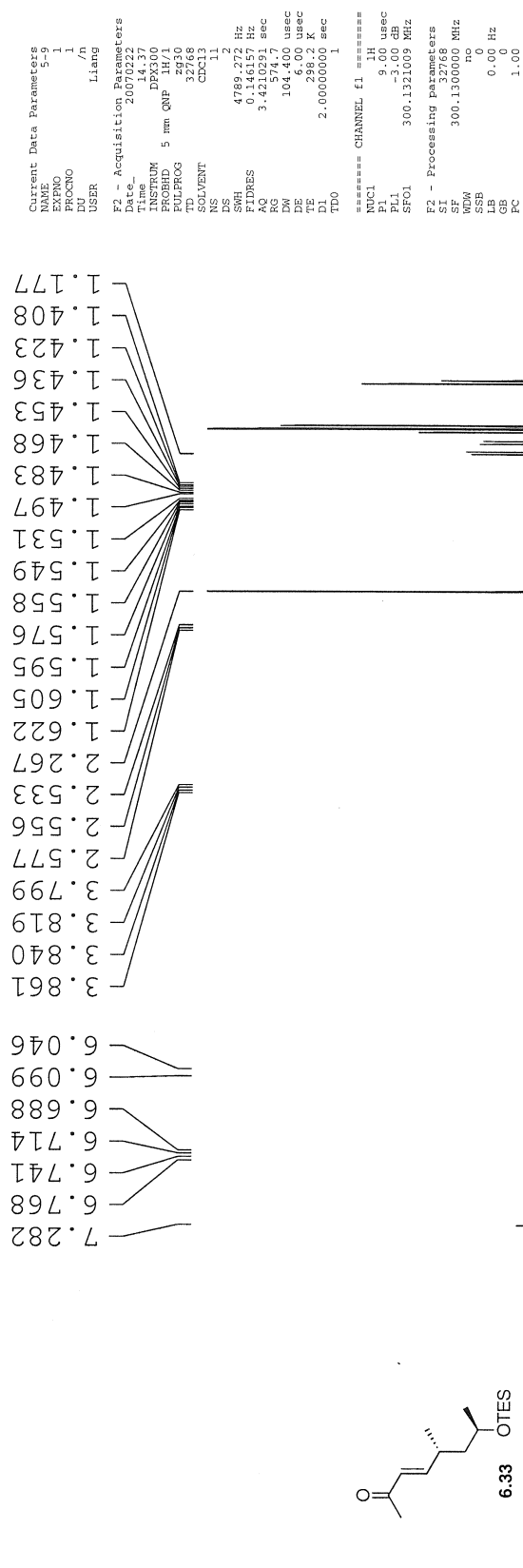




Chemical structure of compound 6.16 is shown below the spectrum. The structure is a complex molecule featuring a central carbon atom bonded to a hydroxyl group (OH), a TESO group, a carboxylic acid group (CO<sub>2</sub>ODMB), and a side chain containing a TESO group, a carboxylic acid group (CO<sub>2</sub>OTBS), and a methyl group. The chemical structure is labeled 6.16.



## 6.16







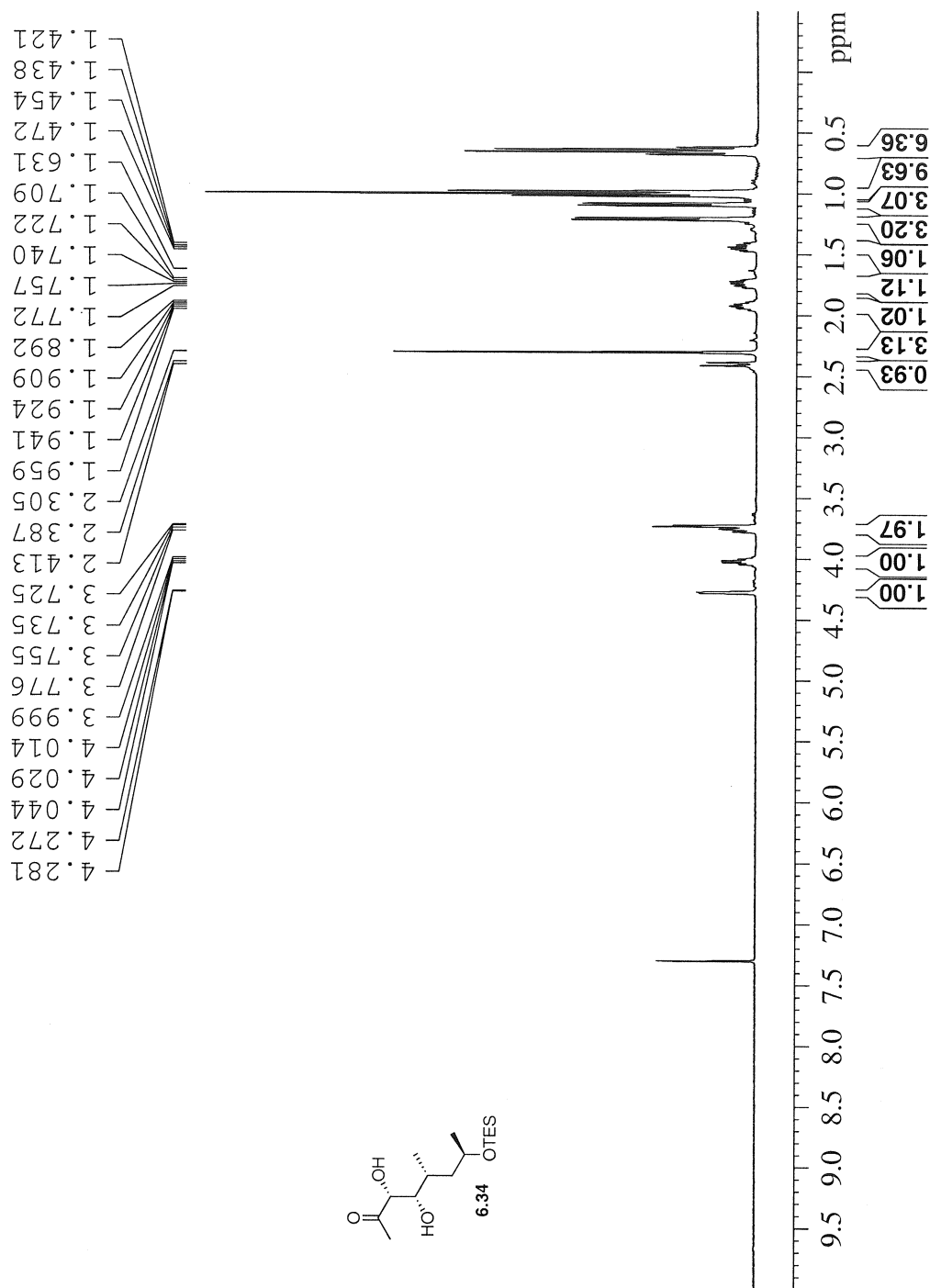
Current Data Parameters	
NAME	5-34-2
EXPNO	3
PROCNO	1
DU	/n
USER	Liang
F2 - Acquisition Parameter	
Date_	20070417
Time	16:34
INSTRUM	BXA400
PULPROG	zgpg30
F2	5 mm BBO
TD	45536
SOLVENT	CDCl3
NS	1184
DS	4
SWH	25125.629 Hz
FIDRES	0.333387 Hz
AQ	1.3042164 s
RG	4597.6
DW	19.900 us
DE	26.00 us
TE	300.2 K
DELTA1	0.1500001 s
deltat1	0.03000000 s
DELTA	0.05000000 s
TD0	1
===== CHANNEL f1 =====	
NUC1	13C
P1P1	7.80 us
PL1	-3.00 dB
SPFO1	100.5936591 MHz
===== CHANNEL f2 =====	
CPDPRG2	waltz16
NUC2	1H
P2P2	135.00 us
PCPD2	17.40 dB
PL12	17.40 dB
PL12	17.40 dB
PL12	17.40 dB
PL13	17.40 dB
SPFO2	400.0116000 MHz
F2 - Processing parameters	
SI	32768
WDW	EM
SSB	100.5825950 MHz
LB	EN
GB	3.00 Hz
PC	0
SC	1.40

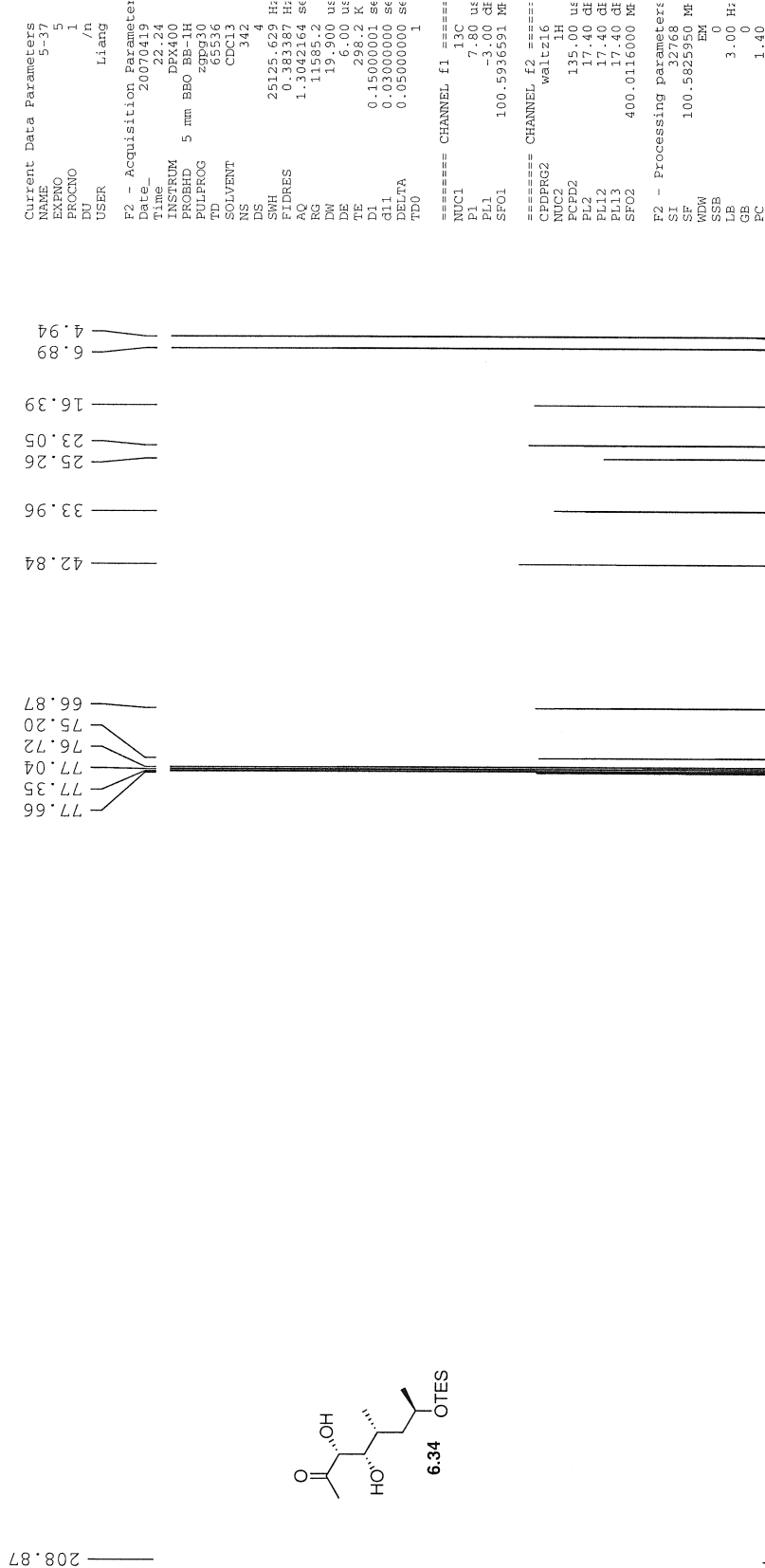
Current Data Parameters  
 NAME 3-3  
 EXPNO 1  
 PROCNO 1  
 DU /n  
 USER Liang

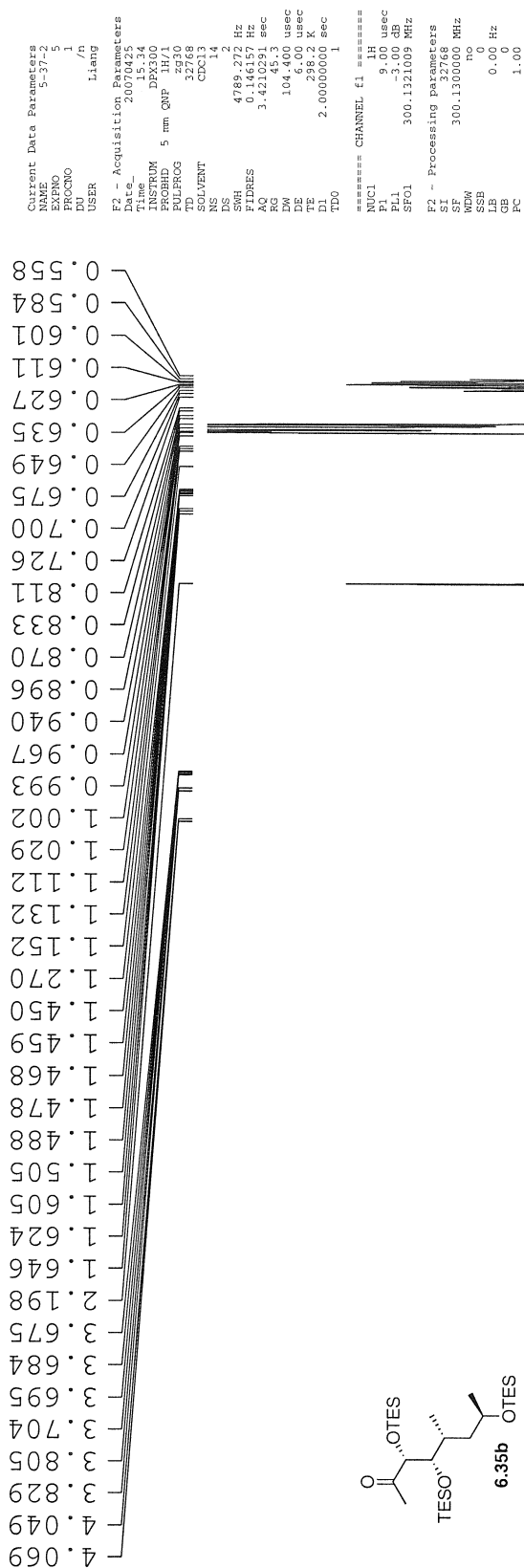
F2 - Acquisition Parameters  
 Date\_ 20070419  
 Time 21.48  
 INSTRUM spect  
 PULPROG zgpg30  
 TD 32768  
 SOLVENT ctd13  
 NS 32  
 DS 2  
 SWH 6410.256 Hz  
 FIDRES 0.195625 Hz  
 AQ 2.5559540 sec  
 RG 327.68  
 DW 78.000 usec  
 DE 6.00 usec  
 TE 298.2 K  
 D1 2.0000000 sec  
 D10 1

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 14.70 usec  
 PL1 0.00 dB  
 SFO1 400.0128001 MHz

F2 - Processing parameters  
 SI 32768  
 SF 400.0100000 MHz  
 WDW no  
 SSB 0  
 LB 0  
 GB 0  
 PC 1.00







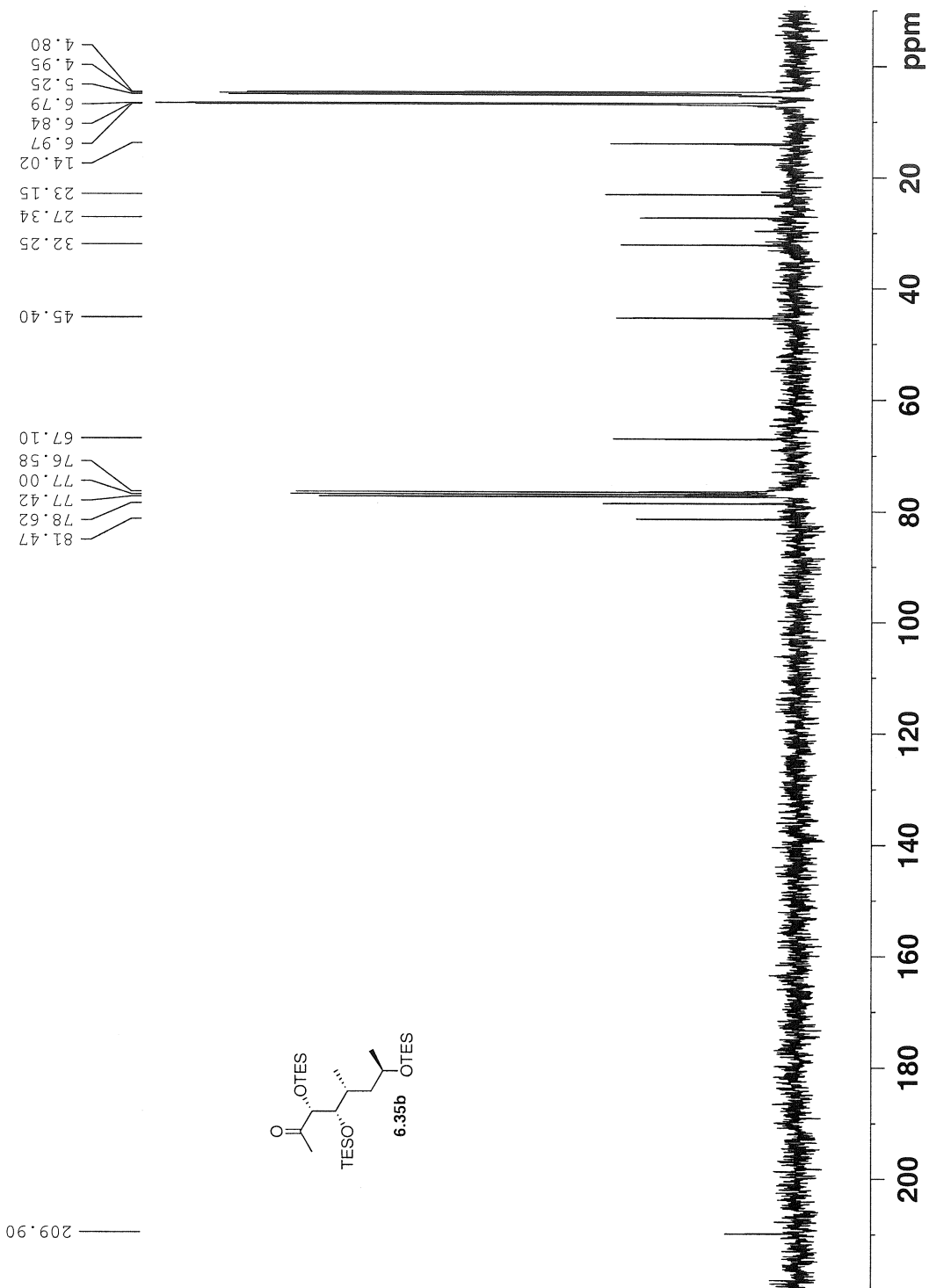
Current Data Parameters  
NAME 5-37-2  
EXPNO 8  
PROCNO 1  
DU /n  
USER Liang

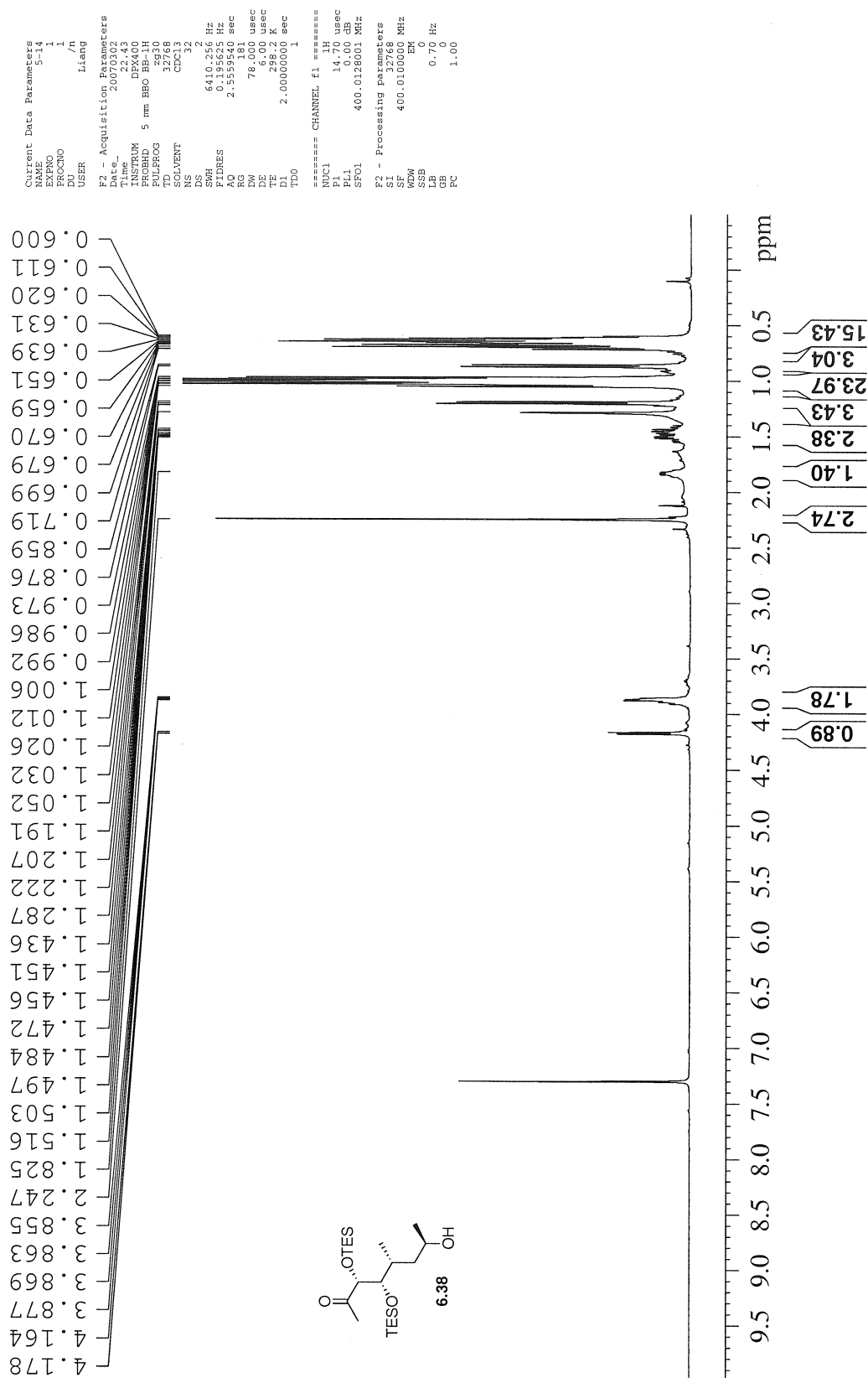
F2 - Acquisition Parameter  
Date\_ 20070425  
Time 16.46  
INSTRUM DPX300  
PROBHD 5 mm QNP 1H/1  
PULPROG zgpg30  
TD 65536  
SOLVENT CDC13  
NS 116  
DS 4  
SWH 18832.393 Hz  
FIDRES 0.287366 Hz  
AQ 1.7400308 sec  
RG 9195.2  
DW 26.550 us  
DE 6.00 us  
TE 298.2 K  
D1 0.15000001 sec  
d11 0.03000000 sec  
DELTA 0.05000000 sec  
TD0 1

===== CHANNEL f1 =====  
NUC1 13C  
P1 8.80 us  
PL1 -3.00 dB  
SFO1 75.4760505 MHz

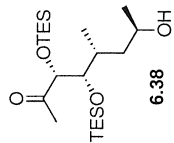
===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 80.00 us  
PL2 -3.00 dB  
PL12 17.55 dB  
PL13 17.55 dB  
SFO2 300.1312005 MHz

F2 - Processing parameters  
SI 32768  
SF 75.4677490 MHz  
WDW EM  
SSB 0  
LB 3.00 Hz  
GB 0  
PC 1.40





209.28



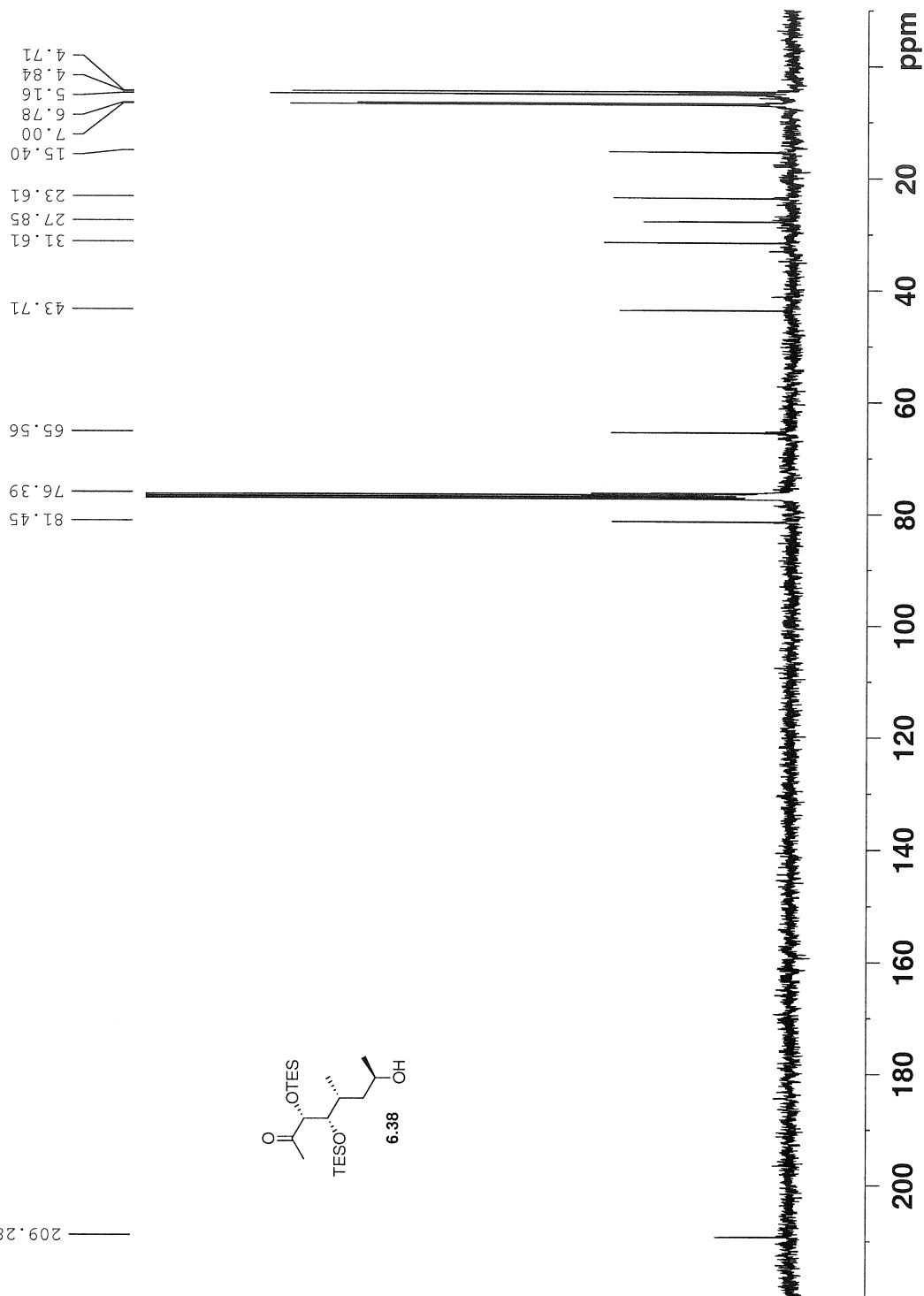
Current Data Parameters  
 NAME 5-38  
 EXPNO 1  
 PROCNO 1  
 DU /n  
 USER Liang

F2 - Acquisition Parameters  
 Date\_ 20070424  
 Time 18.56  
 INSTRUM DFX400  
 PROBHD 5 mm BBO BB-1H  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT  
 NS 740  
 DS 4  
 SWH 25135.627 Hz  
 FIDRES 0.383387 Hz  
 AQ 1.3042164 s  
 RG 4597.6  
 DW 19.900 us  
 DE 6.00 us  
 TE 298.2 K  
 D1 0.15000001 s  
 d11 0.03000000 s  
 DELTA 0.05000000 s  
 TDO 1

===== CHANNEL f1 =====  
 NUC1 13C  
 P1 7.80 us  
 PL1 3.00 dB  
 SFO1 100.5936591 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 135.00 us  
 PL2 17.40 dB  
 PL12 17.40 dB  
 PL13 17.40 dB  
 SFO2 400.0116000 MHz

F2 - Processing parameters  
 SI 32768  
 SF 100.5825950 MHz  
 WDW EM  
 SSB 0  
 LB 3.00 Hz  
 GB 0  
 PC 1.40

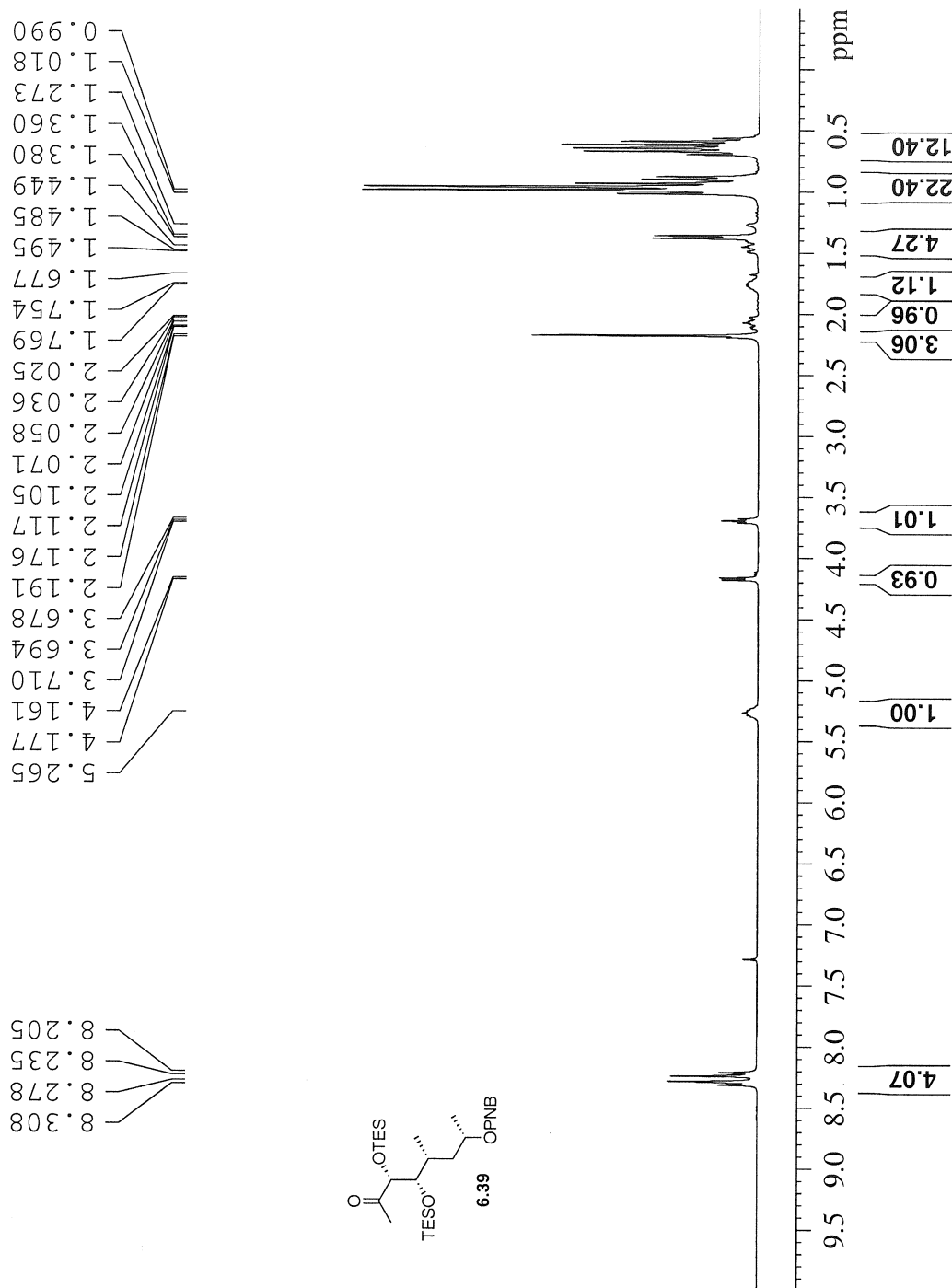


Current Data Parameters  
 NAME 5-2  
 EXPNO 1  
 PROCNO 1  
 DU /n  
 USER Liang

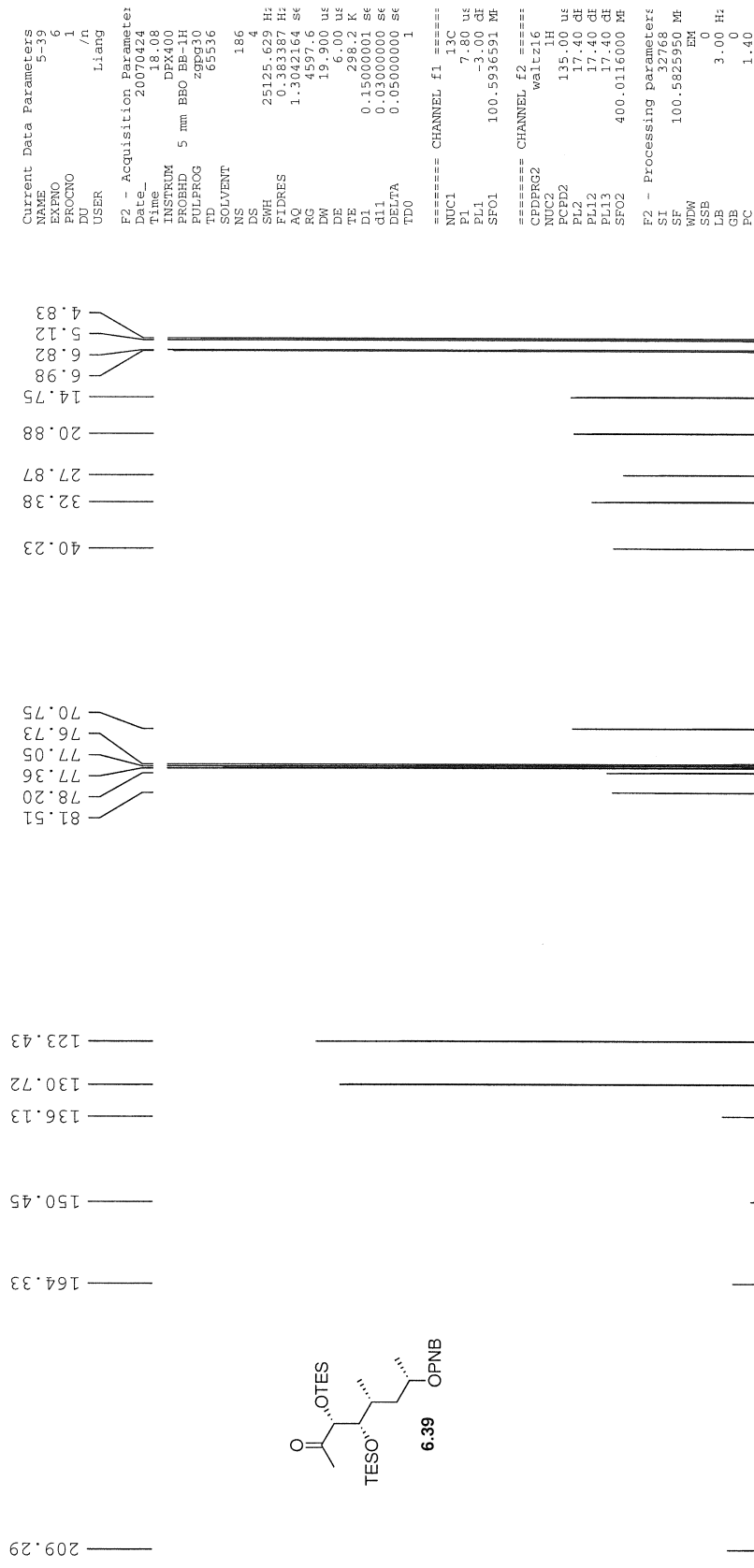
F2 - Acquisition Parameters  
 Date\_ 20070424  
 Time 17.33  
 INSTRUM spect  
 PROBD 5 mm QNP 1H/1  
 PULPROG zg30  
 TD 32768  
 SOLVENT CDCl3  
 NS 32  
 DS 2  
 SMH 4789.272 Hz  
 FIDRES 0.146157 Hz  
 AQ 3.4210291 sec  
 RG 327.68  
 DW 104.400 usec  
 DE 6.00 usec  
 TE 298.2 K  
 D1 2.0000000 sec  
 D10 1

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 9.00 usec  
 PL1 -3.00 dB  
 SFO1 300.1321009 MHz

F2 - Processing parameters  
 SI 32768  
 SF 300.1300000 MHz  
 MMW no  
 SSB 0  
 LB 0  
 GB 0  
 PC 1.00





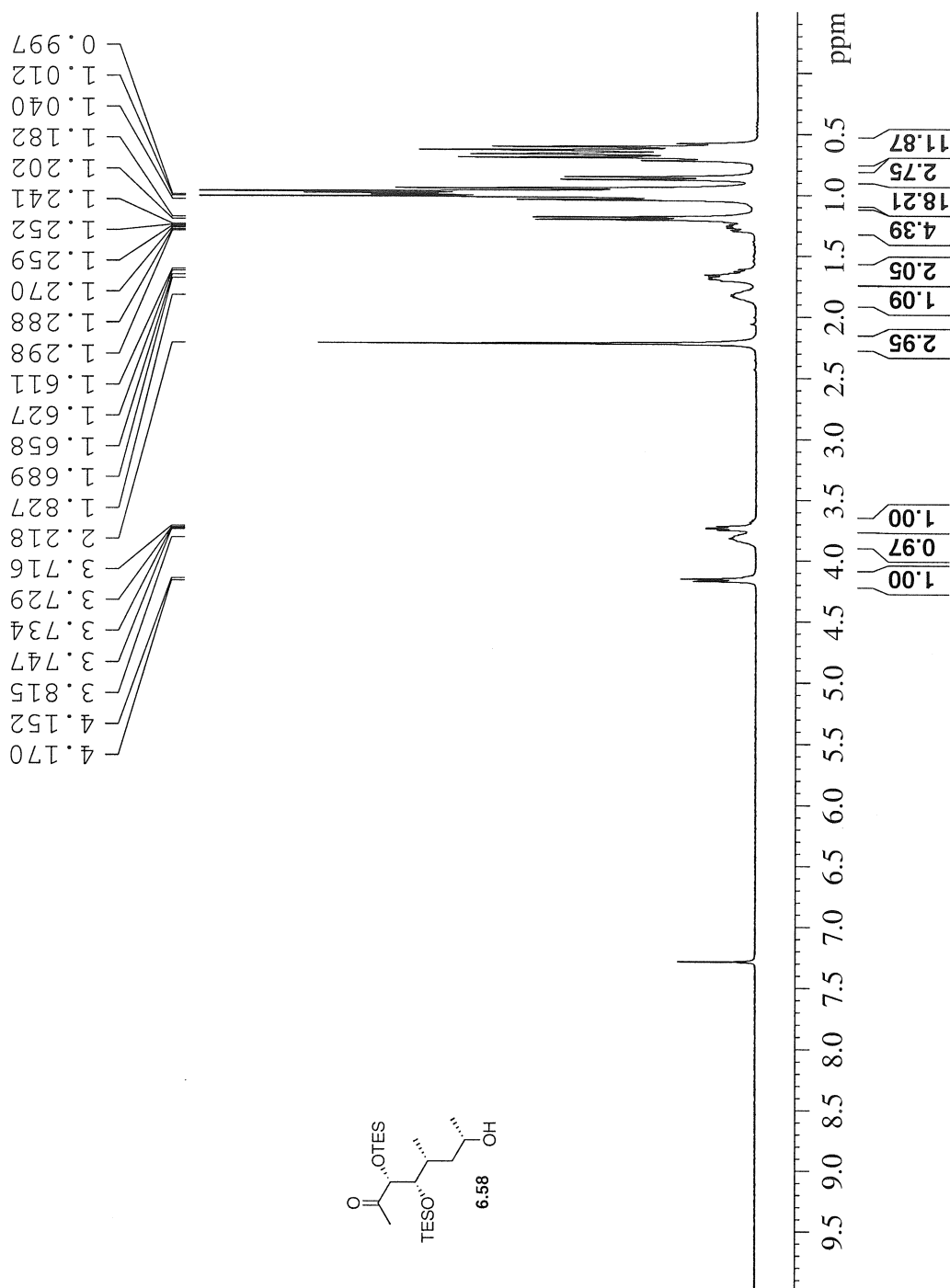


Current Data Parameters  
 NAME 5-40  
 EXPNO 2  
 PROCNO 1  
 DU /n  
 USER Liang

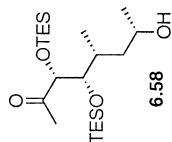
F2 - Acquisition Parameters  
 Date\_ 20070426  
 Time 11.14  
 INSTRUM spect  
 PROBRD 5 mm QNP 1H/1  
 PULPROG zg30  
 TD 32768  
 SOLVENT CDCl3  
 NS 12  
 DS 2  
 SMH 4789.272 Hz  
 FIDRES 0.146157 Hz  
 AQ 3.42109 sec  
 RQ 1.137  
 LW 104.400 usec  
 DE 6.00 usec  
 TE 298.2 K  
 D1 2.0000000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 9.00 usec  
 PL1 -3.00 dB  
 SFO1 300.1321009 MHz

F2 - Processing parameters  
 SI 32768  
 SF 300.1300000 MHz  
 WDW no  
 SSB 0  
 GB 0.00 Hz  
 GSS 0  
 PC 1.00



210.08



4.78  
5.16  
6.79  
6.97  
15.22  
24.41  
27.61  
32.88  
43.79  
66.19  
76.58  
77.01  
77.43  
78.06  
81.51

Current Data Parameters  
NAME 5-40  
EXPNO 1  
PROCNO 1  
DU /n  
USER Liang

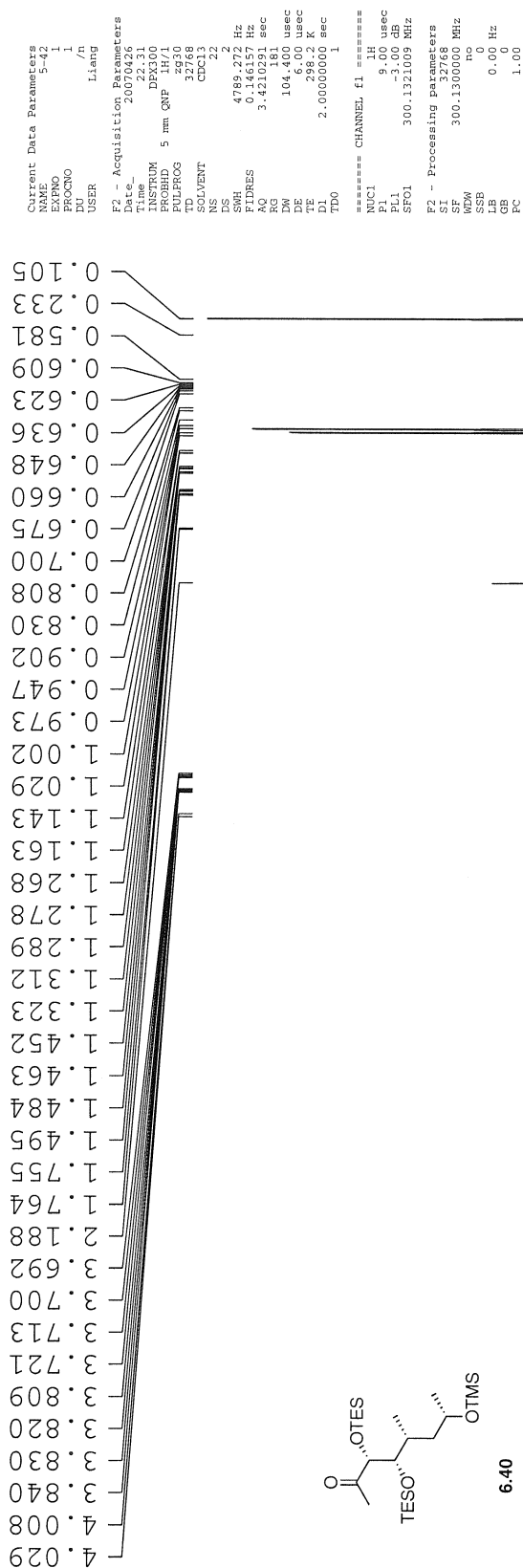
F2 - Acquisition Parameters  
Date\_ 20070426  
Time 11.26  
INSTRUM DFX300  
PROBHD 5 mm QNP 1H/1  
PULPROG zgpg30  
TD 65536  
SOLVENT CDCl3  
NS 204  
DS 4  
SWH 18832.363 Hz  
FIDRES 0.287360 Hz  
AQ 1.7400308 s  
RG 9195.2  
DW 26.550 us  
DE 6.00 us  
TE 298.2 K  
D1 0.15000001 s  
d11 0.03000000 s  
DELTA 0.05000000 s  
TDO 1

===== CHANNEL f1 =====  
NUC1 13C  
P1 8.80 us  
PL1 -3.00 dB  
SFO1 75.4760595 MHz

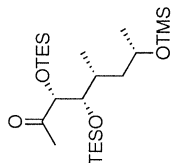
===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 80.00 us  
PL2 -3.00 dB  
PL12 17.55 dB  
PL13 17.55 dB  
SFO2 300.1312005 MHz

F2 - Processing parameters  
SI 32768  
SF 75.4677490 MHz  
WDW EM  
SSB 0  
LB 3.00 Hz  
GB 0  
PC 1.40

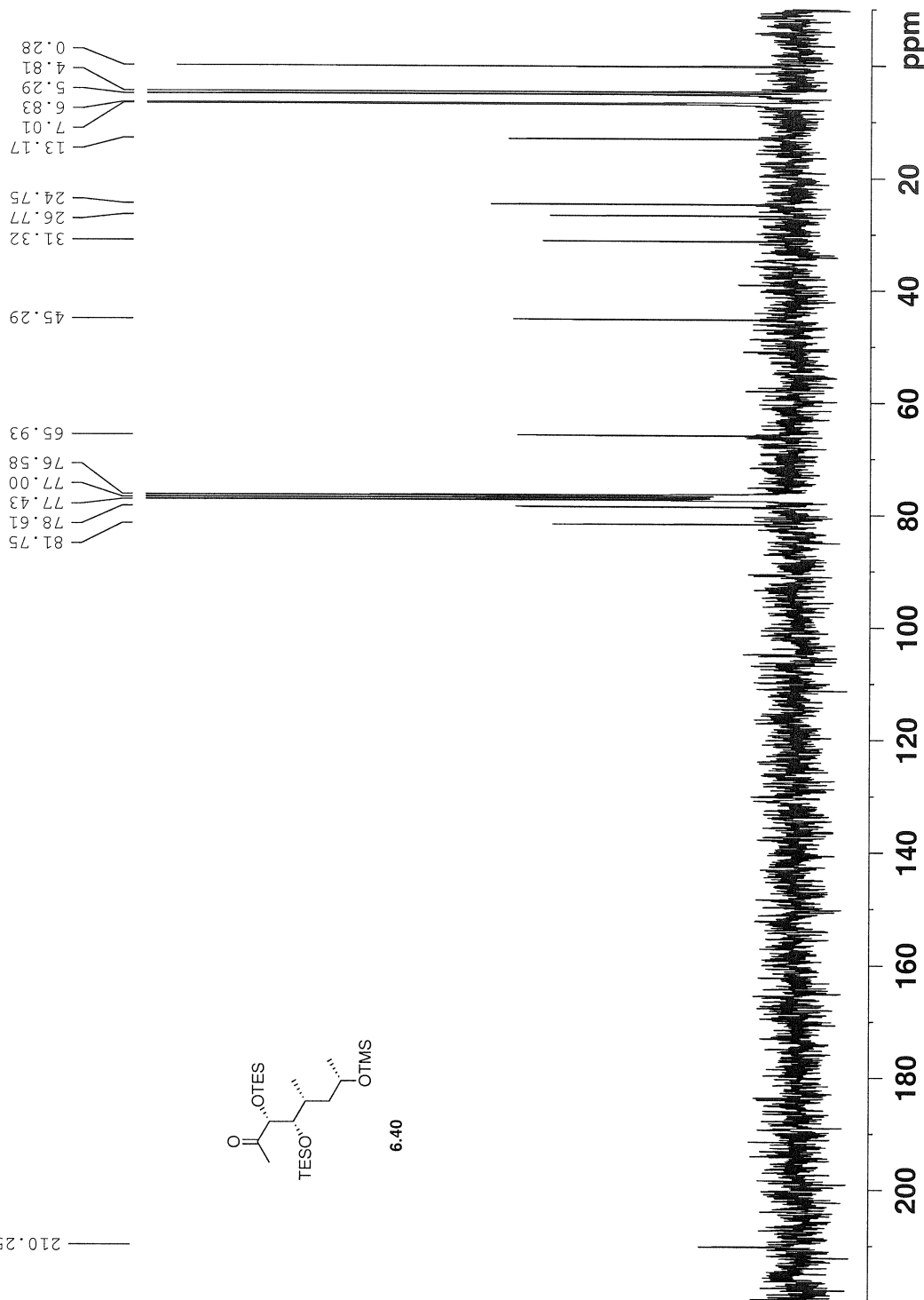
ppm



210.25



6.40



Current Data Parameters  
 NAME 5-43  
 EXNO 1  
 PRGNO 1  
 DU /n  
 USER Liang

F2 - Acquisition Parameter  
 Date\_ 20070426  
 Time 22.45  
 INSTRUM DFX300  
 PROBHD 5 mm QNP 1H/1  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 407  
 DS 4  
 SWH 18932.303 Hz  
 FIDRES 0.287360 Hz  
 AQ 1.7400308 sec  
 RG 9195.2  
 DW 26.550 us  
 DE 6.00 us  
 TE 298.2 K  
 D1 0.15000001 sec  
 d11 0.03000000 sec  
 DELTA 0.05000000 sec  
 TDO 1

===== CHANNEL f1 =====  
 NUC1 13C  
 P1 8.80 us  
 PL1 -3.00 dB  
 SFO1 75.4760505 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 P2 80.00 us  
 PL2 -3.00 dB  
 PL12 17.55 dB  
 PL13 17.55 dB  
 SFO2 300.1312005 MHz

F2 - Processing parameters  
 SI 32768  
 SF 75.4677490 MHz  
 WDW EM  
 SSB 0  
 LB 3.00 Hz  
 GB 0  
 FC 1.40

ppm

20

40

60

80

100

120

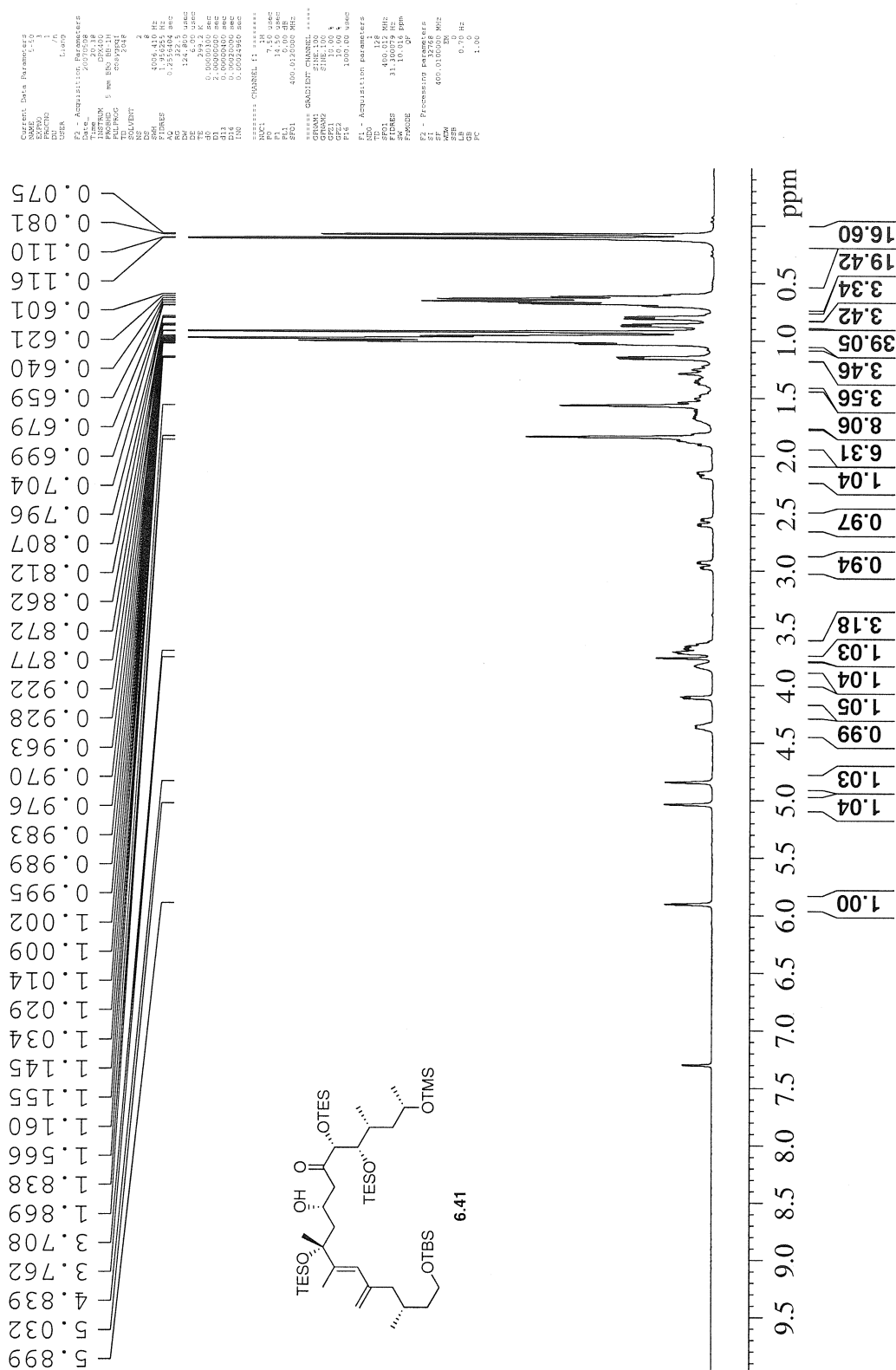
140

160

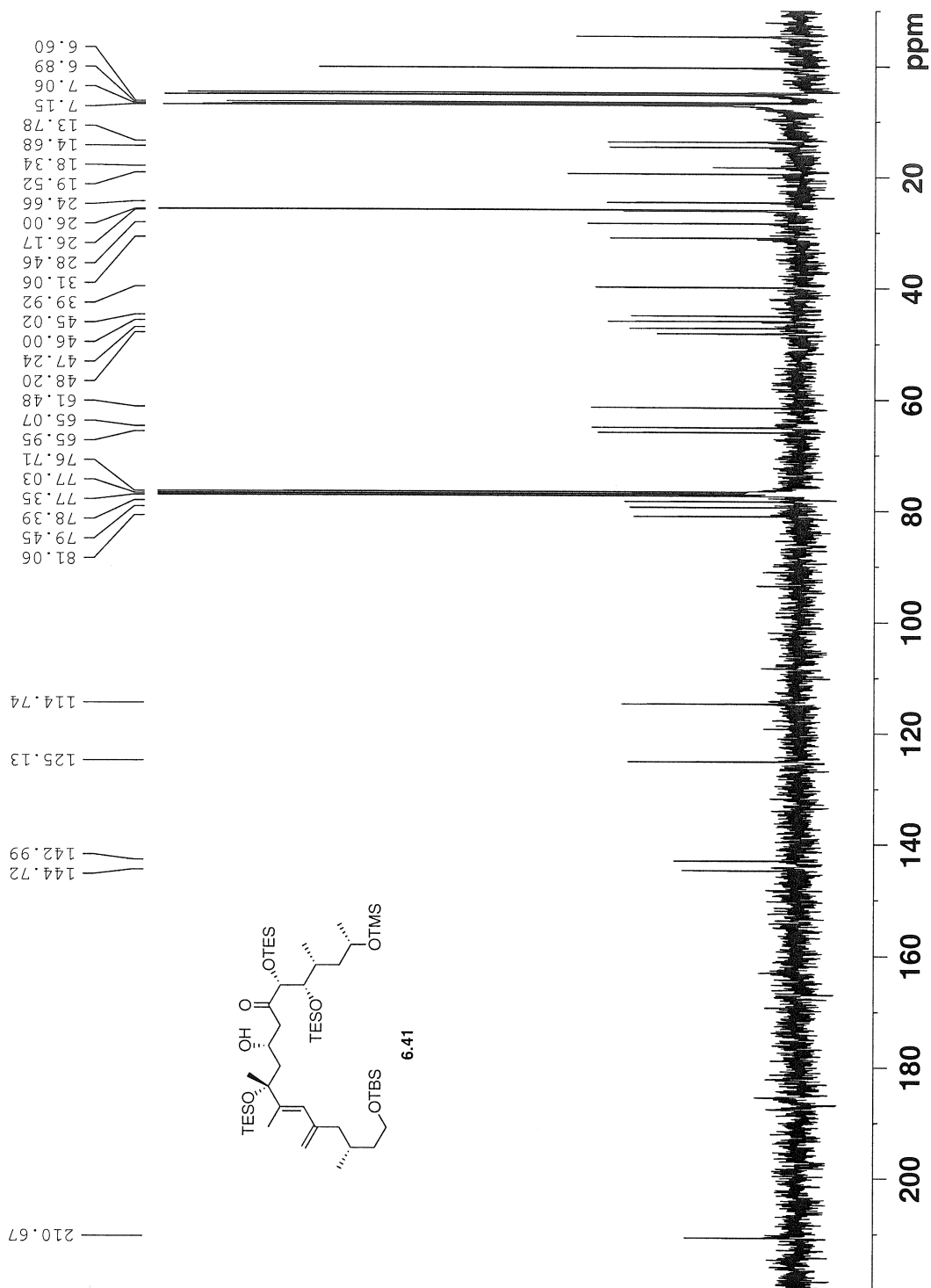
180

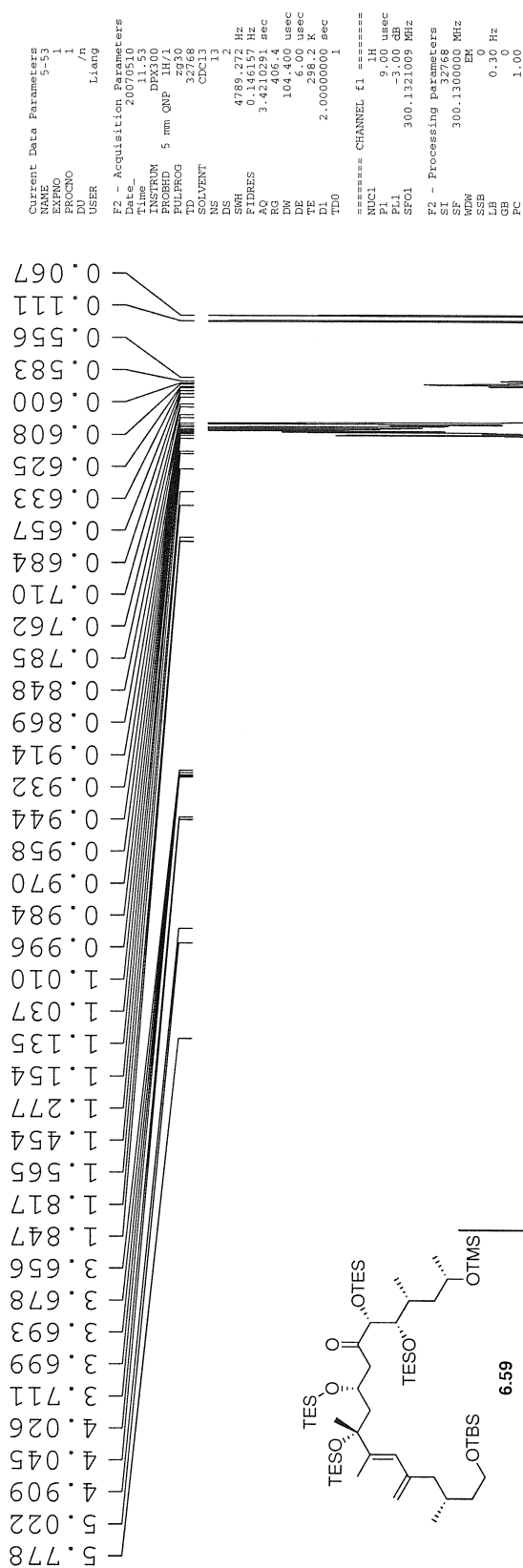
200

328



Current Data Parameters  
NAME 5-50  
EXPNO 3  
PROCNO 3  
USER Liang  
F2 - Acquisition Parameters  
Date\_ 20070508  
Time 20:38  
PULPROG zgpg30  
PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
TD 2048  
SOLVENT  
NS 8  
DS 2  
SWH 4006.410 Hz  
FIDRES 1.956255 Hz  
AQ 0.2556404 sec  
RG 322.5  
DE 124.000 usec  
TE 299.2 K  
d0 0.00000300 sec  
d1 2.00000000 sec  
d13 0.0000400 sec  
d15 0.0000000 sec  
d16 0.0000000 sec  
d17 0.00024960 sec  
===== CHANNEL f1 =====  
NUC1 1H  
P1 7.10 usec  
PL1 0.00 dB  
SFO1 400.0120000 MHz  
===== GRADIENT CHANNEL =====  
GUNIT 100  
GENAM2 SINE 100  
GP21 10.00 %  
GP22 10.00 %  
P16 1000.00 usec  
F1 - Acquisition parameters  
NUC2 128  
TD 128  
SFO2 400.012 MHz  
FIDRES 31.300079 Hz  
AQ 0.2556404 sec  
DE 124.000 usec  
TE 299.2 K  
===== GRADIENT CHANNEL =====  
GUNIT 100  
GENAM2 SINE 100  
GP21 10.00 %  
GP22 10.00 %  
P16 1000.00 usec  
F2 - Processing parameters  
SI 32768  
SF 100.5825950 MHz  
WDW EM  
SSB 0  
LB 3.00 Hz  
GB 0  
PC 1.40







```

Current Data Parameters
NAME      5-53
EXPNO     3
PROCNO    2
DU        /n
USER      Liang

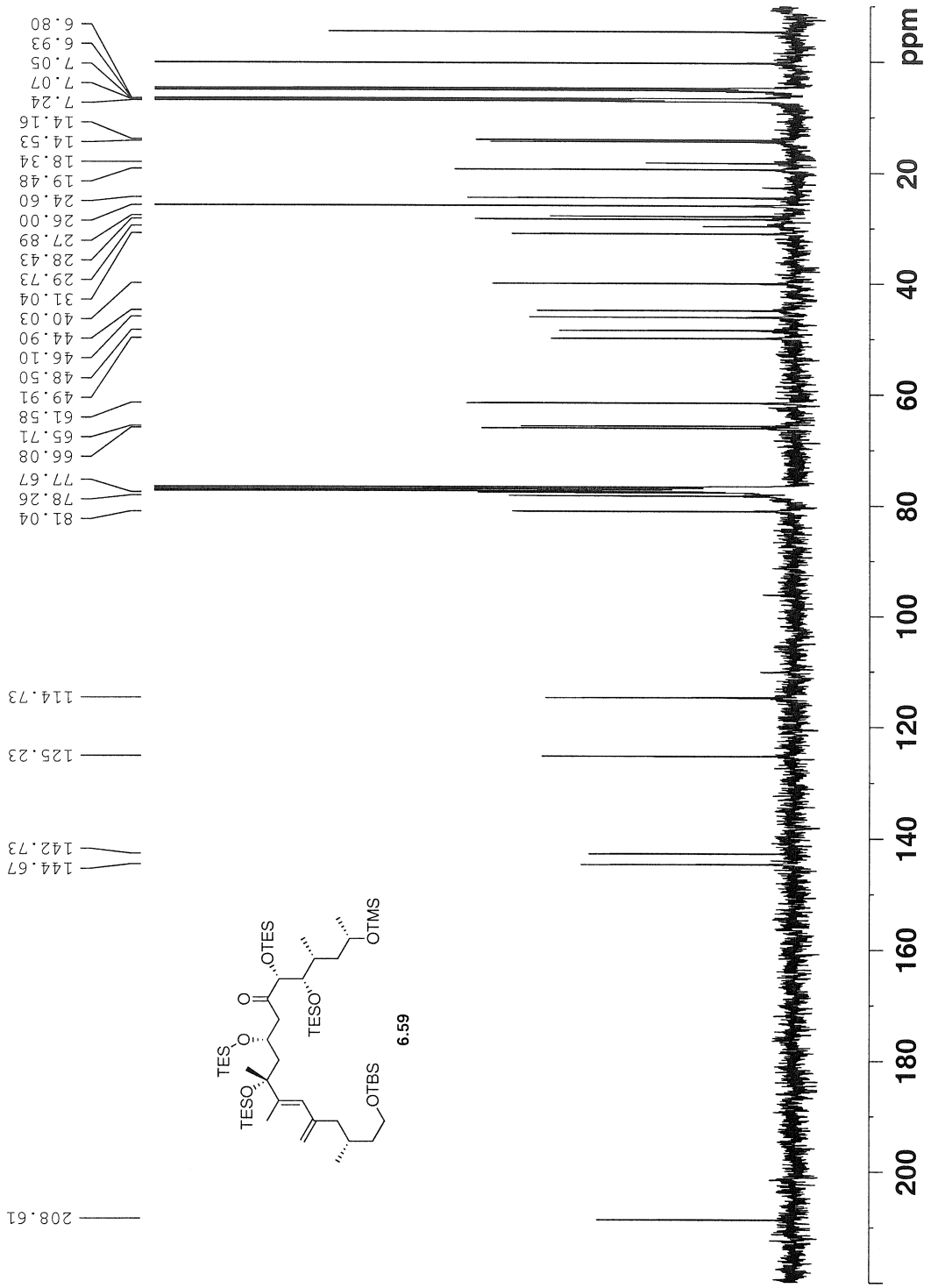
F2 - Acquisition Parameters
Date_     20070510
Time      17.30
INSTRUM   DFX400
PROBHD    5 mm BBO BB-1H
PULPROG   zgpg30
TD         65536
SOLVENT    CDCl3
NS         1994
DS         4
SWH        25125.624 Hz
FIDRES     0.383387 Hz
AQ         1.3042164 sec
RG         4597.6
DW         19.900 us
DE         6.00 us
TE         298.2 K
D1         0.15000001 sec
d11        0.03000000 sec
DELTA     0.05000000 sec
TD0        1

===== CHANNEL f1 =====
NUC1       13C
P1         7.80 us
PL1        -3.00 dB
SFO1       100.5936591 MHz

===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
PCPD2      135.00 us
PL2        17.40 dB
PL12       17.40 dB
PL13       17.40 dB
SFO2       400.0116000 MHz

F2 - Processing parameters
SI          32768
SF         100.5825950 MHz
WDW         EM
SSB         0
LB          0
GB          0
PC         1.40

```

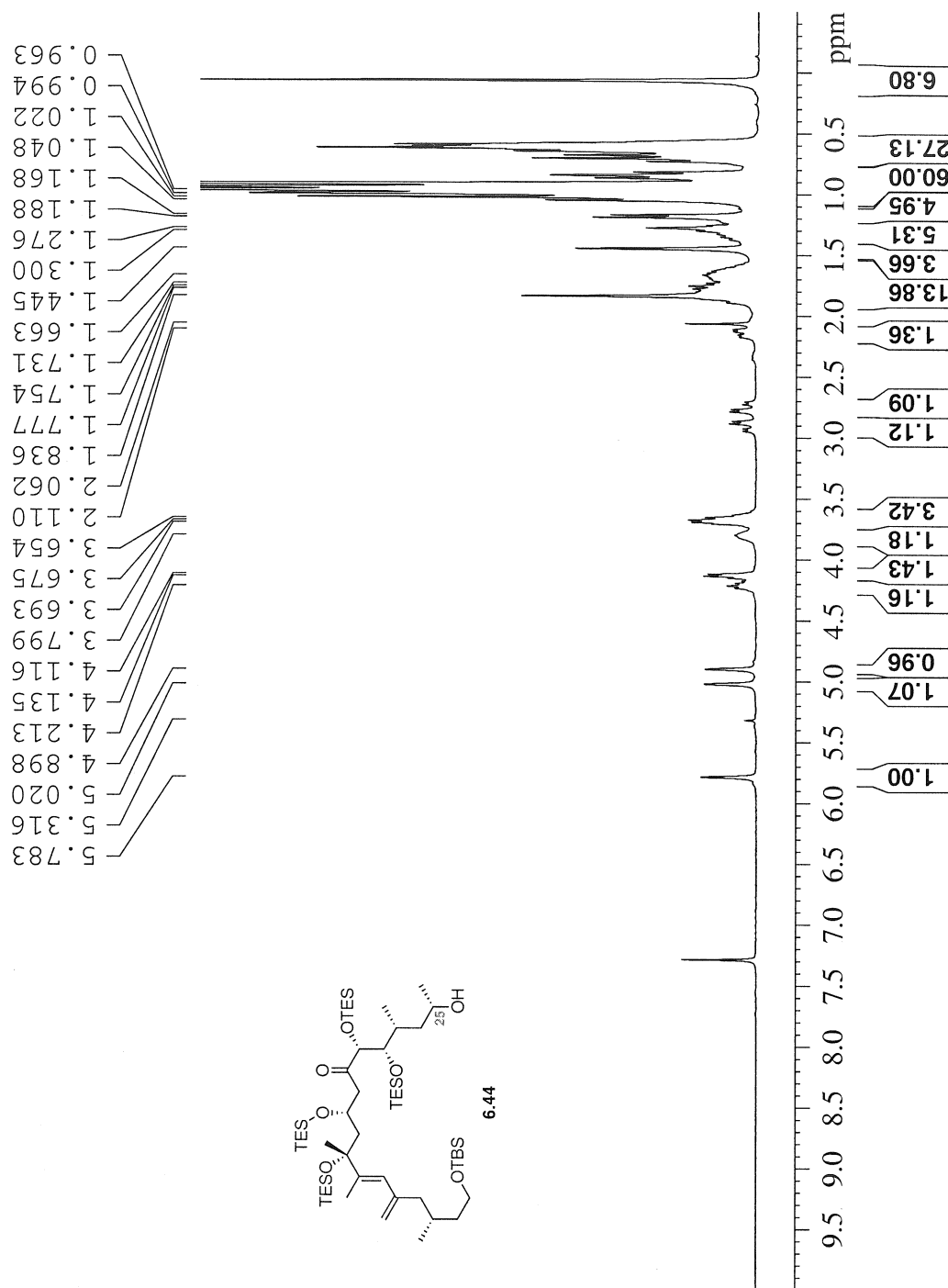


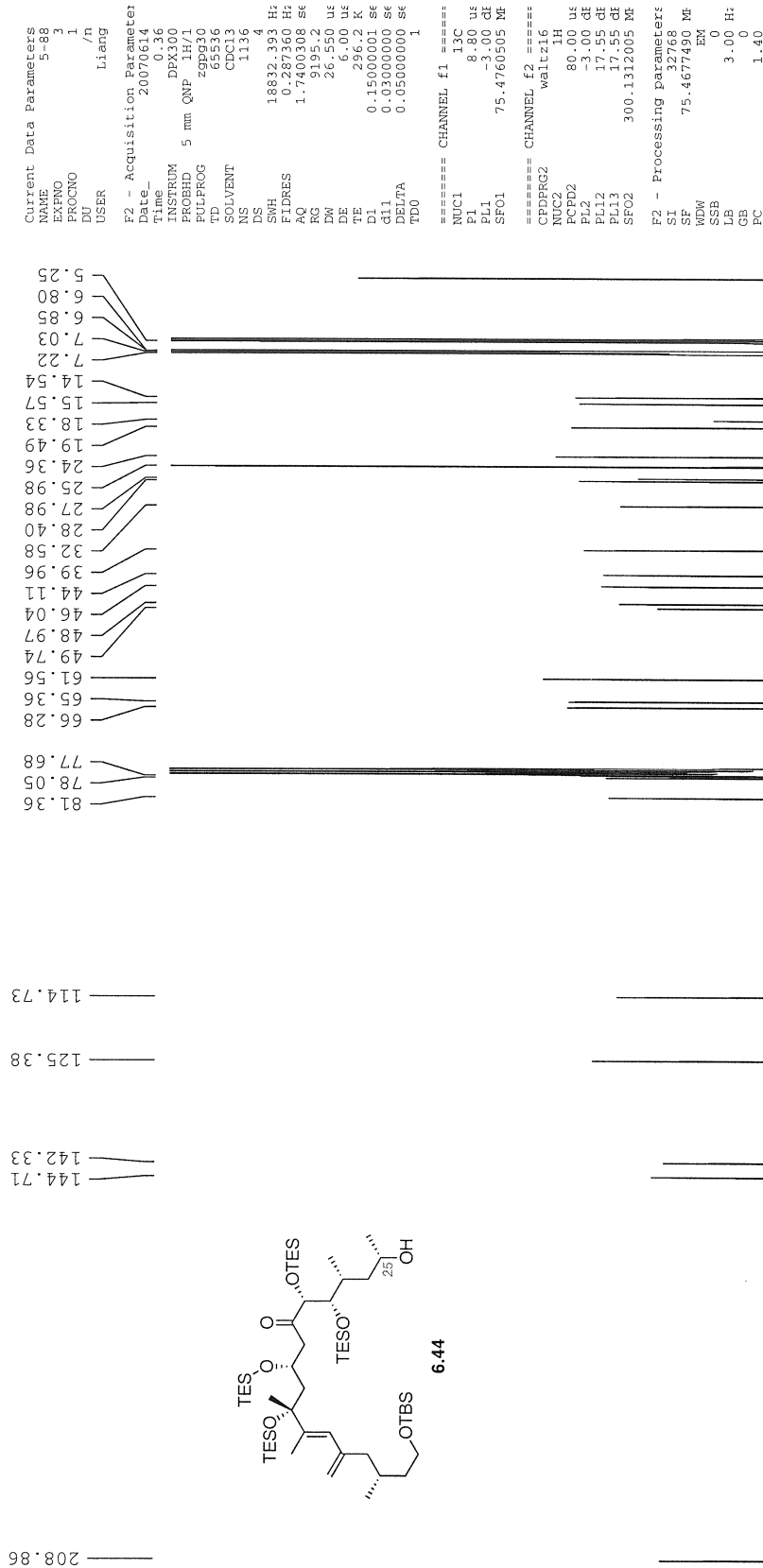
Current Data Parameters  
 NAME 3-88  
 EXPNO 1  
 PROCNO 1  
 DU /n  
 USER Liang

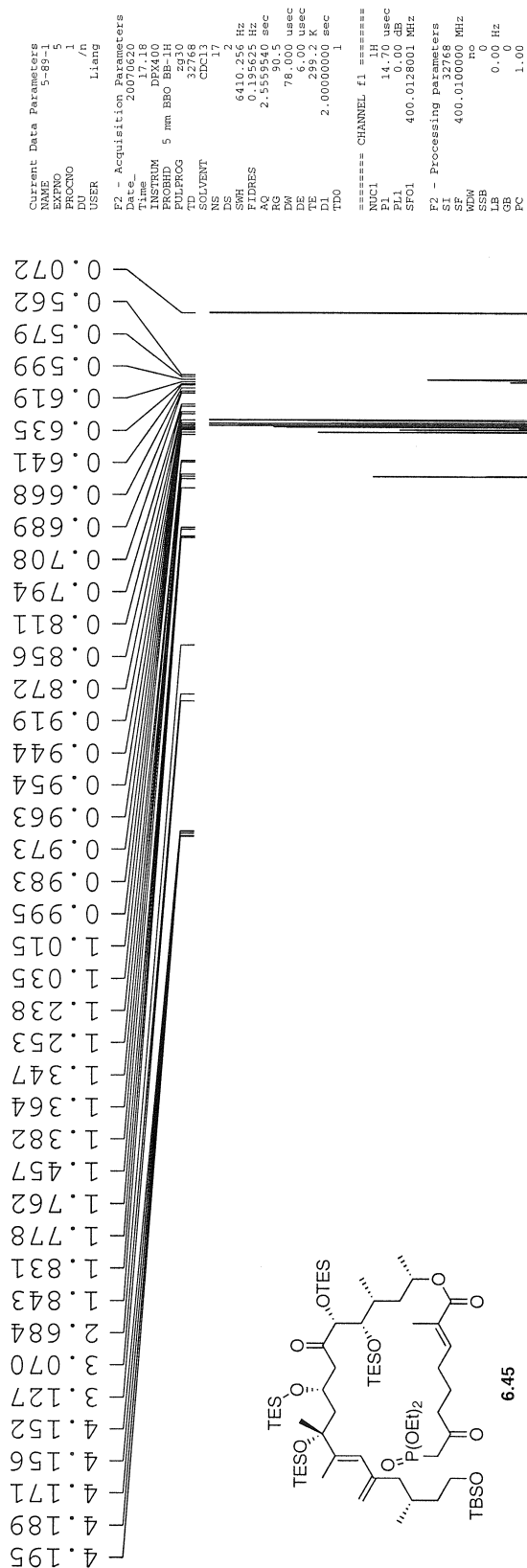
F2 - Acquisition Parameters  
 Date\_ 20070613  
 Time 23:44  
 INSTRUM spect  
 PULPROG zgpg30  
 TD 32768  
 SOLVENT CDCl3  
 NS 32  
 DS 2  
 SWH 4789.272 Hz  
 FIDRES 0.146157 Hz  
 AQ 3.4210291 sec  
 RG 655.36  
 DW 104.400 usec  
 DE 6.00 usec  
 TE 296.2 K  
 D1 2.0000000 sec  
 D10 1

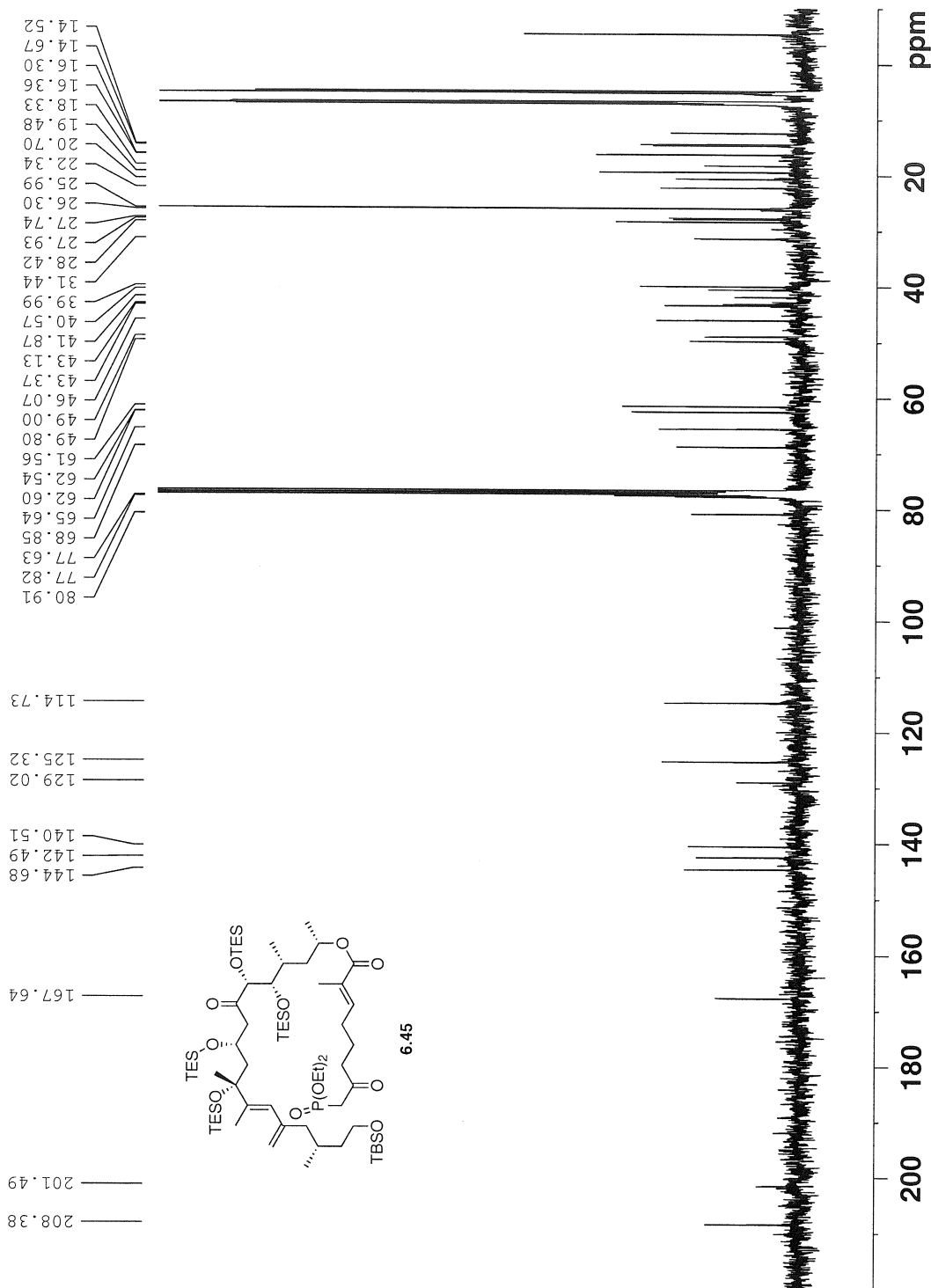
===== CHANNEL f1 =====  
 NUC1 1H  
 P1 9.00 usec  
 PL1 -2.00 dB  
 SFO1 300.1321009 MHz

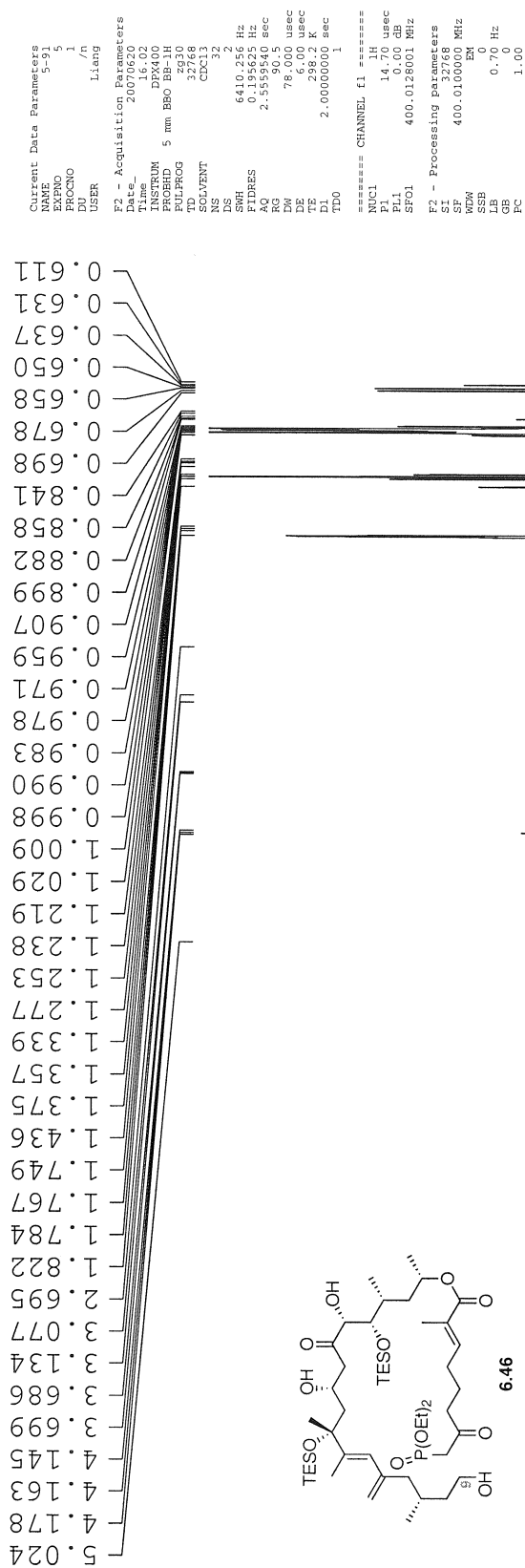
F2 - Processing parameters  
 SI 32768  
 SF 300.1300000 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



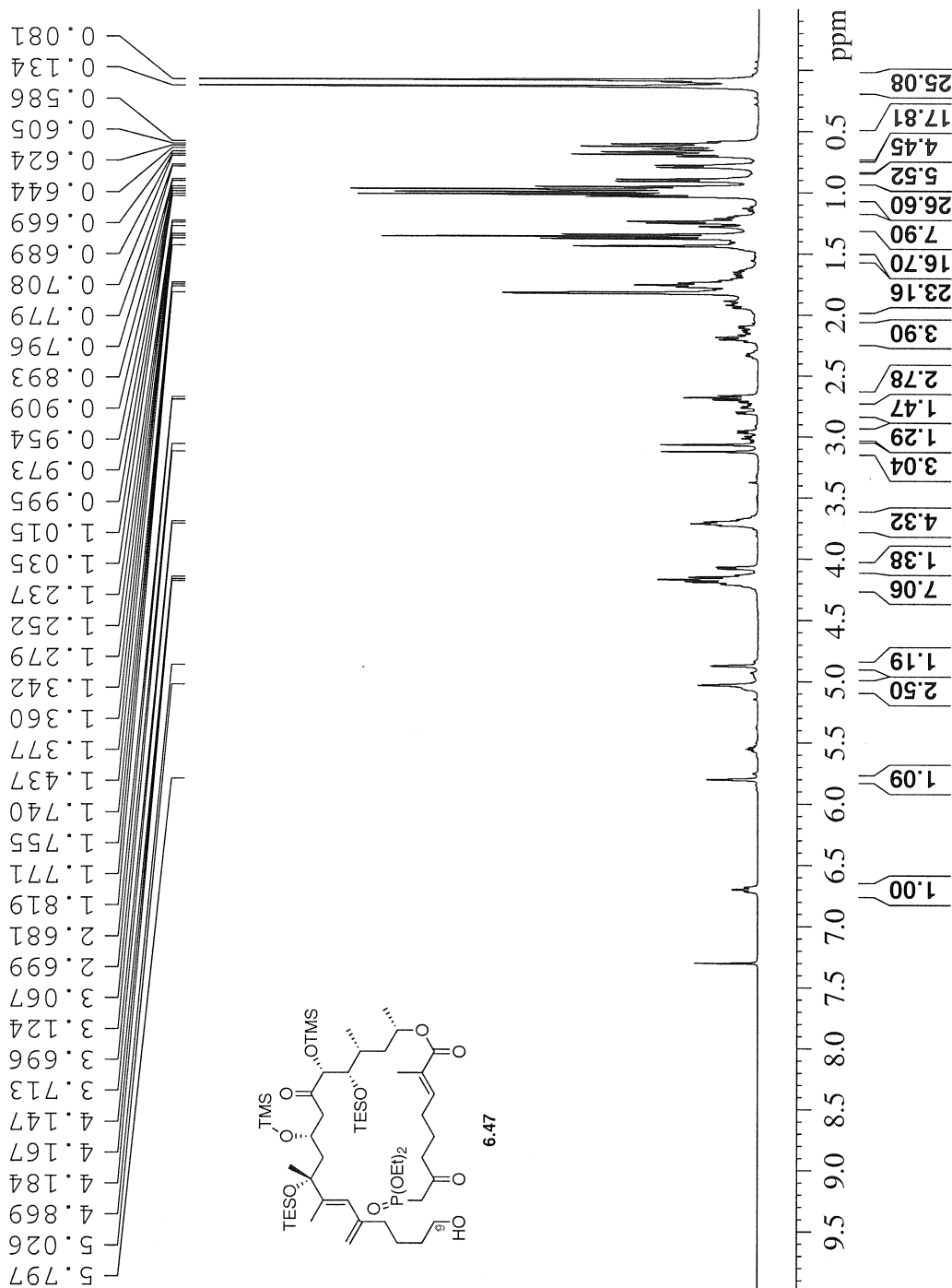




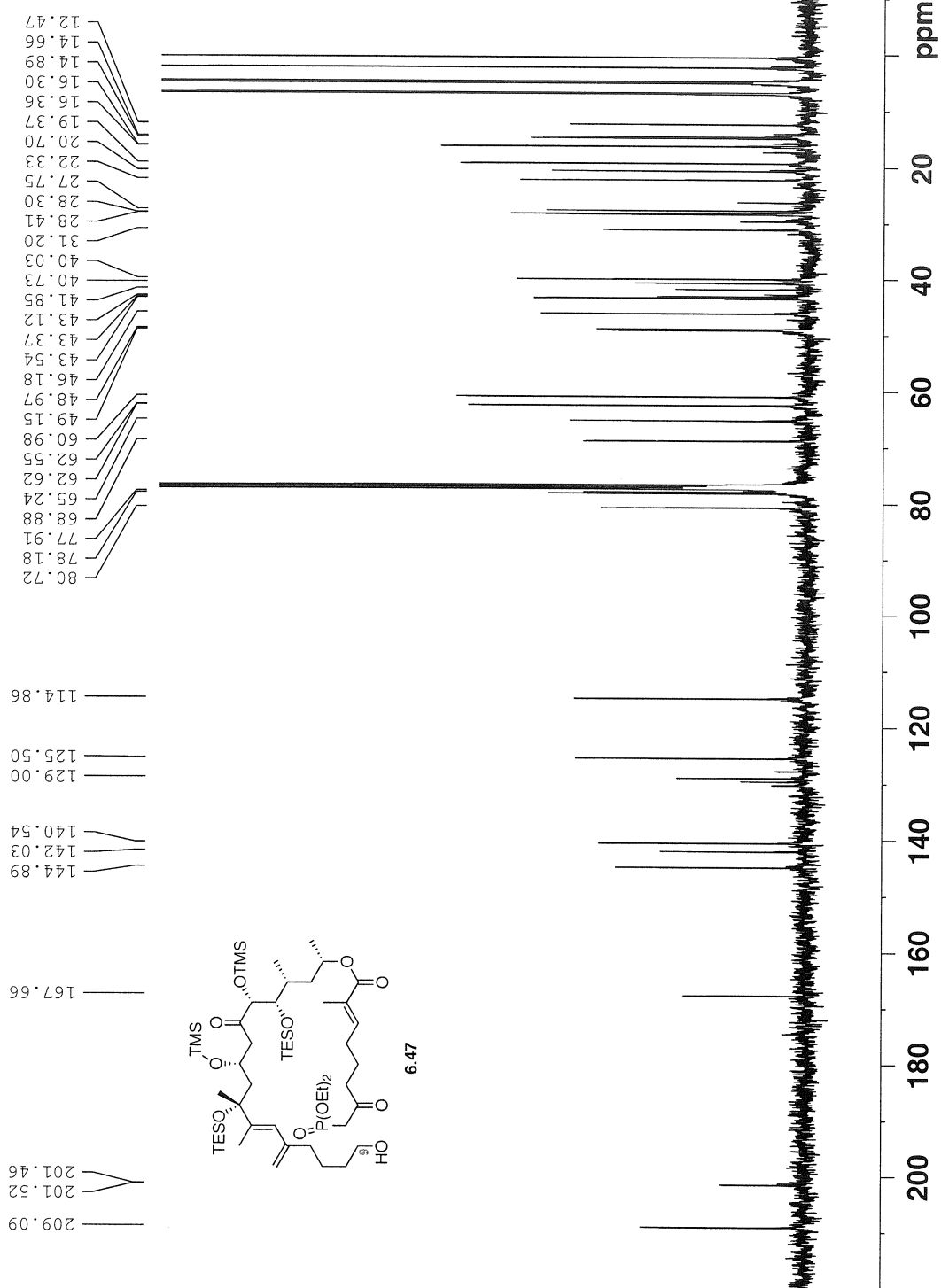


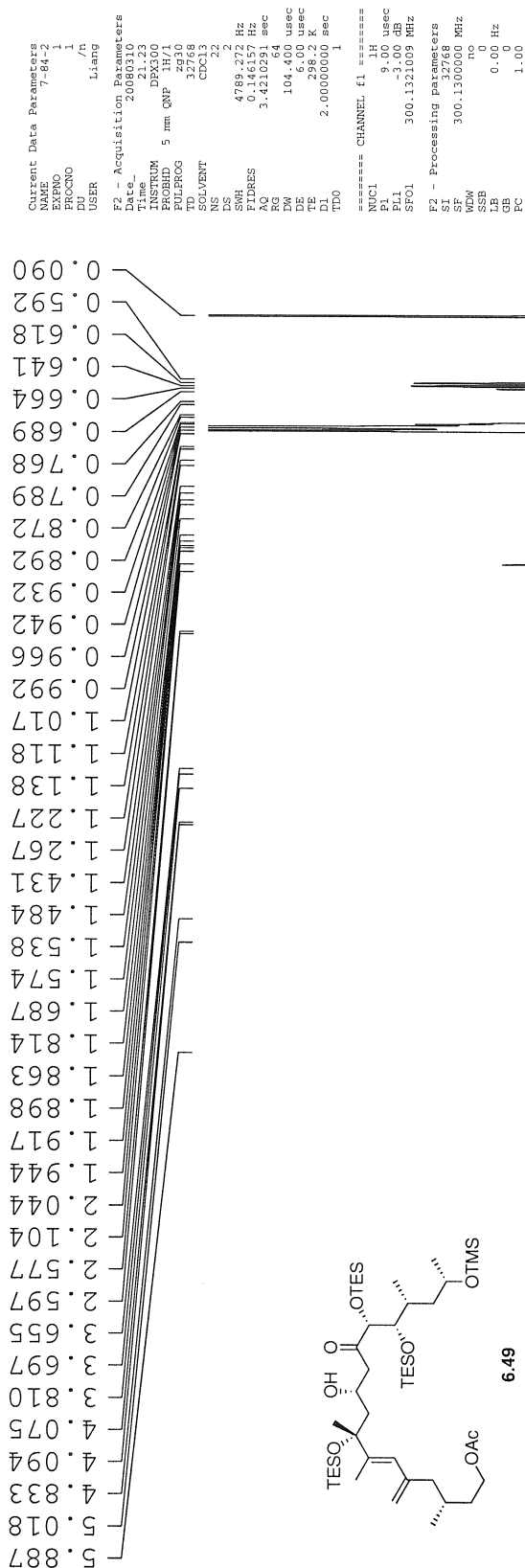




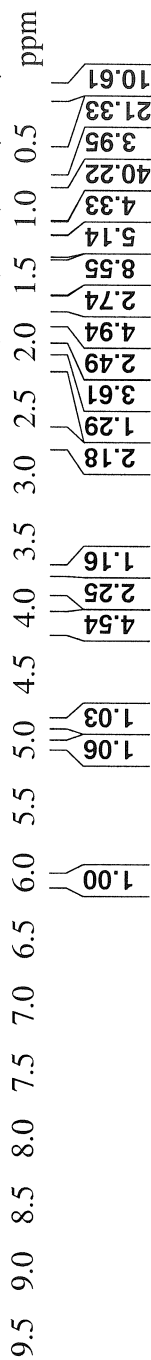
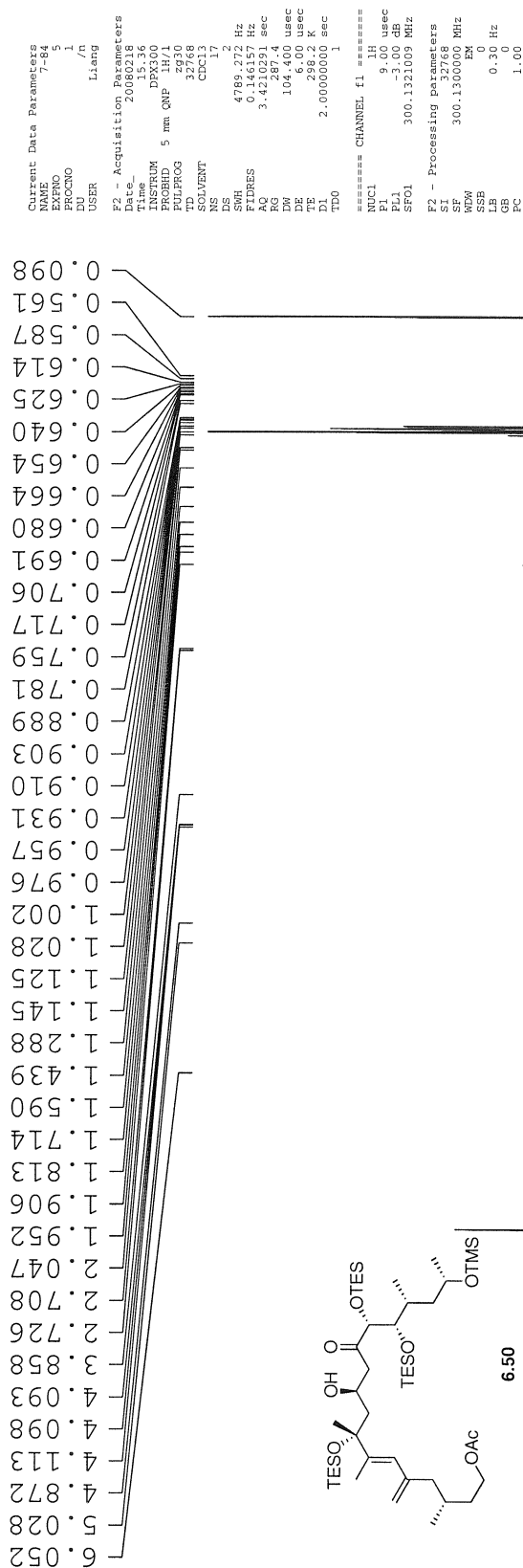


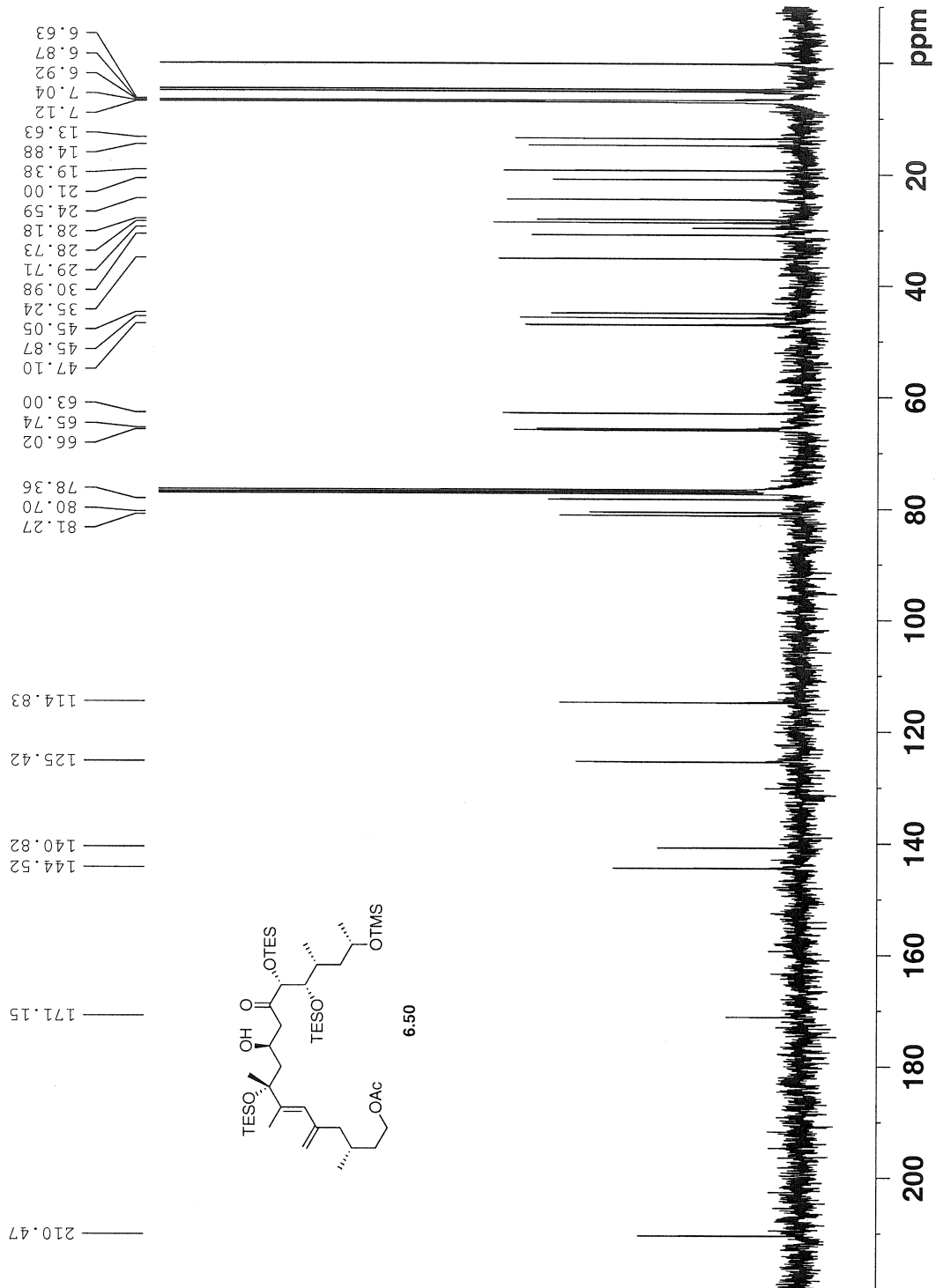












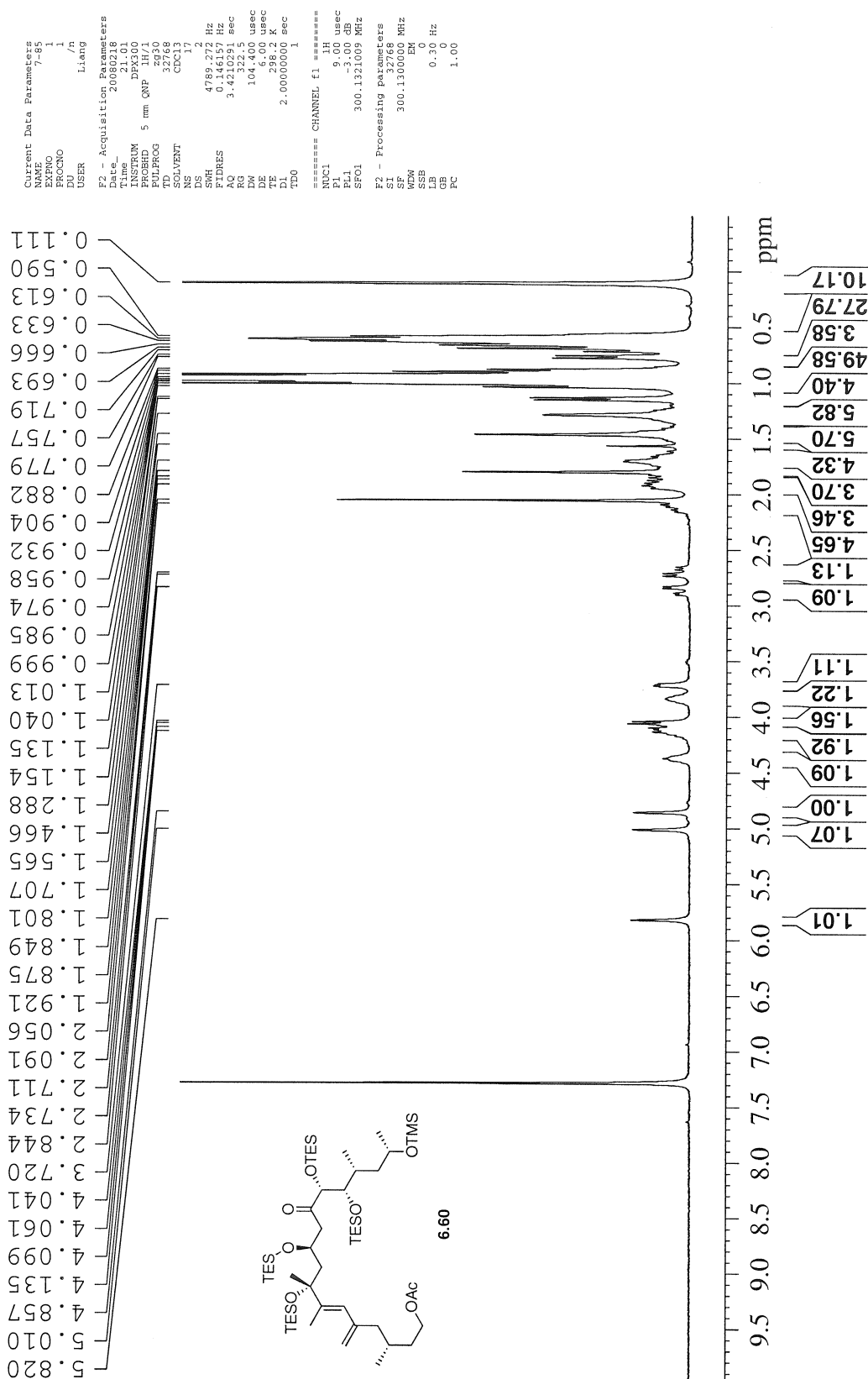
Current Data Parameters  
NAME 6-91  
EXPNO 1  
PROCNO 1  
DU /n  
USER Liang

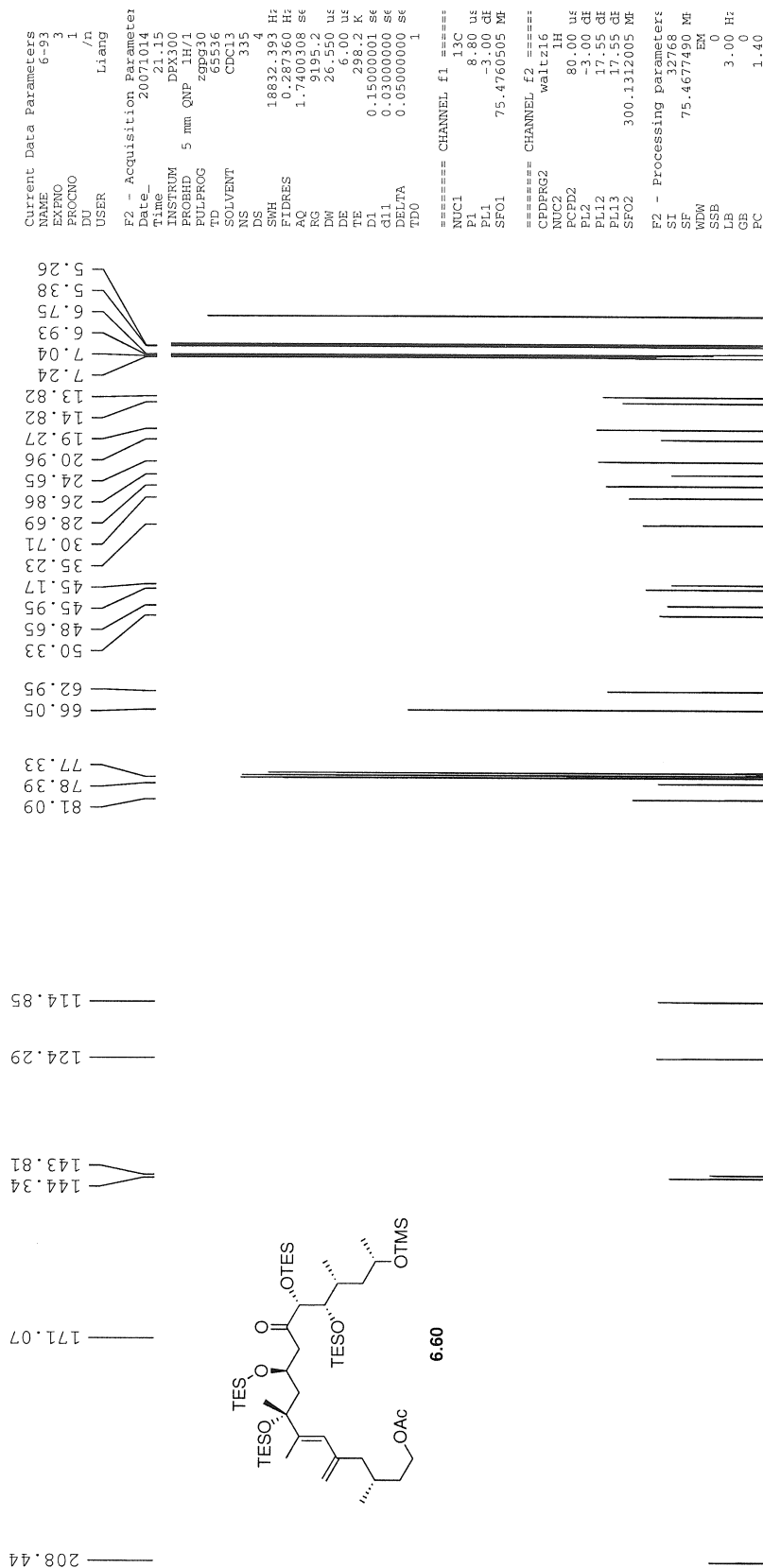
F2 - Acquisition Parameters  
Date\_ 20071011  
Time 18.28  
INSTRUM DFX400  
PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
TD 65536  
SOLVENT  
NS 253  
DS 4  
SWH 25125.695 Hz  
FIDRES 0.383387 Hz  
AQ 1.3042164 s  
RG 4597.6  
DW 19.900 us  
DE 6.00 us  
TE 299.2 K  
D1 0.15000001 s  
d11 0.03000000 s  
DELTA 0.05000000 s  
TD0 1

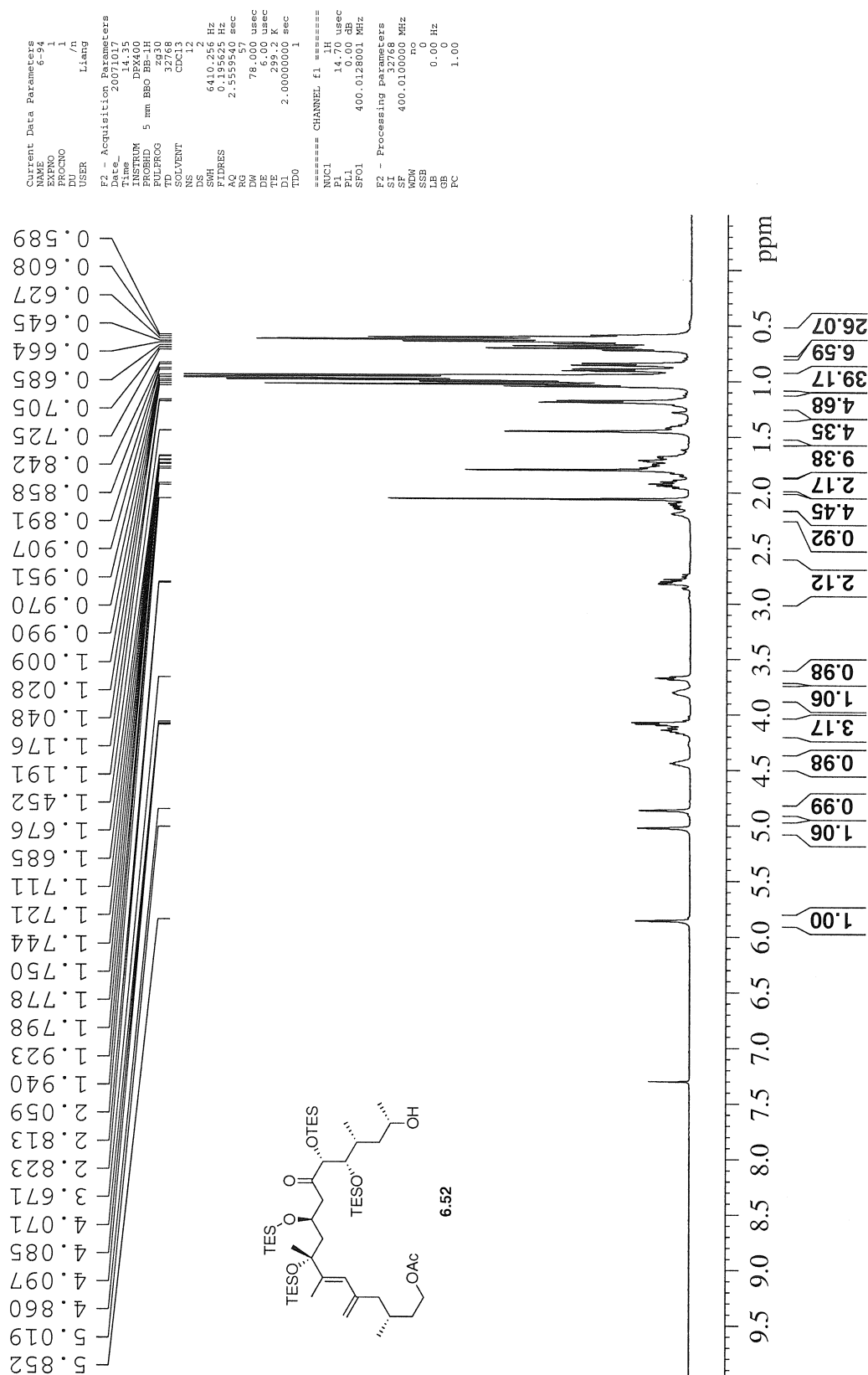
===== CHANNEL f1 =====  
NUC1 13C  
P1 7.80 us  
PL1 -3.00 dB  
SFO1 100.5936591 MHz

===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 135.00 us  
PCPD2 17.40 dB  
PL2 17.40 dB  
PL13 17.40 dB  
SFO2 400.0116000 MHz

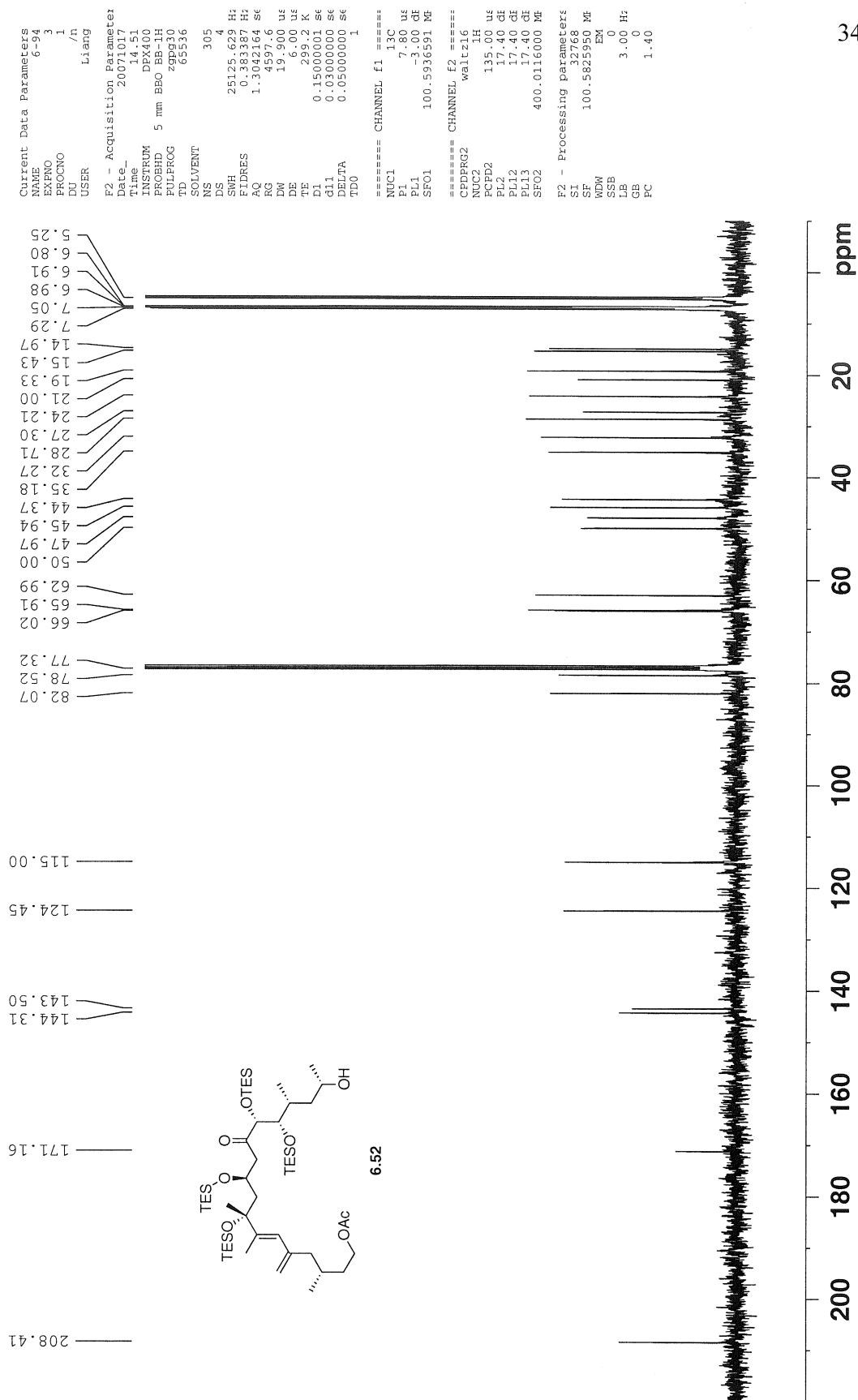
F2 - Processing parameters  
SI 32768  
SF 100.5825950 MHz  
WDW EM  
SSB 0  
LB 3.00 Hz  
GB 0  
PC 1.40

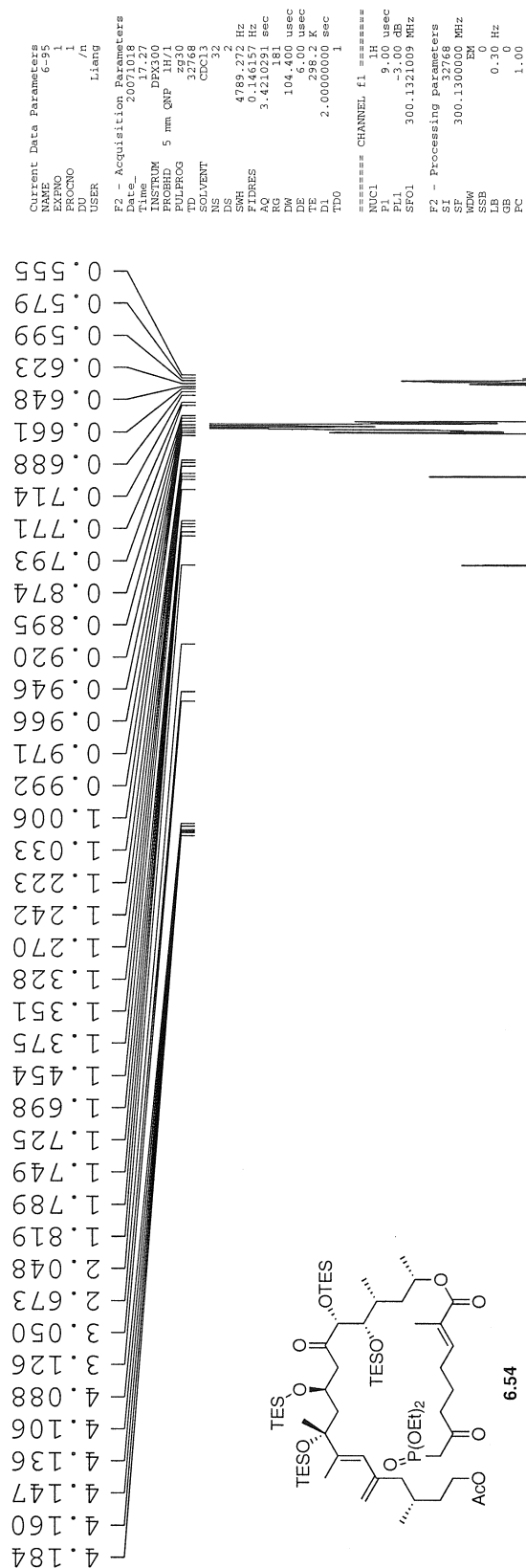


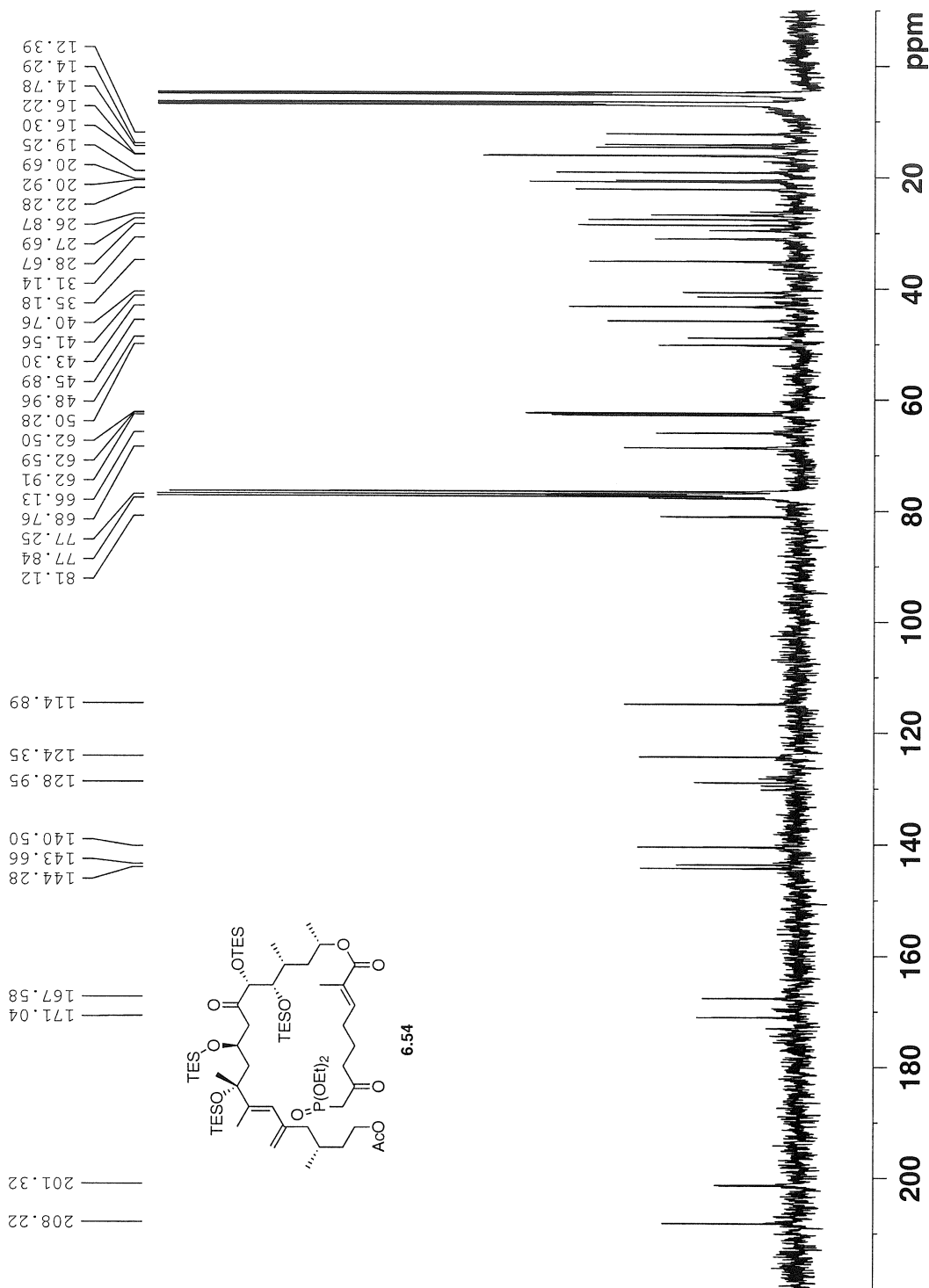


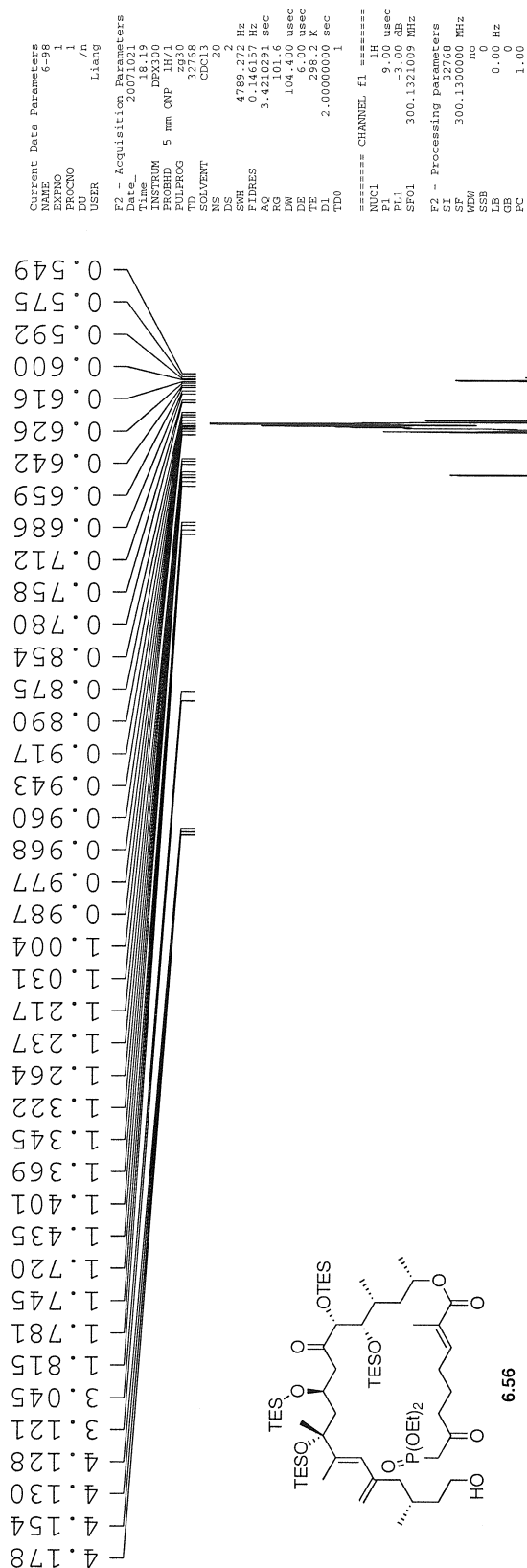


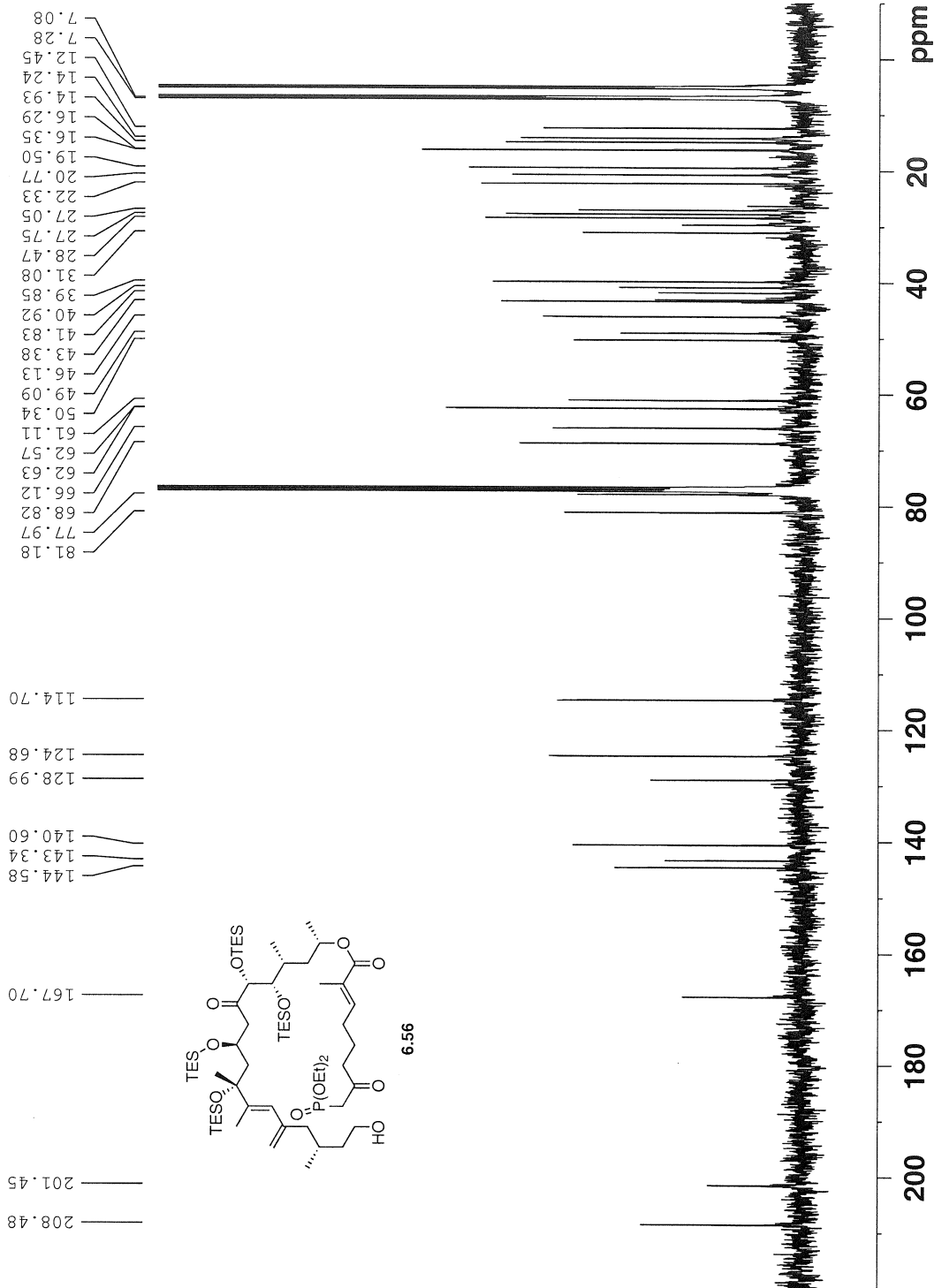


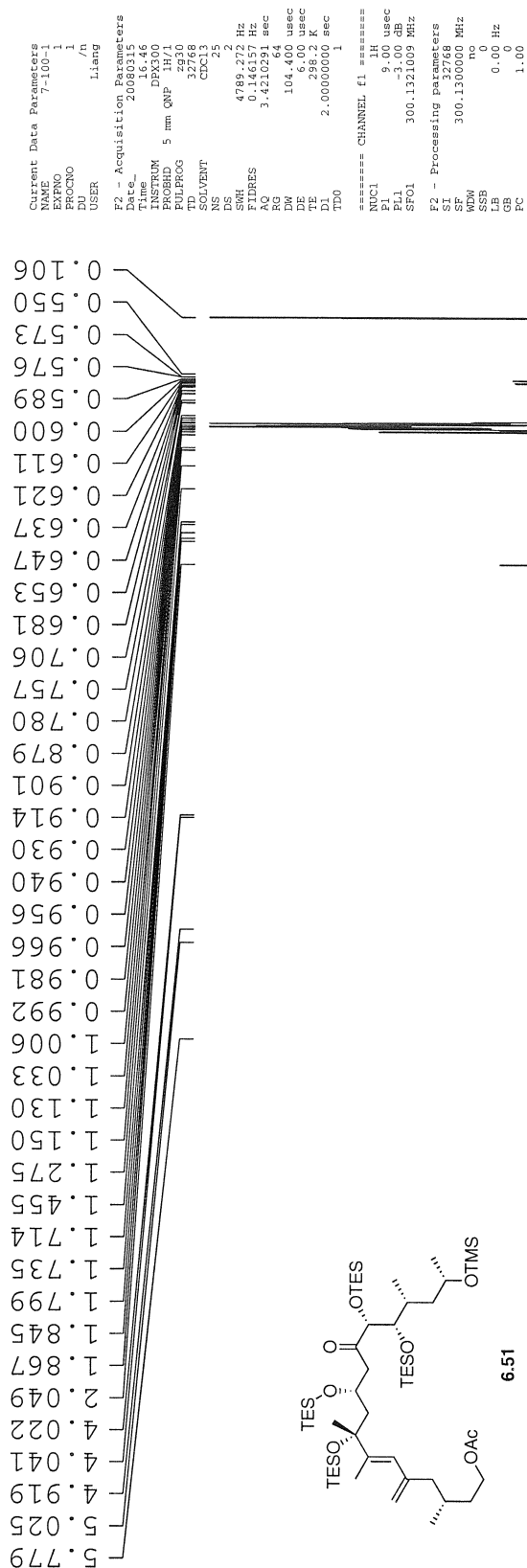












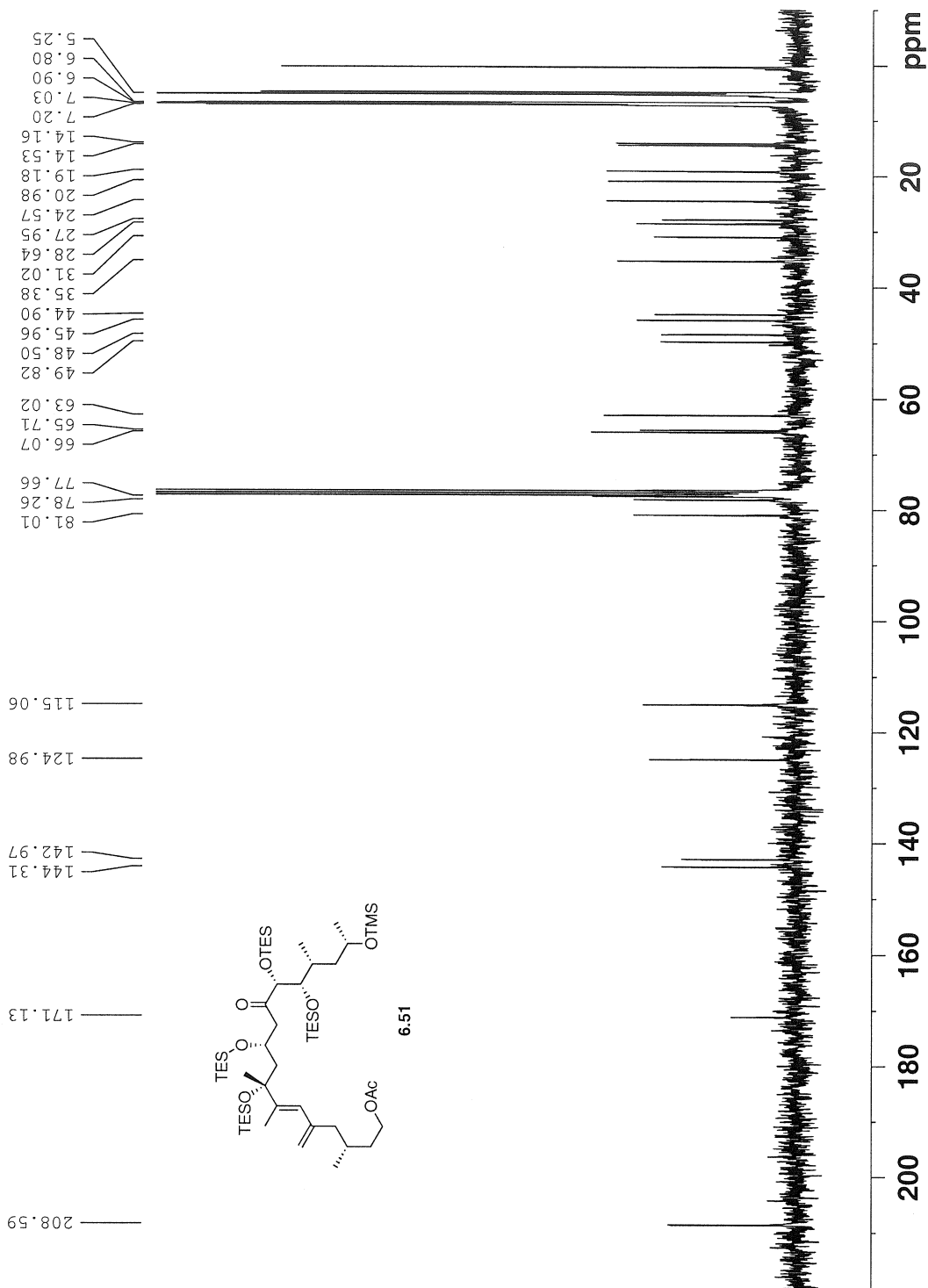
Current Data Parameters  
NAME 7-100-1  
EXPNO 3  
PROCNO 1  
DU /n  
USER Liang

F2 - Acquisition Parameters  
Date\_ 20080315  
Time 17.04  
INSTRUM DPX300  
PROBHD 5 mm QNP 1H/1  
PULPROG zgpg30  
TD 65536  
SOLVENT CDCl3  
NS 513  
DS 4  
SWH 18832.383 Hz  
FIDRES 0.287360 Hz  
AQ 1.7400308 s  
RG 9195.2  
DW 26.550 us  
DE 6.00 us  
TE 298.2 K  
D1 0.15000001 s  
d11 0.03000000 s  
DELTA 0.05000000 s  
TD0 1

===== CHANNEL f1 =====  
NUC1 13C  
P1 8.80 us  
PL1 -3.00 dB  
SFO1 75.4760505 MHz

===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 80.00 us  
PL2 -3.00 dB  
PL12 17.55 dB  
PL13 17.55 dB  
SFO2 300.1312005 MHz

F2 - Processing parameters  
SI 32768  
SF 75.4677490 MHz  
WDW EM  
SSB 0  
LB 3.00 Hz  
GB 0  
PC 1.40



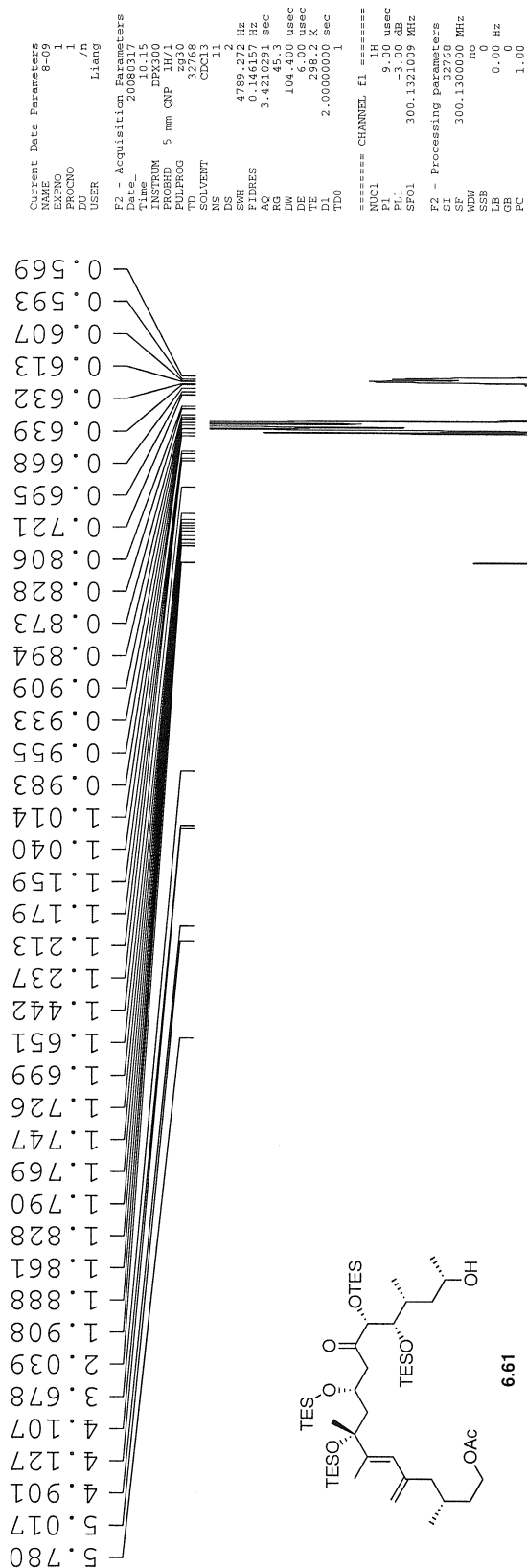
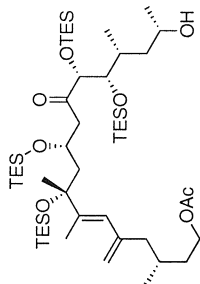




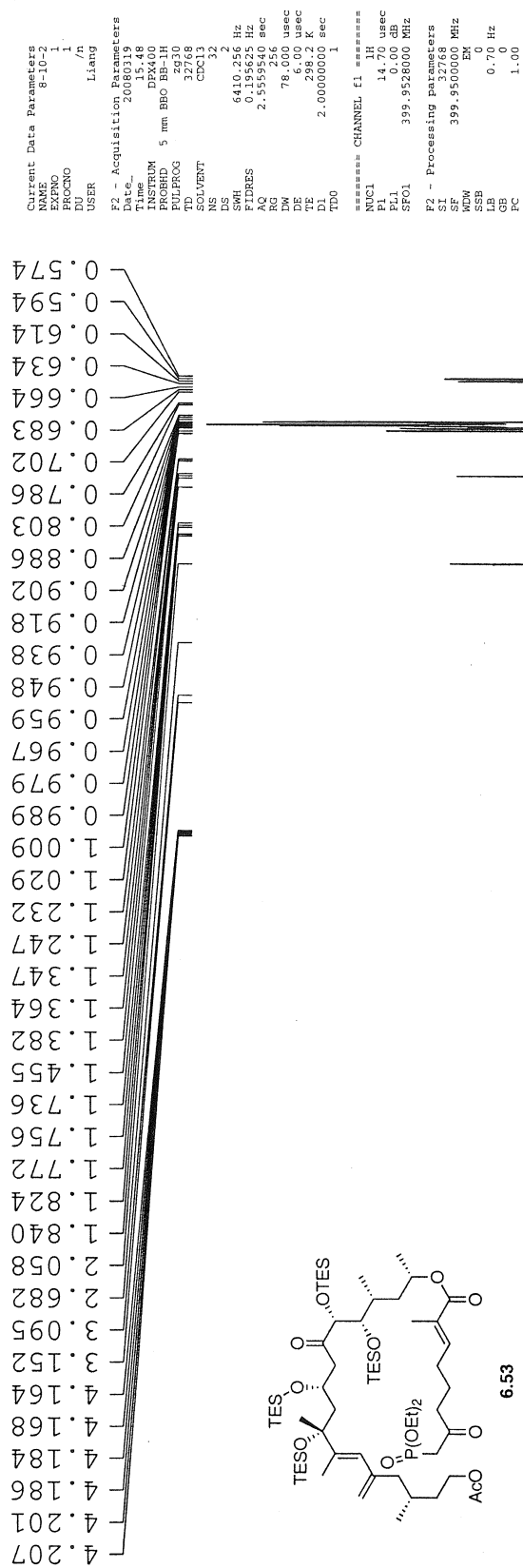
Figure 1 is a line graph illustrating the evolution of the average number of nodes per cluster over 100 iterations for various values of  $\alpha$ . The x-axis represents the iteration number (0 to 100), and the y-axis represents the average number of nodes per cluster (0 to 208.82). The graph shows that as  $\alpha$  increases, the average number of nodes per cluster generally increases and stabilizes at higher values. For example, the line for  $\alpha = 0.08$  stabilizes around 171.15, while the line for  $\alpha = 0.82$  stabilizes around 208.82.

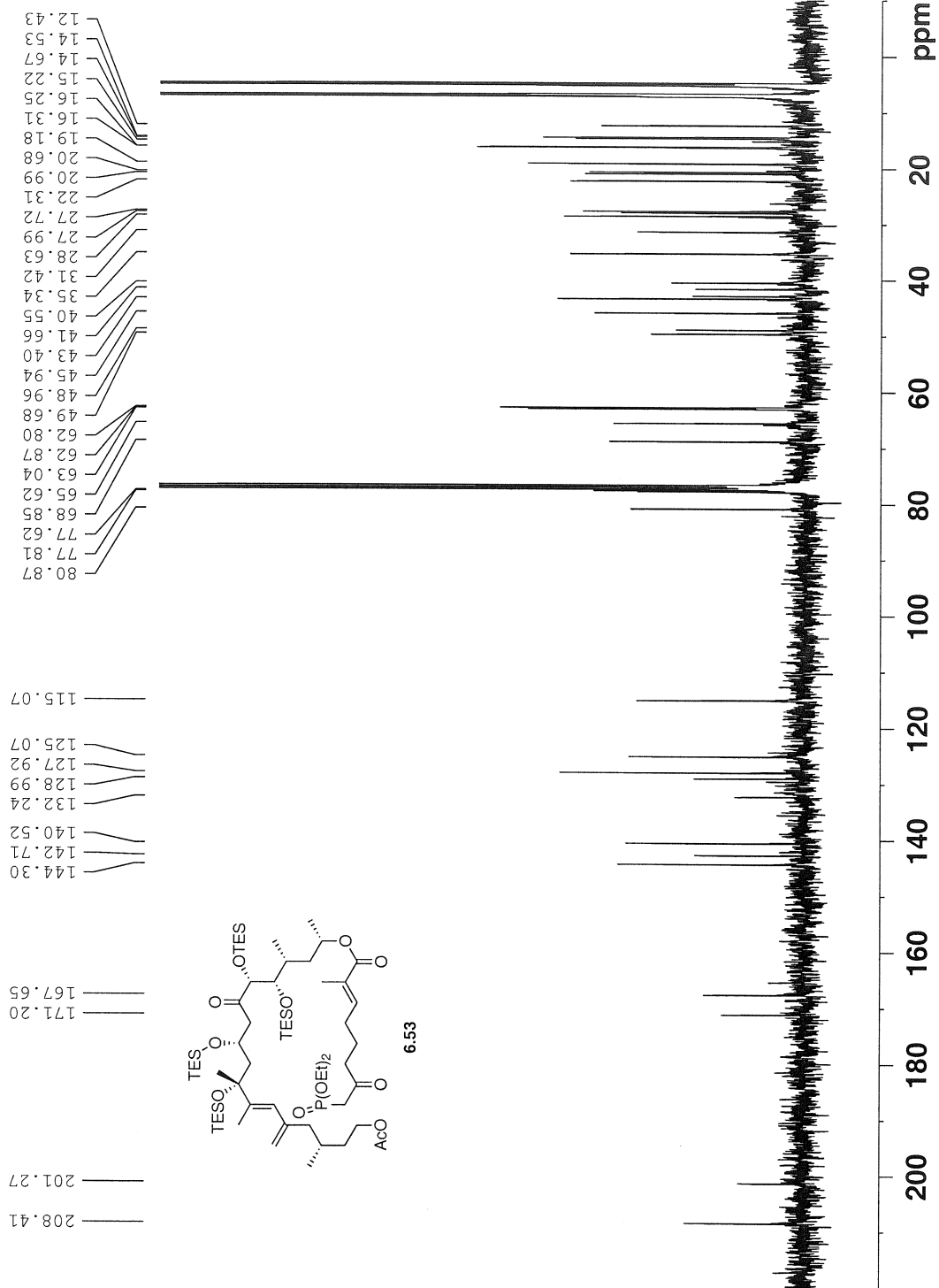
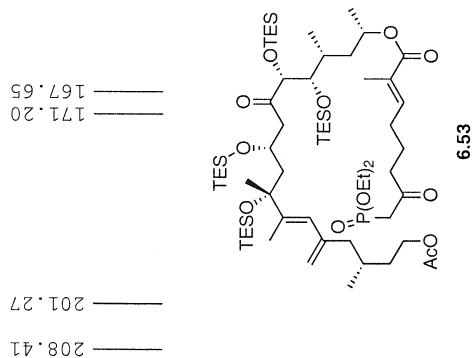
Iteration	$\alpha = 0.08$	$\alpha = 0.16$	$\alpha = 0.37$	$\alpha = 0.70$	$\alpha = 0.82$
0	171.15	144.39	142.60	125.16	115.07
10	171.15	144.39	142.60	125.16	115.07
20	171.15	144.39	142.60	125.16	115.07
30	171.15	144.39	142.60	125.16	115.07
40	171.15	144.39	142.60	125.16	115.07
50	171.15	144.39	142.60	125.16	115.07
60	171.15	144.39	142.60	125.16	115.07
70	171.15	144.39	142.60	125.16	115.07
80	171.15	144.39	142.60	125.16	115.07
90	171.15	144.39	142.60	125.16	115.07
100	171.15	144.39	142.60	125.16	115.07



## 6.61







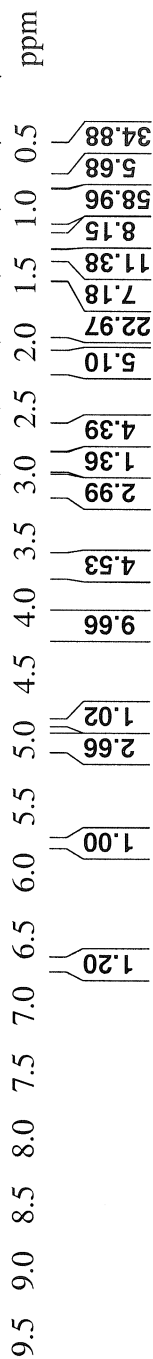
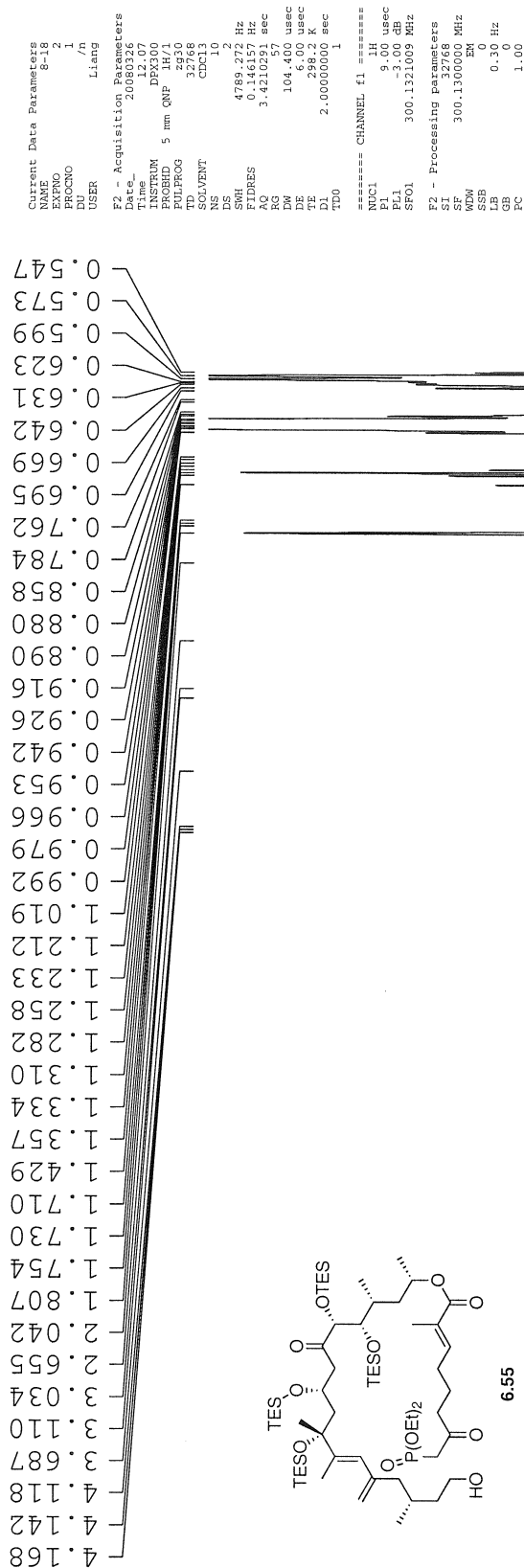
Current Data Parameters  
NAME 8-10-2  
EXPNO 4  
PROCNO 1  
DU /n  
USER Liang

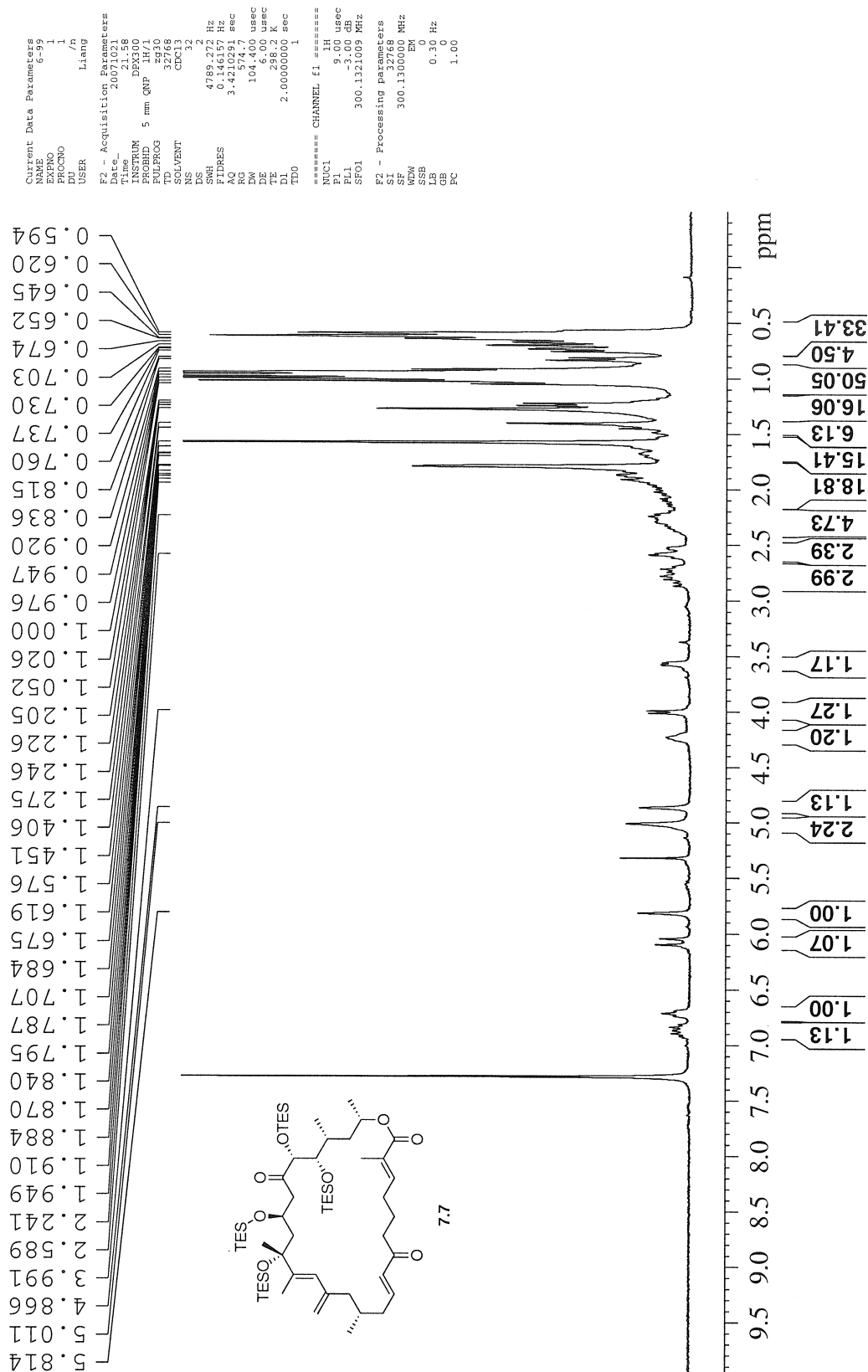
F2 - Acquisition Parameters  
Date\_ 20080319  
Time 18.56  
INSTRUM DFX400  
PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
TD 65536  
SOLVENT  
NS 710  
DS 4  
SWH 25195.629 Hz  
FIDRES 0.383287 Hz  
AQ 1.3042164 sec  
RG 16384  
DE 19.900 us  
TE 298.2 K  
D1 0.15000001 sec  
D11 0.03000000 sec  
DELTA 0.05000000 sec  
TD0 1

===== CHANNEL f1 =====  
NUC1 13C  
P1 7.80 us  
PL1 -3.00 dB  
SFO1 100.5785700 MHz

===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 135.00 us  
PL2 17.40 dB  
PL12 17.40 dB  
PL13 17.40 dB  
SFO2 399.9516000 MHz

F2 - Processing parameters  
SI 32768  
SF 100.5675080 MHz  
WDW EM  
SSB 0  
LB 3.00 Hz  
GB 0  
PC 1.40

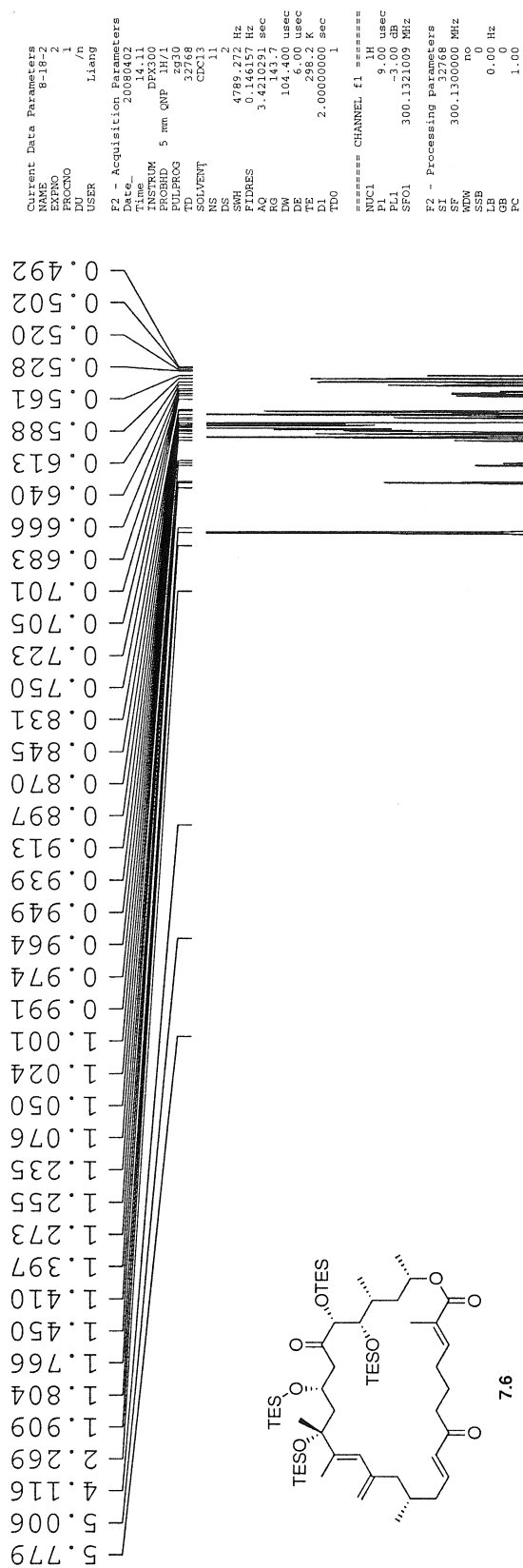


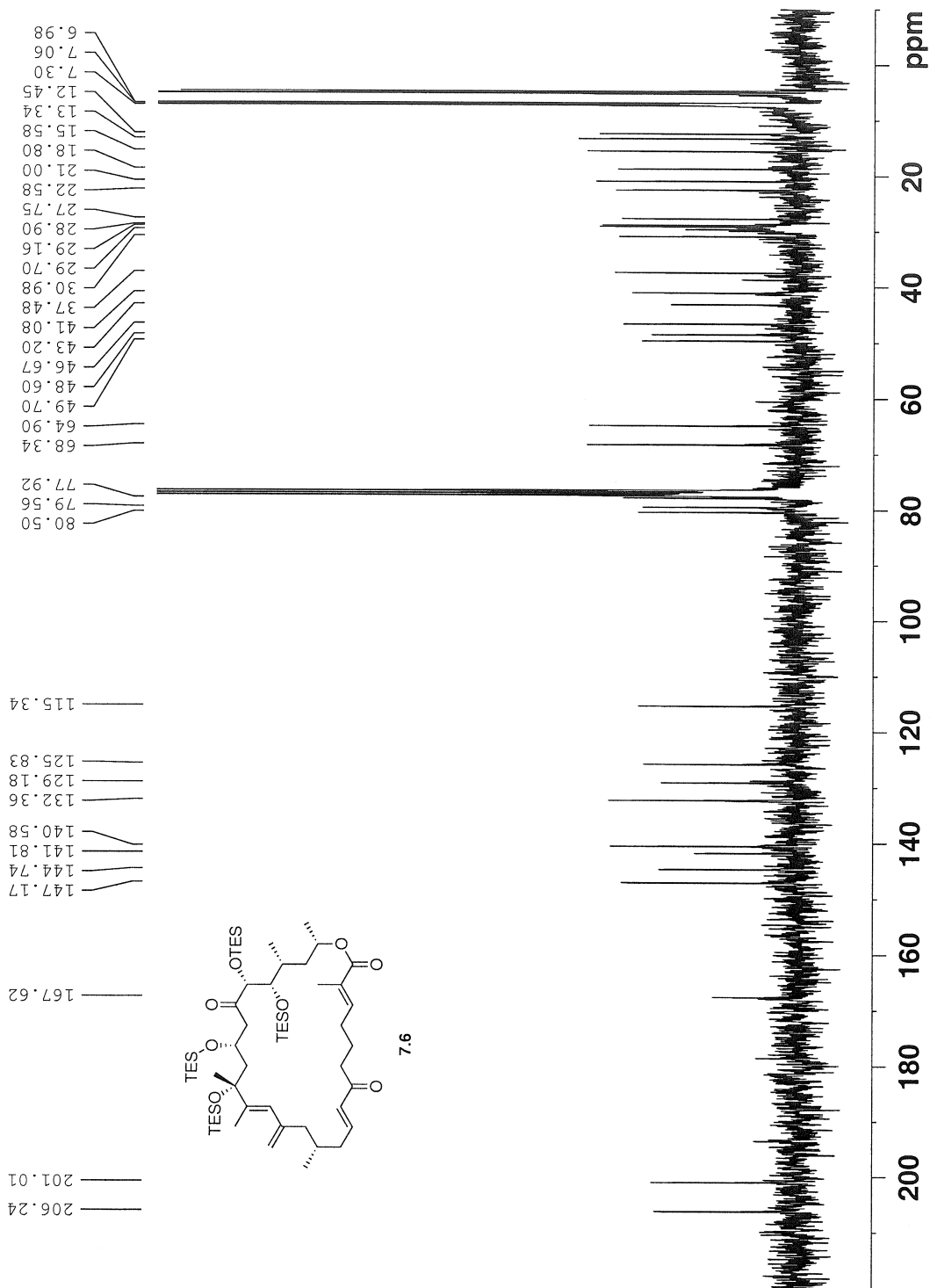


Chemical structure of compound 7.7 is shown in the bottom left. The structure is a macrocyclic ketone with a long chain containing two double bonds, a ketone, and a TES-protected alcohol. The macrocycle is substituted with a TES-protected alcohol and a TES-protected ketone.

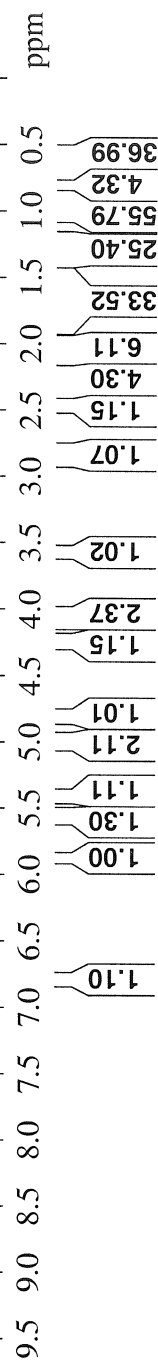
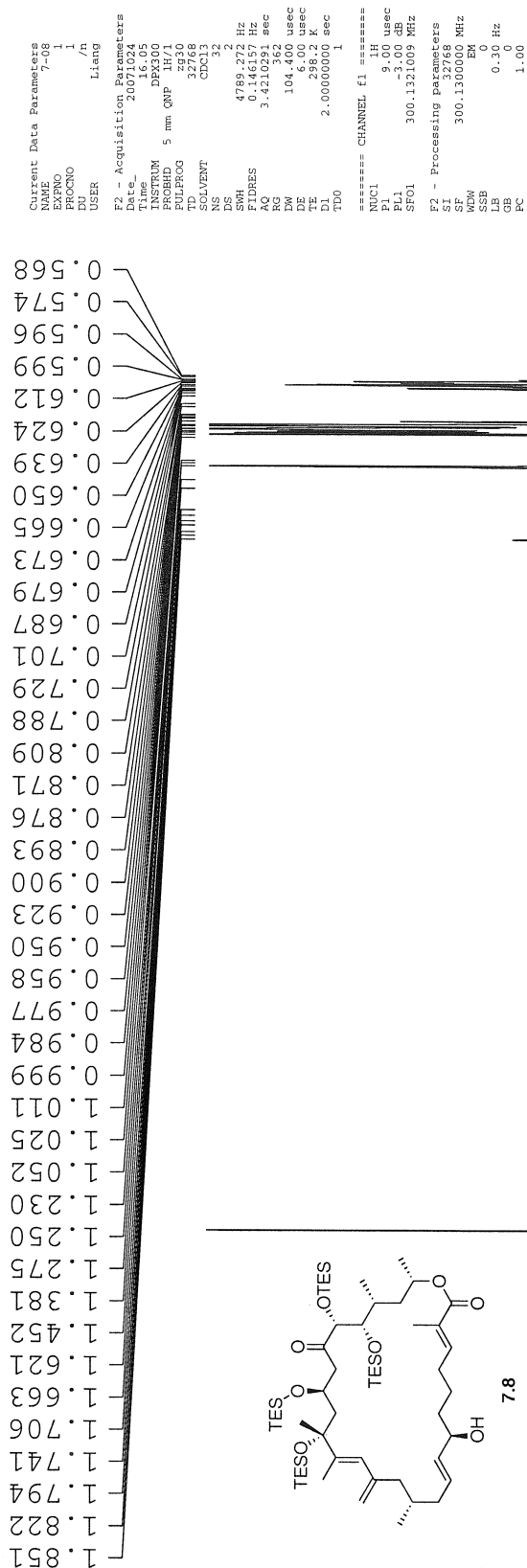
**13C NMR Spectrum Data (ppm):**

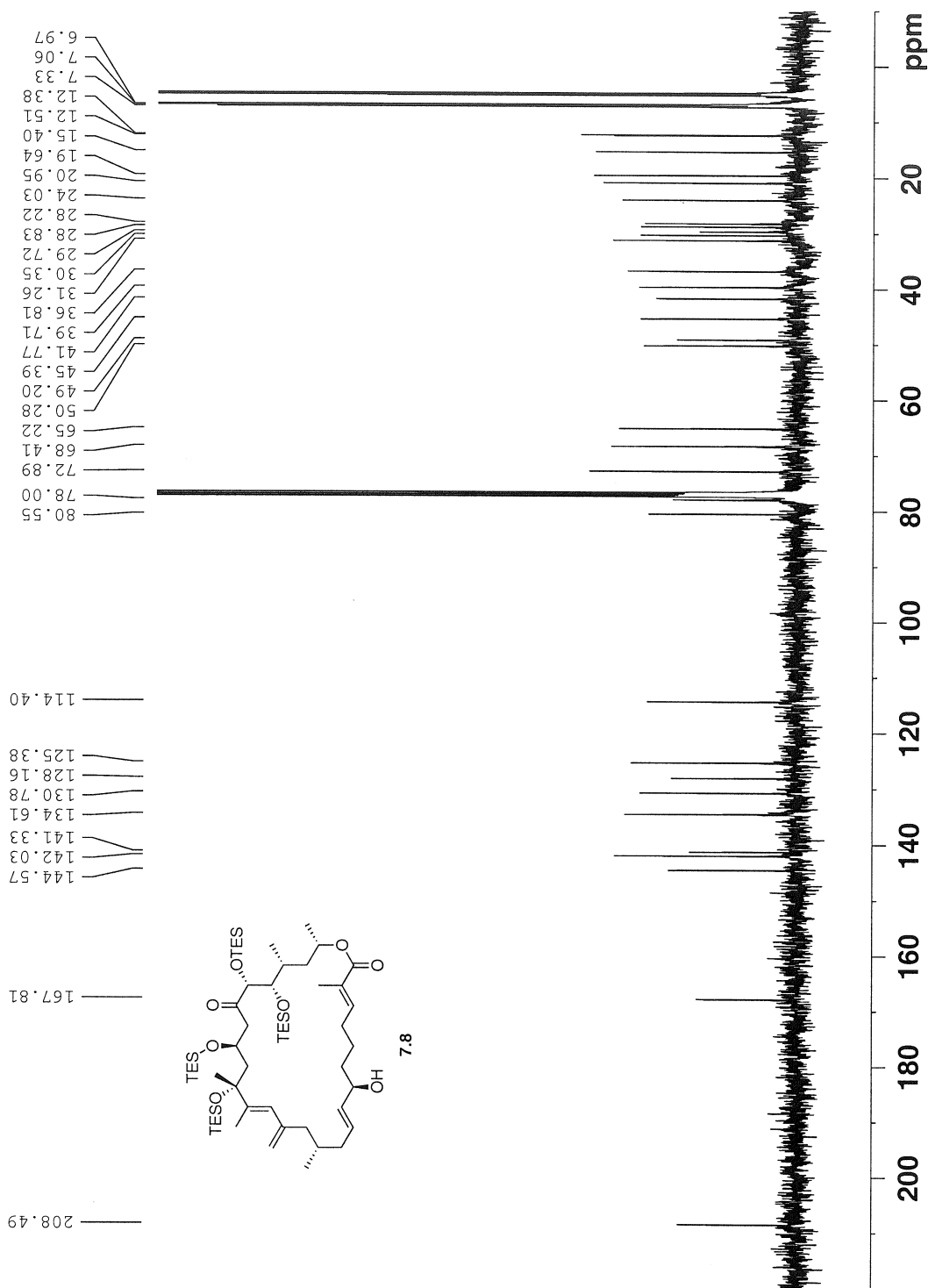
Chemical Shift (ppm)
208.11
200.98
167.61
147.32
144.03
142.81
140.77
132.36
129.29
124.66
115.11
80.56
77.38
77.35
77.04
76.72
68.49
65.14
50.56
48.96
46.09
41.72
40.19
37.15
31.12
30.76
27.77
27.39
23.13
20.99
19.66
15.32
12.81
12.48
7.29
7.07
7.04
6.92

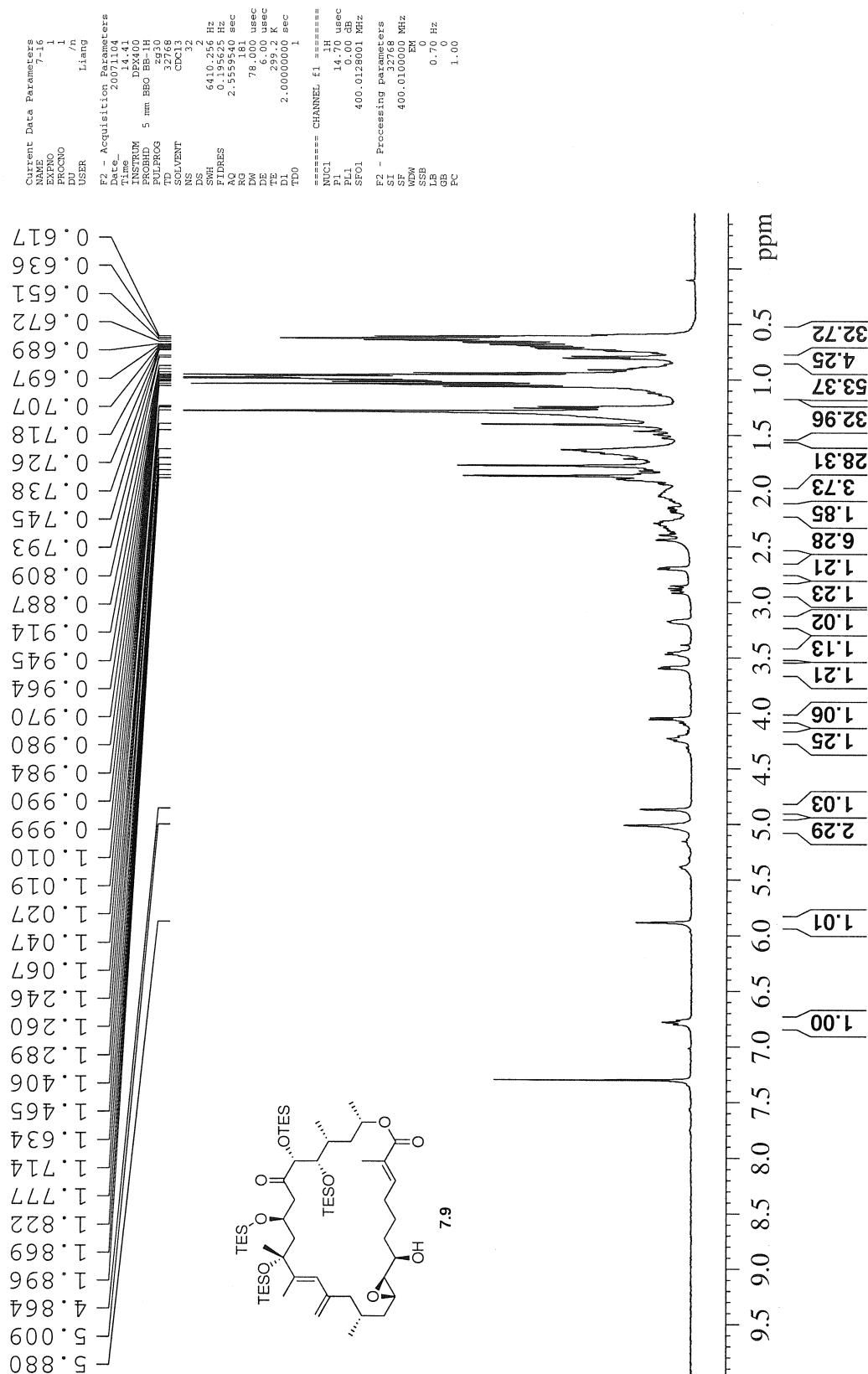


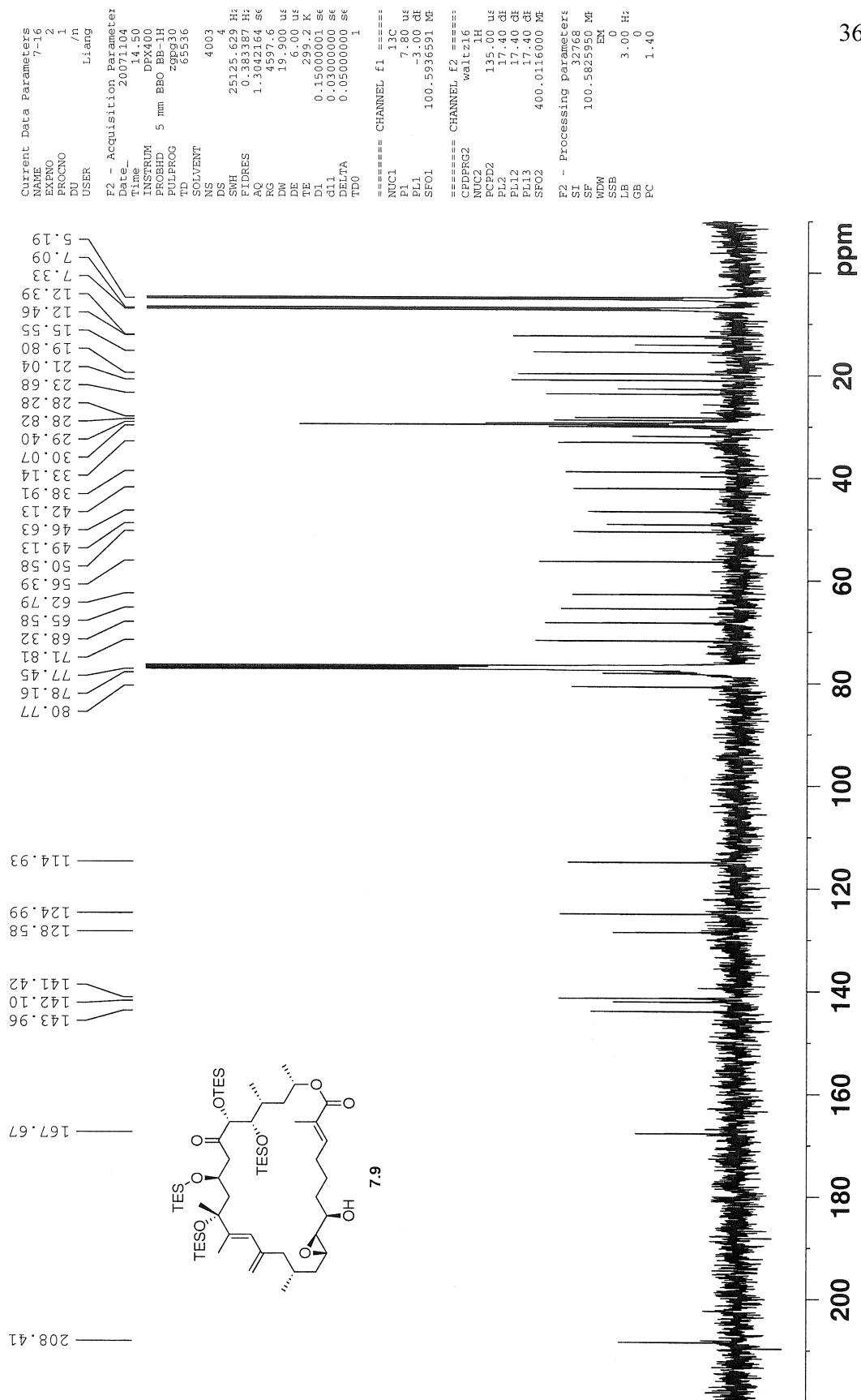


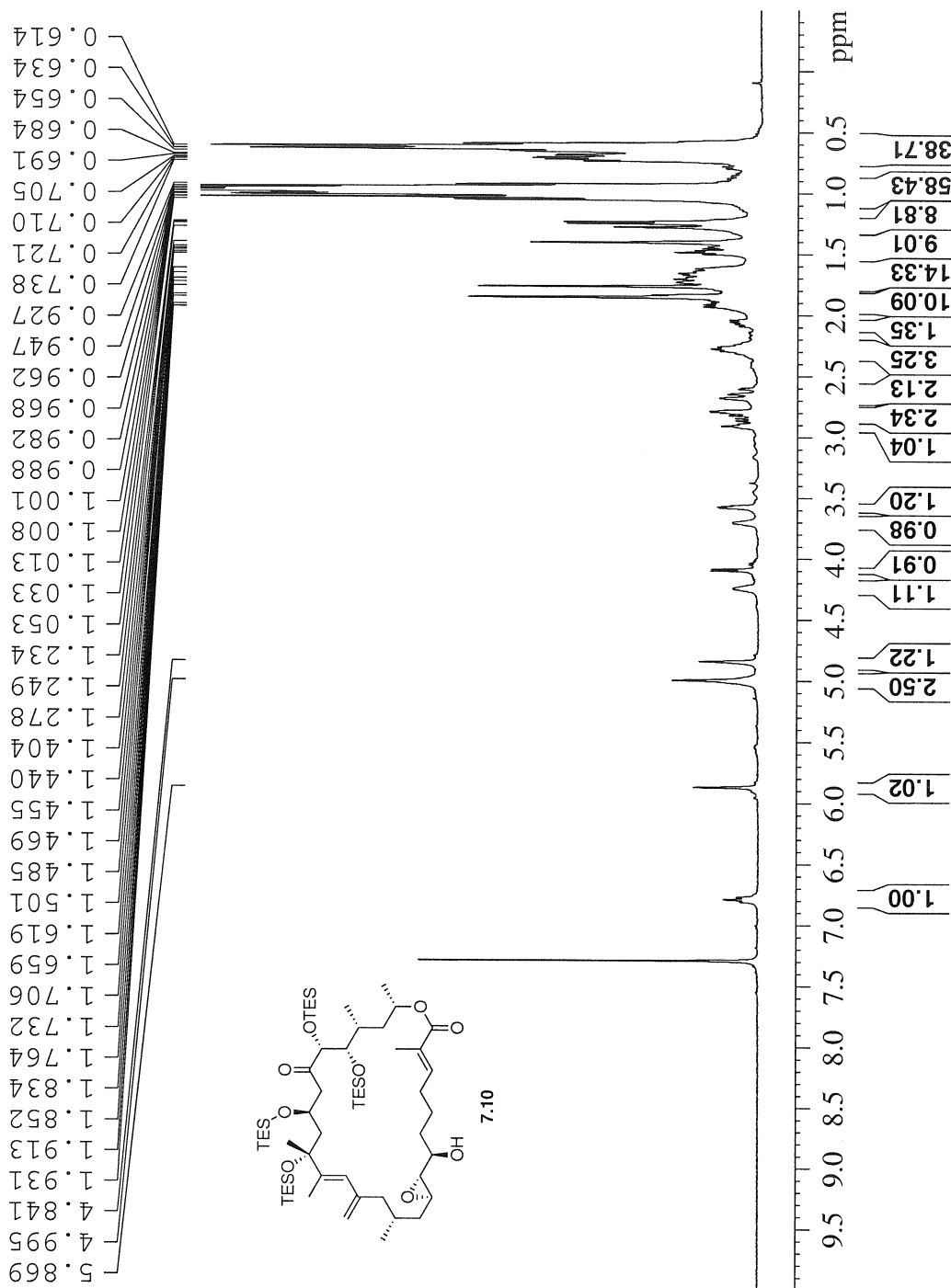


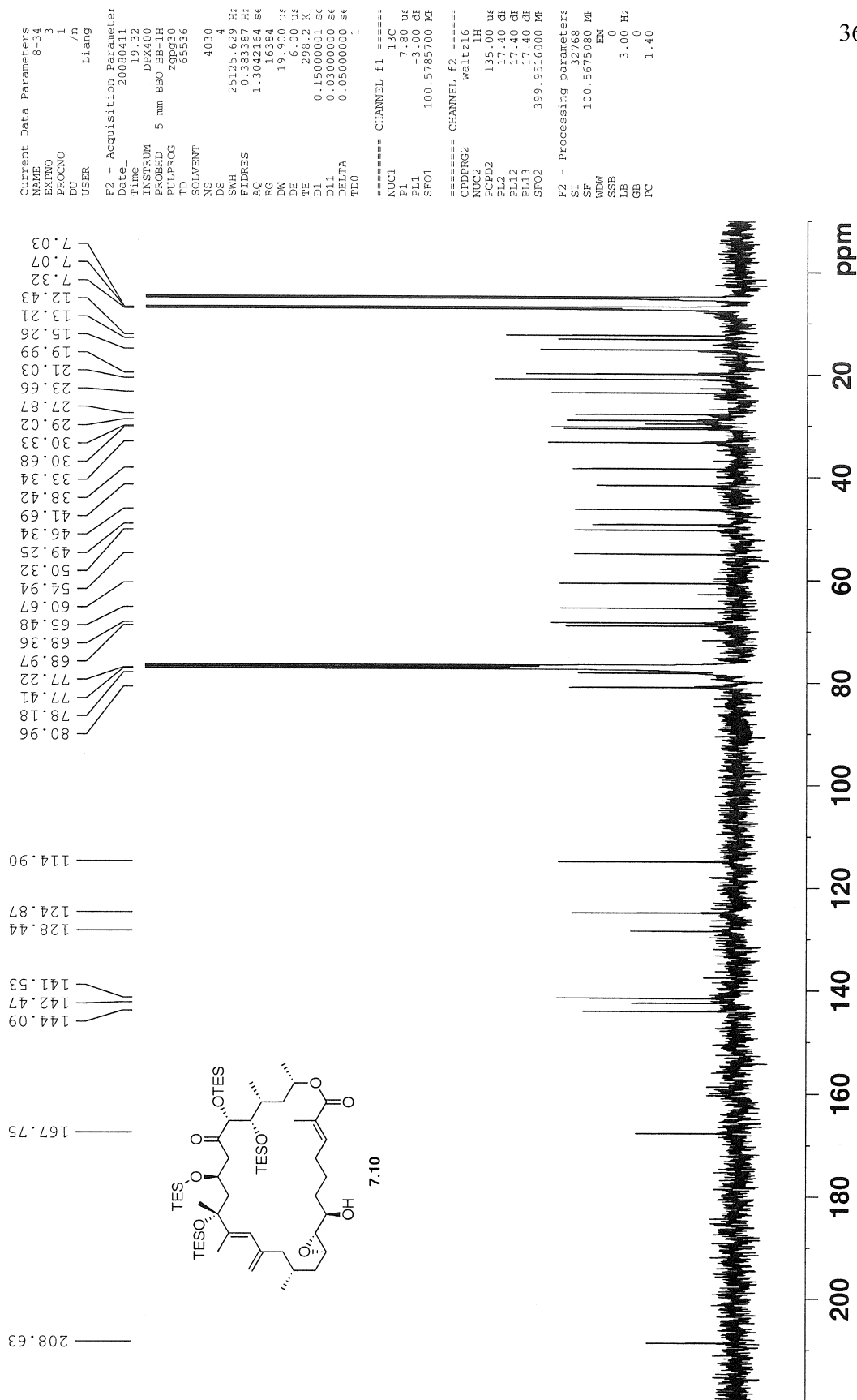


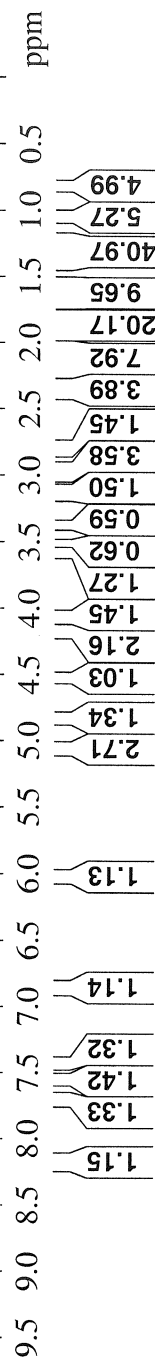
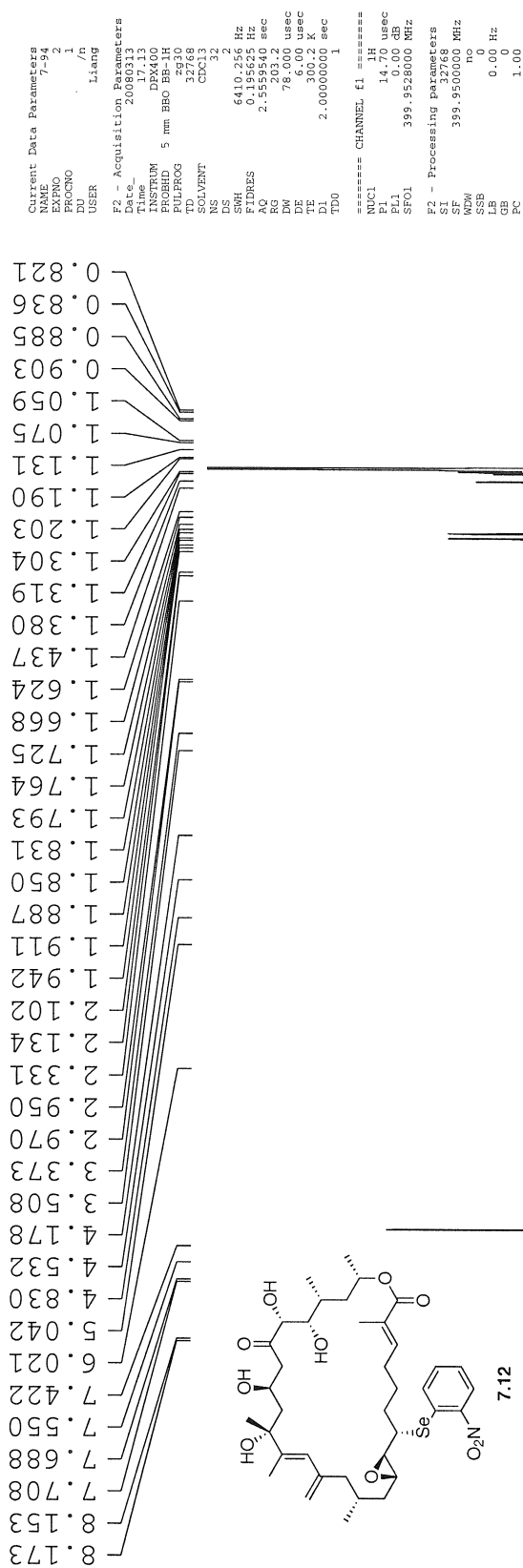


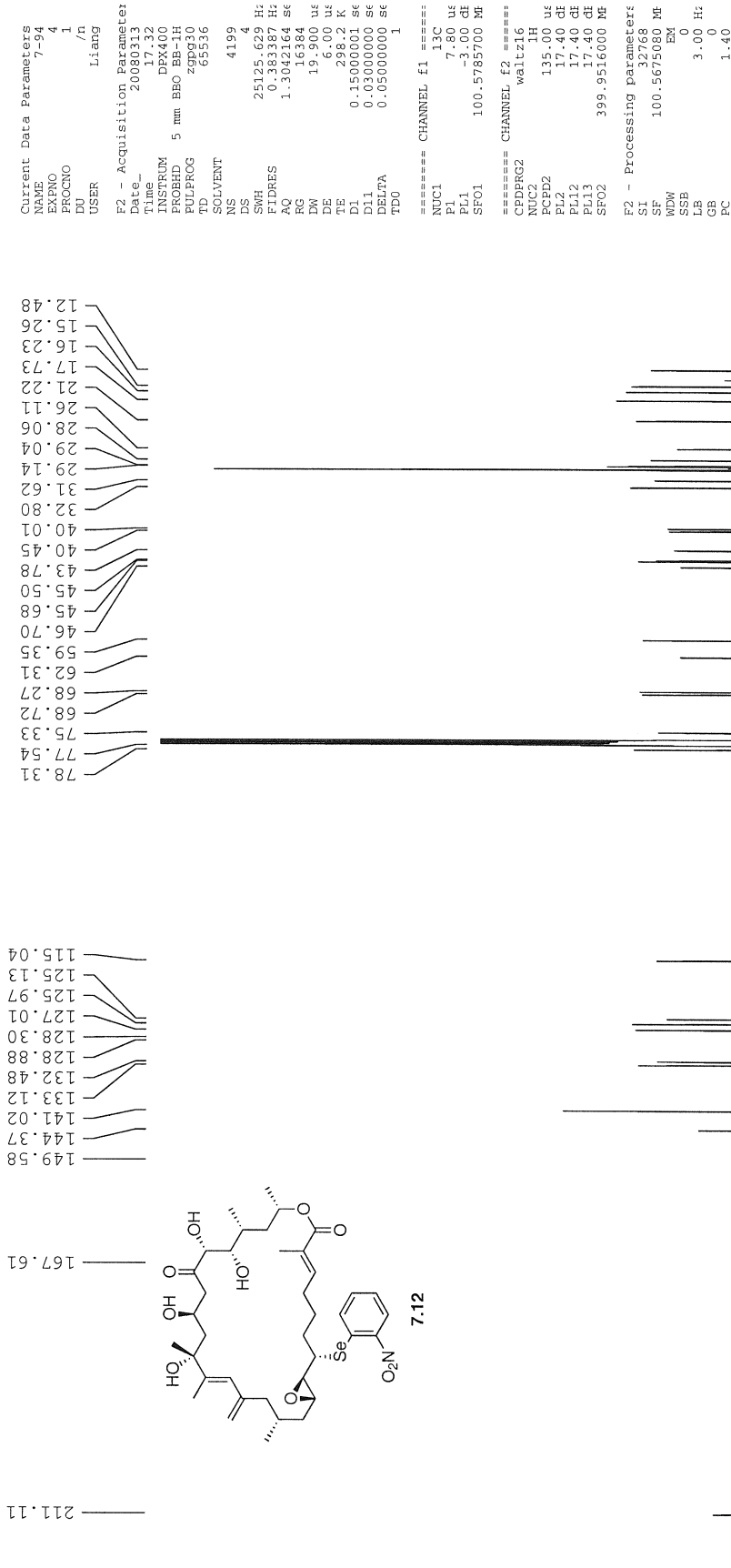




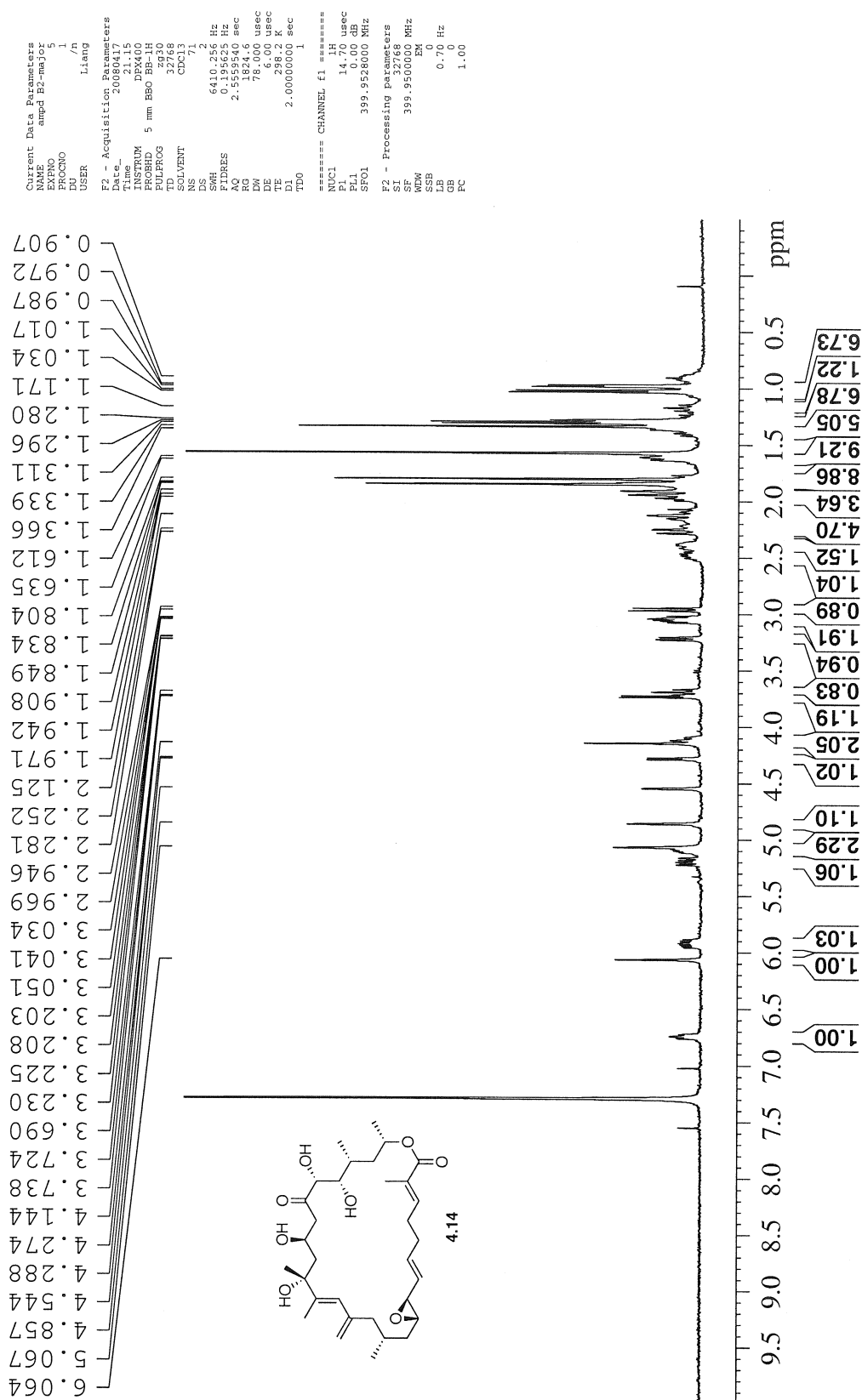


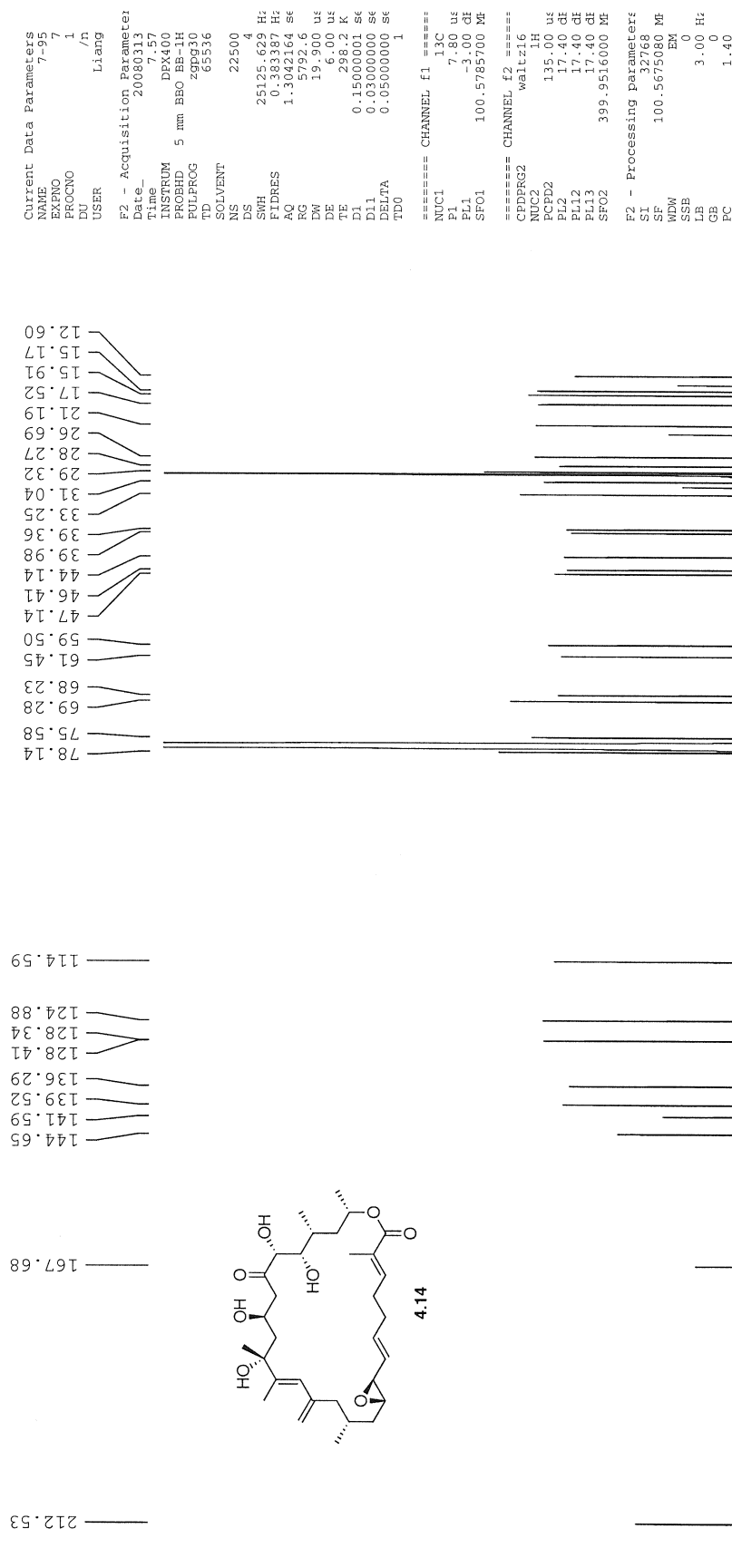


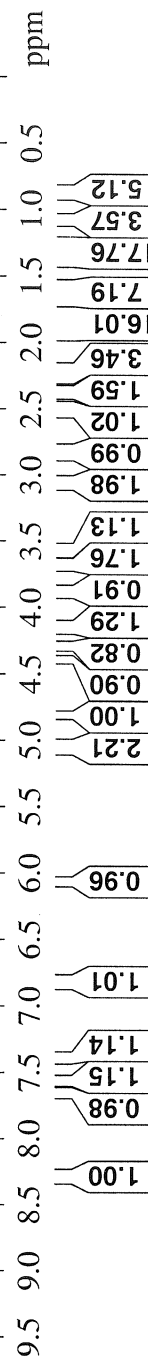
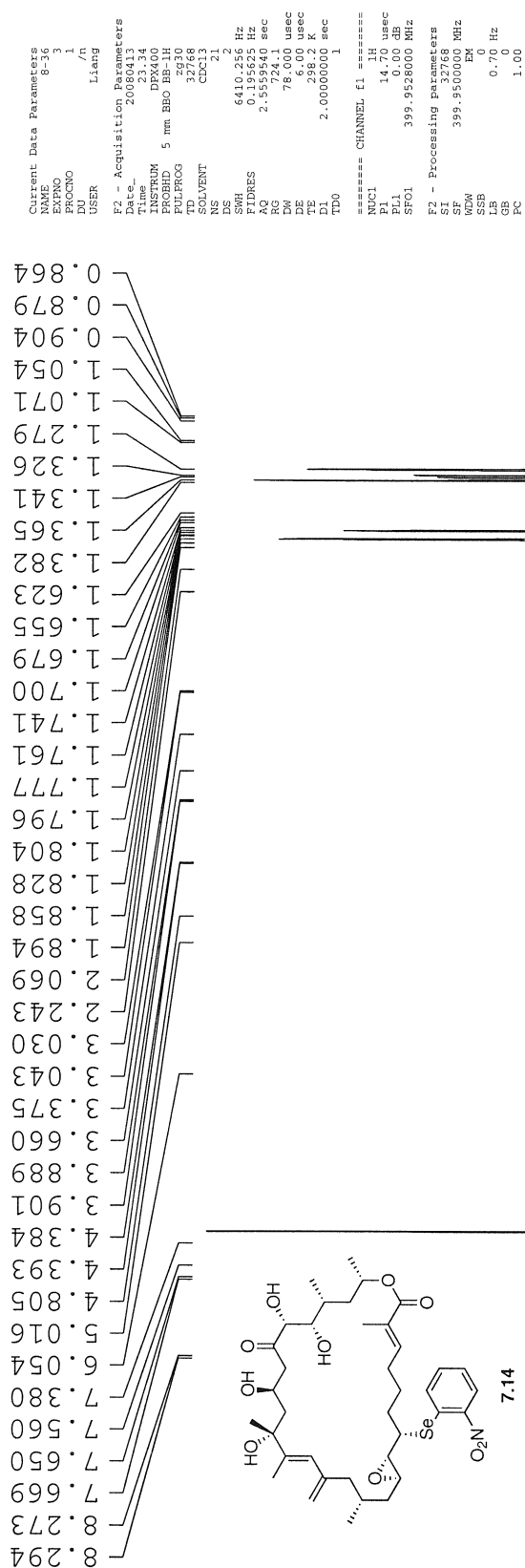












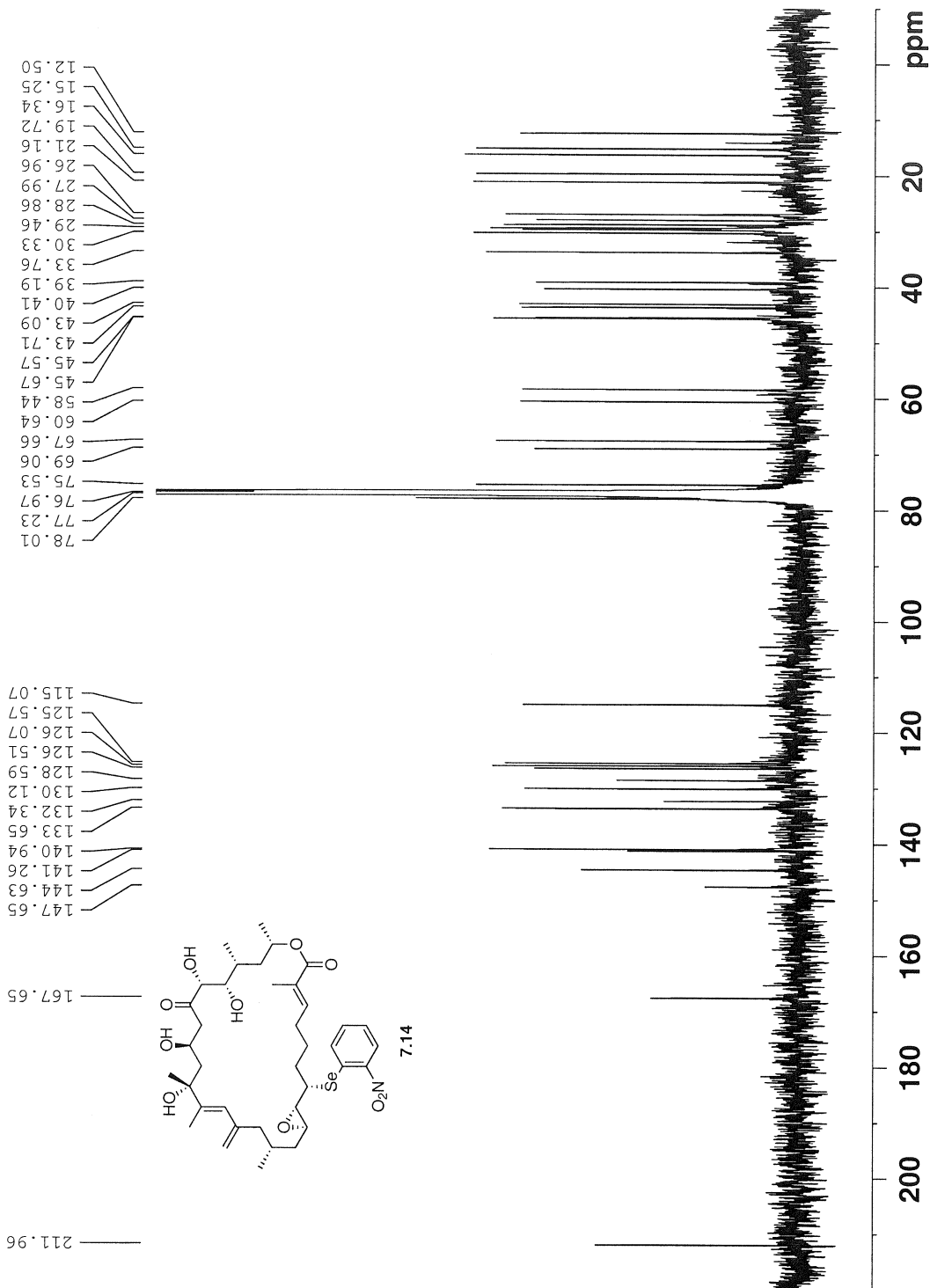
Current Data Parameters  
NAME 8-36  
EXPNO 5  
PROCNO 1  
DU /n  
USER Liang

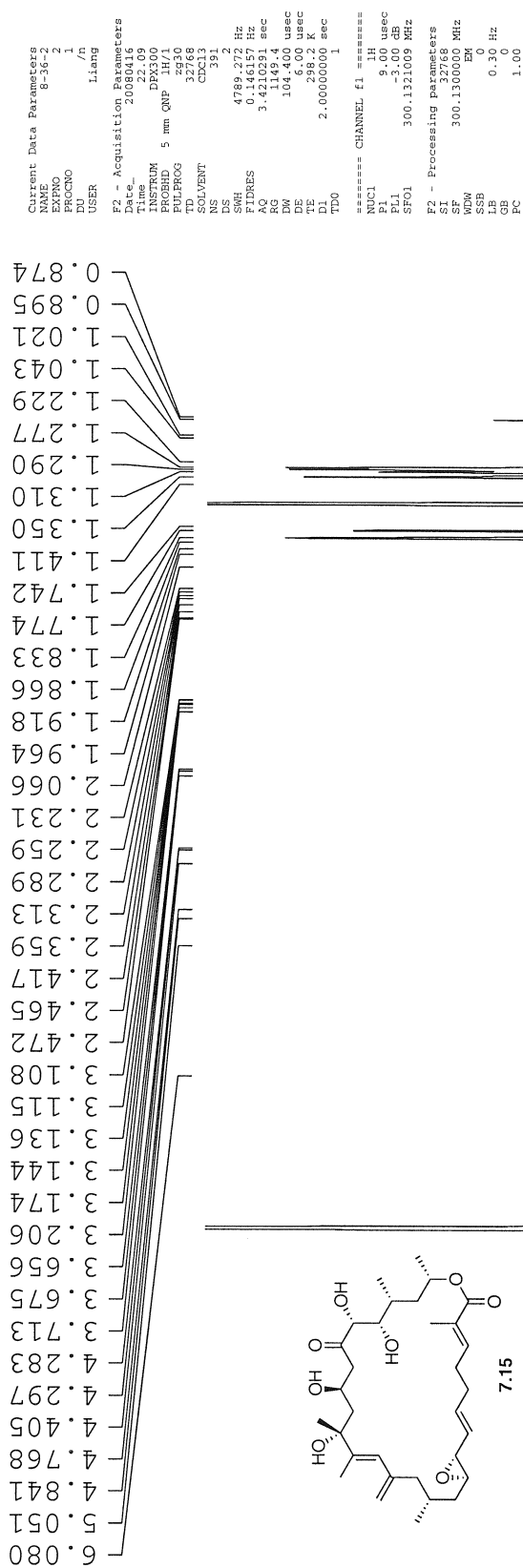
F2 - Acquisition Parameters  
Date\_ 20080413  
Time 23:50  
INSTRUM DFX400  
PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
PC 65536  
SOLVENT NS  
DS 19205  
SWH 25125.629 Hz  
FIDRES 0.383387 Hz  
AQ 1.3042164 s  
RG 16384  
DW 19.900 us  
DE 6.00 us  
TE 298.2 K  
D1 0.15000001 s  
D11 0.03000000 s  
DELTA 0.05000000 s  
TD0 1

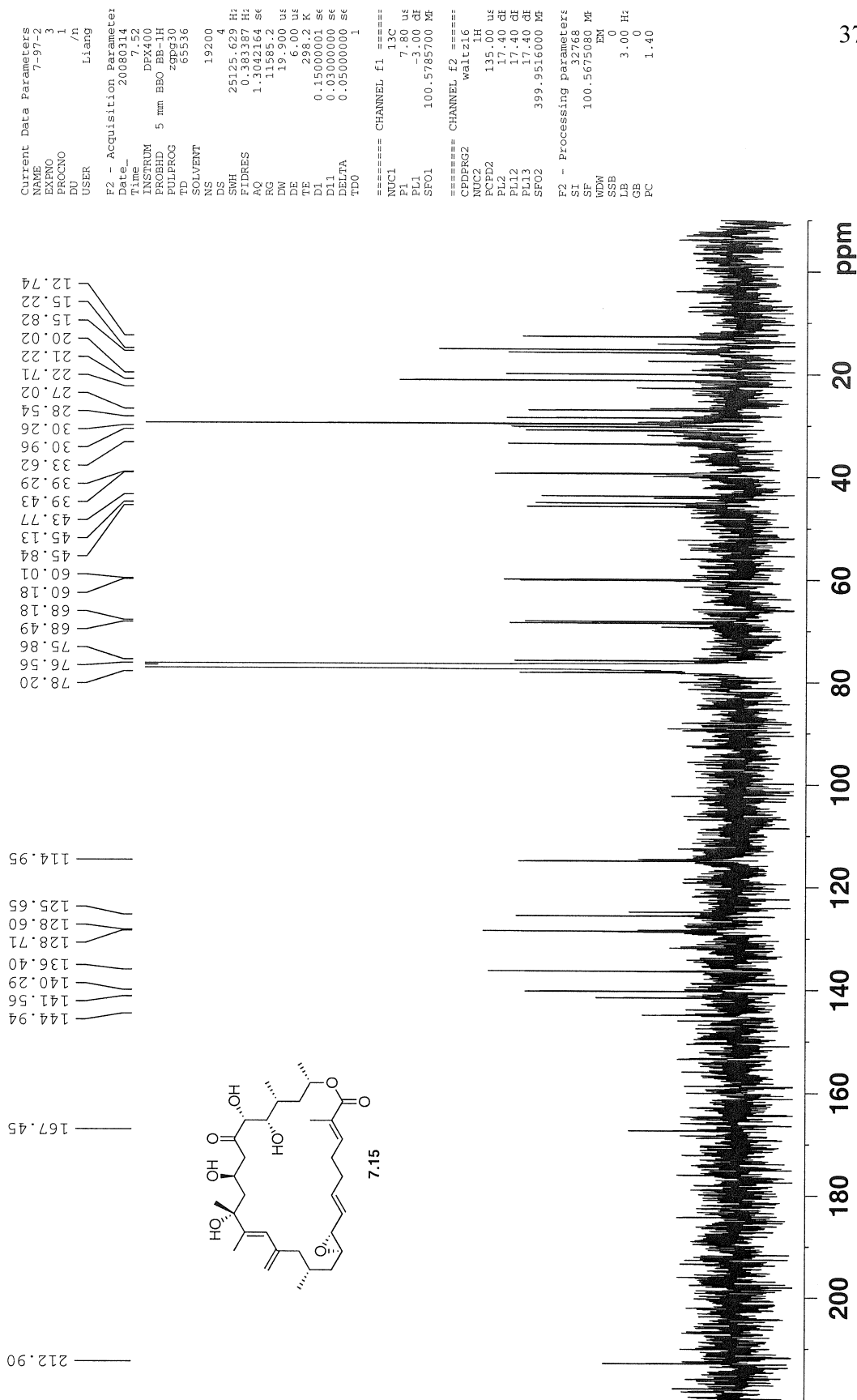
===== CHANNEL f1 =====  
NUC1 <sup>13</sup>C  
P1 7.80 us  
PL1 -3.00 dB  
SFO1 100.5785700 MHz

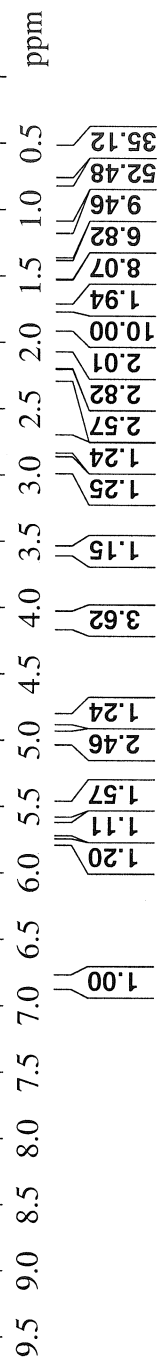
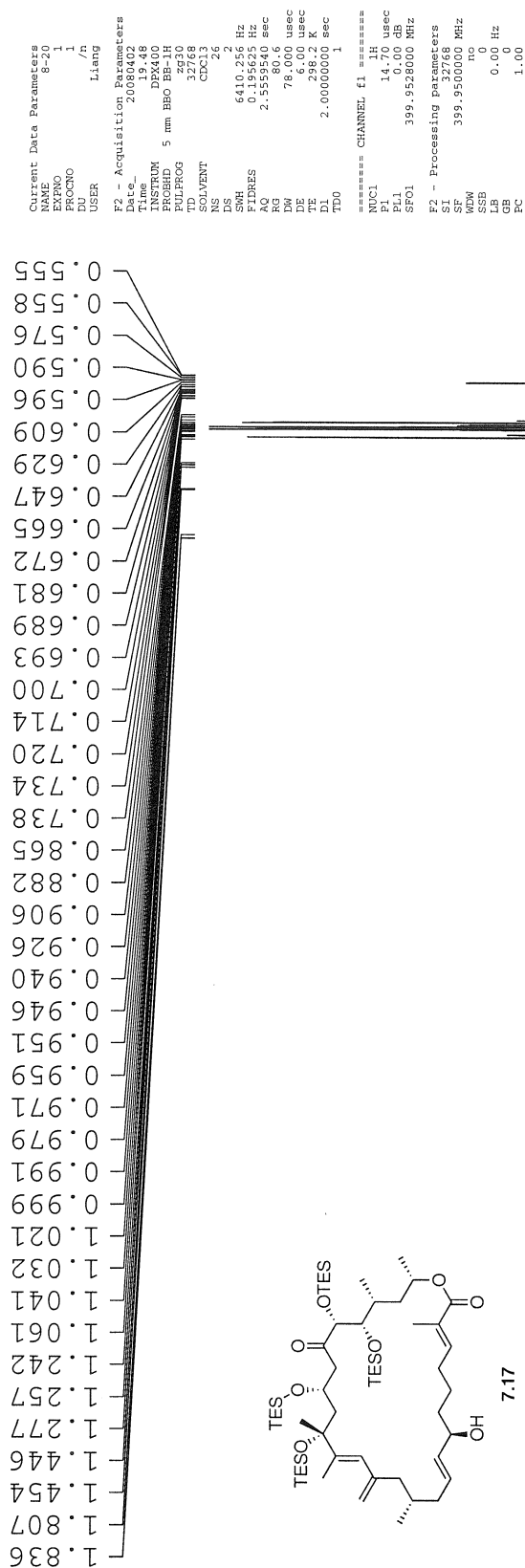
===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 <sup>1</sup>H  
PCPD2 135.00 us  
PL2 17.40 dB  
PL12 17.40 dB  
PL13 17.40 dB  
SFO2 399.9516000 MHz

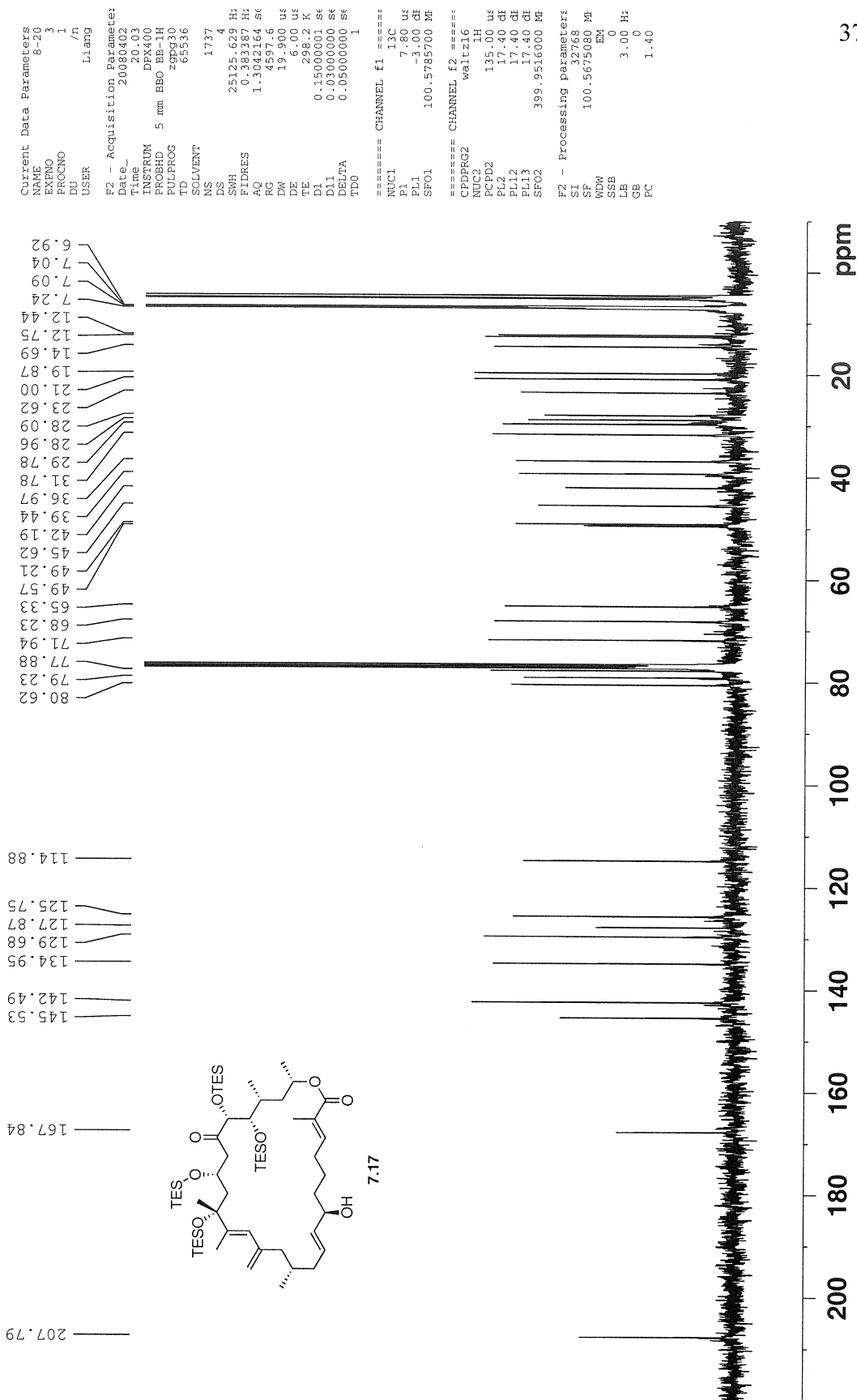
F2 - Processing parameters  
SI 32768  
SF 100.5675080 MHz  
WDW EM  
SSB 0  
CB 3.00 Hz  
CF 0  
PC 1.40





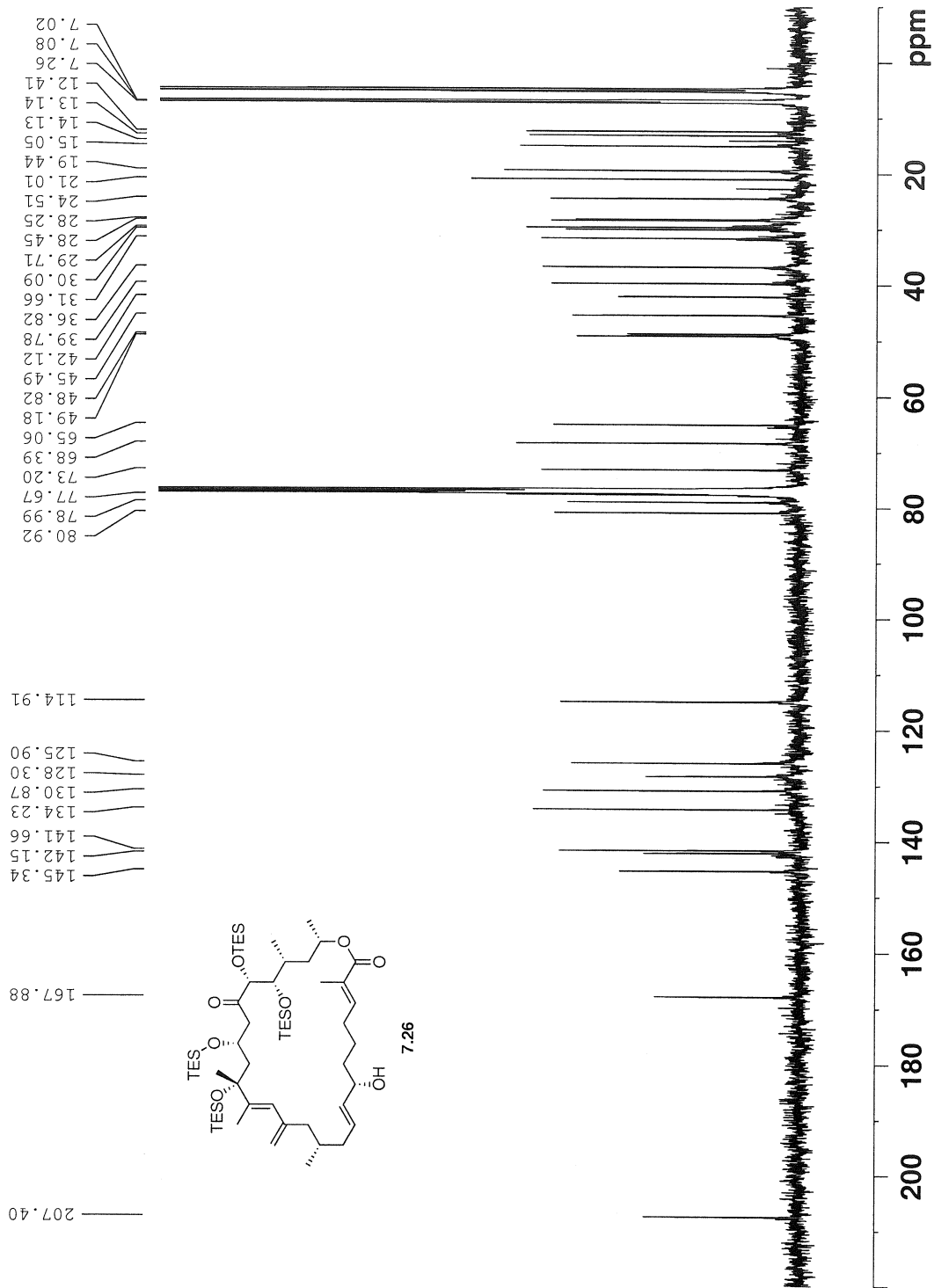


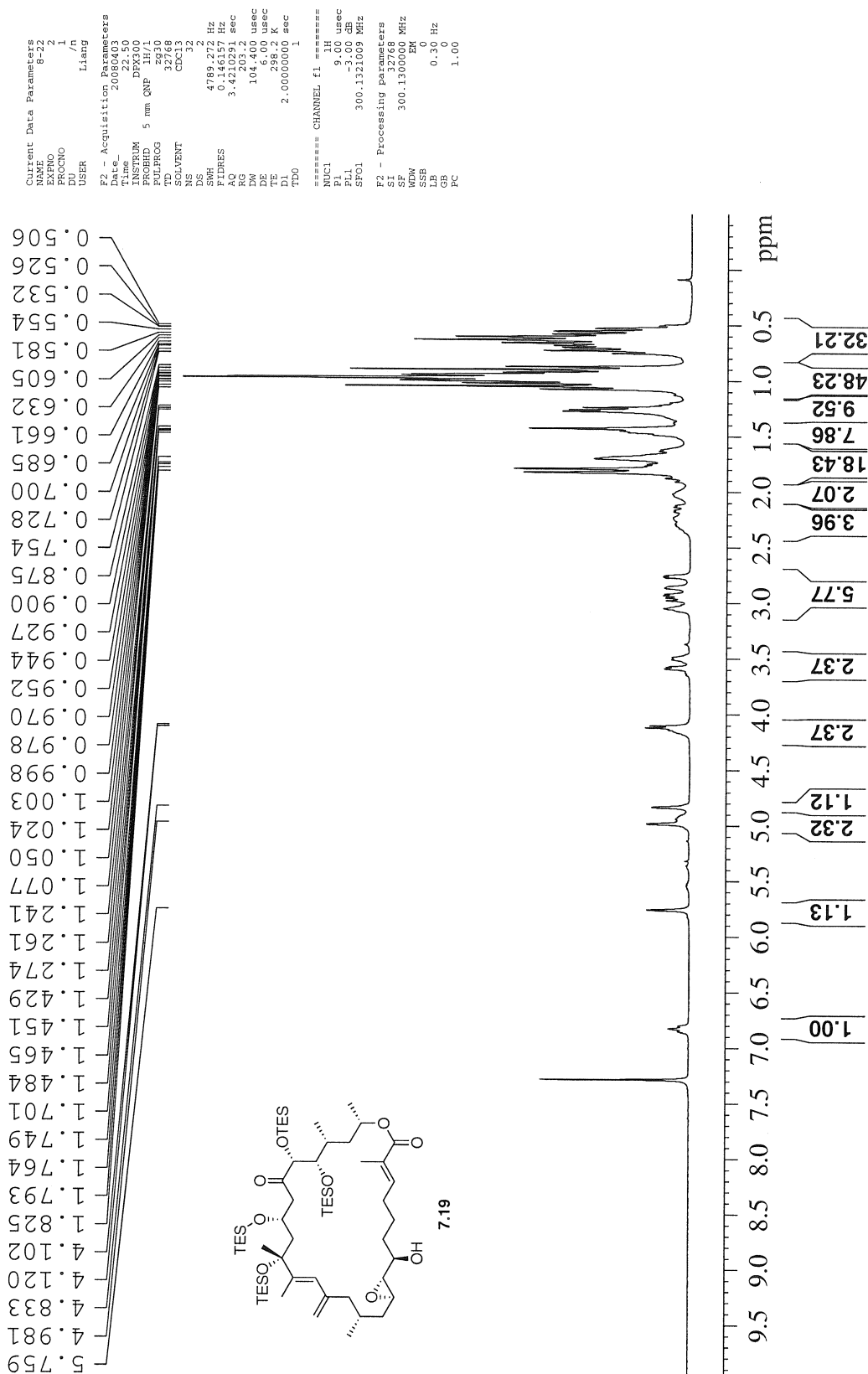


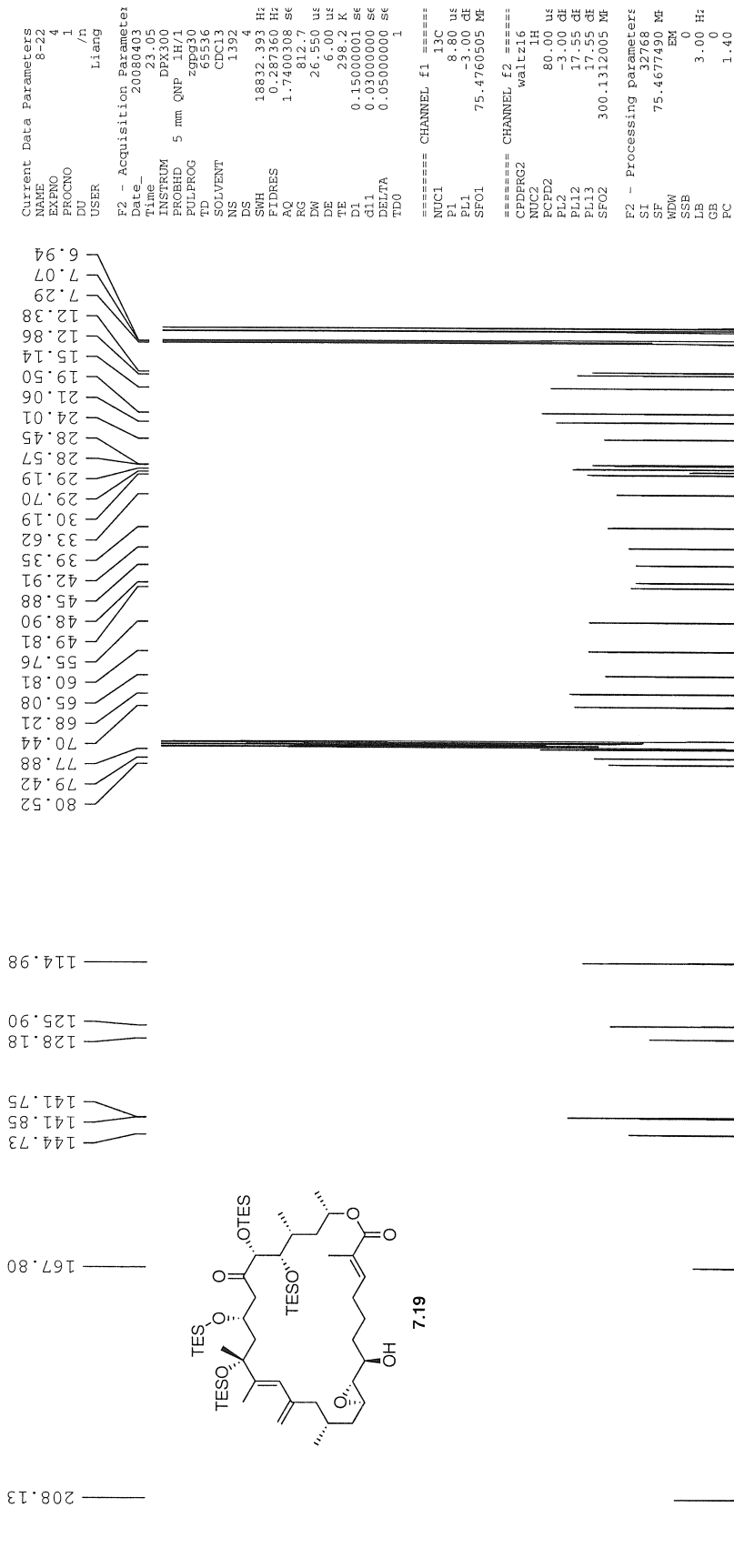


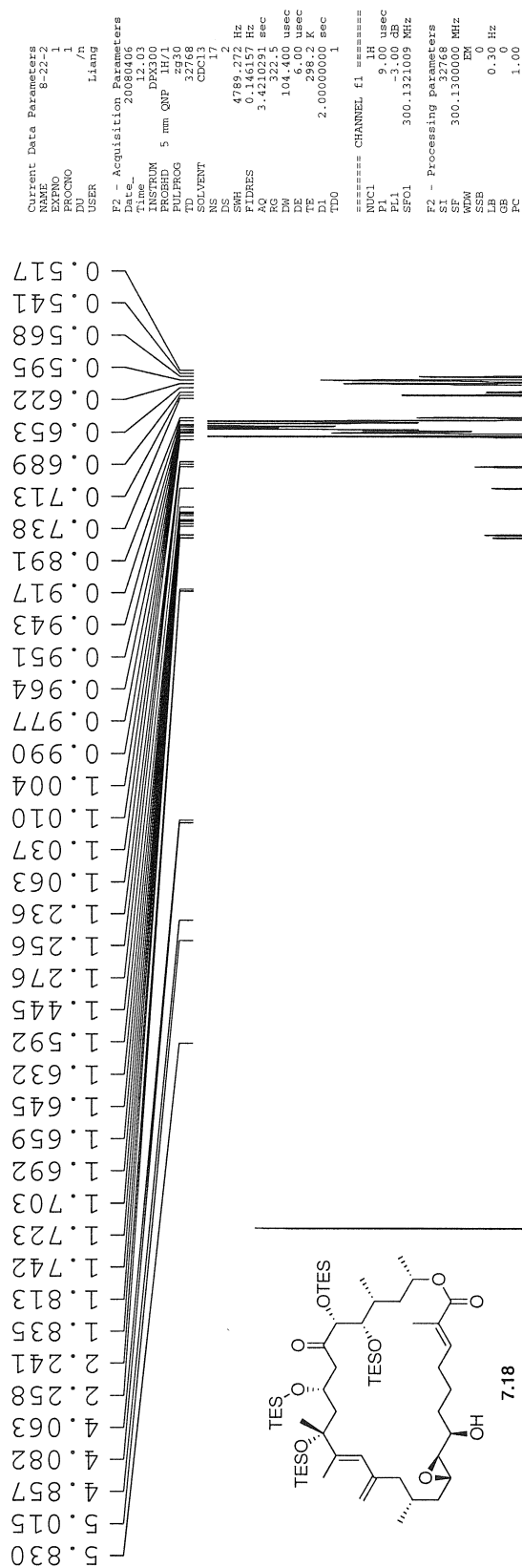




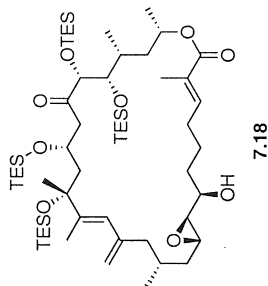


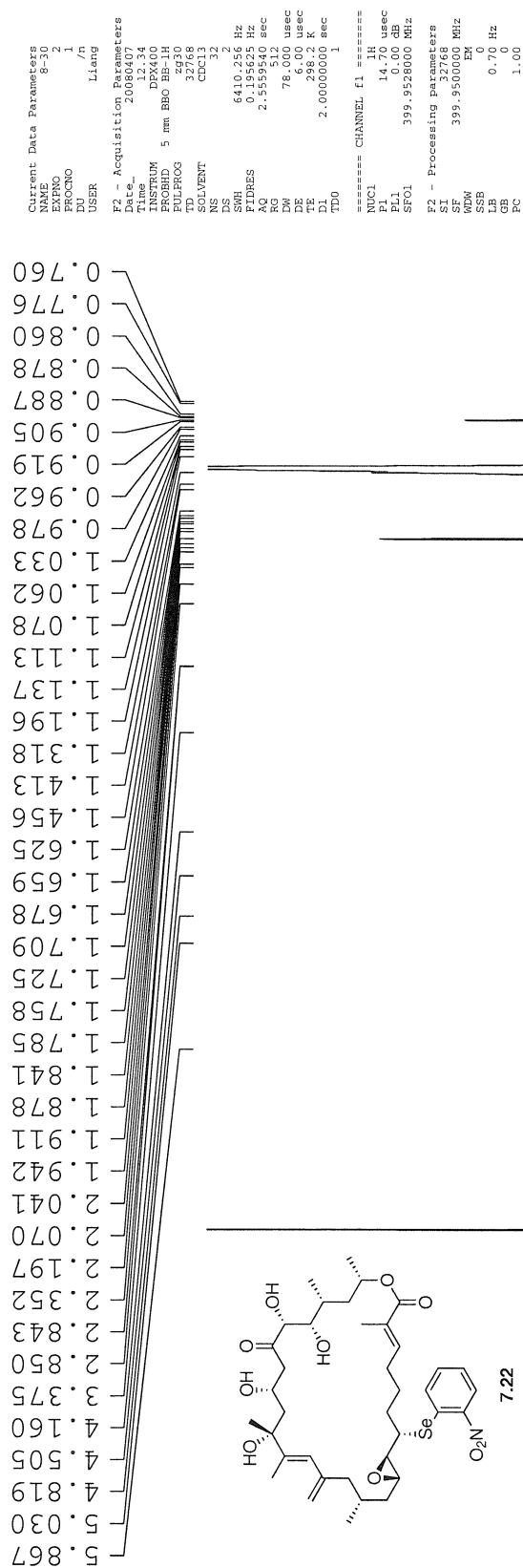


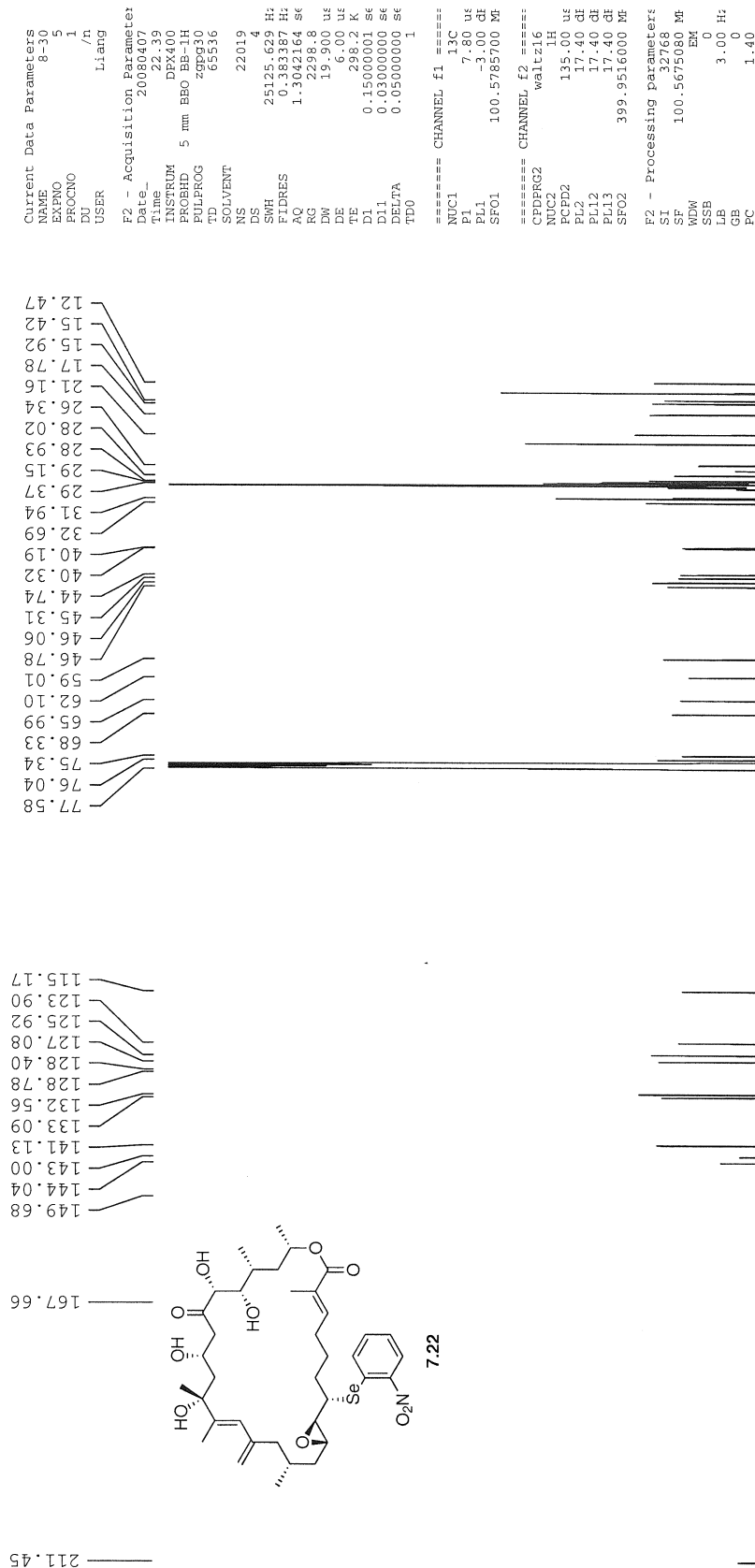




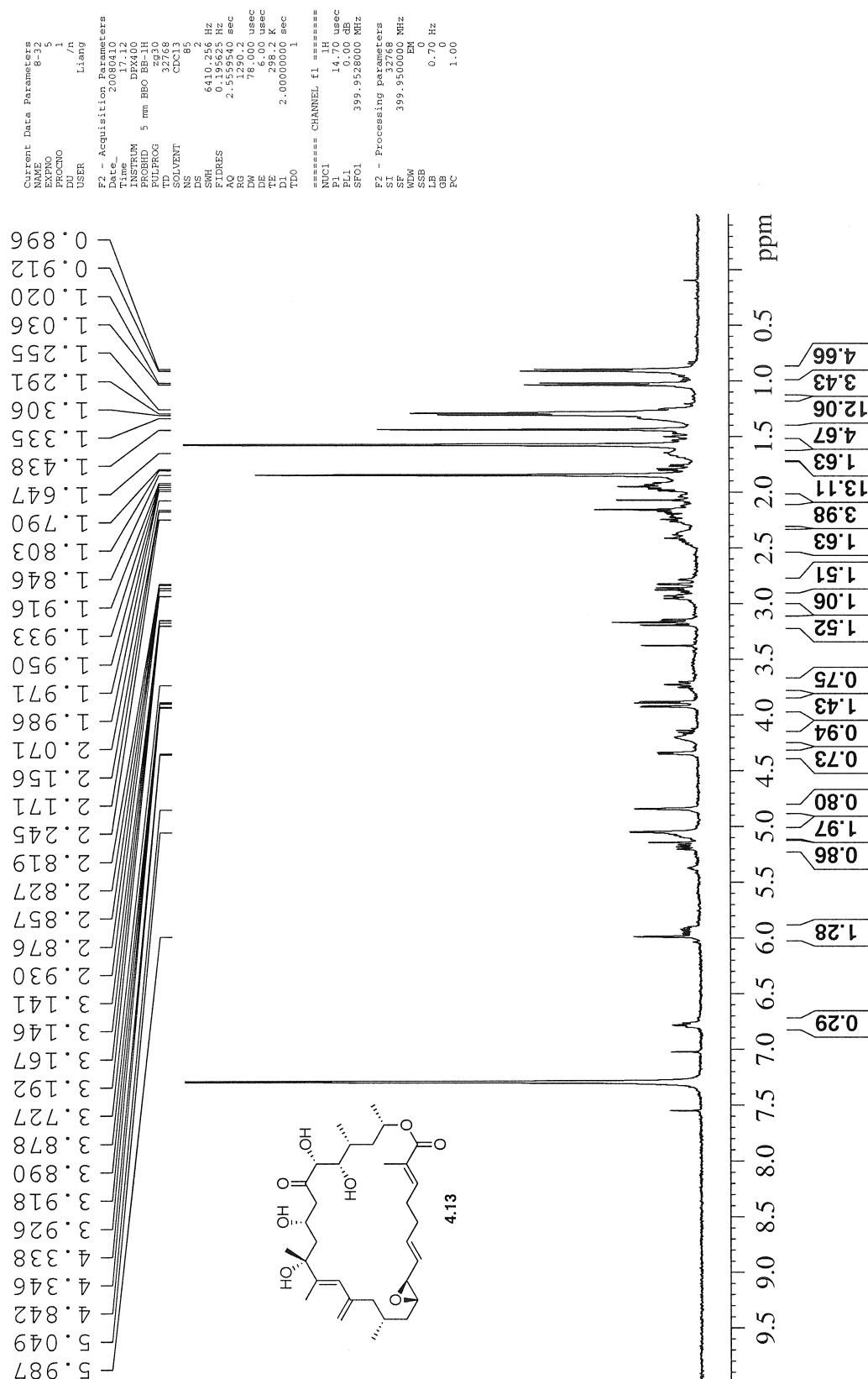
Chemical structure of compound 7.18 is shown in the bottom right corner. The structure is a complex macrocyclic molecule featuring a central ring system with various functional groups, including a carboxylic acid, a hydroxyl group, and a vinyl group. The structure is labeled with 'TESO' and 'TES' groups, indicating the presence of these substituents.

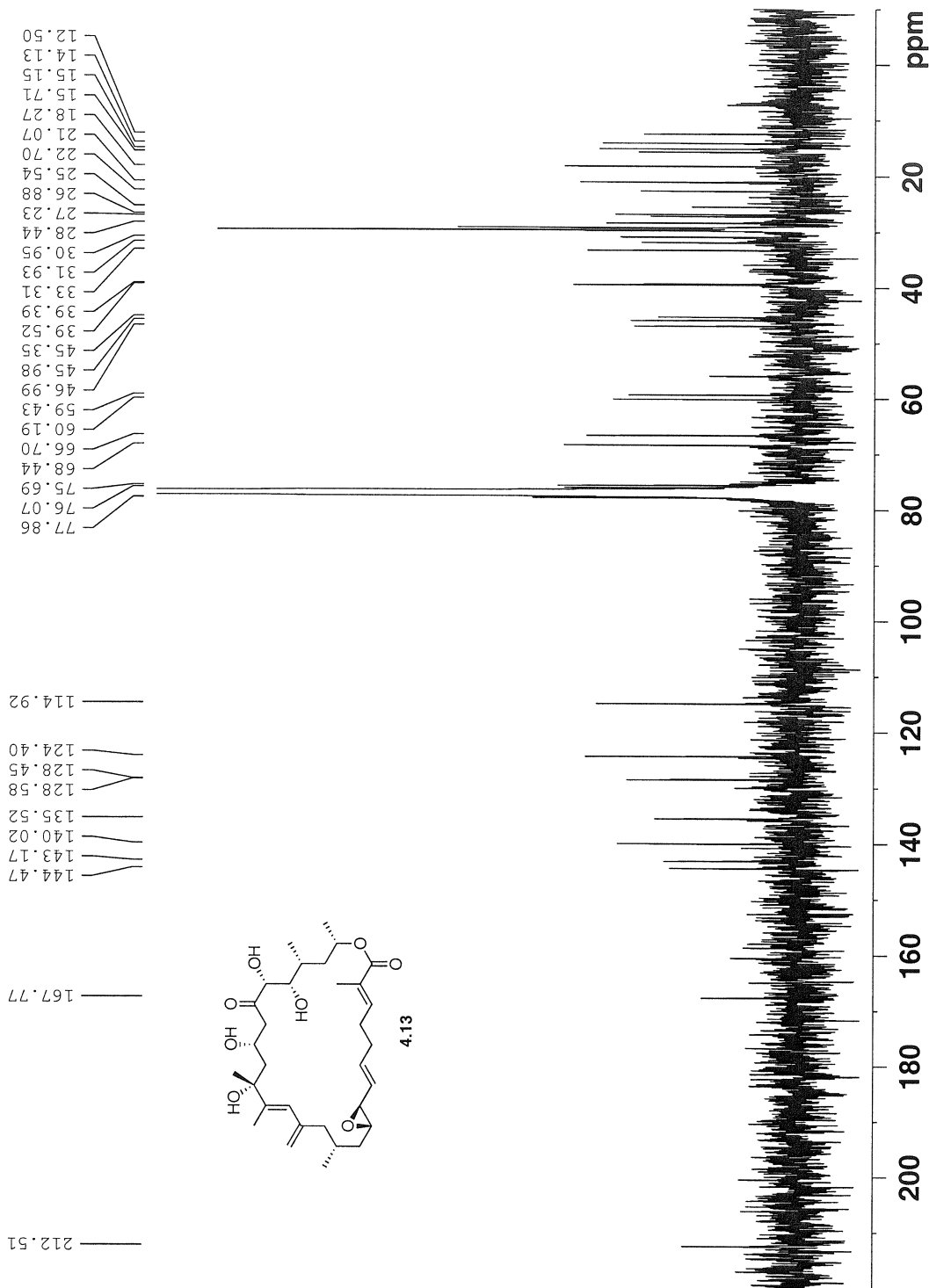


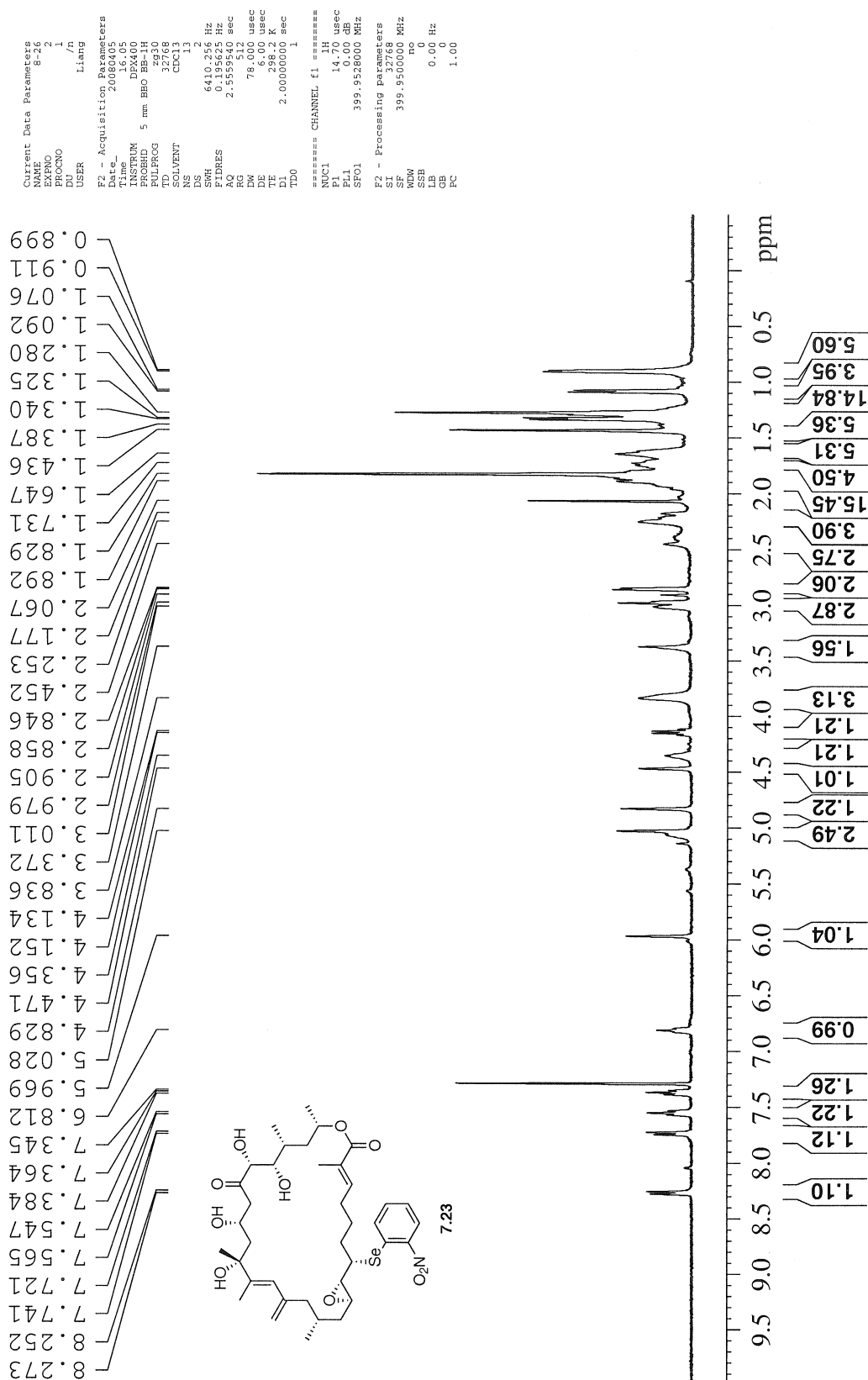












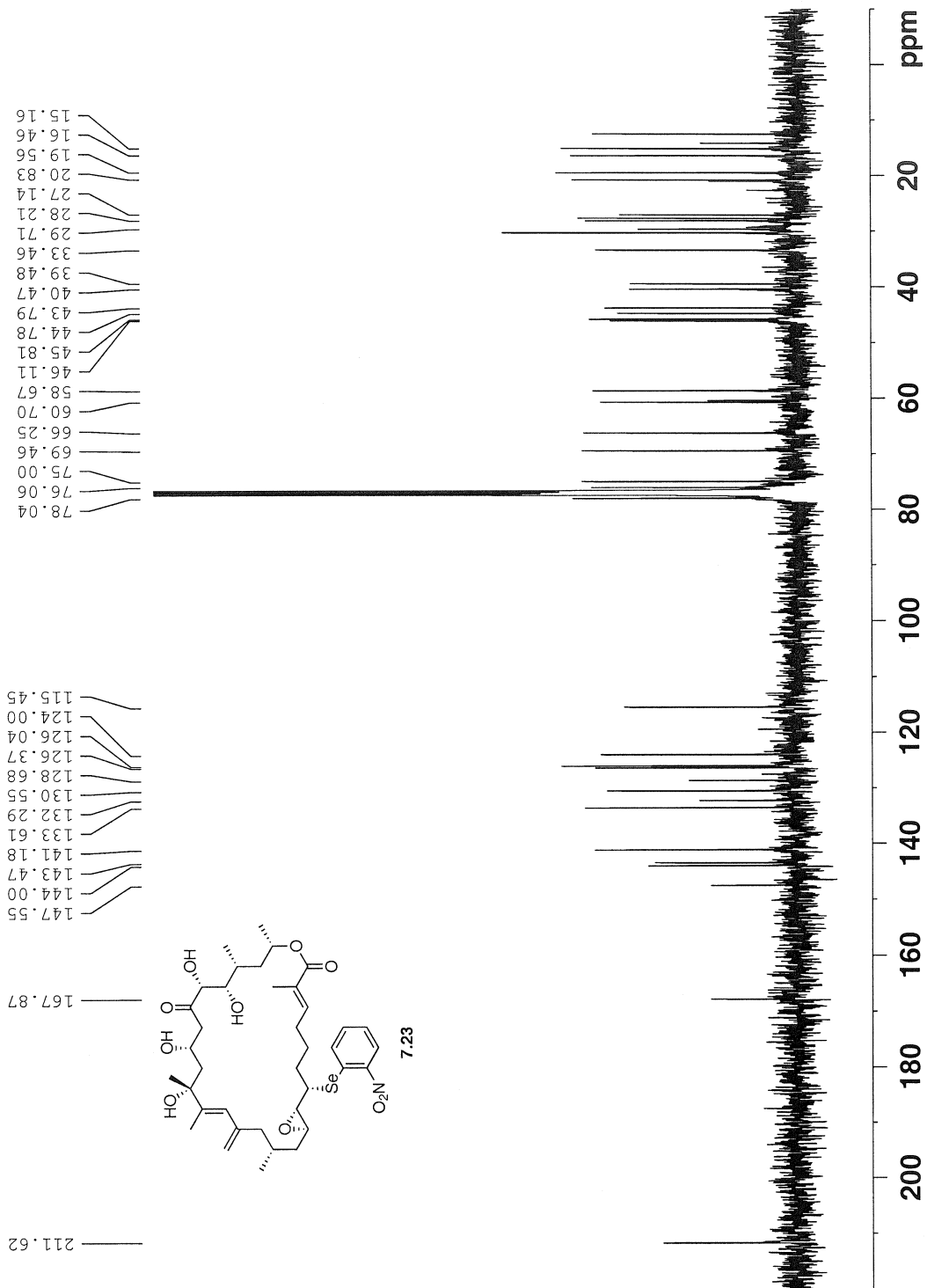
Current Data Parameters  
NAME 8-26  
EXPNO 4  
PROCNO 1  
DU /n  
USER Liang

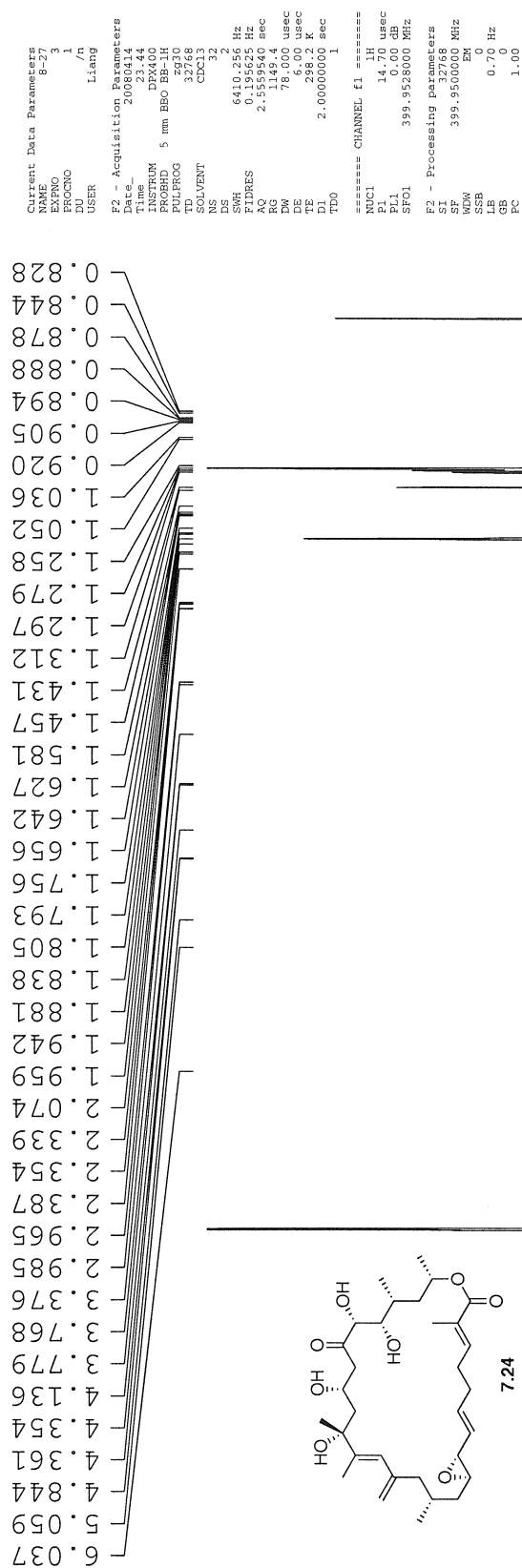
F2 - Acquisition Parameters  
Date\_ 20080405  
Time 16.55  
INSTRUM DFX400  
PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
TD 65536  
SOLVENT  
NS 3726  
DS 4  
SWH 25125.628 Hz  
FIDRES 0.383387 Hz  
AQ 1.3042164 s  
RG 4096  
DW 19.900 us  
DE 6.00 us  
TE 298.2 K  
D1 0.15000001 s  
D11 0.03000000 s  
DELTA 0.05000000 s  
TD0 1

===== CHANNEL f1 =====  
NUC1 13C  
P1 7.80 us  
PL1 -2.00 dB  
SFO1 100.5785700 MHz

===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 135.00 us  
PL2 17.40 dB  
PL12 17.40 dB  
PL13 17.40 dB  
SFO2 399.9516000 MHz

F2 - Processing parameters  
SI 32768  
SF 100.5675080 MHz  
WDW EM  
SSB 0  
LB 3.00 Hz  
GB 0  
PC 1.40





Current Data Parameters  
NAME 8-27  
EXPNO 5  
PROCNO 1  
DU /n  
USER Liang

F2 - Acquisition Parameters  
Date\_ 20080414  
Time 23.58  
INSTRUM DFX400  
PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
TD 65536  
SOLVENT  
NS 18937  
DS 4  
SWH 25125.629 Hz  
FIDRES 0.383387 Hz  
AQ 1.3042164 s  
RG 6502  
DW 19.900 us  
DE 6.00 us  
TE 298.2 K  
D1 0.15000001 s  
D11 0.03000000 s  
DELTA 0.05000000 s  
TD0 1

===== CHANNEL f1 =====  
NUC1 13C  
P1 7.80 us  
PL1 3.00 dB  
SFO1 100.5785700 MHz

===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 135.00 us  
PL2 17.40 dB  
PL12 17.40 dB  
PL13 17.40 dB  
SFO2 399.9516000 MHz

F2 - Processing parameters  
SI 32768  
SF 100.5675080 MHz  
WDW EM  
SSB 0  
LB 3.00 Hz  
GB 0  
PC 1.40

