## AN ABSTRACT OF THE THESIS OF

Brittany A. Beebe for the degree of Master of Science in Fisheries Science presented on September 24, 2019.

Title: Evaluating Fish Rescue as a Drought Adaptation Strategy for Imperiled Coho Salmon: A Life-Cycle Modeling Approach

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Projected intensification of drought as a result of climate change may reduce the capacity of streams to rear fish, exacerbating the challenge of recovering ESA-listed salmon populations. Without management intervention, some stocks will likely go extinct as stream drying and fragmentation reduce juvenile survival to unsustainable levels. To offset drought-related mortality, fish rescue programs have proliferated, whereby juvenile salmonids are captured and transferred to offsite rearing facilities. However, efficacy of this potential conservation tool remains poorly understood.I developed a life cycle model to examine the implications of fish rescue on coho salmon abundance across serial life stages. The model operates under the assumption that fish rescue improves juvenile survival but may decrease smolt-toadult return rates, eitherthrough lower marine survival or increased straying as a result of offsite rearing. The simulation model examines scenarios across various quantities of rescued fish, time in captivity, drought severity, and reduced smolt-to-
adult return rates. My results indicate that rescue increased adult returns and lowered extinction risk, particularly for fish captively reared for 1 year but decreased adult returns and increased extinction risk for fish reared only over summer when smolt-to-adult return rates were low. The findings suggest that fish rescue with long holding periods of approximately 1 year function more like a stock enhancement program than a drought adaptation tool, so its potential effects on adult returns should be evaluated with caution.
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Evaluating Fish Rescue as a Drought Adaptation Strategy for Imperiled Coho Salmon: A Life-Cycle Modeling Approach
by
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## APPROVED:

Major Professor, representing Fisheries Science

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I understand that my thesis will become part of the permanent collection of Oregon State University libraries. My signature below authorizes release of my thesis to any reader upon request.

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## Introduction

Myriad studies have attempted to predict biological response to climate change (Hijmans \& Graham, 2006; Cheung et al., 2010; Schindler \& Hilborn, 2015), yet many fail to provide tangible options to ameliorate the impacts of climate change on imperiled species (Wainwright \& Weitkamp, 2013). Rising temperatures, changes in the timing and intensity of precipitation, and habitat loss can cause shifts in species distribution, life history events, population dynamics, extinction risk, and interspecificinteractions (Mawdsley, O'Malley \& Ojima, 2009). With the rate of climate change outpacing the ability of many species to adapt and disperse (Hoffmann \& Sgrò, 2011), there is an urgent need for new tools and adaptation strategies to combat pervasive environmental changes that threaten species viability. The extent and degree of impact differs among regions (Dore, 2005; Kundzewicz et al., 2008), so conservation options should be tailored to local conditions. Here, I evaluate an emerging strategy for reducing the impacts of drought on an imperiled salmonid species.

Climate adaptation strategies are classified into four types: habitat protection and management, species management, monitoring and planning, and law and policy (Mawdsley et al., 2009). Current management tools use a combination of these general strategies with actions such as habitat protection and restoration, captive rearing, and assisted migration. Habitat protection and restoration often prioritize activities to conserve refugia (Beier \& Brost, 2010) and reduce environmental stressors (e.g., planting trees to shade streams or removing species that transpire excessively) (Katz et al., 2007). Captive rearing has been used to maintain genetic diversity (Brown \& Day, 2002; Bowkett, 2009) or breed new traits (van Oppen et al., 2015) and has likely saved some species from extinction (Lockwood et al., 2005); however, it
can also lead to domestication selection and individuals that fare poorly in the wild (Araki, Cooper \& Blouin, 2007; Tenger-Trolander et al., 2019). Further, captive rearing is often an "ark" strategy that is meant to save animals from temporary threats but not necessarily an altered future (Bowkett, 2009). Lastly, assisted migration is the action of translocating organisms to areas that are predicted to have more suitable habitat in the future, but are currently inaccessible because they are fragmented or beyond the dispersal potential of the organism. Assisted migration at larger scales and beyond current species ranges is highly controversial and rarely practiced in conservation (Hewittet al., 2011). Smaller scale applications that relocate individuals (i.e., problem bears, beached whales, entrapped fishes) to more suitable areas within the local habitat matrix are common and often go largely unnoticed (Higgins \& Bradford, 1996; Linnell et al., 1997). For example, species may be reintroduced to portions of their native range where they have been extirpated (Sarrazin \& Barbault, 1996), or introduced to portions where they have never been present but would likely fare well (Galloway et al., 2016). Perhaps the most ubiquitous example of small scale species translocations is fish salvage, the act of collecting fish from areas that become fragmented or dewatered by human activities and releasing them into adjacent habitats (Nagrodski et al., 2012). Salvaged fish are typically released immediately but in some instances are held in artificial rearing facilities for longer periods of time (Lopez Arriaza et al., 2017). Salvage may be implemented episodically, such as with instream construction activities, or may operate on a seasonal basis, coinciding with regularly scheduled water drawdowns (KTVZ, 2017).

While fish salvage is meant to save fish from threats that last only days and assisted migration to expand species range is a response to future climatic states, there is a relative lack
of management interventions at intermediate timescales. Most stressors are seasonal, but the topic of seasonally assisted migration is not broadly recognized or assessed. In highly seasonal environments, where the largest impacts of climate change are likely to be associated with altered annual extremes rather than changes in mean conditions (Jaeger, Olden \& Pelland, 2014), it is possible that relocating individuals during stressful portions of the year could allow populations to persist where they would otherwise be predicted to go extinct. However, this intermediate scale of intervention (i.e., seasonal rescue within contemporary species ranges) remains largely unexplored in the field of climate change adaptation.

Here, I explore the potential for monthly-to-annual rearing and translocations within species ranges to mitigate the effects of seasonal drying on stream-dwelling fish. Riverine ecosystems exhibit extreme seasonal variation that will likely dictate species response to climate change. For example, precipitation and runoff interact with watershed characteristics such as geomorphology and land cover to regulate the influx of water into the stream channel and dictate the temporal pattern of discharge (Poff et al., 1997). Streamflow can be relatively constant (e.g., groundwater dominated streams), but generally, flow varies seasonally by several orders of magnitude (e.g., precipitation dominated streams), causing aquatic ecosystems to oscillate between contracted and expanded states (Lake, 2003; Humphries \& Baldwin, 2003). The magnitude of contraction varies among regions and climatic zones, but streams with intermittent flow are common, especially in headwaters and regions with distinct wet and dry seasons (Larned et al., 2010; Woelfle-Erskine, Larsen \& Carlson, 2017). Reduced water input and increased water withdrawal can intensify stream contraction, resulting in severe and prolonged dryness. During the contraction process, riffle habitat sections often dry
first, transforming streams into an assortment of isolated pools (Stanley, Fisher \& Grimm, 1997; Labbe \& Fausch, 2000; Hwan \& Carlson, 2016). In addition to reduced habitat availability, sustained drought conditions can increase extremes of physicochemical conditions, such as temperature and dissolved oxygen (Magoulick \& Kobza, 2003) and even lead to seasonal pool drying. In rain-dominated watersheds of the Pacific Northwest, intermittent streams provide summer rearing habitat for culturally valued species (Everest, 1973; May \& Lee, 2004; Wigington et al., 2006). Projected intensification of drought conditions in the region may increase the duration and severity of stream fragmentation as summers are projected to become hotter and drier (Mote \& Salathé, 2010). This will likely decrease summer minimum flow conditions throughout the region (Hamlet et al., 2013), reduce the rearing capacity of many streams (Harvey, Nakamoto \& White, 2006; Falke et al., 2011), and exacerbate the challenge of recovering ESA-listed populations (Crozier et al., 2019).

In response to current and projected threats, there is growing interest in developing strategies to mitigate the effects of stream drying on salmonid populations. A potentially controversial implementation of assisted movement and rearing, called fish rescue, has been used to combat drought-induced mortality in intermittent streams. Fish rescue differs from the more typical fish salvage operations. Instead of immediately transporting fish to other habitats, fish rescue holds fish for an extended period of time before releasing them back to the stream. The California Department of Fish and Wildlife (CDFW) has used this strategy in the Russian River Basin in central California for the past few years, rescuing thousands of stranded juvenile coho and chinook salmon and steelhead each year (CDFW, personal communication). Rescued coho salmon are transported to a hatchery for use in a broodstock program or reared until
streamflow improves. Likewise, the Carmel River Steelhead Association has also implemented fish rescue in California, collecting fish from fragmented habitat during low flows (May-Oct) and rearing them in a specialized facility until release the following winter (Lopez Arriaza et al., 2017). Fish rescue programs aim to increase overall survival by circumventing life stages with low survival, but the length of time fish are held and the number of fish rescued can influence the effects of rescue. Rescue often occurs over summer months when streamflow is low, but if habitat is limited for subsequent life stages, then returning re scued fish to the stream may increase fish density beyond a sustainable threshold and decrease survival. Additionally, captively reared fish may exhibit altered phenotypes (Kihslinger, 2006), which could additionally lowertheir subsequent survival compared to wild (i.e., non-rescued) counterparts. If a large percentage of fish are rescued, the effects of rescue are magnified, underscoring the need to better understand the consequences of fish rescue.

To evaluate the efficacy of fish rescue, I developed a simulation model to explore how drought-induced fragmentation and seasonal fish rescue interact to affect salmon population dynamics. Specifically, I developed a life-cycle model to examine abundance of returning adults and extinction risk across various levels of rescue intervention, drought severity, and smolt-toadult return rates. The model is flexible enough for use in systems with a range of environmental conditions and different rescue interventions. To showcase my life-cycle model, I use it to evaluate one of the most prominent fish rescue programs in the Pacific Northwest. The program, Northwest Wild Fish Rescue (NWFR), removes tens of thousands of ESA-listed juvenile coho salmon (Oncorhynchus kisutch) from stream reaches that are susceptible to drying and rears them in captivity until smoltification (approximately 1 year). My life-cycle
model will enable managers to assess the trade-offs associated with fish rescue and identify the range of scenarios when intervention may be an appropriate action in a specific watershed. Here, I compare the abundance of returning adults and extinction risk resulting from differing rescue scenarios to answer the following questions:

1. Under what ecological conditions does rescue increase adult returns and lower extinction risk compared to scenarios without fish rescue?
2. Can fish rescue effectively operate as a drought mitigation tool by maintaining adult returns across various levels of drought severity?
3. How are the results of fish rescue sensitive to model parameters that represent ecological context (survival in wild, population size, drought severity, etc.) or the costs of rescue (hatchery-like effects on post-rescue survival)?

## Methods

## Model Overview

Coho salmon have complex life cycles and exhibit ontogenetic habitat shifts, so survival during one life stage, such as juvenile oversummersurvival, may or may not influence population productivity (Mason, 1976). Life-cycle models are widely used in salmon conservation to explore life stage-specificimpacts on population dynamics (Moussalli \& Hilborn, 1986; Nickelson \& Lawson, 1998; Oosterhout et al., 2005; Scheuerell et al., 2006). I developed a life-cycle model to examine the effects of fish rescue across varying levels of intervention, drought severity, and assumptions regarding the effects of captive rearing on smolt-to-adult survival. The model operates under the assumption that fish rescue improves
juvenile survival but may decrease smolt-to-adult return rates, either through lower marine survival (Jonsson, Jonsson \& Hansen, 2003; Saloniemi et al., 2004; Jokikokko et al., 2006) or increased straying as a result of captive rearing (Keefer \& Caudill, 2014). Additionally, I assume that drought reduces summer rearing capacity, thus reducing survival of wild fry (Harvey et al., 2006; Hwan et al., 2018).

## Case Study

Located within the Cascade Mountain foothills of the Pacific Northwest, the East Fork (EF) Lewis River, originates in the Gifford Pinchot National Forest and flows westward, emptying into the Lewis River which flows into in the Columbia River. The EF Lewis Riverwatershed is approximately $610 \mathrm{~km}^{2}$ and consists of small urban and rural communities. It is labeled as a Tier 1, key watershed by the President's Forest Plan because it provides, or is expected to provide, high quality fish habitat and water quality (USFS, 1995). However, land use activity throughout the watershed (e.g., agriculture, logging, and mining) has degraded habitat and increased stream temperatures in the lowerEF Lewis River Basin (National Marine Fisheries Service, 2013). Juvenile coho salmon are primarily distributed throughout tributaries in the lower EF Lewis River and in side- and off-channel habitat in the main stem river. Two streams within the lower EF Lewis Riversub-basin, Mason Creek and its tributary, Tsugawa Creek, are used as a case study for which the model is applied. These rain-dominated intermittent streams in southwestern Washington are dominated by the region's Mediterranean climate, which is characterized by wet, mild winters and dry, warm summers. They flow approximately 19 km in total length, exclusively through private land. Elevation of the creeks range from 5-210 m above
sea level and there are relatively few slopes > 4\%, with the steepest areas occurring abruptly as a result of beaver dams and log jams. By late summer, much of these streams (>50\%) have become fragmented.

## Northwest Wild Fish Rescue

From May through July, ~15,000-32,000 coho salmon fry, along with some steelhead and cutthroat juveniles, are collected from tributaries throughout the East Fork Lewis River (Mason Creek, Tsugawa Creek , Rock Creek, Mill Creek) and Salmon Creek sub-basins (Mill Creek). The fish are then reared for approximately 8-12 months in spring-fed raceways and released in March or April the following spring. Fish are released into 12 different tributaries within the EF Lewis River and Salmon River sub-basins, but not necessarily back into the same stream from which they were removed. In the model, I represent NWFR in one scenario of parameter combinations but also explore scenarios representing a range of conditions.

## Model Application

Although I use NWFR as a case study, the model is flexible, representing a range of environmental conditions and rescue interventions that can be applied to a variety of systems and rescue programs. I explored numerous combinations of freshwater rearing capacities for summer and winter, the range of which were influenced by the study system yet are applicable to other systems exhibiting similar capacities. Likewise, the range of rescue levels (i.e., nu mber of fish rescued) was influenced by the study program but combined with differing captive rearing durations can still be applied to a variety of rescue programs, not just NWFR. The only
parameter that is specific to the study system is the capacity for eggs (i.e., total spawning habitat, discussed in spawner to fry section below). In the simulations, spawning habitat was not a limiting factor, which supports the common assumption that only a small number of coho salmon adults are needed to fully "seed" a stream (Quinn, 2018). Consequently, model results may be less applicable to systems where spawning habitat is a constraint on production.

## Coho Salmon Life Cycle

The model framework imitates a three-year coho salmon life cycle. Coho salmon in the Lower Columbia River (LCR) spawn in tributaries during late November through early January (Hirose, 1983). After incubating in the gravel over winter, fry emerge in spring and hold in slow margins or off-channel habitat until late spring to early summer when they move into thalweg habitat which serves as refuge during low flow periods (Hodges \& Magoulick, 2011). During the winter, parr are thought to move back into slow, off-channel habitat which protect them from high flows (Bustard \& Narver, 1975; Hartman \& Brown, 1987; Nickelson et al., 1992). Winter has often been cited as the factor limiting smolt production (Nickelson, 1998), though some argue that summer rearing capacity limits production (Bradford, Taylor \& Allan, 1997;

Grantham et al., 2012). The following spring, smolt migrate downstream to the ocean. Generally, coho salmon rear in marine waters for 1.5 years before returning to spawn in their natal streams during fall or early winter (Quinn, 2018). A portion of male adult salmon are precocious males (known as jacks) that return to spawn after spending less than one year in the ocean. Spawning surveys conducted between 2002-2005 in Oregon found that jacks comprise $3-23 \%$ of all male spawners but jack proportions vary among populations within the LCR
(Suring, Brown \& Moore, 2006). To reduce model complexity, all spawning coho in the model were assumed to be age 3 given that the effects of cohort dynamics on resilience was beyond the scope of the model.

The model framework reflects the basic life cycle of coho salmon and simulates abundance across five serial life stages: spawner, fry, parr, smolt, and adult (Table 1). Wild fish move sequentially through each life stage and are subjected to corresponding survival processes. I simulated fish rescue as an alternative pathway between the fry and smolt life stages and examined two captive rearing durations: short- and long-term rescue. For the shortterm scenario, fish are held in captivity only during the season of wet ted habitat contraction, which was considered as the summer fry-to-parr survival stage. For long-term rescue, fish are held for roughly a year through both the fry-to-parr (i.e., summer) and the parr-to-smolt (i.e., winter) stages (Figure 1).

Table 1. State variables in the life-cycle model.

| State Variables | Description |
| :--- | :--- |
| Fry | Fish that hatch and survive through first spring in natal stream and are <br> then rescued |
| Parr | Rescued fish surviving through first summer in captivity (short-term <br> rescue only) <br> Parr <br> W |
| Smolt $_{r}$ | Wild fish surviving through first summer in natal stream <br> Fish surviving rescue activities, including time in captivity, up until <br> release as smolts |
| Smolt | Wild fish surviving through first winter in freshwater; before seaward <br> migration |
| Adult | Fish surviving seaward migration and first summer in marine |
| Spawner | environment <br> Rescued and wild adults surviving a full year in marine environment <br> from end of firstsummer and migrating back to natal stream to spawn |



Figure 1. Box and arrow diagram of the life-cycle model with long-term rescue depicted in orange.

## Model Functions

Survival was represented by the transition of individuals from one life stage to the next. The transition between the spawner and fry life stages include $d$ fecundity in addition to survival. The survival probabilities for each transition (explained below) were used in a binomial distribution to incorporate demographic stochasticity to the simulated number of individuals at the subsequent life stage (Nickelson \& Lawson, 1998). Binomial distributions are expressed as

$$
\begin{equation*}
N_{s+1} \sim \operatorname{Binom}\left(N_{s}, S_{s}\right) \tag{1}
\end{equation*}
$$

where, $S_{s}$ is the probability of surviving from life stage $s$ to life stage $s+1, N_{s}$ is the number of individuals at life stage $s$, and $N_{s+1}$ is the number of individuals at life stage $s+1$.

Three general methods were used to calculate survival. The Beverton-Holt function was used for freshwater survival (Moussalli \& Hilborn, 1986), a beta distribution was used for marine survival (Hill, Botsford \& Hastings, 2003), and a constant parameter value was used for survival in captivity (NWFR, personal communication).

## Freshwater Survival

Mean survival within the freshwater portion of the model (from spawner to smolt) was density-dependent (Jonsson, Jonsson \& Hansen, 1998) and represented by sequential Beverton-Holt functions. The classic density-dependence equation is expressed as

$$
\begin{equation*}
\bar{N}_{S+1}=\frac{N_{S}}{\frac{1}{p_{S}}+\frac{N_{S}}{K_{S}}} \tag{2}
\end{equation*}
$$

where, $N_{s}$ is the number of individuals at life stage $s, P_{s}$ is the productivity from life stage $s$ to life stage $s+1$, and $K_{s}$ is the habitat capacity between life stage $s$ to life stage $s+1$ (Moussalli \& Hilborn, 1986). With simple algebra, the equation can be rearranged and solved for survival, where $X_{s}$ is the survival from life stage $s$ to life stage $s+1$.

$$
\begin{equation*}
X_{S}=\frac{\bar{N}_{S+1}}{N_{S}}=\frac{1}{\frac{1}{p_{S}}+\frac{N_{S}}{K_{S}}} \tag{3}
\end{equation*}
$$

## Marine Survival

Mean survival within the marine portion of the model (from smolt to spawner) was density-independent (Jonsson et al., 1998). These state transitions used a beta distribution to produce survival probability (Hill et al., 2003). Beta distributions are expressed as

$$
\begin{align*}
& X_{S} \sim \operatorname{Bet} a(\alpha, \beta)  \tag{4}\\
& \alpha=\left(\frac{\bar{x} *(1-\bar{x})}{\sigma^{2}}-1\right) * \bar{x} \\
& \beta=\left(\frac{\bar{x} *(1-\bar{x})}{\sigma^{2}}-1\right) *(1-\bar{x})
\end{align*}
$$

where, $\bar{x}$ is the mean survival from life stage $s$ to life stage $s+1$ and $\sigma$ is the standard deviation of the survival estimates from life stage $s$ to life stage $s+1$ (Johnson, Kotz \& Balakrishman, 1995). Both $\bar{x}$ and $\sigma$ are calculated from gathered literature values (Table 2).

## Captive Survival

The controlled captive environment eliminates mortality from starvation and predation resulting in high survival rates. Northwest Fish Rescue reports a $98 \%$ survival rate (NWFR, personal communication), which was used in the simulations. This survival level was comparable to the $95.3 \%$ mean survival rate observed in four hatcheries in Washington (WDFW, unpublished data); however, other fish rescue captive rearing facilities have had issues
with cannibalism or constraints of drought on fish rearing capacity, which may result in lower survival rates (Lopez Arriaza et al., 2017).

Table 2. Model parameters, theirvalues, and references for these values.

| Parameter | Description | Value | Reference(s) |
| :---: | :---: | :---: | :---: |
| $f$ | Fecundity | 2500 |  |
| $s$ | Sex ratio | 0.5 |  |
| $p_{e}, p_{s}, p_{w}$ | Egg, summer, and winter maximum productivity | 0.429, 0.26, 0.9 | Nickelson, 1998; <br> Bradford et al., 2000 |
| $K_{e}, K_{S}, K_{w}$ | Egg, summer, and winter capacity | $\begin{aligned} & 902,500 ; 5,000- \\ & 45,000 ; 5,000- \\ & 45,000 \end{aligned}$ | Seetext (spawnerto fry; fry to parr; parr to smolt) |
| $\bar{x}_{c}$ | Mean captivity survival | 0.98 | NWFR, personal communication |
| $\bar{x}_{w e}$ | Mean wild early marine survival | 0.098 | Bradford 1995 |
| $\bar{x}_{r e}$ | Mean rescued early marine survival | $\begin{aligned} & 0.0196,0.0392, \\ & 0.0588,0.0784, \\ & 0.0980 \end{aligned}$ | See text (rescued fish) |
| $\bar{x}_{l}$ | Mean late marine survival (all fish) | 0.15 |  |
| $\sigma_{r e}$ | Rescued early marine survival standard deviation | 0.01 |  |
| $\sigma_{w e}$ | Wild early marine survival standard deviation | 0.01 |  |
| $\sigma_{l}$ | Late marine survival standard deviation (all fish) | 0.01 |  |
| $E$ | Encounter probability at low fry density | 0.4 |  |

## Spawner to Fry

Simulations were initialized using an arbitrary spawner abundance of 500 based loosely on the number of coho salmon spawners that would be expected in a stream the size of Mason and Tsugawa Creeks. To calculate egg abundance, the spawner abundance in each generation
was multiplied by the sex ratio ( 0.5 proportion of females) and fecundity ( 2500 eggs/female). This egg abundance was then used in the Beverton-Holt equation as the $N_{s}$ value and used to calculate the number surviving to fry. The egg capacity ( $K_{e}$ ) used in the Beverton-Holt equation is calculated by multiplying stream length for the focal system (19 km) by a regional estimate for maximum spawner density (19 female/km; Bradford, Myers \& Irvine, 2000) and fecundity (2500 eggs/female). This value remained unchanged throughout the simulations. Thus, the quantity of spawning habitat, (i.e., capacity for egg-to-fry rearing) was representative of a subcomponent of a catchment consisting of small tributary streams.

## Fry to Parr

Simulated fry that were not diverted to the rescue pathway reared over summer in the natural environment.I explored a range of parameters that represent different levels of rearing capacity due to either different levels of drought or intrinsic differences among streams in their summer habitat and hydrology. Mean summer rearing capacities ( $K_{s}$ ), ranging from 5000 to 45000 in increments of 5000, were used in the simulations and represented severe to minimal constraints of habitat contraction on survival, respectively (Table 2). The range of values were consistent with those observed in Mason and Tsugawa Creeks during the summer of 2017 (WDFW, unpublished data).

## Parr to Smolt

Parr reared over winter to reach the smolt life stage. To make direct comparisons between summer and winter limitations, winter rearing capacity ( $K_{w}$ ) used the same range of
values ( $K_{w}=5000$ to 45000 ) that were used for summer rearing capacity. A winter rearing capacity was randomly chosen from this range and used in the model for each simulation.

## Smolt to Adult

Early marine survival of wild fish ( $\bar{x}_{w e}$ ) encompassed the time of downstream migration through the first summer in the ocean. For this transition, I drew survival probabilities from a beta distribution (eq. 4-6) with mean early marine survival of $9.8 \%$ (Bradford, 1995) for each time step.

## Adult to Spawner

The state transition between adult and spawner encompassed the second year in the marine environment, from the end of the first summer through upstream migration to spawn. A single late marine survival ( $\bar{x}_{l}$ ) was drawn from a beta distribution with a mean of $15 \%$ for each time step and applied to all fish, rescued and wild.

## Rescued Fish

Fish rescue removed fish from the stream at the fry life stage. At this point, rescued fry move to the parr life stage via short-term captivity survival or directly into the smolt life stage via long-term captivity survival. The short-term rescue scenario replaces the summer survival applied to wild fish, and the long-term rescue scenario circumvents both the summerand winter survival applied to wild fish (Figure 1). While captivity survival is much higher than either summer or winter survival experienced by wild counterparts, rearing in artificial conditions
(such as hatcheries) has been shown to alter fish behavior (Nickelson, 1986; Berejikian et al., 1995) and survival (Leider et al., 1990) in subsequent life stages once returned to the natural environment. Although not bred in captivity, rescued fish spend much of their freshwater life stages in artificial conditions, and I assumed this could negatively affect smolt-to-adult return rates. Since it is unknown how rescuing fish impacts later life stages, I explored a range of survival penalties based on empirical estimates of how marine survival differs between hatchery and wild fish. Reported estimates of marine survival of hatchery fish compared to wild fish range from ~37-100\% (Jonsson et al., 2003; Kallio-Nyberg et al., 2004; Jokikokko et al., 2006; Hyvärinen \& Rodewald, 2013). The highest comparative survival (i.e., 100\%) occurred for captively reared fish released as parr (Jokikokko et al., 2006), while the lowest comparative survival was associated with fish released as 1 year-olds smolts (Jonsson et al., 2003). Saloniemi et al. (2004) reported that wild smolts had 4.5 times highersurvival than hatchery-reared counterparts, and considering the wild marine survival rate of 0.098 , this represents $\sim 22 \%$ comparative marine survival for hatchery fish. Lower values could represent the combined effects of post-release mortality, reduced marine survival and reduced homing. In the model, early marine survival of rescued fish was calculated by multiplying early marine survival of wild fish by a penalty value. I explored penalties of $0.2,0.4,0.6,0.8$ and 1 , where 0.2 was the most severe penalty and 1 represented no penalty. Though it is possible that the magnitude of penalty increases with the duration of rescue, I did not explore this potential relationship.

## Rescue Level

The attempted rescue level remained constant for every generation of a given simulation, ranging from 5000 to 35000 in increments of 10000 . Since fry abundance may fall below the attempted rescue level, I used a functional response to model the realized rescue level as a function of fry abundance. I used a capped linear (i.e., "hockey-stick") functional response, in which $40 \%$ of the total fry are captured, up to the attempted rescue level (i.e., the capacity for captive rearing). In other words, when fry abundance was less than the captive rearing capacity, $40 \%$ of the available fry were rescued. This avoids rescuing an unrealistic percentage of available fry.

## Model Output

Two main response metrics were used to explore the impact of fish rescue on coho salmon population dynamics: spawner abundance and extinction risk. These metrics are commonly used for various salmonid life-cycle models within the Columbia River Basin (Zabel et al., 2013). Spawner abundance was calculated by taking the geometric mean of spawner abundance across all generations of a given simulation. Recruitment variability tends to be lognormal as a result of the multiplicative survival framework, so geometricmean is commonly used in life cycle models to avoid placing undue weight on outliers and to betterexplain the central tendency of the dataset (Zabel et al., 2013). Extinction risk was calculated as the percentage of generations within a simulation in which spawner abundance falls below an extinction threshold. Following the standards set by Zabel et al. (2013), the extinction threshold was set at 50 spawners.

## Scenarios Modeled

The effects of fish rescue were examined by varying rearing duration (i.e., short, long) and limiting season (i.e., the degree to which production is limited by summer or winter rearing capacity). For each scenario (outlined in table 3), rescue level and drought condition were allowed to vary. Rescue levels were setto 0, 5000, 15000, 25000, and 35000. No rescue simulations (rescue level $=0$ ) served as a baseline to which the other four rescue simulations were compared. Drought conditions were explored by adjusting the summer rearing capacity $\left(K_{s}\right)$. For low drought conditions, the capacity was higher, and for more severe drought, the capacity was lower.

Table 3. Five sets of parameter combinations are run for each rescue level (none, 5000, 15000, 25000,35000 ). Rearing duration refers to length of time in captivity, short-term or long-term, and limiting season is the season that exhibits the least amount of available habitat. Note that for no rescue simulations, rearing duration does not apply because no fish are rescued.

| Scenario | Rearing Duration | Limiting Season |
| :--- | :--- | :--- |
| 1 | short | winter |
| 2 | short | summer |
| 3 | long | winter |
| 4 | long | summer |

Each complete life cycle followed one generation through the three-yearlifespan.

Simulations spanned 33 generations (99 years), and 10,000 replicate simulations were run for each rescue level. Otherlife-cycle models in the Columbia River Basin project simulations for 100 years (Zabel et al., 2013), and I aimed for a similarly long projection timeframe. This allows for comparison among results from the different models, which is particularly helpful for
comparing extinction risks. Since several parameters were chosen at random from statistical distributions, 10,000 simulations per rescue level ensured that a sufficient number of simulations were run for each combination of parameter values to accurately summarize the central tendency of stochastic processes and explore parameter space. All modeling was done using Program R (R Core Team, 2017).

## Results

## Relative Changes in Abundance

My model explored how the positive effects of fish rescue propagated through the coho salmon life-cycle under alternative scenarios of seasonal habitat capacity and the effects of captive rearing on marine survival. I found that increasing freshwater survival through fish rescue translated into higher adult returns across most of the parameter space I explored (Figure 2). In the small watershed my simulation represented, the median adult coho salmon returns without fish rescue was 62 fish (range $=0-152$ ). In contrast, at an intermediate rescue level (collecting 15,000 fry) and intermediate penalty value (40\% reduction in marine survival), I observed the following results. For short-term rescue in summer-limited simulations, the median of adult returns among simulations was 132 fish (range $=45-198$ ), representing a 112.9\% increase in abundance compared to no-rescue simulations in the respective modeling scenario. The median adult returns for short-term rescue in winter-limited simulations was 87 fish (range $=24-190$ ), an $86 \%$ increase. For long-term rescue in summer-limited simulations, the median adult return was 225 (range $=154-327$ ) a $262.9 \%$ increase. Long-term rescue in winterlimited simulations produced a median of 227 fish (range= 160-329), a $266.1 \%$ increase. While
fish rescue increased adult returns in most contexts explored, I also found that it could have a negative effect on returns if rescue was short-term (i.e., only for summer) and the penalty on subsequent adult survival was substantial (i.e., captive reared fish had one-fifth the marine survival of wild fish) (Figure 3).


Figure 2. Geometric mean adult returns across increasing drought severity and rescue level for long-term rescue with summer-limited rearing capacity and intermediate penalty (i.e., 0.6).


Figure 3. Geometric mean adult returns across rescue level and colored by penalty ( 0.2 is most severe penalty) for each of four scenarios: long-term rescue with summer limitation, long-term rescue with winterlimitation, short-term rescue with summer limitation, and short-term rescue with winter limitation. The median value of no rescue simulations is represented by a dashed line.

## Relative Changes in Extinction Risk

The positive effect of fish rescue on freshwatersurvival almost always translated to decreased extinction risk and only increased this risk in short-term rescue simulations (Figure 4). In the small tributary I simulated, extinction risk without fish rescue ranged from 0-100\% with a median of $18.2 \%$ and mean of $38.0 \%$ for winter-limited simulations and a median of $21.2 \%$ and mean of $39.8 \%$ for summer-limited simulations. For long-term simulations, all combinations of rescue levels and penalty values had a $0 \%$ median extinction risks. For shortterm simulations, increased extinction risk occurred for all rescue levels with penalty of 0.2. For
summer-limited, short-term rescue with penalty of 0.2, median extinction risk was $36.4 \%$ for rescue level of 5000 and $57.6 \%$ for rescue levels of 15000,25000 , and 35000 . For winterlimited, short-term rescue with penalty of 0.2 , median extinction risk was $69.7 \%, 90.9 \%, 87.9 \%$, and $93.9 \%$ for rescue levels of 5000, 15000, 25000, and 35000, respectively. A higher median extinction risk compared to no rescue simulations also occurred for winter-limited, short-term simulations when the penalty was 0.4 and rescue level was 15000 and 25000. Extinction risk in these scenarios was $27.3 \%$ and $19.7 \%$, respectively.


Figure 4. Median extinction risk across rescue level and colored by penalty for each of four scenarios: long-term rescue with summer limitation, long-term rescue with winter limitation, short-term rescue with summer limitation, and shot-term rescue with winter limitation. The median value of no rescue simulations is represented by a dashed line.

Long-term rescue scenarios exhibited extinction risk $>50 \%$ for $1.1 \%$ of summer-limited simulations and $0.9 \%$ for winter-limited systems. Forshort-term rescue scenarios, extinction risk $>50 \%$ occurred for $10.1 \%$ and $27.2 \%$ of summer- and winter-limited simulations, respectively. Comparatively, no rescue scenarios exhibited extinction risk >50\% for $40.3 \%$ of summer-limited simulations and $37.6 \%$ of winter-limited simulations.

## Effect of Rescue Penalty

Rescue penalty, when expressed as the marine survival rate of rescued fish relative to that of wild fish, had a near linear effect on adult returns (Figure 5). The slope of this relationship was larger for long-term rescue (regardless of the season of habitat limitation) and less positive forshort-term rescue (within this scenario it was least positive for winter-limited simulations). Short-term rescue scenarios resulted in fewer adult returns compared to longterm rescue scenarios, and simulations with high penalty (i.e., 0.2 and 0.4 ) resulted in negative effects on adult returns (Figure 3). Thus, the effect size of the penalty was strongest in the longterm rearing scenario, but the effects were most important in the short-term rearing scenario because it reduced abundance below that of no rescued scenarios (i.e., fish rescue can have a net negative effect).


Figure 5: Median adult returns across penalty level and colored by rescue scenarios. Dashed horizontal line is median adult returns for all no rescue simulations.

## Effect of Rescue Level

Rescue level (i.e., captive rearing capacity assuming a $40 \%$ collection efficiency) interacted with the duration of rescue and penalty level to affect abundance. For long-term rescue, rescue level had a positive effect on adult returns that were strongly dampened by increased penalty. For short-term rescue, the positive effect of rescue level was weaker and became negative when penalty was high (i.e., 0.2). For short-term rescue in winter-limited systems, the effect of rescue level was only positive when penalty was low (>0.6).

## Discussion

Water overallocation and climate change are exacerbating the duration and severity of stream fragmentation, threatening the productivity and viability of salmonid populations. In basins where water scarcity issues are unlikely to be resolved, there is an urgent need for management interventions that ameliorate the effects of drought on fish survival. I developed a simulation model to explore how seasonally assisted movement to captive rearing facilities could function as a drought adaptation tool. My findings indicate that fish rescue could have strong positive effects on adult returns and population viability, yet they also identify potential risks (i.e., forshort-term rescue if rescue strongly reduces subsequent marine survival). Further, due to the manner by which fish rescue affects adult returns in relation to drought severity, I question whether fish rescue actually functions as a climate adaptation tool or whetherit would more accurately be described as stock enhancement, which is already ubiquitous for salmon stocks across the Pacific rim (Mahnkenl et al., 1998). I would expect a climate adaptation tool to lessen the negative effect of drought on fish abundance, making the slope of this relationship less negative. However, the primary effect of fish rescue was on the intercept of this relationship, meaning fish rescue boosted returns across all drought levels (including no drought) and did not do so in proportion to the severity of drought (Figure 2). Fish rescue functioned most closely to a drought adaptation tool when summer capacity was limiting, and fish were held for summeronly. This short-term rescue could produce negative effects on fish returns and population viability if winter capacity was limiting and the effects of captive rearing on marine survival were strongly negative (Figure 6). This worst-case scenario is not implausible; there is rarely data to confirm that drought-impacted streams are actually limited
by summer rather than winter rearing habitat and the negative effects of captive rearing on fitness may be stronger than I anticipated due to mechanisms beyond the scope of my model, such as intergenerational effects of captive rearing (Araki et al. 2007). The strongest positive effect of fish rescue occurred when fish were held long-term, over both summer and winter, in systems where summer capacity was limiting. Captive rearing over two seasons increases survival so strongly compared to that of the wild that it offset even the harshest scenarios of marine survival penalty that were considered. In summary, fish rescue could be an effective way to maintain higher returns of salmon in the face of exacerbated stream drying, but its effect on population dynamics more closely resembles that of hatchery supplementation rather than drought mitigation and could have negative effects in certain contexts that are difficult to rule out.

Model results reflect the particularly strong effect size of captive rearing on freshwater survival. In the wild, freshwatersurvival is density dependentand can be rather low due to high mortality during the post-emergence transition to feeding (Armstrong \& Nislow, 2006), during periods of summer base flow (Hwan et al. 2018), and during periods of winter food limitation (Biro et al. 2005) or flooding (Bell, Duffy \& Roelofs, 2001). Survival in the wild varies, but recent work found summersurvival rates of $\sim 25 \%$ for coho salmon in drought-fragmented streams (Hwan et al. 2018) and a variety of winterstudies report survival rates ranging from ~5-60\% (Quinn \& Peterson, 1996; Solazzi et al., 2000; Ebersole et al., 2006, 2009; Wigington et al., 2006). Thus fish rescue likely generates a $\sim 4$-fold increase in survival for summer-only programs, and potentially a> 10-fold increase in cumulative survival for year-round rescue programs. The extent to which this dramatic increase in freshwater survival translates into
increased adult returns depended on three interacting factors: (1) the penalty of rescue on
survival at subsequent life stages (2) the captive rearing duration (i.e., summer only vs. summer and winter), and (3) the season that limited rearing capacity (i.e., summervs, winter-limited).


Figure 6. The difference in adult returns for short-term, moderate rescue (15,000 fish) and no rescue simulations. Kratio represents seasonal limitations, where values $<0.5$ indicate a summer-limited system, and values $>0.5$ indicate a winter-limited system. Penalty refers to the reduction in marine survival as a consequence of rescue. The only negative effect of rescue (darkest two colors below the white line) occurred for the most severe penalties ( $\sim 0.4-0.2$ ) where adult returns for rescue simulations were less than those for no rescue.

Fish rescue had the strongest effect when fish were held for summer and winter (i.e., one year). This was not only because survival rates were elevated across an additional life stage, but also because the captive reared fish were released after the density depe ndent portion of the life cycle. Releasing fish after summer rearing in captivity causes them to compete with wild fish during the winter. This can reduce population productivity if the rescued fish have substantially lower survival as adults, particularly if overwinter habitat is limiting, in which case summer rescue increases the overabundance of fish headed into the winter survival bottleneck and effectively replaces wild fish with captive reared indi viduals that have lower fitness as adults. It is plausible that a stream could exhibit conspicuous summer fragmentation yet also have poor overwinter habitat that is the actual bottleneck to production. Indeed the survival rates reported by Hwan et al. (2018) for drought years in fragmented streams exceed many published values for winter survival. The negative effects of fish rescue that we re observed would likely be stronger if overwinter survival was size-dependent, as is often reported (Quinn \& Peterson, 1996; Ebersole et al., 2006; Pess et al., 2011). Fish have the physiological capacity to feed at much higher levels than they achieve in the wild (Armstrong \& Schindler, 2011), so captive rearing with ad libitum feeding can dramatically increase growth and condition. It is possible that summer rescue results in large individuals that outcompete wild fish during winter, but I did not considersuch scenarios because I know of no studies measuring how seasonal captivity affects size-dependent winter survival.

While there is strong debate over the role of hatcheries in salmon conservation, fish rescue programs have not attracted similar controversy, partly because of theirsmall scale, but likely also because they are putatively drought-adaptation programs rather than tools for
fisheries enhancement. However, the plot of adult returns as a function of drought severity and rescue level (Figure 2 ) suggests that the quantitative effects of fish rescue are not consistent with drought adaptation. For example, in the case of long-term rescue in a summer-limited system (i.e., the scenario that Northwest Fish Rescue may represent), rescuing fish always increases returns, but it does not reduce the negative effect of drought on returns. This is because fish rescue creates a novellife-cycle pathway that is insensitive to the constraints of freshwater rearing but does not improve conditions for fish in the natural life-cycle pathway. Even at rescue levels lowerthan those currently in practice by existing rescue programs such as NWFR, captive survival dominates the life-cycle pathway and captive-reared individuals become a greater proportion of the population, such that the threat of drought to wild fish makes a small reduction in total fish abundance. Thus, fish rescue would more accurately be described as a form of hatchery enhancement to fish production, rather than a drought mitigation tool. In the mid-Columbia River Basin, hatchery programs are categorized as eitherintegrated conservation programs, safety-net programs, or harvest augmentation programs (Hillman et al., 2013). I argue that fish rescue emulates the integrated conservation type program as it increases production and more closely aligns with this hatchery-type program than a drought mitigation tool.

The severity of drought impacts varies among locations and through time, and managers will likely weigh the costs and benefits of rescue differently depending on their local context. When fish become stranded or isolated in poor quality habitat, simply relocating fish to adjacent continuous habitat may be a low cost and low risk option. This approach is appealing because it minimizes fish handling and reduces any known or unknown consequences of
captive rearing. However, it may not be effective if summerrearing capacity is limiting because moving fish into already occupied areas will increase density and could reduce growth and survival if these areas are density dependent. However, this method could be effective if fry spatial distributions are strongly limited by dispersal and there are underutilized habitats that translocation could help fill to capacity. Intermittent streams contain crucial rearing habitat for juvenile salmonids (Wigington et al., 2006) and pockets of fragmented habitat may retain sufficient size and quality (Hwan et al., 2018), in which case, intervention may not be needed. More research on the fate of fish in fragmented habitats and the ability of fish to disperse across networks prior to stream drying would help elucidate the potential benefits of in-stream translocations.

If continuous, non-fragmented habitat is not readily available, captively rearing fish during the summer low flow conditions could be the only rescue option. This strategy is intuitively attractive because it reduces time in captivity, but it could also exacerbate density dependence if winter rearing capacity is limiting, which is often difficult to determine. There is uncertainty overhow rescued fish interact with wild fish, though there is some indication that captively reared fish may exhibit greater size, aggression, and dominance that outweighs any prior resident advantage of naturally-reared fish (Rhodes \& Quinn, 1998). However, a study with Atlantic salmon suggests that naturally-reared wild fish may dominate over wild fish that have been hatchery-reared (Metcalfe, Valdimarsson \& Morgan, 2003). The extent of domestication on dominance behavior depends on the captive rearing environment (Ruzzante, 1994). Additionally, minimizing hatchery time may reduce behavioral differences between wild
and hatchery-reared fish (Jackson \& Brown, 2011), and captive rearing duration may influence the magnitude of rescue effects.

Alternatively, holding fish for both summer and winter may be an option when both summer and winter rearing capacity are low due to water overallocation and habitat degradation. This strategy effectively acts as a low and high flow rescue program and had the highest numerical response in adult returns. However, long-term rescue may increase survival and capacity far above natural levels, and may lead to multiple generations affected by captive rearing. Araki et al. (2007) found that fish raised for one generation in a hatchery had 62.5\% of the fitness of wild fish, dropping 37.5\% per captive-reared generation. If similartrends exist for rescued fish, then continually rescuing the offspring of previously rescued fish could magnify the negative effects of rescue. This may manifest in lowerantipredator response (Jackson \& Brown, 2011), increased stray rates (Schroeder, Lindsay \& Kenaston, 2001), and lower reproductive success (Thériault et al., 2011). Rescue of high fractions of the population could also inhibit natural selection for phenotypes that increase survival in the wild during drought (e.g., by migrating to different summerhabitats) (Everest, 1973), but rescue may keep a population viable until conservation efforts can resolve the underlying limitation to natural production.

Fish rescue is a tempting strategy for managers responsible for dwindling salmonid populations threatened by ecological drought. The sight of ESA-listed fish dying en masse as fragmented pools dry up makes inaction seem irresponsible. However, this history of conservation is filled with decisions that seemed urgently needed at the time, but ultima tely proved ineffective and often difficult to reverse. I hope that the model results illustrate the
potential benefits and risks of fish rescue to managers considering this intervention and help them interpret the results of existing programs. Managers should expect that fish rescue can increase adult returns, supporting the claims made by groups such as Northwest Fish rescue. However, as Lopez Arriaza et al. (2017) found, rescue can lead to lead to the majority of individuals being captively reared and poses risks to the wild populations if captively reared fish have reduced fitness and compete with wild fish. Fish rescue for multiple seasons functions more like a hatchery enhancement program than a drought adaptation tool, so its potential effects on adult returns should be evaluated with caution. However, as ecological drought intensifies due to climate change, some stocks will likely go extinct without management intervention and forms of fish rescue may be the only way to method available for maintaining the viability and genetic diversity of these populations.

## Bibliography

Araki H., Cooper B. \& Blouin M.S. (2007). Genetic Effects of Captive Breeding Cause a Rapid, Cumulative Fitness Decline in the Wild. Science 318, 100-103.
https://doi.org/10.1126/science. 1145621
Armstrong J.B. \& Schindler D.E. (2011). Excess digestive capacity in predators reflects a life of feast and famine. Nature 476, 84-87. https://doi.org/10.1038/nature10240

Armstrong J.D. \& Nislow K.H. (2006). Critical habitat during the transition from maternal provisioning in freshwaterfish, with emphasis on Atlantic salmon (Salmo salar) and brown trout (Salmo trutta). Journal of Zoology 269, 403-413. https://doi.org/10.1111/j.1469-7998.2006.00157.x

Beier P. \& Brost B. (2010). Use of Land Facets to Plan for Climate Change: Conserving the Arenas, Not the Actors: Land Facets for Climate-Change Planning. Conservation Biology 24, 701-710. https://doi.org/10.1111/j.1523-1739.2009.01422.x

Bell E., Duffy W.G. \& Roelofs T.D. (2001). Fidelity and Survival of Juvenile Coho Salmon in Response to a Flood. Transactions of the American Fisheries Society 130, 450-458

Berejikian B.A., Tezak E.P., Schroder S.L., Flagg T.A. \& Knudsen C.M. (1995). Competitive Differences between Newly Emerged Offspring of Captive-Reared and Wild Coho Salmon. 8

Bowkett A.E. (2009). Recent Captive-Breeding Proposals and the Return of the Ark Concept to Global Species Conservation. Conservation Biology 23, 773-776. https://doi.org/10.1111/j.1523-1739.2008.01157.x

Bradford M.J. (1995). Comparative review of Pacific salmon survival rates. Canadian Journal of Fisheries and Aquatic Sciences 52, 1327-1338. https://doi.org/10.1139/f95-129

Bradford M.J., Myers R.A. \& Irvine J.R. (2000). Reference points for coho salmon ( Oncorhynchus kisutch) harvest rates and escapement goals based on freshwater production. Canadian Journal of Fisheries and Aquatic Sciences 57, 677-686. https://doi.org/10.1139/cjfas-57-4-677

Bradford M.J., Taylor G.C. \& Allan A.A. (1997). Empirical Review of Coho Salmon Smolt Abundance and the Prediction of Smolt Production at the Regional Level. 16

Brown C. \& Day R.L. (2002). The future of stock enhancements: lessons for hatchery practice from conservation biology. Fish and Fisheries 3, 79-94. https://doi.org/10.1046/j.14672979.2002.00077.x

Bustard D.R. \& Narver D.W. (1975). Preferences of Juvenile Coho Salmon ( Oncorhynchus kisutch ) and Cutthroat Trout ( Salmo clarki ) Relative to Simulated Alteration of Winter Habitat. Journal of the Fisheries Research Board of Canada 32, 681-687. https://doi.org/10.1139/f75-087

Cheung W.W.L., Lam V.W.Y., Sarmiento J.L., Kearney K., Watson R., Zeller D., et al. (2010). Large-scale redistribution of maximum fisheries catch potential in the global ocean under climate change: CLIMATE CHANGE IMPACTS ON CATCH POTENTIAL. Global Change Biology 16, 24-35. https://doi.org/10.1111/j.1365-2486.2009.01995.x

Crozier L.G., McClure M.M., Beechie T., Bograd S.J., Boughton D.A., Carr M., et al. (2019). Climate vulnerability assessment for Pacific salmon and steelhead in the California Current Large Marine Ecosystem. PLOS ONE 14, e0217711. https://doi.org/10.1371/journal.pone. 0217711

Dore M.H.I. (2005). Climate change and changes in global precipitation patterns: What do we know? Environment International 31, 1167-1181. https://doi.org/10.1016/j.envint.2005.03.004

Ebersole J.L., Colvin M.E., Wigington P.J., Leibowitz S.G., Baker J.P., Church M.R., et al. (2009). Hierarchical Modeling of Late-Summer Weight and Summer Abundance of Juvenile Coho Salmon across a Stream Network. Transactions of the American Fisheries Society 138, 1138-1156. https://doi.org/10.1577/T07-245.1

Ebersole J.L., Wigington P.J., Baker J.P., Cairns M.A., Church M.R., Hansen B.P., et al. (2006). Juvenile Coho Salmon Growth and Survival across Stream Network Se asonal Habitats. Transactions of the American Fisheries Society 135, 1681-1697. https://doi.org/10.1577/T05-144.1

Everest F.H. (1973). Everest 1973 Ecology and management of summer steelhead in the Rogue River.pdf. Oregon State Game Commission, Corvallis, OR.

Falke J.A., Fausch K.D., Magelky R., Aldred A., Durnford D.S., Riley L.K., et al. (2011). The role of groundwater pumping and drought in shaping ecological futures for stream fishes in a dryland river basin of the western Great Plains, USA. Ecohydrology 4, 682-697. https://doi.org/10.1002/eco. 158

Galloway B.T., Muhlfeld C.C., Guy C.S., Downs C.C. \& Fredenberg W.A. (2016). A Framework for Assessing the Feasibility of Native Fish Conservation Translocations: Applications to Threatened Bull Trout. North American Journal of Fisheries Management 36, 754-768. https://doi.org/10.1080/02755947.2016.1146177

Grantham T.E., Newburn D.A., McCarthy M.A. \& MerenlenderA.M. (2012). The Role of Streamflow and Land Use in Limiting OversummerSurvival of Juvenile Steelhead in

California Streams. Transactions of the American Fisheries Society 141, 585-598. https://doi.org/10.1080/00028487.2012.683472

Hamlet A.F., Elsner M.M., Mauger G.S., Lee S.-Y., Tohver I. \& Norheim R.A. (2013). An Overview of the Columbia Basin Climate Change Scenarios Project: Approach, Methods, and Summary of Key Results. Atmosphere-Ocean 51, 392-415. https://doi.org/10.1080/07055900.2013.819555

Hartman G.F. \& Brown T.G. (1987). Use of Small, Temporary, Floodplain Tributaries by Juvenile Salmonids in a West Coast Rain-Forest Drainage Basin, Carnation Creek, British Columbia. Canadian Journal of Fisheries and Aquatic Sciences 44, 262-270. https://doi.org/10.1139/f87-035

Harvey B.C., Nakamoto R.J. \& White J.L. (2006). Reduced Streamflow Lowers Dry-Season Growth of Rainbow Trout in a Small Stream. Transactions of the American Fisheries Society 135, 998-1005. https://doi.org/10.1577/T05-233.1

Hewitt N., Klenk N., Smith A.L., Bazely D.R., Yan N., Wood S., et al. (2011). Taking stock of the assisted migration debate. Biological Conservation 144, 2560-2572.
https://doi.org/10.1016/j.biocon.2011.04.031
Higgins P.S. \& Bradford M.J. (1996). Evaluation of a Large-Scale Fish Salvage to Reduce the Impacts of Controlled Flow Reduction in a Regulated River. North American Journal of Fisheries Management 16, 666-673. https://doi.org/10.1577/15488675(1996)016<0666:EOALSF>2.3.CO;2

Hijmans R.J. \& Graham C.H. (2006). The ability of climate envelope models to predict the effect of climate change on species distributions. Global Change Biology 12, 2272-2281. https://doi.org/10.1111/j.1365-2486.2006.01256.x

Hill M.F., Botsford L.W. \& Hastings A. (2003). The effects of spawning age distribution on salmon persistence in fluctuating environments. Journal of Animal Ecology 72, 736-744. https://doi.org/10.1046/j.1365-2656.2003.00745.x

Hillman T.W., Kahler T., Mackey G., Muraukas J., Murdoch A., Murdoch K., et al. (2013). Monitoring and evaluation plan for PUD hatchery programs. Wenatchee, Washington.

Hirose P. (1983). Lower Columbia River coho spawning ground surveys, 1967-82. Oregon Department of Fish and Wildlife.

Hodges S.W. \& Magoulick D.D. (2011). Refuge habitats for fishes during seasonal drying in an intermittent stream: movement, survival and abundance of three minnow species. Aquatic Sciences 73, 513-522. https://doi.org/10.1007/s00027-011-0206-7

Hoffmann A.A. \& Sgrò C.M. (2011). Climate change and evolutionary adaptation. Nature 470, 479-485. https://doi.org/10.1038/nature09670

Humphries P. \& Baldwin D.S. (2003). Drought and aquatic ecosystems: an introduction. Freshwater Biology 48, 1141-1146. https://doi.org/10.1046/j.1365-2427.2003.01092.x

Hwan J.L. \& Carlson S.M. (2016). Fragmentation of an Intermittent Stream During Seasonal Drought: Intra-Annual and Interannual Patterns and Biological Consequences: Stream Contraction During Seasonal Drought. River Research and Applications 32, 856-870. https://doi.org/10.1002/rra. 2907

Hwan J.L., Fernández-Chacón A., Buoro M. \& Carlson S.M. (2018). Dry season survival of juvenile salmonids in an intermittent coastal stream. Canadian Journalof Fisheries and Aquatic Sciences 75, 746-758. https://doi.org/10.1139/cjfas-2017-0026

Hyvärinen P. \& Rodewald P. (2013). Enriched rearing improves survival of hatchery-reared Atlantic salmon smolts during migration in the River Tornionjoki. Canadian Journalof Fisheries and Aquatic Sciences 70, 1386-1395. https://doi.org/10.1139/cjfas-2013-0147

Jackson C.D. \& Brown G.E. (2011). Differences in antipredator behaviour between wild and hatchery-reared juvenile Atlantic salmon ( Salmo salar ) under seminatural conditions. Canadian Journal of Fisheries and Aquatic Sciences 68, 2157-2166. https://doi.org/10.1139/f2011-129

Jaeger K.L., Olden J.D. \& Pelland N.A. (2014). Climate change poised to threaten hydrol ogic connectivity and endemicfishes in dryland streams. Proceedings of the National Academy of Sciences 111, 13894-13899. https://doi.org/10.1073/pnas. 1320890111

Johnson N.L., Kotz S. \& Balakrishman N. (1995). Continuous Univariate Distributions, 2nd edn. Wiley Series in Probability and Mathematical Statistics, New York.

Jokikokko E., Kallio-Nyberg I., Saloniemi I. \& Jutila E. (2006). The survival of semi-wild, wild and hatchery-reared Atlantic salmon smolts of the Simojoki River in the Baltic Sea. Journal of Fish Biology 68, 430-442. https://doi.org/10.1111/j.0022-1112.2006.00892.x

Jonsson N., Jonsson B. \& Hansen L. (1998). The relative role of density-dependent and densityindependent survival in the life cycle of Atlantic salmon Salmo salar. Journal of Animal Ecology 67, 751-762

Jonsson N., Jonsson B. \& Hansen L.P. (2003). The marine survival and growth of wild and hatchery-reared Atlantic salmon: Growth and survival of salmon. Journal of Applied Ecology 40, 900-911. https://doi.org/10.1046/j.1365-2664.2003.00851.x

Kallio-Nyberg I., Jutila E., Saloniemi I. \& Jokikokko E. (2004). Association between environmental factors, smolt size and the survival of wild and reared Atlantic salmon from the Simojoki River in the Baltic Sea. Journal of Fish Biology 65, 122-134. https://doi.org/10.1111/j.0022-1112.2004.00435.x

Katz S.L., Barnas K., Hicks R., Cowen J. \& Jenkinson R. (2007). Freshwater Habitat Restoration Actions in the Pacific Northwest: A Decade?s Investment in Habitat Improvement. Restoration Ecology 15, 494-505. https://doi.org/10.1111/j.1526-100X.2007.00245.x

Keefer M.L. \& Caudill C.C. (2014). Homing and straying by anadromous salmonids: a review of mechanisms and rates. Reviews in Fish Biology and Fisheries 24, 333-368. https://doi.org/10.1007/s11160-013-9334-6

Kihslinger R.L. (2006). Early rearing environment impacts cerebellargrowth in juvenile salmon. Journal of Experimental Biology 209, 504-509. https://doi.org/10.1242/jeb. 02019

KTVZ (2017). Fall fish rescue set to return on upper Deschutes. KTVZ
Kundzewicz Z.W., Mata L.J., Arnell N.W., DöLI P., Jimenez B., Miller K., et al. (2008). The implications of projected climate change for freshwater resources and their management. Hydrological Sciences Journal 53, 3-10. https://doi.org/10.1623/hysj.53.1.3

Labbe T.R. \& Fausch K.D. (2000). Dynamics of IntermittentStream Habitat Regulate Persistence of a Threatened Fish at Multiple Scales. Ecological Applications 10, 1774-1791

Lake P.S. (2003). Ecological effects of perturbation by drought in flowing waters. Freshwater biology 48, 1161-1172

Larned S.T., Datry T., Arscott D.B. \& Tockner K. (2010). Emerging concepts in temporary-river ecology. Freshwater Biology 55, 717-738. https://doi.org/10.1111/j.13652427.2009.02322.x

Leider S.A., Hulett P.L., Loch J.J. \& Chilcote M.W. (1990). Electrophoretic comparison of the reproductive success of naturally spawning transplanted and wild steelhead trout through the returning adultstage. Aquaculture 88, 239-252. https://doi.org/10.1016/0044-8486(90)90151-C

Linnell J.D.C., Aanes R., Swenson J.E., Odden J. \& Linnell J.D.C. (1997). Translocation of carnivores as a method for managing problem animals: a review. Biodiversity and Conservation 6, 1245-1257. https://doi.org/10.1023/B:BIOC.0000034011.05412.cd

Lockwood M.A., Griffin C.P., Morrow M.E., Randel C.J. \& Silvy N.J. (2005). SURVIVAL, MOVEMENTS, AND REPRODUCTION OF RELEASED CAPTIVE-REARED ATTWATER'S

PRAIRIE-CHICKEN. Journal of Wildlife Management 69, 1251-1258. https://doi.org/10.2193/0022-541X(2005)069[1251:SMAROR]2.0.CO;2

Lopez Arriaza J., Boughton D.A., Urquhart K. \& Mangel M. (2017). Size-conditional smolting and the response of Carmel River steelhead to two decades of conservation efforts. PLOS ONE 12, e0188971. https://doi.org/10.1371/journal.pone. 0188971

Magoulick D.D. \& Kobza R.M. (2003). The role of refugia for fishes during drought: a review and synthesis. Freshwater Biology 48, 1186-1198. https://doi.org/10.1046/j.13652427.2003.01089.x

Mahnkenl C., Ruggerone G., Waknitzl W. \& Flaggl T. (1998). A Historical Perspective on Salmonid Production from Pacific Rim Hatcheries. 1, 38-53

Mason J.C. (1976). Response of Underyearling Coho Salmon to Supplemental Feeding in a Natural Stream. The Journal of Wildlife Management 40, 775. https://doi.org/10.2307/3800576

Mawdsley J.R., O’Malley R. \& Ojima D.S. (2009). A Review of Climate-Change Adaptation Strategies for Wildlife Management and Biodiversity Conservation. Conservation Biology 23, 1080-1089. https://doi.org/10.1111/j.1523-1739.2009.01264.x

May C.L. \& Lee D.C. (2004). The Relationships among In-Channel Sediment Storage, Pool Depth, and Summer Survival of Juvenile Salmonids in Oregon Coast Range Streams. North American Journal of Fisheries Management 24, 761-774. https://doi.org/10.1577/M03073.1

Metcalfe N.B., Valdimarsson S.K. \& Morgan I.J. (2003). The relative roles of domestication, rearing environment, prior residence and body size in deciding territorial contests between hatchery and wild juvenile salmon. Journalof Applied Ecology 40, 535-544. https://doi.org/10.1046/j.1365-2664.2003.00815.x

Mote P.W. \& Salathé E.P. (2010). Future climate in the Pacific Northwest. Climatic Change 102, 29-50. https://doi.org/10.1007/s10584-010-9848-z

Moussalli E. \& Hilborn R. (1986). Optimal Stock Size and Harvest Rate in Multistage Life History Models. Canadian Journal of Fisheries and Aquatic Sciences 43, 135-141. https://doi.org/10.1139/f86-014

Nagrodski A., Raby G.D., Hasler C.T., Taylor M.K. \& Cooke S.J. (2012). Fish stranding in freshwater systems: Sources, consequences, and mitigation. Journal of Environmental Management 103, 133-141. https://doi.org/10.1016/j.jenvman.2012.03.007

National Marine Fisheries Service (2013). Lower Columbia River salmon and steelhead ESA recovery plan

Nickelson T.E. (1998). A habitat-based assessment of coho salmon production potential and spawner escapement needs for Oregon coastal streams. Oregon Department of Fish and Wildlife Portland.

Nickelson T.E. (1986). Influences of upwelling, ocean temperature, and smolt abundance on marine survival of coho salmon (Oncorhynchus kisutch) in the Oregon Production Area. Canadian Journal of Fisheries and Aquatic Sciences 43, 527-535

Nickelson T.E. \& Lawson P.W. (1998). Population viability of coho salmon, Oncorhynchus kisutch, in Oregon coastal basins: application of a habitat-based life cycle model. Canadian Journal of Fisheries and Aquatic Sciences 55, 2383-2392

Nickelson T.E., Rodgers J.D., Johnson S.L. \& Solazzi M.F. (1992). Seasonal changes in habitat use by juvenile coho salmon (Oncorhynchus kisutch) in Oregon coastal streams. Canadian Journal of Fisheries and Aquatic Sciences 49, 783-789

Oosterhout G.R., Huntington C.W., Nickelson T.E. \& Lawson P.W. (2005). Potential benefits of a conservation hatchery program for supplementing Oregon coast coho salmon ( Oncorhynchus kisutch ) populations: a stochastic model investigation. Canadian Journal of Fisheries and Aquatic Sciences 62, 1920-1935. https://doi.org/10.1139/f05-080
van Oppen M.J.H., OliverJ.K., Putnam H.M. \& Gates R.D. (2015). Building coral reef resilience through assisted evolution. Proceedings of the National Academy of Sciences 112, 23072313. https://doi.org/10.1073/pnas. 1422301112

Pess G.R., Kiffney P.M., Liermann M.C., Bennett T.R., Anderson J.H. \& Quinn T.P. (2011). The Influences of Body Size, Habitat Quality, and Competition on the Movement and Survival of Juvenile Coho Salmon during the Early Stages of Stream Recolonization. Transactions of the American Fisheries Society 140, 883-897. https://doi.org/10.1080/00028487.2011.587752

Poff N.L., Allan J.D., Bain M.B., Karr J.R., Prestegaard K.L., Richter B.D., et al. (1997). The Natural Flow Regime. BioScience 47, 769-784. https://doi.org/10.2307/1313099

Quinn T.P. (2018). The behavior and ecology of Pacific salmon and trout, 2nd edn. University of Washington Press, Seattle, WA.

Quinn T.P. \& Peterson N.P. (1996). The influence of habitat complexity and fish size on overwinter survival and growth of individually marked juvenile coho salmon ( Oncorhynchus kisutch ) in Big Beef Creek, Washington. Canadian Journal of Fisheries and Aquatic Sciences 53, 1555-1564. https://doi.org/10.1139/f96-092

R Core Team (2017). R: A language and environment forstatistical computing. Foundation for Statistical Computing, Vienna, Austria.

Rhodes J. \& Quinn T. (1998). Factors affecting the outcome of territorial contests between hatchery and naturallyreared coho salmon parr in the laboratory. Journalof Fish Biology 53, 1220-1230

Ruzzante D.E. (1994). Domestication effects on aggressive and schooling behavior in fish. Aquaculture 120, 1-24. https://doi.org/10.1016/0044-8486(94)90217-8

Saloniemi I., Jokikokko E., Kallio-Nyberg I., Jutila E. \& Pasanen P. (2004). Survival of reared and wild Atlantic salmon smolts: size matters more in bad years. ICES Journal of Marine Science 61, 782-787. https://doi.org/10.1016/j.icesjms.2004.03.032

Sarrazin F. \& Barbault R. (1996). Reintroduction: challenges and lessons for basic ecology. Trends in Ecology \& Evolution 11, 5

Scheuerell M.D., Hilborn R., Ruckelshaus M.H., Bartz K.K., Lagueux K.M., Haas A.D., et al. (2006). The Shiraz model: a tool for incorporating anthropogenic effects and fish-habitat relationships in conservation planning. Canadian Journal of Fisheries \& Aquatic Sciences 63, 1596-1607. https://doi.org/10.1139/F06-056

Schindler D.E. \& Hilborn R. (2015). Prediction, precaution, and policy under global change. Science 347, 953-954. https://doi.org/10.1126/science. 1261824

Schroeder R.K., Lindsay R.B. \& Kenaston K.R. (2001). Origin and Straying of Hatchery Winter Steelhead in Oregon Coastal Rivers. 11

Solazzi M.F., Nickelson T.E., Johnson S.L. \& Rodgers J.D. (2000). Effects of increasing winter rearing habitat on abundance of salmonids in two coastal Oregon streams. 57, 9

Stanley E.H., Fisher S.G. \& Grimm N.B. (1997). Ecosystem Expansion and Contraction in Streams. BioScience 47, 427-435. https://doi.org/10.2307/1313058

Suring E.J., Brown E.T. \& Moore K.M.S. (2006). Lower Columbia River Coho Status Report 2002 2004: Population abundance, distribution, run timing, and hatchery influence.

Tenger-TrolanderA., Lu W., Noyes M. \& Kronforst M.R. (2019). Contemporary loss of migration in monarch butterflies. Proceedings of the National Academy of Sciences 116, 1467114676. https://doi.org/10.1073/pnas. 1904690116

Thériault V., Moyer G.R., Jackson L.S., Blouin M.S. \& Banks M.A. (2011). Reduced reproductive success of hatchery coho salmon in the wild: insights into most likely mechanisms:

FITNESS OF HATCHERY COHO SALMON IN THE WILD. Molecular Ecology 20, 1860-1869. https://doi.org/10.1111/j.1365-294X.2011.05058.x

USFS (1995). Gifford Pinchot National Forest land and resource management plan. United States Forest Service. Vancouver, WA

Wainwright T.C. \& Weitkamp L.A. (2013). Effects of Climate Change on Oregon Coast Coho Salmon: Habitat and Life-Cycle Interactions. Northwest Science 87, 219-242. https://doi.org/10.3955/046.087.0305

Wigington P., Ebersole J., Colvin M., Leibowitz S., Miller B., Hansen B., et al. (2006). Coho salmon dependence on intermittent streams. Frontiers in Ecology and the Environment 4, 513-518. https://doi.org/10.1890/1540-9295(2006)4[513:CSDOIS]2.0.CO;2

Woelfle-Erskine C., Larsen L.G. \& Carlson S.M. (2017). Abiotichabitat thresholds for salmonid over-summer survival in intermittent streams. Ecosphere 8, e01645. https://doi.org/10.1002/ecs2.1645

Zabel R., Cooney T., Jordan C., Carmichael R.W., Jonasson B.C., Sedell E.W., et al. (2013). Lifecycle models of salmonid populations in the interior Columbia River Basin

