

AN ABSTRACT OF THE DISSERTATION OF

Tomas Vergel C. Jamir for the degree of Doctor of Philosophy in Oceanography presented on June 30, 1998. Title: Distribution, Seasonal Variation and Community Structure of the Demersal Trawl Fauna of Ragay Gulf, Philippines.

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Abstract approved: _____

William G. Pearcy

The demersal fish community structure of Ragay Gulf, Burias Pass, Ticao Pass and Northern Samar Sea, Philippines (RABUTINOS) was studied from data obtained from the surveys carried out by RV Sardinella during the wet northeast monsoon, dry intermonsoon and wet southwest monsoon seasons. Also included were samples taken during the dry early maturity stages of the strong 1982-1983 El Niño-Southern Oscillation event.

Detrended Correspondence Analysis (DCA), Cluster Analysis, Two-Way Indicator Species Analysis (TWINSpan) and correlation of DCA Axis 1 with depth, temperature, salinity and oxygen implemented by the PC-ORD and SAS programs revealed depth as the principal environmental gradient along which faunal changes occur. Fisher's Protected Least Significant Difference tests and multiple linear regression analysis associated the observed depth zonation pattern with species assemblages distributed above the thermocline (warm water fauna), within the thermocline (cool water fauna) and below the thermocline (cold water fauna). These corresponds to regions shallower than 90 m, between 90 to 150 m, and deeper than 150 m, respectively.

Within each of these major depth zones, the important determinants of fish assemblage structure include substrate characteristics (rocky-coraline, sandy, muddy); proximity to major rivers (salinity and/or turbidity gradient); depth (shallower or deeper than 35 m); biotic interactions (predation, competition, recruitment) and availability of

food. Seasonal ordination and classification based on species abundance and occurrences revealed eight gradually overlapping species site groups and five species groups separable by distinct demersal fish assemblages, depth ranges, substrate types and water mass characteristics.

Except for seasonal variability in distribution, the fish assemblages of RABUTINOS were relatively stable in terms of species composition, relative abundance and dominance. Two-thirds of the top 20 most abundant species/taxa remained as common components of the seasonal species assemblages. The main environmental perturbations associated with the 1982-'83 El Niño event included significant declines in precipitation, elevated salinity and temperature levels, and early spring-like conditions that resulted in the drastic reduction in mean seasonal catch rates and number of demersal fish assemblage groups by half. These remaining assemblages contained species/taxa that were well adapted to the harsh and shifting environmental conditions typical of their estuarine and thermocline habitats.

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Distribution, Seasonal Variation and Community Structure of the Demersal Trawl
Fauna of Ragay Gulf, Philippines

by

Tomas Vergel C. Jamir

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31 JULY '98

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DISTRIBUTION, SEASONAL VARIATION AND COMMUNITY STRUCTURE OF THE DEMERSAL TRAWL FAUNA OF RAGAY GULF, PHILIPPINES

INTRODUCTION

Over the last four decades, commercial exploitation of Southeast Asia's demersal fishery resources has been confined primarily to shallow coastal waters less than 50 m deep (Pauly, 1988). Recently, however, the depletion of nearshore stocks and advances in mechanized fishing technology have shifted commercial fishing interests towards the economic potential of deeper waters. The meteoric rise of the Gulf of Thailand demersal trawl fishery and the subsequent decline of its resource base (Pauly, 1988) is a classic example of the serious ecological, political and economic consequences of unfettered overcapitalization in the fishing industry.

Effective management of the fishery not only requires knowledge of the species composition of assemblages but also the interactions among species and their responses to the environment. Despite its recognized shortcomings the single-species stock model remains the basis for most of today's management decisions (Brander, 1988; Sainsbury, 1988). This is partly due to the marine scientist's need to know about the autecology and biology of single-species stocks (Pauly, 1982) and to the slow progress in the development of multispecies models that can be used as guides in the formulation of management strategies (Brander, 1988; Sainsbury, 1982).

A number of problems arise in the stock assessment and management of diverse temperate and tropical fisheries based on the single species approach. As the diversity of the species within the catch increases, the assessment of each individual's state of exploitation becomes more difficult and complicated. In this situation, the consequence of attempting to manage the fishery by regulating only a few of the main species become

less predictable (Brander, 1988). Single species studies often ignore differences in yield and overall species composition of different areas (Brander, 1988). To a fisheries manager, however, these differences are important, as the value of the catch in most fisheries are strongly influenced by its species composition (Pauly and Mines, 1982; Sainsbury, 1988).

Few fisheries operate on a single species or stock, especially in the tropics, where multispecies fisheries are predominant (Russ, 1991). Fishing operation directly influences the community by the removal of individuals or indirectly through habitat modification (Sainsbury, 1988; Russ, 1991). It is now recognized that unmanaged growth in this industry not only causes the decline of target species but also leads to ecosystem overfishing (Pauly, 1979, 1988). This phenomenon is recognized worldwide from its trail of widespread alterations in the species composition of most exploited fish communities, e.g., the *Balistes* (Family: Balistidae) population explosion that displaced the sciaenid community from Senegal to Nigeria (Longhurst and Pauly, 1987) and the *Loligo* (Family: Loliginidae) population outburst that displaced the Leiognathidae population in the Gulf of Thailand (Tiews *et al.*, 1967; Pauly, 1979) and Manila Bay (Ganaden, 1990). Therefore, confident multispecies fisheries management requires an understanding of community structure and dynamics from which (1) the ecological and fishery implications of alternative exploitation strategies may be ascertained (Brander, 1988; Sainsbury, 1988) and (2) undesirable community changes in species composition may be prevented while encouraging desirable ones (Pauly and Murphy, 1982; Sainsbury, 1982; May, 1984; Rothschild, 1983; Sherman and Alexander, 1986).

Besides the extensive studies conducted in northern Australian continental shelves, relatively little is known about the complex structure of demersal fish communities in the tropical western Pacific Ocean (Longhurst and Pauly, 1987). To the author's knowledge, only a few studies of tropical demersal fish community structure by means of multivariate analysis (excluding coral reef areas and lagoons) have been carried out in Southeast Asia, e.g., the Samar Sea (McManus, 1985) and Ragay Gulf, Philippines

(Federizon, 1992), and Malaysia (Chan and Liew, 1986). This deficiency implies that work on the population dynamics of demersal fish stocks in the region is usually conducted blindly using data (e.g., catch/effort, species and size composition) that may come from different communities (Pauly, 1988).

In the tropics, different interacting factors have been attributed to the way demersal fish communities are structured. This includes depth and spatial factors (McManus, 1985; Harris and Poiner, 1990; Ramm *et al.*, 1990; Federizon, 1992), temperature and salinity (Rainer and Munro, 1982; Rainer, 1984; Longhurst and Pauly, 1987; Bianchi, 1991; Sheaves, 1998), substrate type (Rainer and Munro, 1982), proximity to rocky or coral reefs (Watson *et al.*, 1990; Federizon, 1992; Newman *et al.*, 1997), ontogenetic factors (Rainer and Munro, 1982; Weng, 1990; Morin *et al.*, 1992), sensory adaptation (Tejerina-Garro *et al.*, 1998), seasonal cycles (Watson *et al.*, 1990; Ansari *et al.*, 1995); food availability (Weng, 1990; Caddy and Bakun, 1995) and turbidity (Watson *et al.*, 1990; Cyrus and Blaber, 1992; Caddy and Bakun, 1995). The complexity in which fish communities are structured emphasizes the importance of conducting rigorous community analyses on demersal fishes as a prerequisite to ecologically sound resource management.

After comparing the distribution of demersal tropical fishes around the world, Longhurst and Pauly (1987) found relative stability in the fish fauna of the continental shelves, with many of the same families represented over similar substrates and depths. From these environmental factors, they also established four basic fish assemblage categories, i.e., fishes of (1) inshore and estuarine, muddy habitats and turbid waters, (2) sandy deposits and clear waters, (3) rocky reefs (both estuarine and offshore), and (4) coral reef outlier species.

The strong El Niño-Southern Oscillation event in 1982-'83 presented a major, albeit limited, opportunity to see whether Longhurst and Pauly's hypothesis holds true. In this study, the stability of the RABUTINOS demersal fish communities subjected to

exogenous perturbations will be looked at in terms of the following criteria: (1) stability of relative abundance patterns, (2) stability of dominance, (3) stability of species composition, and (4) stability of spatial distribution (Krebs, 1978).

This study investigates the species composition and structure of demersal fish assemblages in Ragay Gulf, Burias Pass, Ticao Pass and Northern Samar Sea (=RABUTINOS) in relation to environmental variables, (wet and dry) seasons, a strong El Niño-Southern Oscillation event and geographic location. More specifically, the main objectives of this research are:

(1) to describe the abundance, structure and composition of demersal fish assemblages at RABUTINOS in relation to other Indo-Pacific forms,

(2) to determine the spatial distribution and temporal variation of fish assemblage patterns in relation to the regular Southeast Asian monsoon seasons and the local effects of the El Niño-Southern Oscillation,

(3) to identify possible key environmental gradients and/or biotic processes that may be responsible for the way the fish assemblages were distributed and structured, and

(4) to determine the stability of tropical fish communities in relation to the regional environmental perturbations brought about by the strong 1982-1983 El Niño-Southern Oscillation event.

This type of synecological study is a necessary step towards understanding multispecies stocks (Caddy and Sharp, 1986) and can be extended to descriptive community dynamics (McManus, 1985) in order to find general patterns of species compositions under given environmental conditions and fishing effort. Comparison of assemblages from similar ecosystems in different areas might also reveal general trends in the community dynamics of tropical shelves (Bianchi, 1990). In addition, this work

could also be useful in fisheries management. For example, species composition of trawl catches from a given area may be roughly anticipated from assemblage maps derived from this analysis, or assemblage boundaries can be used to frame legislation aimed at reducing the impact of growing commercial trawl fisheries on the small-scale non-trawl fisheries (Pauly, 1988).

A. Review of Literature:

1. Community Structure of Tropical Demersal Fishes.

Longhurst and Pauly (1987) determined that the main environmental factors determining what species occur in a given area are the amount of organic mud in the substrate, presence of isolated patches of rocky or coral reefs, occurrence of brackish conditions associated with lagoons and river mouths, and the characteristics of the continental shelf's water mass. Under similar conditions in the tropics, they also asserted that the expected fish fauna is remarkably consistent, with more families being represented in the western Indo-Pacific than in the Atlantic. Essentially four basic categories of fish assemblages can be found in the tropics (Longhurst and Pauly, 1987).

a. *Fish of inshore and estuarine muddy habitats and turbid waters.* This assemblage is dominated by two forms of drums/croakers (family: Sciaenidae) -- fast swimming benthic-pelagic piscivores and the benthic forms with inferior mouth and sometimes with gular barbels. Sciaenid assemblages inhabit the shallow (<15 - 20 m), soft-bottom waters of Southeast Asia (Chong *et al.*, 1990), northern Australia (Blaber *et al.*, 1989; Blaber *et al.*, 1995), west coast of India (Ansari *et al.*, 1995) and western Africa (Fager and Longhurst, 1968). This bathymetric boundary could be deeper than 20 m depending on the extent of riverine influence and warm water mass, e.g., down to 60 m northwest off Guiana and Brazil where large volumes of freshwater are discharged into

the sea by the Orinoco, Amazon and other major South American rivers (Richards, 1955; Lopez, 1964; Longhurst and Pauly, 1987).

b. *Sandy deposits and clear waters*. This assemblage are found farther offshore, where fishes of pink or silvery color with large eyes adapted to clear waters and lighted conditions and feed mostly on benthic epifauna and vagile benthos abound (Longhurst and Pauly, 1987). Included here are the grunts (family: Haemulidae), brems (family: Sparidae), mojarras (family: Gobiidae), large-eyes (family: Priacanthidae), goatfishes (family: Mullidae), threadfin brems (family: Nemipteridae) and slipmouths (family: Leiognathidae) found throughout the tropics (Fager and Longhurst, 1968; Blaber *et al.*, 1994). The last two families, however, are common only in the Indo-Pacific region (Jones, 1985) and are the mainstay of Southeast Asian demersal fisheries (Tiews and Borja, 1965; McManus, 1985; Pauly, 1988). The sandy and cold water regions between 200 - 300 m deep are home to bathypelagic and benthic faunas like the greeneyes (family: Chlorophthalmidae), lizardfishes (family: Synodidae = Synodontidae) and jacks (family: Carangidae) in western Africa (Fager and Longhurst, 1968), northern Australia (Okera, 1982), and in Southeast Asia (Federizon, 1992).

c. *Rocky reefs (both estuarine and offshore)*. There are three principal families that dominate this assemblage, i.e., groupers, snappers and emperors (Longhurst and Pauly, 1987). These families inhabit the windward reef slopes, lagoons and leeward back reefs of the Great Barrier Reef (Newman *et al.*, 1997). Snappers are also known to move upstreams in the tropical estuaries of Australia (Sheaves, 1996a, 1996b) and are abundant in the rocky coralline areas in the Philippines (Warfel and Manacop, 1950; Federizon, 1992), Caribbean (Russ, 1991), or sandy regions in East Africa (Morgan, 1964), Australia (Okera, 1982), and South China Sea (Longhurst and Pauly, 1987).

d. *Outlier species of coral reefs*. There is no single dominant family in this group composed mainly of triggerfishes (Balistidae), pufferfishes (Tetraodontidae) and boxfishes (Ostraciontidae). These families are abundant in areas close to coral reefs, e.g.,

Southeast Asia (Russ, 1991), Great Barrier Reef (Newman *et al.*, 1997), and Equatorial Pacific Islands (Thresher, 1991).

Warfel and Manacop (1950) conducted exploratory trawling surveys of different potential fishing grounds in the Philippines prior to the extensive mechanization of the fishery. Their findings indicated little variation in the biomass and dominance of fish families living within 75 fathoms (140 meters) of the muddy and sandy substrates throughout the country. The dominant species include slipmouths (family: Leiognathidae), lizardfishes (family: Synodidae) and mojaras (family: Gobiidae). Thirty years later, McManus (1985) found similar relative abundance and species/taxa composition in Samar Sea and Carigara Bay, validating Longhurst and Pauly's (1987) thesis regarding the general stability of tropical demersal fish communities.

2. Application of Multivariate Statistics in Community Ecology.

The application and power of multivariate statistical techniques in the analysis of complex, multispecies tropical fish stock has been well documented by researchers from Australia and elsewhere. For example, Bianchi (1991) employed two-way species indicator analysis (TWINSPAN) and detrended correspondence analysis (DECORANA) to identify demersal species groups in Mexico and Costa Rica. Ross and Doherty (1994) used principal components analysis (PCA) to examine the physicochemical properties and species abundance relationships in the Barrier Island, Gulf of Mexico. Canonical discriminant analysis (CDA) was used by Sheaves (1998) to investigate species distributions and abundances of fishes in tropical Queensland estuaries; hierarchical classification by Newman *et al.* (1997) for the Great Barrier Reef and by Rainer and Munro (1982) in northern Australia. Bray-Curtis Ordination (BCO) and principal coordinate analysis (PCO) were used by Long and Poiner (1994) and by Ramm *et al.*, (1990) to analyze fish communities in northern Australia, and by Ansari *et al.* (1995) on the west coast of India.

In the Philippines, multivariate analysis was conducted by McManus (1985) on the Samar Sea demersal trawl fishery using DECORANA and TWINSpan. Federizon (1985) used a combination of DECORANA, TWINSpan, Correspondence Analysis (CA), Non-Metric Multi-Dimensional Scaling (NMDS) and cluster analysis to analyze the fish community structure of Ragay Gulf.

B. Description of the Survey Area:

1. Marine Geography of RABUTINOS.

The waters of Ragay Gulf, Burias Pass, Ticao Pass and Northern Samar Sea form a narrow channel of water oriented along the northwest - southeast direction on Eastern Luzon, Philippines (Figure 1). This body of water is about 180 miles long and 10 to 25 miles wide. This is approximately half the width and the same length as Oregon's Central Willamette Valley. This narrow passageway serves as the primary connection between the strong Kurushio Current and the internal seas of Central Philippines (Wyrtki, 1961). Hence, any anomalous perturbation in the equatorial western Pacific Ocean readily reflects on the physico-chemical properties of RABUTINOS, making the Ragay Gulf system potentially useful as a barometer for detecting or monitoring possible ENSO impacts in the region.

Oriented in a northwest-southeast direction, RABUTINOS is bounded to the north by Viñas River (122°27'E, 13°55'N) and to the south by the 12°N latitude for the purpose of this research. To the east are the Cordillera Mountain range of Luzon and the island of Samar. Separating these two land masses is the narrow San Bernardino Strait, the gateway to the Pacific Ocean. North of San Bernardino Strait is the shallow Sorsogon Bay. The western boundaries include Bondoc Peninsula and the islands of Burias, Ticao and Masbate.

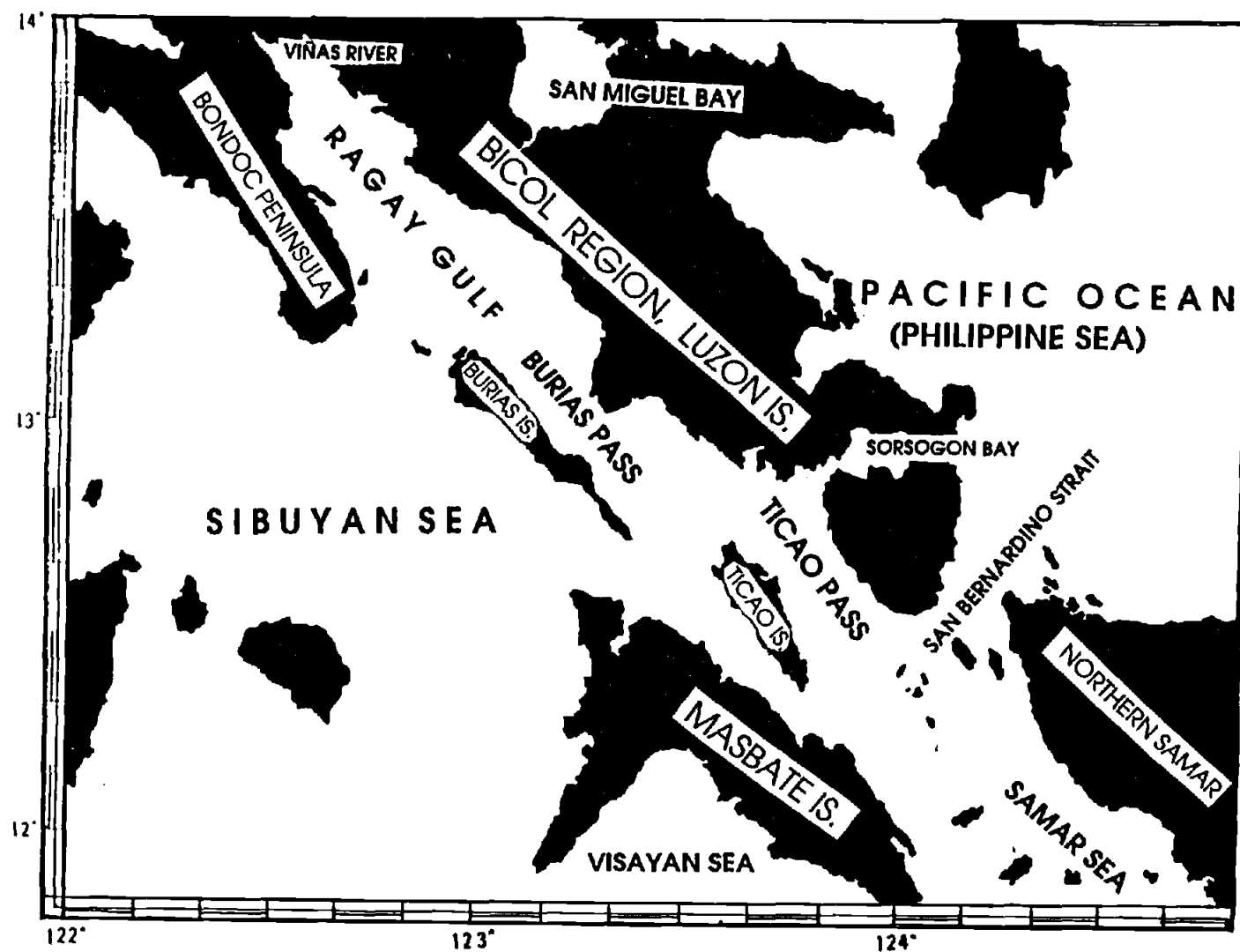


Figure 1. Geographic Map Showing the Location of Ragay Gulf, Burias Pass, Ticao Pass and Northern Samar Sea, Philippines.

Ragay Gulf is a small protected basin approximately 600 m deep at its mid-section (Figure 2). It has two broad sills of about 100 - 130 m deep, with a western and southern opening to the larger Sibuyan Sea Basin. The waters of the western equatorial North Pacific Ocean directly interact with Sibuyan and Sulu Sea Basins through the narrow and deep channels of San Bernardino Strait, Burias Pass, Ticao Pass and western Samar Sea. The Ragay Basin is characterized by relatively wide insular shelf and steep insular slope. Terrigenous mud and riverine sands characterize the sea floor in the vicinity of Viñas River while sandy-silty ooze characterizes the bottom deposits of the deeper sections of the basin farther away from the rivers. Much of Northern Samar Sea is also made up of a wide, gradually sloping insular shelf overlain by riverine mud and sand deposits. This makes the two areas highly suitable for trawling operations.

Narrow shelves, sharp drop-offs and rocky-coralline substrates characterize the eastern coast of Ticao Pass. Complex underwater ridges, deep channels and hard, rocky to sandy substrates mark the seafloor west of San Bernardino Strait. These conditions severely limit the areas available for demersal trawl operation within these two sites.

2. The Southeast Asian Monsoon.

Much of Southeast Asian climate and oceanography are influenced by the regular seasonal patterns of the monsoons (Morgan and Fryer, 1985; Suryanaryana *et al.*, 1992). The strong northeast winds that sometimes exceed 39 kph characterize the northeast monsoon season. It is most fully developed in the months of January - February and is often associated with cool weather and torrential rains in RABUTINOS.

The dry, inter-monsoon transition period follows the northeast monsoon season. Around the months of July - August, the southwest monsoon season reaches its peak with

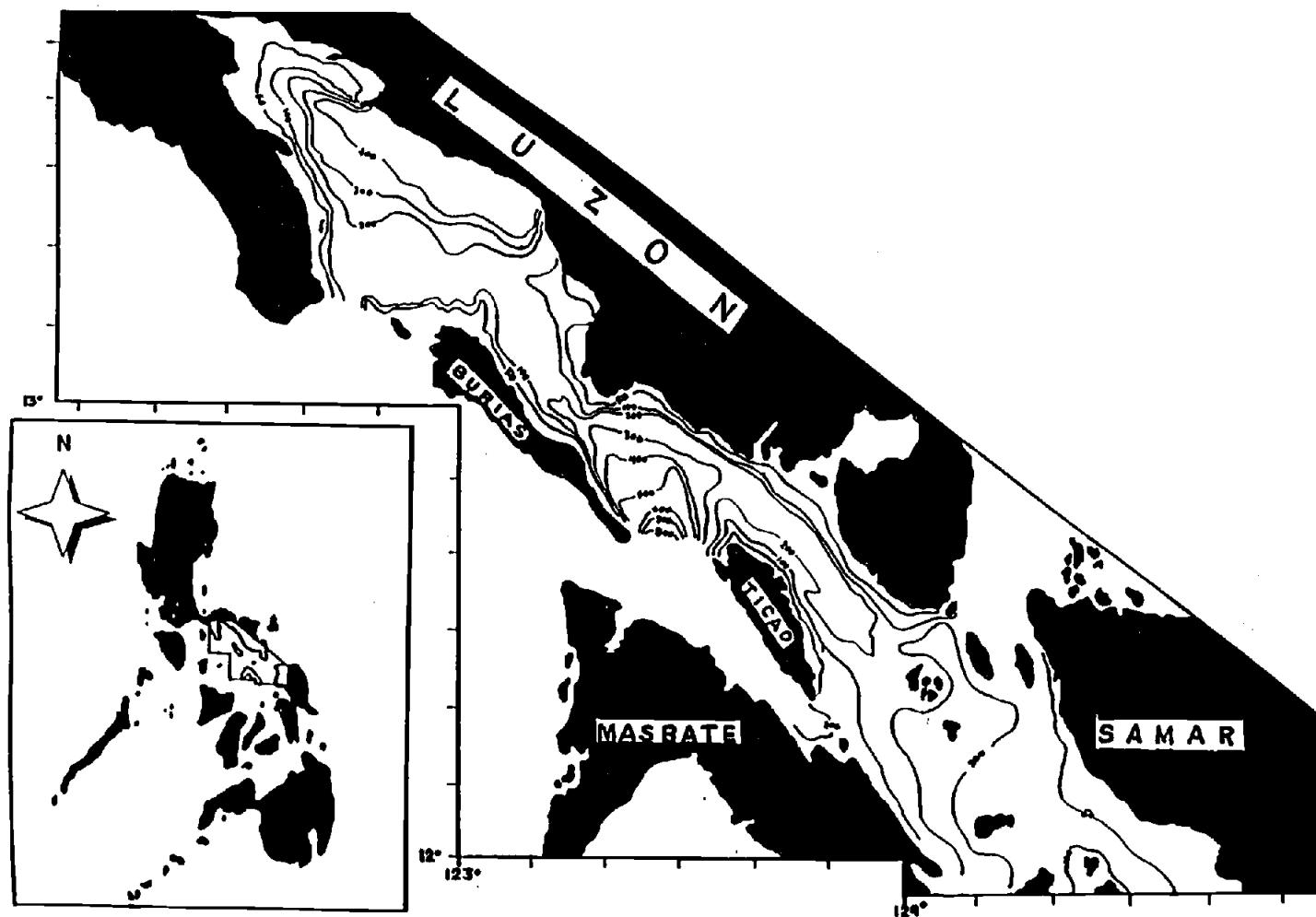


Figure 2. Bathymetric Map of Ragay Gulf, Burias Pass, Ticao Pass and Northern Samar Sea, Philippines. Depths are given in meters.

winds reaching 20-29 kph (Kumagai and Bagarinao, 1983). Warm humid weather punctuated by occasional hurricanes and heavy rainfall typifies the southwest monsoon season.

The wind lulls typical of the inter-monsoon transition phase may have a significant influence on the recruitment of tropical fishes as indicated by the pervasive bi-modal recruitment pattern for most species (e.g., Pauly and Navaluna, 1983; Ingles and Pauly, 1984; Corpuz *et al.*, 1985; Robertson and Duke, 1990). Peak spawning seasons for many species have been found to occur at times of the year when prevailing wind and/or current is at their minimum velocities (Johannes, 1980), i.e., from March - May and October - November.

The 15 year average annual rainfall pattern for the Masbate Island Weather Station is shown in Figure 3. The lowest precipitation levels usually occur during the dry intermonsoon months of March and April. The average monthly precipitation starts to increase thereafter until it reaches a high plateau around the months of July to December. The increase in precipitation during the southwest monsoon months of June to August reflects the passage of typhoons and tropical storms as the Inter-Tropical Convergence Zone (ITCZ) moves into the region.

The high precipitation levels during the northeast monsoon months of September to December is typical on the eastern side of the Philippines and attributed to orographic effects. The sudden drop in precipitation levels around August before spiking up again a month later signals the change in the seasons as the winds shift from the southwest to the northeast. The variation in the mean monthly rainfall pattern in Masbate Island is large, especially during the rainy season.

The monthly average rainfall pattern from January 1979 to December 1984, which includes the period of this study, is shown in Figure 4. The abrupt drop in rainfall around the month of August was very pronounced in all years except in 1981. Precipitation levels

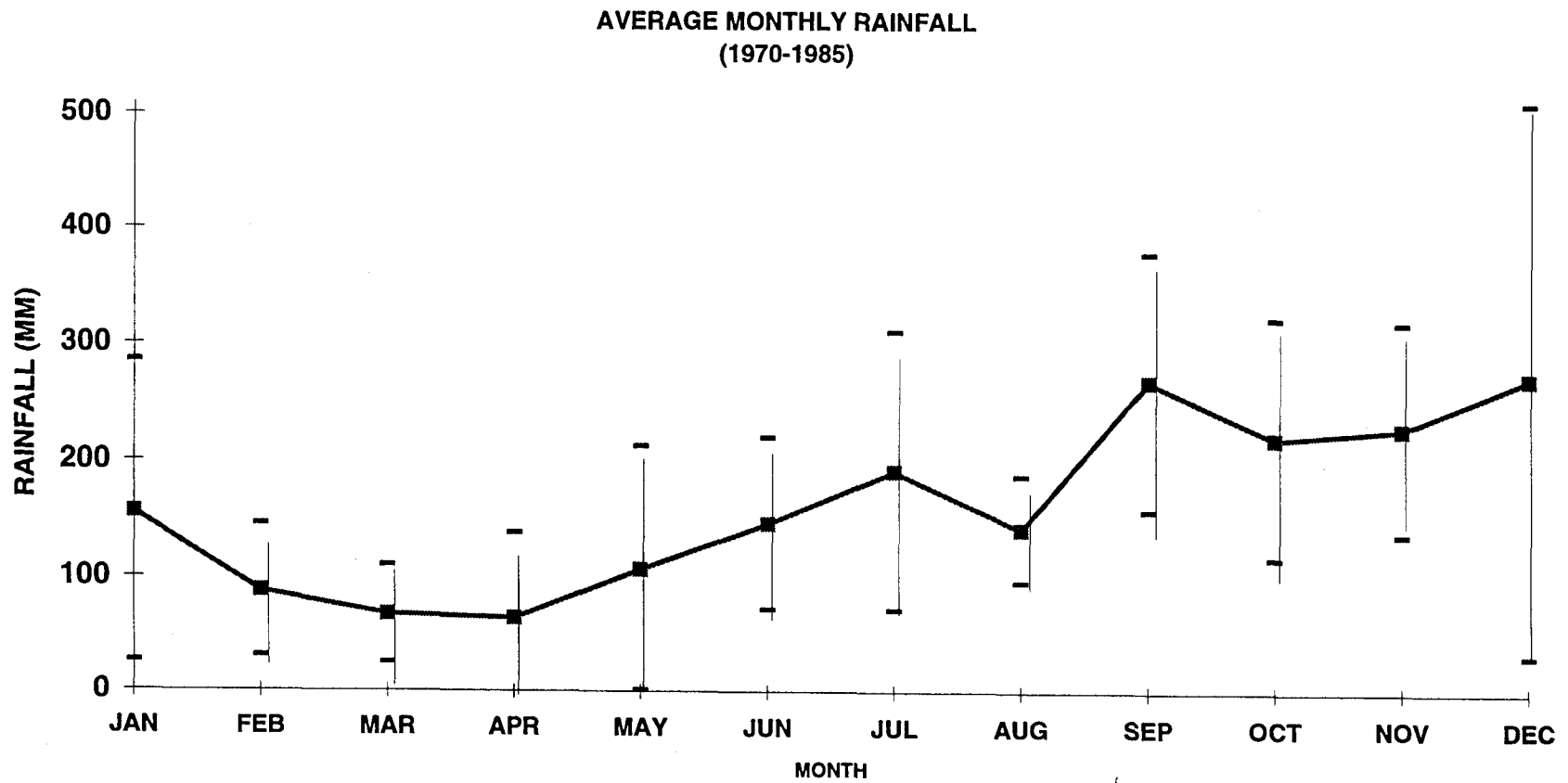


Figure 3. Average Monthly Rainfall Time Series Pattern for the Masbate Island Weather Station.

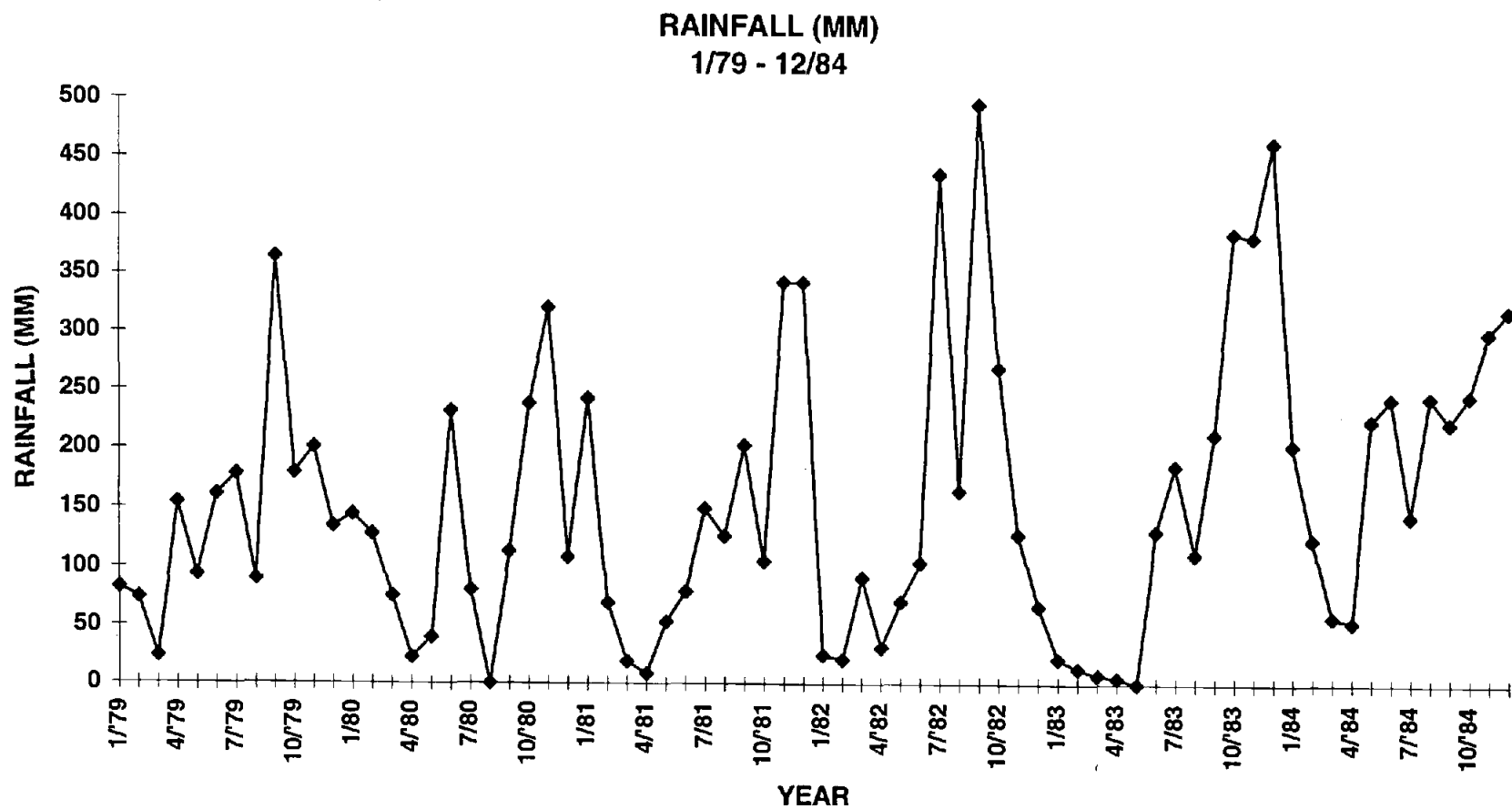


Figure 4. Monthly Rainfall Time Series Pattern from January 1979 to December 1984 for the Masbate Island Weather Station.

were also lowest around the month of April, except during the 1982-1983 ENSO when low precipitation levels were already recorded as early as January 1983 and remained low until May 1983.

Rainfall levels were high during the survey's first two cruises in November and December 1981, as well as in July and September 1982 followed by drought conditions thereafter. As the survey ended in January 1983, the normal northeast monsoon rains never came and the precipitation levels were already below normal values. This eventually developed into conditions that prevailed throughout Southeast Asia over the next five months.

3. Physical Oceanography.

The water masses of the Philippines is characterized by a thick (~50 - 100 m) thermocline located at 100 - 150 m depths (Megia and Villadolid, 1953; Megia and Sebastian, 1951; Wyrski, 1961). This permanent thermal discontinuity is regarded as an effective barrier to the fertilization of the photic zone by the nutrient-rich bottom waters. Hence, fertilization of coastal waters is confined to nutrient inputs from rivers and terrestrial run-offs during the rainy southwest monsoon season and occasional localized upwellings during the northeast monsoon season (Megia and Sebastian, 1951; Megia and Villadolid, 1953; Jacinto, 1983; Suryanarayana *et al.*, 1992).

The oceanographic surveys conducted at the Gulf of Thailand and South China Sea (Wyrski, 1961), Philippine territorial waters (Graham, 1952, 1953); Manila Bay (Megia, 1953), Lingayen Gulf (Sebastian *et al.*, 1959), Samar Sea and Carigara Bay (Labao, 1982), Visayan Sea (Aprieto, 1978), San Miguel Bay (Legasto *et al.*, 1975; Pauly and Mines, 1982), Sorsogon Bay (Ordonez *et al.*, 1975), shallow coastal lagoons of Batangas (Jacinto, 1983; San Diego, 1985); and Ragay Gulf (Mines *et al.*, 1984) all point to the pervasive influence of the seasonal monsoons in structuring the characteristics of

the physical environment. The regular environmental cycle basically revolves around the “dry” and “wet” seasons which exert their influence on the distribution and physical characteristics (e.g., temperature, salinity, dissolved oxygen, nutrients) of the waters in the upper ocean.

During the wet southwest monsoon season, coastal bodies of water throughout the region become “estuarized” similar to the Gulf of Carpentaria in Northern Australia (Longhurst and Pauly, 1987); West Bay of Bengal (Suryanarayana *et al.*, 1992); Lingayen Gulf, Philippines (Sebastian *et al.*, 1959); Manila Bay (Megia, 1953); San Miguel Bay, Philippines (Pauly and Mines, 1982); and shallow coastal lagoons of Batangas, Philippines (Jacinto, 1983; San Diego, 1985). The extent of low salinity water layer varies from 30 - 35 m in the Gulf of Carpentaria and the shallow bays of the Philippines, to 50 m off the coast of India.

The relatively cool, dry, strong and persistent winds characteristic of the northeast monsoon tends to weaken thermal stratification as the low salinity layers disappear and the water column cools down and becomes mixed (Wyrski, 1961; Suryanarayana *et al.*, 1992). Occasional local inversions and “upwelling” (Rochford, 1991) events also take place during this season (Wyrski, 1961; Megia, 1953; Mines *et al.*, 1984).

The fifteen year average monthly temperature pattern is shown in Figure 5. Cool temperatures characterized the northeast monsoon months from December to February. Air temperature warms up again during spring and continues to rise throughout the intermonsoon transition period. Peak temperatures occur during the month of May just before the southwest monsoon rains start to pour. The temperature cools down a little during the rainy months of June to July. Then it plateaus around 28°C during the northern hemisphere summer before starting to cool down again by autumn. The temperature variations from the mean monthly values are small and similar throughout the year.

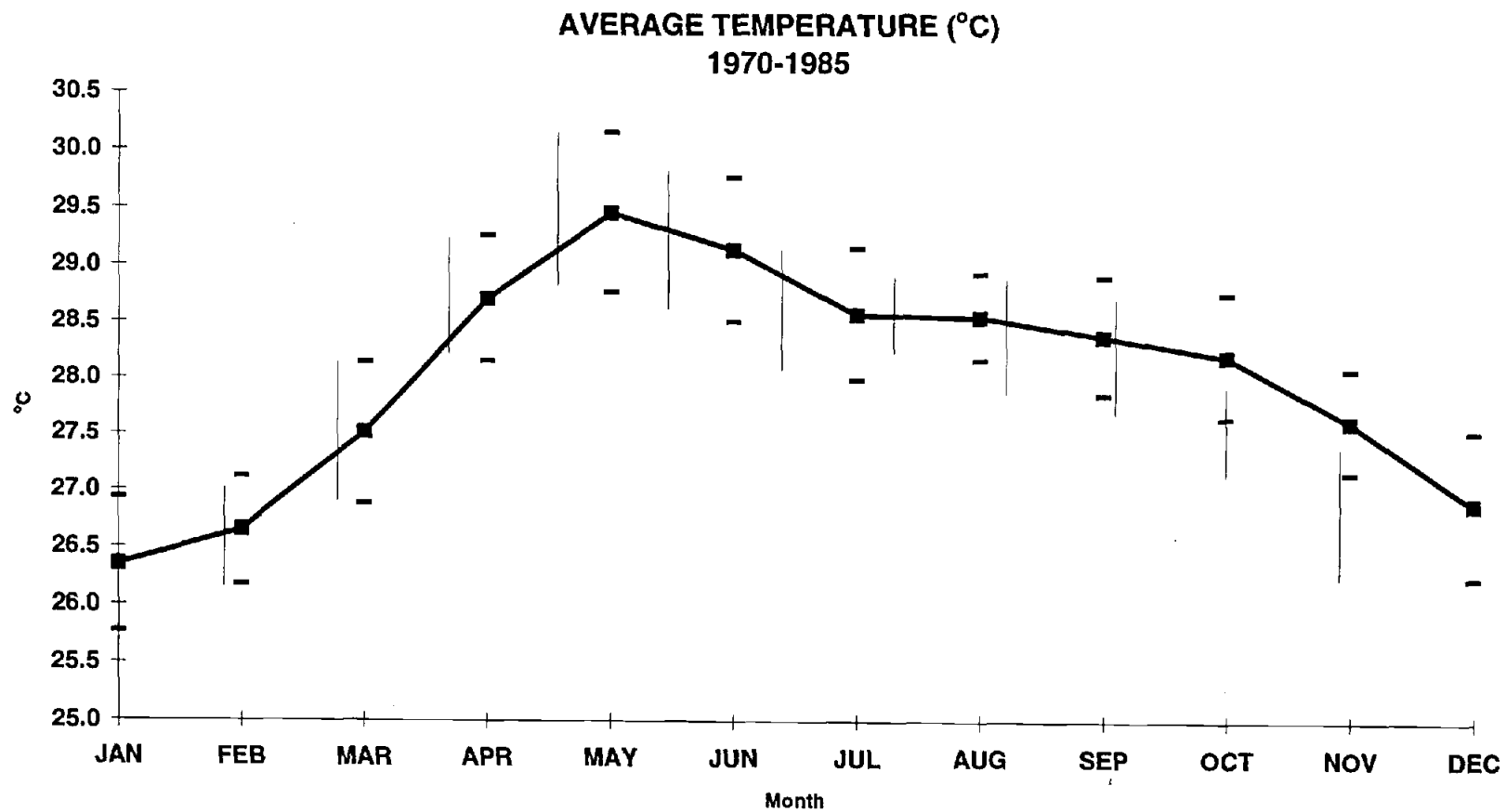


Figure 5. Average Monthly Air Temperature Time Series Pattern for the Masbate Island Weather Station.

The temporal variation in the average monthly air temperature at the Masbate Island Weather Station from January 1979 to December 1984 is shown in Figure 6. The regular seasonal temperature pattern described above was evident in this series, i.e., coolest temperatures in January and warmest in May, except during 1982-1983 and 1984 summer. Summer was relatively cooler and winter was warmer than normal at Masbate Island during the 1982-1983 ENSO event. Summer was also cooler than normal in 1984.

Figure 7 shows the variation in the average annual air temperature at the Masbate Island Weather Station from 1961 to 1996. This diagram shows that two to three years prior to a major ENSO event, (i.e., 1970-71, 1982-83 and 1997-98), the average annual temperature at Masbate Island rose above normal, with the annual average temperature immediately cooling after an ENSO event.

C. El Niño-Southern Oscillation

The relative stability and regular seasonal cycle of the monsoon system in Southeast Asia are periodically perturbed, at increasing regularity (Longhurst and Pauly, 1987), by the global climatic event called El Niño-Southern Oscillation. In the strictest sense, El Niño (EN) originally referred to the local, periodic mild warming of the Peruvian coastal waters around Christmastime, hence, El Niño or Spanish for “little boy” (Cane, 1983; Trenberth 1991; Enfield, 1992). The atmospheric counterpart of El Niño, the Southern Oscillation (SO) was coined by Sir Gilbert Walker for the sea-level pressure “see-saw” or standing wave that he observed between Darwin, Australia (12.4°S, 130.9°E) and Tahiti (17.5°S and 149.6°W) in the South Pacific (Trenberth, 1991; Diaz and Markgraf, 1992). Today, El Niño-Southern Oscillation (ENSO) refers to the large-scale anomalous warming of the Pacific Ocean due to the incursion of warm water from the Western Equatorial Pacific Ocean into Central and/or Eastern Equatorial Pacific Ocean in conjunction with the cessation of equatorial upwelling of cold waters there (Cane, 1983; Rasmusson and Wallace, 1983; Glantz, 1991; and Enfield, 1992).

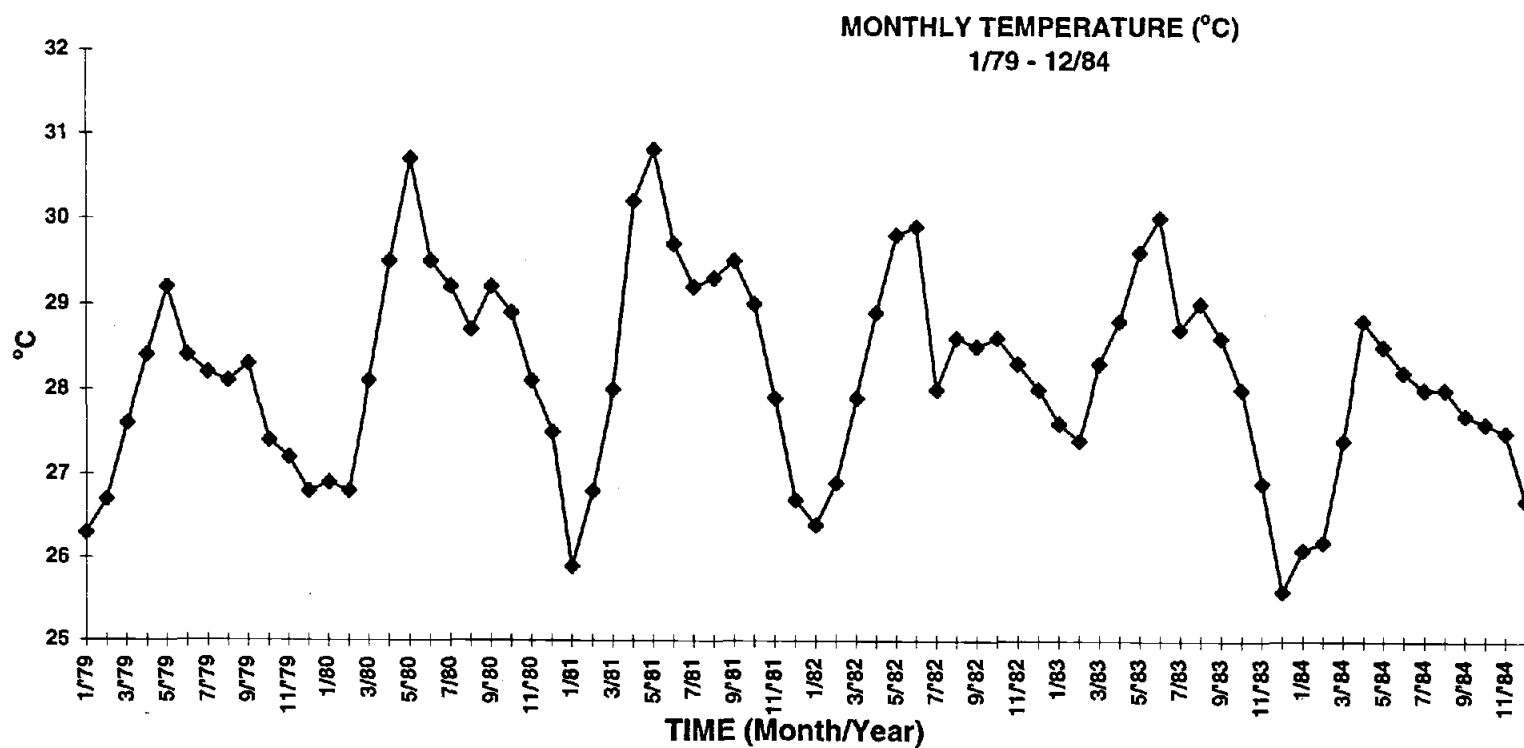


Figure 6. Monthly Air Temperature Time Series Pattern from January 1979 to December 1984 for the Masbate Island Weather Station.

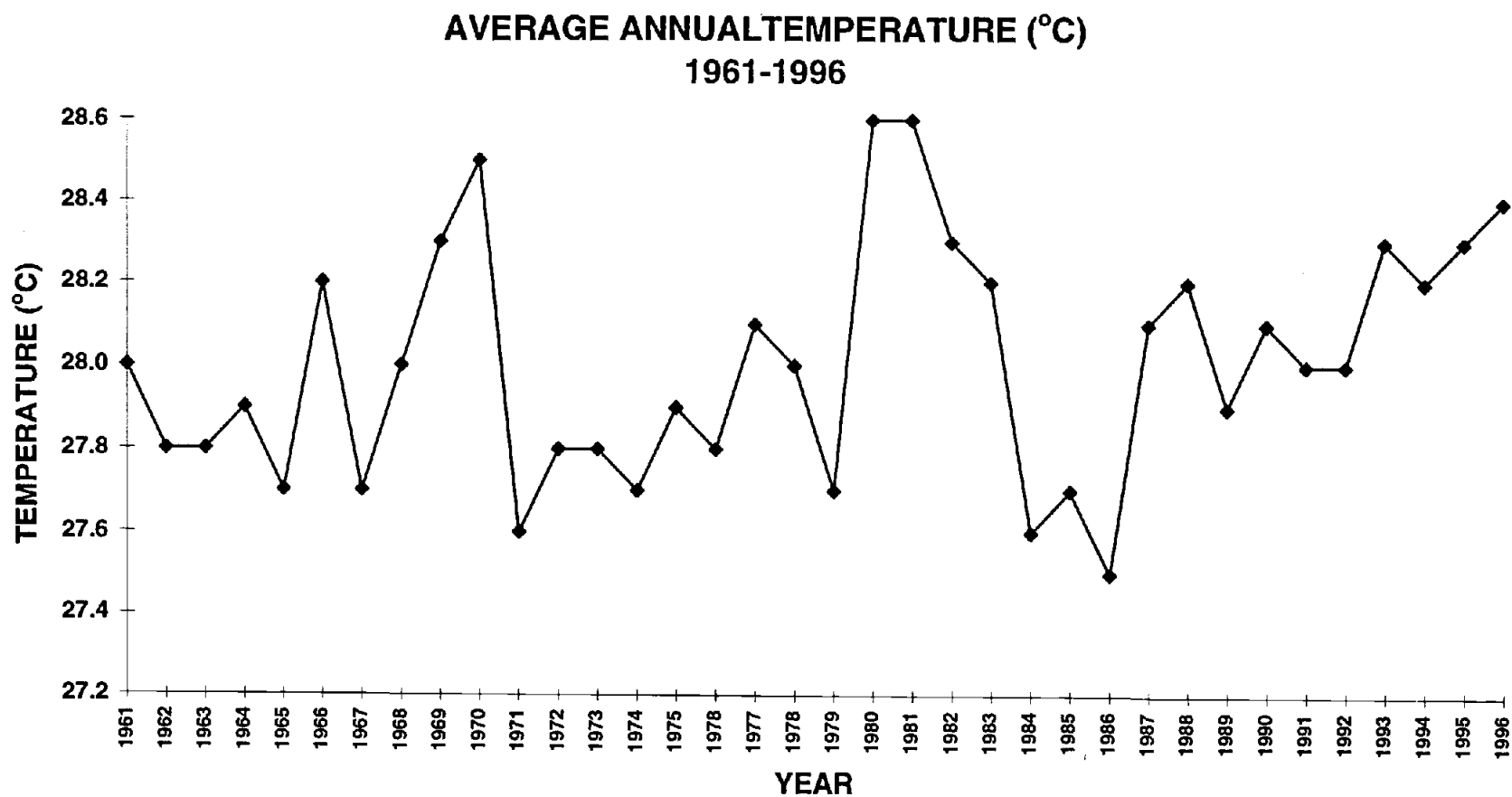


Figure 7. Average Annual Air Temperature Time Series Pattern from 1961 to 1996 for the Masbate Island Weather Station.

The life cycle of a canonical ENSO event can be subdivided into the prelude, onset, event and maturity stages (Cane, 1983; Trenberth, 1991). The start or end of a typical ENSO event occurs around the intermonsoon months of March to May. This period is characterized by the weakest wind and atmosphere-ocean coupling in the western Pacific Ocean (Trenberth, 1991). Following Cane's (1983) designation, the year prior to ENSO is designated as (-1), the ENSO event year as (0) and the year that follows as (+1).

The Prelude Stage, now referred to as La Niña" (Diaz and Kiladis, 1992), is characterized by stronger than average easterlies, higher sea levels, deeper than average thermoclines and anomalously warm sea surface temperature (SST) in the western equatorial Pacific Ocean. By Fall (-1), a significant reversal in the prevailing easterlies and an eastward shift in the Indo-Australian precipitation center marks the Onset Stage of ENSO (Trenberth, 1991; Enfield, 1992). An anomalous band of warm waters also appears in the Equatorial Pacific Ocean and extends across to the South Pacific between 15°S and 30°S (Trenberth, 1991) while the Intertropical Convergence Zone (ITCZ) and South Pacific Convergence Zone (SPCZ) shifts away from their normal positions (Trenberth, 1976).

As the trade winds weaken or reverse directions, waters piled on the western Pacific relaxes forming eastward moving Kelvin waves. Around March (0) to May (0), a tongue of anomalously warm water appears signifying the "peak phase" of the ENSO Event in the eastern Pacific. Later, this merges with the warm SST anomalies that develop in the central Pacific Ocean around August (0) to October (0). ENSO reaches Maturity around the months of December (0) to February (+1), ending the event around March (+1) to April (+1) as SST anomalies collapse and normal wind and ocean conditions prevail. Positive SST anomalies, however, could remain in the central and eastern Pacific through early part of the year before finally collapsing back to normal conditions around June (+1) or later.

The 1982-'83 ENSO event deviated from the canonical ENSO in a number of unique ways (Trenberth, 1991; Cane, 1983). The wind anomalies that occurred around Fall (1981) did not materialize until Spring (1982). By July (1982) to August (1982), the wind anomalies were already sufficiently strong and persistent. Around May (1982), SST anomalies were already noticeable and by August (1982), SST warming was already substantial in the eastern Pacific Ocean. Also, in a reversal of the normal ENSO pattern, mid-oceanic warming did not lag behind the major anomalies at the South American coasts (Trenberth, 1991). Instead, peak SST anomalies were already reached in the western Pacific Ocean sometime in November (1982) to December (1982) while along the South American coasts, they were attained only in January (1983) to February (1983).

Another dramatic swing in the SO around February (1983) to March (1983) signaled the return of the atmosphere to its normal conditions. In the central and eastern equatorial Pacific, however, positive SST anomalies lingered on throughout the early part of 1983, making the 1982-'83 ENSO event the strongest and longest recorded event of the century (Trenberth, 1991; Enfield, 1992).

In the monsoonal system of Asia, ENSO's impact is felt primarily in the alteration of the normal precipitation regime (Trenberth, 1991; Enfield, 1992). The correlation between rainfall, drought and ENSO events was examined by Quinn, *et al.* (1978) for Indonesia and by Allen (1989), Allen *et al.* (1989) and McGregor (1989) for New Guinea. During an ENSO event, the Inter-Tropical Convergence Zone (ITCZ) and the Southern Pacific Convergence Zone (SPCZ) merge together as the two systems shift equatorwards and to the east. This displaces the precipitation center thousands of miles to the southeast and away from the typical monsoon regions of Southeast Asia (Rasmusson and Wallace, 1983; Barnett, 1991; Lau and Shea, 1991). Therefore, instead of a normal "dry" season, year-long extreme drought conditions prevail in Indonesia, the Philippines and northern Australia while catastrophic flooding and strong tropical hurricanes hit the normally arid South Sea Islands to the east (Trenberth, 1991; Enfield, 1992).

The effects of El Niño-Southern Oscillation on fisheries can be substantial. For example, Staples (1983), Vance *et al.* (1985), and Love (1987) conducted extensive studies on the banana prawn fishery of the Gulf of Carpentaria and found the highest correlation between prawn catches and rainfall in the Karumba Region. Strong correlations were also found between the spring (especially November) Southern Oscillation Index (SOI) and total seasonal banana prawn catch in the Gulf of Carpentaria (Staples, 1983; Love, 1987). Longhurst and Pauly (1987) and Raja (1972, 1973) found possible correlation between juvenile *Sardinella longiceps* (Indian oil sardine, family: Clupeidae) abundance in the Kerala-Mysore section of the Indian coast and rainfall during the peak spawning period of the preceding season. Following a catastrophic failure of the monsoon rainfall and the strongest ENSO event since 1826, *S. longiceps* disappeared along the Indian coasts in 1941 and did not recover until 1949 (Longhurst and Pauly, 1987).

MATERIALS AND METHODS

A. Materials:

1. Demersal Fish Samples.

Nine research cruises were completed between November 1981 to January 1983 on board the 411 gross ton research vessel *RV Sardinella* of the College of Fisheries, University of the Philippine in the Visayas (Table 1). The first three cruises were conducted at monthly intervals while the rest were programmed every other month. The sampling period encompassed the two major Southeast Asian monsoon seasons, i.e., the northeast monsoon (winter), southwest monsoon (summer), and the dry intermonsoon period (spring). Also, the last two cruises (November 26 to December 4, 1982 and January 13-20, 1983) serendipitously coincided with the peak and early maturity phases of the 1982-1983 El Niño-Southern Oscillation (ENSO) event (Glynn, 1990; Diaz and Kiladis, 1992). Since ENSO teleconnections are most evident during the northern winter (i.e., at the end of year 0 and into year +1) in the extratropics (Diaz and Markgraf, 1992; Nichols, 1992), the last cruise may shed some information on the potential influence of this worldwide event on Southeast Asian fisheries.

All feasible trawling grounds inside the Ragay Gulf, Burias Pass, Ticao Pass and Northern Samar Sea areas (referred to as RABUTINOS for brevity) were subdivided into 50 m depth sampling strata where the location of the 23 fishing tracks or sampling units (SU) used throughout the survey were selected at random (Figure 8). More closely spaced fishing stations were allotted south of Viñas River inside Ragay Gulf and near the entrance to Sorsogon Bay to account for the expected environmental variability and spatial heterogeneity at the shallowest stratum (i.e., < 50m depth), especially near the major point sources of freshwater (Megia *et al.*, 1953; Sebastian *et al.*, 1959).

Table 1. Dates of the Oceanographic Cruises Conducted Aboard the Research Vessel, *RV Sardinella* at Ragay Gulf, Burias Pass, Ticao Pass and Northern Samar Sea, Philippines.

<u>CRUISE NO.</u>	<u>CRUISE DATE</u>
CR 3	NOVEMBER 7 - 13, 1981
CR 4	DECEMBER 9 - 14, 1981
CR 5	JANUARY 21 - 27, 1982
CR 6	FEBRUARY 28 - MARCH 5, 1982
CR 8	MAY 14 - 28, 1982
CR 9	JULY 21 - 26, 1982
CR 10	SEPTEMBER 24 - 30, 1982
CR 12	NOVEMBER 26 - DECEMBER 4, 1982
CR 13	JANUARY 13 - 20, 1983

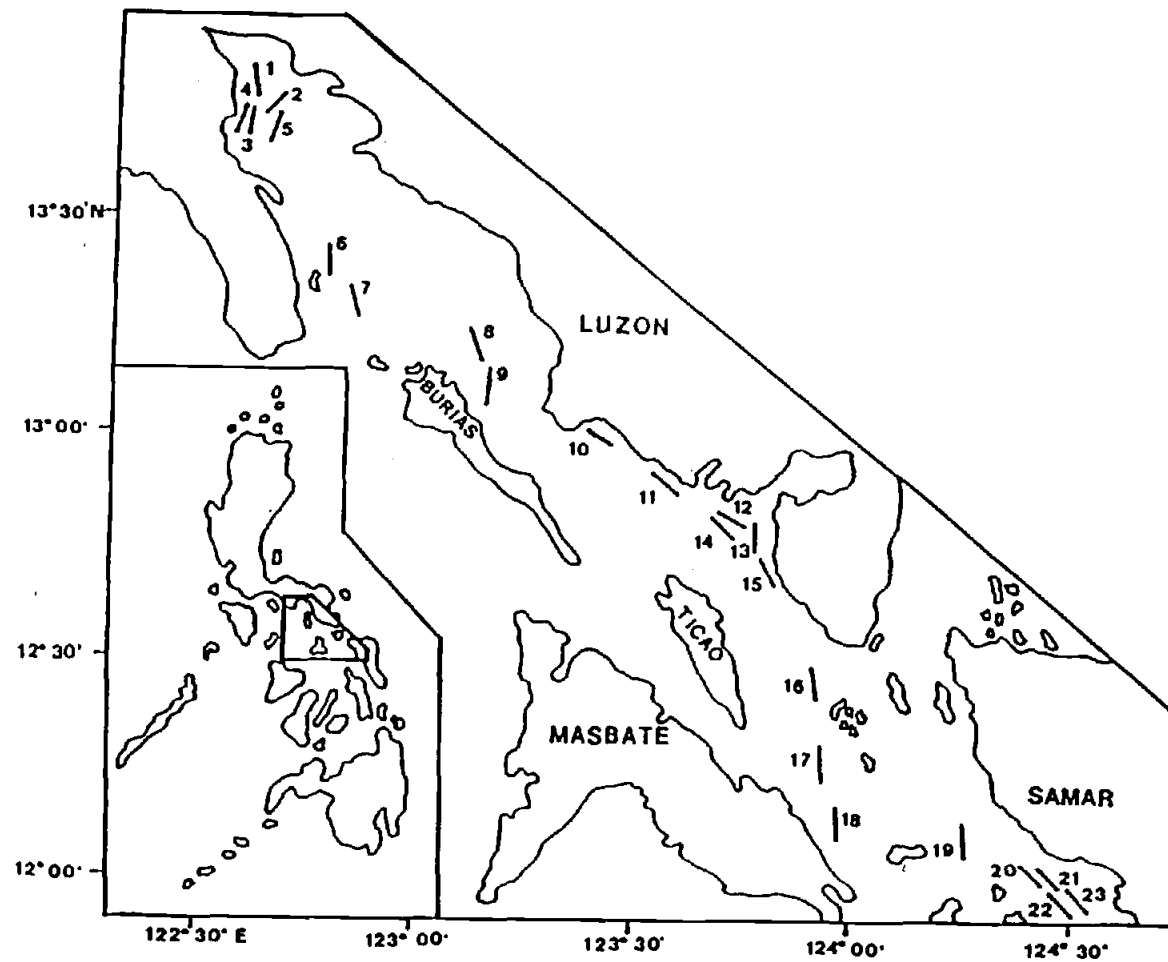


Figure 8. The Location of Fishing Stations (FS) at Ragay Gulf, Burias Pass, Ticao Pass and Northern Samar Sea, Philippines.

Each of the 23 SU's was systematically sampled in every cruise. Due to budgetary constraints, however, fishing stations FS7, FS10, FS12 and FS18 were not sampled on the last cruise. Standard one hour trawl drag was completed at each SU using a German standard 2-seam Engel design otter trawl net with a cod-end stretched mesh size of 50 mm and a headrope length of 35.3 m. Based on earlier test fishing and instrument calibration cruise, the net sonde established the average horizontal and vertical opening of the trawl to be 21 and 5 m respectively when towed at normal speeds of 3.5 knots (6.3 km/h). This gives an average swept area of 0.1323 km^2 per tow.

All trawling tows were conducted during daylight hours following standard trawl sampling and catch-handling procedures, as in Pauly (1983) and Sparre (1985). Briefly, this involves randomly taking subsamples of the catch for sorting, weighing, and identification of fishes down to the species level if possible. Jellyfishes, sponges, corals and sea snakes were noted and discarded (Watson *et al.*, 1990). The total catches of each species was estimated from these subsamples using ratio and proportion. Catches less than 150 kg were counted directly. All catches were converted into catch per hour. The fish identification manuals of Jones and Rosa (1965), Rau and Rau (1980), FAO (1974), and Gjosaeter and Kawaguchi (1980) were used as key taxonomic references. Specimens that were difficult to identify were frozen for later analysis in the laboratory.

At the end of the research project, a total of 200 successful hauls was completed yielding a data set composed of 199 species/taxa from 19 to 23 sampling stations per cruise and 9 sampling periods (cruises). A detailed narrative of the Ragay Gulf survey is given in Sambilay *et al.* (1990).

2. Environmental Samples.

A total of 21 oceanographic stations was systematically sampled during each cruise (Figure 9). At each oceanographic station, *in situ* temperature was measured and

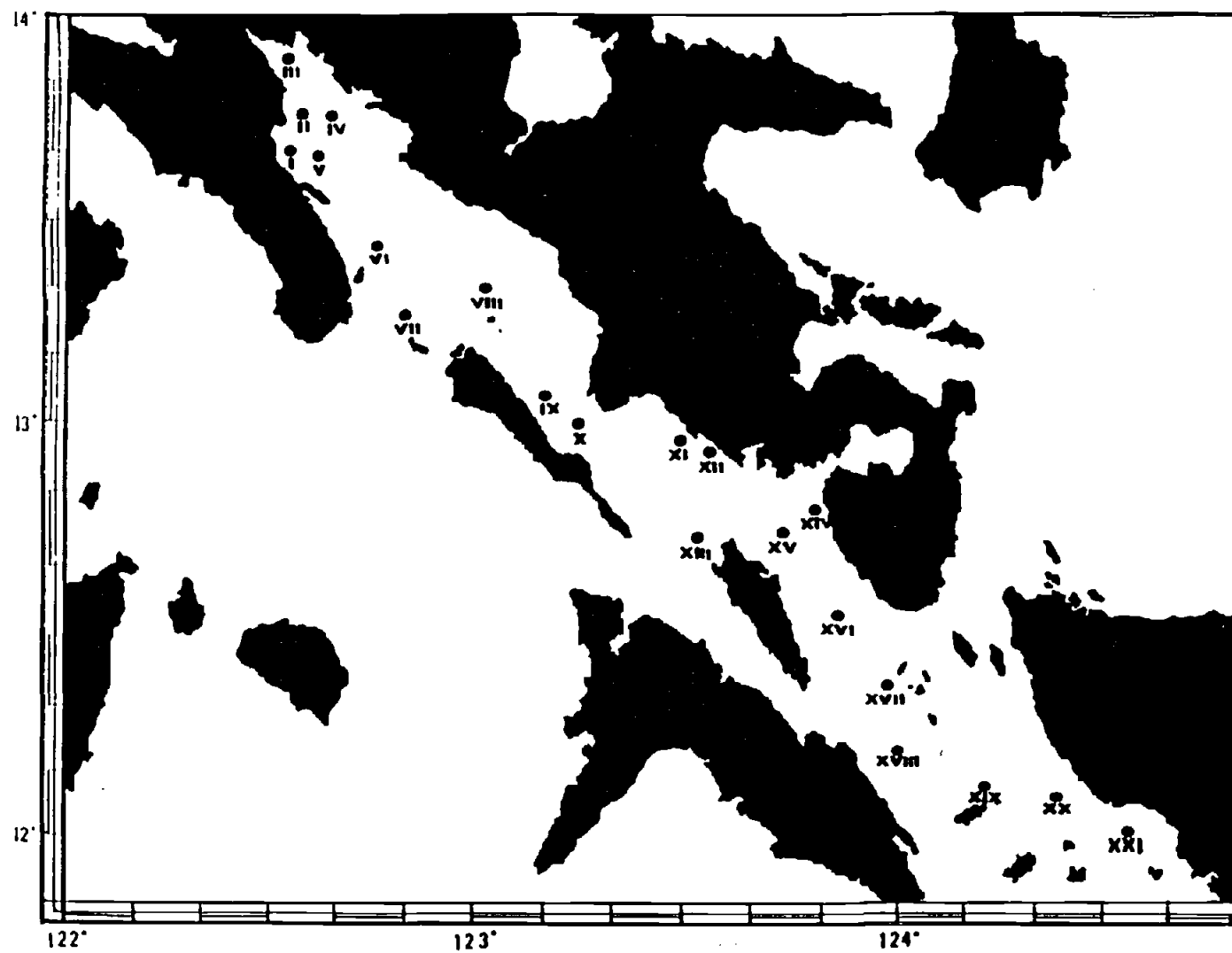


Figure 9. The Location of Oceanographic Stations (OS) at Ragay Gulf, Burias Pass, Ticao Pass and Northern Samar Sea, Philippines.

water samples taken at standard oceanographic depths, i.e., surface, 10, 20, 30, 50, 75, 100, 150, 200, 300, and 500 meters depending on the depth of the particular station. Whenever possible, additional measurements at approximately 1.5 m from the bottom were added to oceanographic stations near the start or end of each fishing track. Environmental characteristics of deep fishing stations were interpolated from the vertical profile(s) of the nearest oceanographic station(s) at similar depths (Bianchi, 1991). Sampling depths were recorded and monitored from the ship's Furuno scientific echosounder Model FE D824 at 1060 Hz and 28/200 Hz variable gain frequency. The ship's position was determined through radar triangulation and the ship's satellite navigation system.

Water temperatures were measured using reversing thermometers attached to Nansen bottles and corrected according to La Fond (1951) and Bialek (1966). Salinity was measured by a Tsurumi-Seiki salinometer periodically calibrated on standard sea water and the Mohr-Knudsen salinity titration procedure (Grashoff, 1976). Dissolved oxygen was determined using the modified Winkler titration method (Grashoff, 1976) and periodically checked with a Horiba water quality checker Model U-7.

All climatological data were derived from observations at the Masbate Island weather station furnished by the Philippine Atmospheric, Geophysical, and Astronomical Services Administration (PAGASA). Substrate types were determined from hydrographic charts published by the Philippine Coast and Geodetic Survey for the area and checked once by taking sediment samples at each fishing stations using a van Veen grab.

B. Methods:

1. General Analytical Procedures.

The analytical strategy outlined by Field *et al.* (1982) for studying multispecies distribution patterns in the oceans was followed with some modifications. This approach separates the search for patterns among the biological variables from attempts to interpret them in terms of the environmental data. This allows the species to “tell their story” without the influence of any previous assumptions about the relationships between the biota and its environment (Day *et al.*, 1971). Each sampling group’s environmental data was subjected to separate statistical tests and the ones that differ significantly were noted as being possible factors responsible for the biotic groups (Field, 1971; Field *et al.*, 1982). All standard multivariate analyses were done on PC-ORD version 2.0 (McCune and Mefford, 1995). Other statistical analyses were run on SAS Institute Statistical Programs (Ray, 1982; Cody and Smith, 1991).

Multivariate ordination and classification can be done on individual (i.e., monthly) cruises (e.g., Lasiak, 1984; McManus, 1985) or on data pooled over a period of two months (e.g., Bianchi, 1991; Blaber *et al.*, 1994), three months (Rainer and Munro, 1982) or more. For example, Ansari *et al.* (1995) subdivided the year into three seasons of four months each, i.e., pre-monsoon, monsoon, and post-monsoon and pooled his data accordingly, while over at the Gulf of Carpentaria in northern Australia, Sheaves (1998) subdivided the year into four seasons of three months each, i.e., pre-wet, wet, post-wet, and dry seasons. Pooling of similar sampling cruises was done to minimize redundancy, simplify analysis, even out noisy data and help reveal underlying ecological relationships without excessively smoothing out important community variations (Gauch, 1982; Digby and Kempton, 1987).

Taking into consideration that excessive agglomeration may smooth out important community variations, an objective approach was taken in deciding which cruises should

be combined together. Since this work is about seasonal fish assemblage patterns, cruise data were pooled based on the classification and ordination of biotic variables rather than on physical climatic subdivisions. This approach was used effectively by Watson *et al.* (1990) and Ansari *et al.* (1995) to reveal cluster groups of cruises that demonstrated seasonal trends.

2. Normal or Q-Type Analysis.

a. *Raw Data.* Q-type analysis is the most common type of analysis in community ecology, where n samples (fishing stations) are grouped according to similarity in s species/taxa or biotic composition (Field *et al.*, 1982; Ludwig and Reynolds, 1988). Biomass (wet weight) was used as the unit of abundance since analysis based on fish weights is more ecologically appropriate (Field *et al.*, 1982; Bianchi, 1991) and relevant to fisheries management than analysis based on numbers (McManus, 1985; Bianchi, 1991; Federizon, 1992; McManus *et al.*, 1996).

A three dimensional primary data matrix of fishing stations (sites) x species x cruise (times) was analyzed in sets of two dimensional data as:

(1) Species (row) x cruise (column) data matrix where each cell contains species abundances averaged over all of the fishing stations that were sampled during each cruise.

(2) Species (row) x stations (column) data matrix where each cell contains the average abundance of each species caught at a given fishing station for all cruises that comprise the given seasonal cluster.

b. *Transformation and Standardization.* The original abundance data were log transformed using the formula:

$$Y_{ij} = \log (X_{ij} + 1),$$

were X_{ij} is the raw data score of the i th species in the j th sample and Y_{ij} is the corresponding transformed score. Field *et al.* (1982), Gauch (1982) and McManus (1985) consider the logarithmic transformation reasonable for this type of data. This transformation has the effect of compressing the upper end of the measurement scale to prevent swamping of other data by the dominance of very large species or large catches of smaller species (Digby and Kempton, 1987; Blaber *et al.*, 1994) and to facilitate comparison between trawl samples (Watson *et al.*, 1990). This transformation was not necessary in TWINSpan analysis since the values were converted to a 1 to 5 scale based on the following lower class limits: 0, 2, 5, 10 and 20 kilograms.

c. *Ordination*. Since there is no one “correct” or ideal ordination or classification system, the most commonly used or highly recommended techniques available were empirically tested first to evaluate which would best represent the actual data set (Gauch, 1982; Krebs, 1989). The different ordination techniques tested and examples of their successful use in ecological research include: *Principal Components Analysis* (McCune, 1988; Ross and Doherty, 1994; Sheaves, 1996; Ponton and Copp, 1997), *Canonical Correlation Analysis* (ter Braak, 1986; Ponton and Copp, 1997), *Reciprocal Averaging* (Hill, 1973; Lasiak, 1984), *Detrended Correspondence Analysis* (Hill, 1979a; Hill and Gauch, 1980; Gauch, 1982; McManus, 1985; Bianchi, 1991; Metzeling, 1993; Marchant, *et al.*, 1994; McManus *et al.*, 1996; Miller and Death, 1997), *Non-Metric Multi-Dimensional Scaling* (Field *et al.*, 1982; Morin *et al.*, 1992; Connolly, 1994; Jones, *et al.*, 1996; Ohman, *et al.*, 1997) and *Bray-Curtis Ordination* (Beals, 1984; Watson, *et al.*, 1990). All analyses were tested using a variety of similarity measures available on the PC-ORD system and on log-transformed *versus* non-transformed data sets.

The different techniques were tested and ranked according to Gauch's (1982) basic performance evaluation criteria, i.e., effectiveness (summarizes the data well and

aids understanding), robustness (ability to produce good results whether a data set involves long or short community gradients, one or more gradients, high or low noise, large or small number of species and samples, etc.), and practicality (computational feasibility, ease of data handling, and interpretability of results).

Principal Components Analysis (PCA) and Canonical Correlation Analysis (CCA) gave the worst results (i.e., points were often clumped around a single spot and produced varying degrees of “arch effect” or quadratic distortion of the first axis onto the second axis due to the ordination’s inability to handle non-linear species response curve). Detrended Correspondence Analysis (DECORANA or DCA) and Reciprocal Averaging (RA) produced similar ordination groupings but RA suffered from extreme arch effect similar to PCA and CCA. Non-Metric Multi-Dimensional Scaling (NMDS) and Bray-Curtis Ordination (BCO) also produced similar patterns, but performance varied with the different seasonal data sets. Most of the ordination techniques were able to extract the major structure in the data and none performed perfectly all the time. After more than 300 test ordination runs on the PCORD program, DECORANA emerged to be the best technique for this particular data set, with BCO and NMDS ranked second.

DECORANA is a modification of Reciprocal Averaging (RA) developed to eliminate the “arch effect” through simple trend removal along successive axes, with optional rescaling of axes to remove compression of points and the distortion of relative distances at either end of the ordination (Digby and Kempton, 1987; Bianchi, 1991). Detrended Correspondence Analysis (DECORANA) is useful in ecological studies because it only assumes a simple unimodal species response curve (ter Braak and Prentice, 1988). Also, it does not assume linear relationships between species abundances and environmental variables like most other ordination techniques (Bianchi, 1991).

The DECORANA options on PC-ORD that were used in this study were:

Downweighting of Rare Species = yes

Rescale Axes = yes (default)

Rescaling Threshold = 0 (default)

Number of Segments = 26 (default)

DCA Output: List of Residuals, Eigenvalue, Scores, Graphs, Correlations

d. *Classification*. Evaluation procedures similar to the ordination techniques were also tested on two popular classification techniques in ecology: *Cluster Analysis* (Rainer and Munro, 1982; Lasiak, 1984; Ramm, *et al.*, 1990; Morin, *et al.*, 1992; Courtney *et al.*, 1995; Jones, *et al.*, 1996) and Two-Way Indicator Species AnalYsis or *TWINSPAN* (Hill, 1973; Hill, 1979b; Gauch, 1982; Bianchi, 1991; Metzeling, 1993; McManus, 1985; McManus, *et al.*, 1996). Both techniques gave comparable results. Since cluster analysis was the preferred method for classifying smaller numbers of objects (Digby and Kempton, 1987), it was used to classify the nine cruises into their respective seasonal groupings.

Cluster analysis is a special kind of hierarchical classification system based on a similarity matrix that involves grouping objects into distinctive subsets (Jain and Dubes, 1988). The Relative Euclidean Dissimilarity measure (RED) was the similarity measure used as it corrects and maintains the metric qualities of the Euclidean measure (Ludwig and Reynolds, 1988). The Unweighted Pair-Group Method using Arithmetic Averages (UPGMA) was used to objectively determine the seasonal grouping of cruises. The UPGMA algorithm computes the average similarity or dissimilarity of a candidate sampling unit to an extant cluster by weighting each sampling unit in that cluster equally regardless of its structural subdivision. Monte Carlo simulation studies proved UPGMA at *par* with single-link clustering when the data were perturbed (Cunningham and Ogilvie, 1972) and better than Ward's method on clusters of unequal sizes (Milligan and Isaac, 1980).

The PCORD - CLUSTER options used were:

Cluster Distance Measure = Relative Euclidean

Group Linkage Method = 4 (i.e., Group Average or UPGMA)

Dendogram Options = Log Transform Dendogram Scale, Single space

TWINSPAN was adopted for the classification of SU's and species into their respective site and species groupings because of its compatibility with DECORANA ordination. Both DECORANA and TWINSPAN are based on correspondence analysis which makes it possible to compare directly the classification from TWINSPAN and the ordination along the first axis of DECORANA (Digby and Kempton, 1987; Bianchi, 1991). TWINSPAN also produces a sorted, two-way community table in which species and stations are arranged along the major gradients within the data (Belbin, 1991; Blaber, *et al.*, 1994). This makes the interpretation of the *r*-mode (species/taxa groupings based on their distribution and abundance on similar stations) and *q*-mode (station groupings based on similar species composition) dendograms easier. Furthermore, the selection of TWINSPAN and DECORANA as the main classification and ordination techniques allow the direct comparison of this work with those of Federizon (1992) for Ragay Gulf and McManus (1985) for Samar Sea.

TWINSPAN (Hill, 1979b) is a polythetic, divisive classification technique that begins with all samples together in a single cluster before being successively divided into a hierarchy of smaller and smaller clusters (Gauch, 1982). The basic procedure starts with an ordination of the samples by Reciprocal Averaging. Species that characterize the axis extremes (i.e., the most dissimilar species) are emphasized to polarize the samples before division of the principal axis near the middle. The process is repeated on the two sample subsets until each cluster has no more than a chosen minimum number of members. In TWINSPAN, importance values are not used directly but are converted to a scale based on lower class limits.

Each seasonally pooled data set was classified by TWINSPAN and the SU clusters for each season were initially assigned a temporary name, e.g., a1, a2, a3, etc. for season X; b1, b2, b3, etc. for season Y; etc. When the number of groups formed within each seasonal classification becomes large, however, it is not always obvious which pairs

of groups from two or more classifications match. TWINSpan and correspondence ordination of the seasonal SU clusters were implemented to classify, order and match each seasonal classification category into a common or "standardized" scale (Gauch, 1982; Digby and Kempton, 1987). Hence, if the same cluster was initially assigned two different temporary names based on its appearance on two different seasons, on the second round of classification, they will be grouped together under the same cluster because of their perfect similarity. Once all of the clusters were delineated in this manner, the temporary names initially given to each of the seasonal SU clusters were then changed into their new standardized group names.

The TWINSpan options on PCORD used in this study were:

Pseudospecies Cut Levels = default

Maximum Number of Indicators per Division = 5 (default)

Maximum Level of Divisions = 6 (default)

Minimum Group Size for Division = 5 (default)

Maximum Number of Species in the Final Tabulation = 100 (default)

Instead of arbitrarily choosing a single similarity threshold to define cluster membership, more easily interpretable and ecologically reasonable clusters were obtained by "cutting joints" on the dendrogram (Helvey and Smith, 1985; Digby and Kempton, 1987; Dennis and Bright, 1988).

e. *Indicator Species*. After summarizing the fish assemblage patterns with the two complementary classification and ordination diagrams, the difference in species which caused the patterns were determined from the raw data (Field *et al.*, 1982). Since removing rare species has little effect on the results of community analysis (Stephenson and Cook, 1980; Long and Poiner, 1994), the top 20 most abundant species (60%-100% of the weight of all species/taxa in each group) were used as the basis for describing each group (Federizon, 1992; Cyrus and Blaber, 1992; Sheaves, 1998). More specifically, a group was qualitatively defined based on its five most dominant species/taxa or family,

its overall species richness (i.e., total number of species), and its exclusive members in the top 20 most abundant species/taxa (Robertson and Blake, 1990; Prochazka, 1998). Here a species/taxa is considered "exclusive" to the group if it only occurs in that particular group's top 20 most abundant species/taxa list of 20 taxa. In all, a total of 90 species/taxa (45% of total) were included in the top 20 list for each season. The diversity of the communities is represented by species richness which is equivalent to the total number of species/taxa comprising the group.

3. Inverse or R-Type Analysis.

The transpose of the q -type data matrix was used in the complementary r -type analysis (i.e., grouping of species based on their abundance in similar stations). Similar procedures and techniques outlined above were applied. The analysis of species groups in relation to the environmental data is more complex in the r -type analysis and simple tests for significant differences between groups are not appropriate (Field *et al.*, 1982). Therefore, its use was limited primarily to the analysis of species grouping derived from the TWINSpan two-way ordered table (Helvey and Smith, 1985; Dennis and Bright, 1988).

4. Relating Environmental Data to the Community Groups.

a. *Environmental Correlations.* The relationship between station groups and environmental variables was analyzed using the DECORANA joint plot option contained in the GRAPH menu of the PCORD program package. This also provides the option of correlating the ordination axes with environmental variables (depth and bottom temperature, salinity, and dissolved oxygen). Partial correlation and Pearson's rank correlation coefficients were calculated among the environmental variables and also

between each variable and the scores of the ordination axis. A significance level of $\alpha \leq 0.05$ was selected as the cut-off level for analysis.

b. *Comparison of Group Means.* Following Field *et al.*'s (1982) procedure, all observations on each environmental variable (e.g., salinity) of one SU group were compared with the corresponding observations of other SU groups. The ones that differ significantly were noted as being possible factors responsible for the biotic groups. Multiple comparison of means was analyzed using Analysis of Variance (ANOVA) and Fisher's Protected Least Significant Difference (PLSD) technique (Ray, 1982; Cyrus and Blaber, 1992). Fisher's PLSD test takes into account the unequal variances of each treatment (Day and Quinn, 1989; Cyrus and Blaber, 1992) and also offers more protection against incorrect inference, *vis-à-vis*, the simple Least Significant Difference (LSD) method. This is achieved simply by adding the restriction that the F-test for equal means must be significant at the 5% level (Ray, 1982), which fixes the experimental error rate at approximately five percent.

c. *Multiple Regression of Fish Abundance on Environmental Variables.* In the absence of evidence to the contrary, (e.g., inter/intra-specific competition and predation, ontogenetic behavior, symbiosis), it is reasonable to assume that environmental factors determine, directly or indirectly, the level of fish abundance. In these circumstances the use of regression analysis is more appropriate than a simple correlation coefficient concerned only with degrees of interdependence (Quinn, 1980; Greig-Smith, 1983; Cyrus and Blaber, 1992; Blaber *et al.*, 1995). Forward Stepwise multiple linear regression (Ray, 1982; Tabachnick and Fidell, 1989) was used to determine the direction and degree of relationship between the properties of the overlying water mass at a given fishing station and the observed abundance of each species/taxon. Bottom temperature, salinity, dissolved oxygen concentration and depth were the variables used to estimate the y values, i.e., the seasonal abundance of fishes expressed as average catch (wet weight) per hour.

A variable was included in the model only if it increases R^2 by 10% or more, maintains an overall significance level of $\alpha \leq 0.05$ or better, and retains a fairly acceptable Mallows's C_p statistics. A value of $C_p > p$ is evidence of biased estimation of parameters due to dropping of variables from the complete model while values of $C_p < p$ results from strong collinearity among one or more of the environmental variables (Philippi, 1993). Here, p is the number of variables in the model including the intercept. Since the regression models are only designed to explore possible relationships and not for prediction purposes, the application of Mallows's C_p criteria and significance testing were more relaxed (Philippi, 1987). For Mallows's $C_p < p$, Studenmund and Cassidy (1987) suggested doing nothing if the multicollinearity has not decreased t-scores to the point of insignificance. If Mallows's $C_p > p$, additional variable(s) were considered, provided the resulting model retained an acceptable R^2 and significance level.

5. Sampling Errors and Limitations.

Shallow-water (< 20 m) communities were not sampled adequately due to the draft limitations of the research vessel and the constraints imposed by existing fisheries regulations. Demersal trawls are both size- and species-selective, however it was impossible to adjust for this type of selectivity without knowing the behavior of most species and/or the real age/size structure of populations (Bianchi, 1991). In shallow waters (20 - 40 m deep), many typically pelagic species were also caught. At this depth, it was difficult to differentiate the demersal fishes from the small pelagics that feed on the bottom and also inhabit this zone as both groups have a much closer relationship in these shallow waters than offshore (Watson *et al.*, 1990; Bianchi, 1991; Blaber *et al.*, 1994). Therefore, pelagic species were also included in the analysis whenever they occurred in the samples. The same applies to the mesopelagic fishes caught by the trawl while carrying out diurnal vertical migrations in the deeper part of the shelf and upper slope.

RESULTS

A. Grouping of Cruises:

The natural grouping of the different cruises are shown in Figure 10. The cruises were clustered based on the similarity of their component fish assemblages. This process revealed four distinct groups of sampling cruises corresponding to the Southeast Asian dry and wet monsoon seasons. The result indicates seasonality in the composition and abundances of the demersal trawl fauna of RABUTINOS.

Wet Northeast Monsoon Season. This group is composed of Cruises 3 (November 1981), 4 (December 1981) and 5 (January 1982) and corresponds to the northern hemisphere winter months. Cool and wet maritime air mass brought about by strong and persistent winds coming from the northeast characterize this Asiatic monsoon.

Dry Intermonsoon Period. Cruises 6 (February-March 1982), 8 (May 1982) and 9 (July 1982) comprise this group that coincides with the spring transition and northern hemisphere spring season. Light breeze, very warm temperatures and relatively dry climatic conditions are typical during this period.

Wet Southwest Monsoon Season. This group consists of Cruises 10 (September 1982) and 12 (November-December 1982) corresponding to the northern hemisphere summer and fall seasons. Warm and rainy weather conditions punctuated by the passage of typhoons characterize the climate during these months.

Dry ENSO Event composed of Cruise 13 (January 1983) represents the peak and early maturity stages of the strong 1982-1983 ENSO event. Instead of joining the Northeast monsoon cluster, both the TWINSpan and Cluster Analysis separated Cruise 13 as totally different from the rest, indicating the uniqueness of the event.

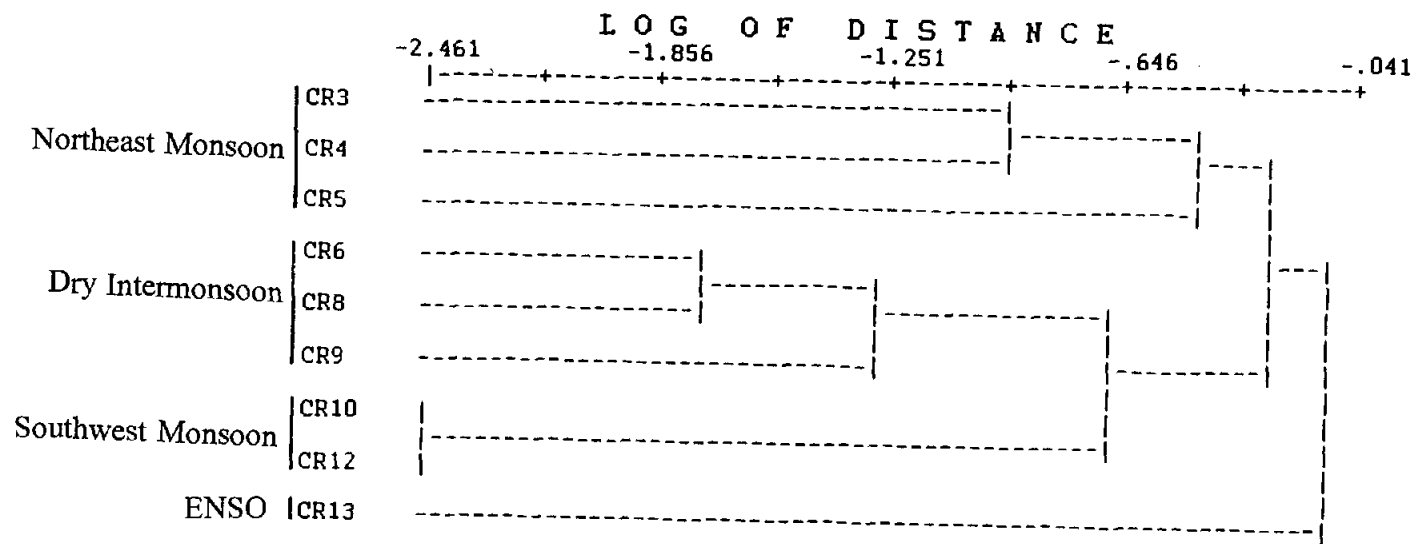


Figure 10. Seasonal Allocation of Cruises Based on Cluster Analysis of Ragay Gulf, Burias Pass, Ticao Pass and Northern Samar Sea Fish Abundance Data.

B. General Description of Trawl Catch:

This section presents the top 20 most abundant (by weight) species or taxa for the whole area and sampling cruises conducted at RABUTINOS. More than half of the 199 species/taxa is rare, each occurring in fewer than 20 of the 200 hauls. Despite the multispecies nature of tropical fisheries, about 60% to 100% of the catch is usually represented by just the top 10 or top 20 species which makes this list very valuable in describing and monitoring the development of any fishery. Both the overall aggregate species list and the seasonal trends in species abundances are included in this section for descriptive and comparative purposes. A list of the species arranged by families is provided in Appendix I.

1. Top 20 Most Abundant Species By Weight.

Table 2 lists the twenty most abundant (by wet weight) species or taxa caught by the sampling gear at RABUTINOS for the whole sampling period. Although this list represents only 11% of the total number of species/taxa (185) in the trawl samples, they account for approximately 3/4 (28 metric tons) of the all the samples caught (44 metric tons). Species dominance was not very pronounced. None of the species/taxa excessively dominated the top 20 most abundant species pool, with relative difference among adjacent species/taxa of 2% or less. Even the share of the top three species/taxa were comparable at 7% to 8%, although their combined abundances accounts for 22%.

At the family/group level, more than half of the catch came from the 18 families represented by the 20 most abundant species/taxa. They include: Tetraodontidae, Balistidae (*Abalistes stellaris*), Monacanthidae (*Alutera monoceros*), Diodontidae, Carangidae (*Carangoides speciosus*, *Decapterus muruadsi*), Centriscidae (*Aeliscus strigatus*), (Dasyatidae), Leiognathidae (*Leiognathus bindus*, *L. splendens*), Lethrinidae (*Lethrinus lentjan*), Mullidae (*Upeneus sulphureus*), Ariommidae (*Ariomma indica*),

Table 2. Top 20 Most Abundant (Wet Weight) Species/Taxa at RABUTINOS for all Cruises and Fishing Stations.

RANK	CODE	SPECIES	N	Total Weight (kgs)	%	Cumulative Frequency
1	ARIOMA	<i>Arioma indica</i>	207	3422.42	7.73%	7.73%
2	TRICHI	<i>Trichiurus haumela</i>	207	3202.82	7.23%	14.96%
3	LEOBIN	<i>Leiognathus bindus</i>	207	3201.80	7.23%	22.19%
4	DIODON	Diodontidae	207	2261.28	5.11%	27.30%
5	DASYAT	Dasyatidae	207	2035.21	4.60%	31.89%
6	CRXSPE	<i>Caranx speciosus</i>	184	1479.93	3.34%	35.23%
7	PRIAMA	<i>Priacanthus macracanthus</i>	207	1376.37	3.11%	38.34%
8	DECAMU	<i>Decapterus muruadsi</i>	207	1362.30	3.08%	41.42%
9	UPESUL	<i>Upeneus sulphureus</i>	207	1338.74	3.02%	44.44%
10	SAURUN	<i>Saurida undosquamis</i>	207	1209.98	2.73%	47.17%
11	TETRAO	Tetraodontidae	207	1163.43	2.63%	49.80%
12	LEOSPL	<i>Leiognathus splendens</i>	207	1083.46	2.45%	52.25%
13	ABALIS	<i>Abalistes stellaris</i>	207	839.15	1.89%	54.14%
14	LOLIGO	Loligo sp.	207	799.22	1.80%	55.95%
15	AELISC	<i>Aeliscus strigatus</i>	115	645.08	1.46%	57.40%
16	LTRILE	<i>Lethrinus lentjan</i>	184	603.04	1.36%	58.77%
17	ALUTMO	<i>Alutera monoceros</i>	207	560.77	1.27%	60.03%
18	SCOMME	<i>Scomberomorus commersonii</i>	207	498.25	1.13%	61.16%
19	LOPHII	Lophiidae	207	487.82	1.10%	62.26%
20	SHARKS	Sharks	207	484.08	1.09%	63.35%
		OTHERS				36.65%
		TOTAL (wt.)		44,286 kgs.		100.00%
		TOTAL (No. of Species)		185		

Trichiuridae (*Trichiurus haumela*), Lophiidae, Priacanthidae (*Priacanthus maculatus*), Synodontidae = Synodidae (*Saurida undosquamis*), sharks, Scombridae (*Scomberomorus commerson*), and the squids, Loliginidae (*Loligo* spp.).

2. Seasonal Trends in Species Abundance.

In order to determine if the four temporal groups derived by cluster analysis were distinct, the top 20 most abundant species/taxa were compared in this section (Figure 11). A total of 40 species/taxa belonging to 28 families was included in the combined top 20 most abundant species/taxa list. The habitat (depth) designations were derived from the TWINSPAN table discussed in later sections.

a. *Northeast Monsoon Season (NE).* The top five species/taxa during the wet northeast monsoon season include: Dasyatidae and Mobulidae (rays), *Ariomma indica* (Indian drift fish, Family: Ariommidae), *Lutjanus bojar* (snapper, Family: Lutjanidae) and Tetraodontidae (pufferfish). Together, they account for 22% of the catch while the top 20 listed species/taxa comprise 54% of the catch. The relative percentage abundance of the species/taxa comprising the catch during this season was relatively even with 17 families represented in the top 20 list. There were five exclusive members in this list, i.e., Mobulidae (rays), *Lutjanus bojar* (Family: Lutjanidae), *Selar crumenophthalmus* (bigeye scads, Family: Carangidae), *Epinephelus guttatus* (grouper, Family: Serranidae), and Ostraciidae (boxfishes). If the ENSO repetition (EN) is not included, the number of exclusive species/taxa is seven, with the addition of *Lethrinus opercularis* (emperors, Family: Lethrinidae) and *Saurida tumbil* (common saury, Family: Synodidae).

b. *Intermonsoon Period (INT).* During the dry intermonsoon period, the top five species/taxa include *Leiognathus bindus* (orange ponyfish, Family: Leiognathidae), *Ariomma indica* (Indian driftfishes, Family: Ariommidae), Diodontidae (pufferfish), *Carangoides speciosus* (jack) and *Decapterus muruadsi* (round scad, Family:

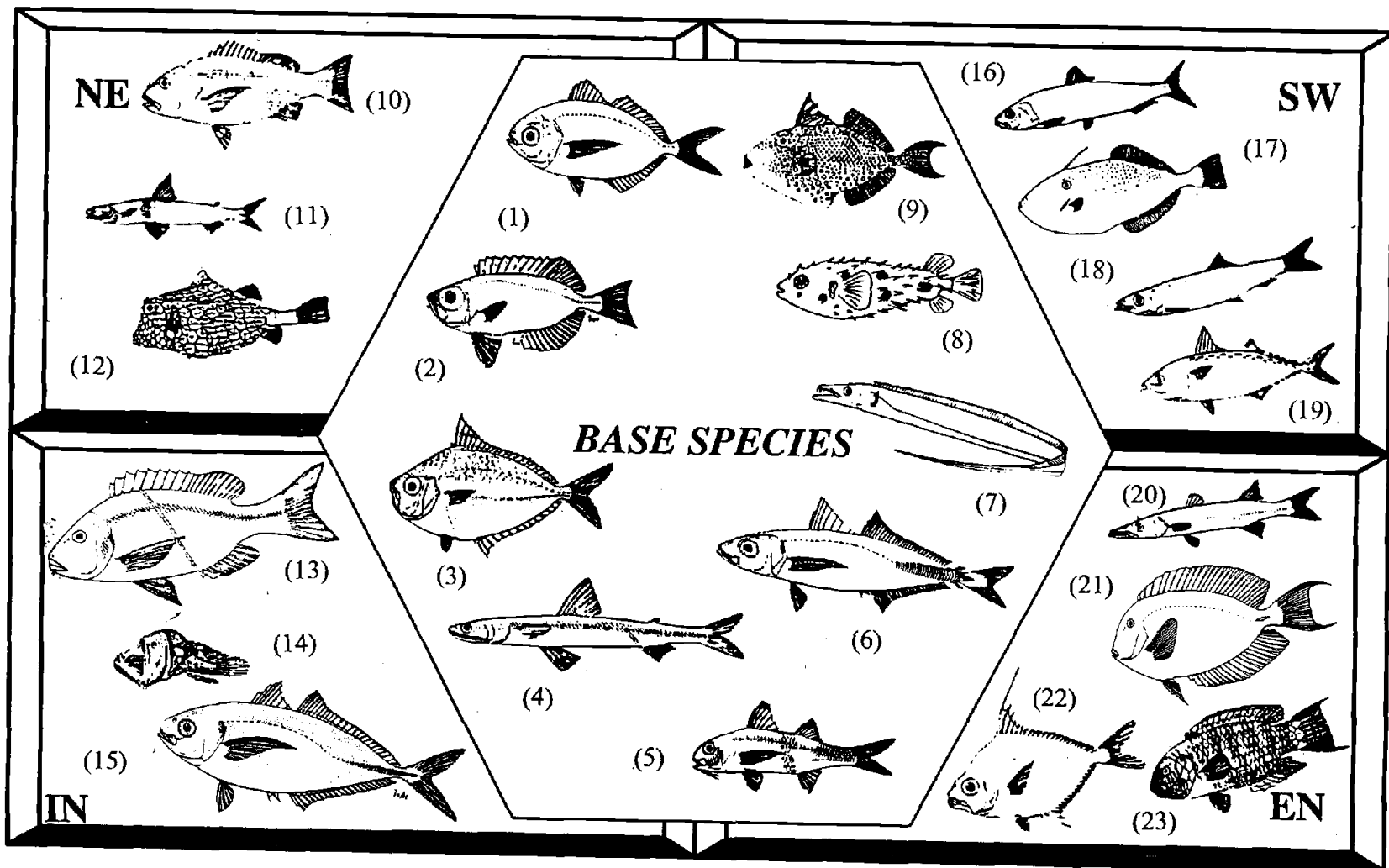


Figure 11. Seasonal Changes in the Top 20 Most Abundant Species/Taxa of RABUTINOS. (Numbers inside the parenthesis refer to the species' name in the index).

	SPECIES	Habitat	NE			INT			SW			EN		
			Mean	%	Rank	Mean	%	Rank	Mean	%	Rank	Mean	%	Rank
NE	<i>Dasyatidae</i>	D/U	12.52	5.21	1	7.64	2.84	9	6.39	2.97	8	19.81	10.72	2
	<i>Mobulidae</i>	D	10.87	4.52	2									
	<i>Ariomma indica</i> (14)	D	9.79	4.08	3	23.18	8.62	2	12.77	5.93	2	31.03	16.79	1
	<i>Lutjanus bojar</i> (10)	S	9.73	4.05	4									
	<i>Tetraodontidae</i>	S/C	8.74	3.64	5	6.82	2.54	11						
	<i>Priacanthus macracanthus</i> (2)	D	8.64	3.59	6	8.10	3.01	6	4.79	2.23	11			
	<i>Leiognathus bindus</i> (3)	S	8.20	3.41	7	31.52	11.72	1	10.34	4.8	4	2.70	1.46	15
	<i>Saurida undosquamis</i> (4)	D	6.92	2.88	8	7.80	2.9	8	3.77	1.75	13	2.89	1.56	13
	<i>Upeneus sulphureus</i> (5)	S	6.75	2.81	9	5.69	2.12	13	7.63	3.54	6	6.85	3.71	5
	<i>Decapterus muruadsi</i> (6)	D	6.28	2.61	10	9.20	3.42	5	5	2.32	10	3.47	1.88	12
	<i>Trichiurus haumela</i> (7)	D	5.85	2.43	11	6.58	2.45	12	46.97	21.8	1	7.89	4.27	4
	<i>Diodontidae</i> (8)	S	5.49	2.29	12	18.90	7.03	3	9.8	4.55	5	6.76	3.66	6
	<i>Loligo</i> sp.	D/U	5.19	2.16	13				1.82	0.84	20	6.73	3.64	7
	<i>Lethrinus opercularis</i>	S/C	4.04	1.68	14							3.79	2.05	10
	<i>Selar crumenophthalmus</i>	D/U	3.90	1.62	15									
	<i>Epinephelus guttatus</i>	S/C	3.65	1.52	16									
	<i>Carangoides speciosus</i>	S/C	3.62	1.51	17	10.73	3.99	4	12.01	5.57	3			
	<i>Saurida tumbil</i> (11)	D/U	3.61	1.50	18							3.63	1.97	11
	<i>Abalistes stellaris</i> (9)	S/C	3.57	1.49	19	4.30	1.6	16	5.76	2.67	9	1.97	1.07	20
	<i>Ostraciidae</i> (12)	S/C	3.37	1.40	20									
INT	<i>Aeliscus strigatus</i>	S/C				7.86	2.92	7						
	<i>Leiognathus splendens</i>	S				7.04	2.62	10	2.74	1.27	16	16.32	8.83	3
	<i>Lethrinus lentjan</i>	S/C				4.89	1.82	14						
	<i>Seriola grandis</i>	D				4.78	1.78	15						
	<i>Lophiidae</i> (14)	D				3.84	1.43	17						
	<i>Selaroides leptolepis</i> (15)	S				3.48	1.3	18						
	<i>Rhinobatidae</i>	D				3.26	1.21	19						
SW	<i>Lutjanus lineolatus</i>	S/C				3.20	1.19	20	7.02	3.26	7			
	<i>Sardinella longiceps</i> (14)	S							3.92	1.82	12			
	<i>Alutera monoceros</i> (17)	S/C							3.27	1.52	14			
	<i>Dussumiera acuta</i> (16)	S							2.83	1.31	15			
	<i>Centriscus scutatus</i>	S/C							2.63	1.22	17	2.32	1.25	17
	<i>Rastrelliger brachysoma</i> (19)	S							2.28	1.06	18			
EN	<i>Alectis ciliaris</i>	S							1.9	0.88	19			
	<i>Sphyræna obtusata</i> (20)	S										4.04	2.18	8
	<i>Seriolina nigrofasciata</i>	D/U										4.01	2.17	9
	<i>Gymnocaesio gymnoptera</i>	S										2.71	1.47	14
	<i>Acanthurus</i> sp. (21)	S/C										2.68	1.45	16
	<i>Leiognathus leuciscus</i> (22)	S										2.13	1.15	18
	<i>Labridae</i> (23)	S/C										2.00	1.08	19
	Weight (kg.)		240.30			268.92			215.41			184.80		
	Total No. of Species		178			170			150			133		

Legend: S = Shallow D = Deep C = Coralline U = Ubiquitous

NE = Northeast Monsoon Season

INT = Intermonsoon period

SW = Southwest Monsoon Season

EN = ENSO Early Maturity Stage

Figure 11. Seasonal Changes in the Top 20 Most Abundant Species/Taxa of RABUTINOS - Index to Species (continued).

Carangidae). Together they comprise 35% of this season's catch while the top 20 listed species/taxa account for 64%. Although *Leiognathus bindus* had a higher percentage share in abundance (12%) than during the NE monsoon, the contribution of each species/taxa was still relatively even. There were six exclusive members to this group, i.e., *Aeliscus strigatus* (shrimpfish, Family: Centriscidae), *Lethrinus lentjan* (red spot emperor, Family: Lethrinidae), *Seriola grandis* (trevally, Family: Carangidae), Lophiidae (goosefish), *Selaroides leptolepis* (Family: Carangidae), and Rhinobatidae (guitarfish). A total of 16 families was represented in the top 20 species list.

c. *Southwest Monsoon Season (SW)*. The rainy southwest monsoon season had *Trichiurus haumela* (hairtails, family: Trichiuridae), *Ariomma indica* (Indian driftfish, family: Ariommidae), *Carangoides speciosus* (jack, family: Carangidae), *Leiognathus bindus* (orange ponyfish, family: Leiognathidae) and Diodontidae in the top 20 most abundant species/taxa list. All five made up 43% of the total catch during this season while the top 20 species/taxa on the list comprise 71%. A total of 16 families were represented by the top 20 species/taxa with *Trichiurus haumela* (family: Trichiuridae) dominating the catch (22%), followed by other taxa with relatively even percentage abundances. Five species were exclusive to this season, i.e., two sardine species, *Sardinella longiceps* (Indian oil sardine) and *Dussumiera acuta* (rainbow sardine, family: Clupeidae); *Alutera monoceros* (unicorn filefish, family: Balistidae); *Rastrelliger brachysoma* (short-bodied mackerel, family: Scombridae); and *Alectis ciliaris* (pennantfish, family: Carangidae). *Centriscus scutatus* (shrimpfish, family: Centriscidae) of the ENSO group is not included in this category.

d. *ENSO Early Maturity Stage (EN)*. The dry ENSO early maturity period has the following species/taxa in the top 5 most abundant list: *Ariomma indica* (Indian driftfish, family: Ariommidae), Dasyatidae (ray), *Leiognathus splendens* (black-tipped ponyfish, family: Leiognathidae), *Trichiurus haumela* (hairtail, family: Trichiuridae) and *Upeneus sulphureus* (yellow goatfish, family: Mullidae). Together they accounted for 44% of the period's catch, while the top 20 species/taxa contributed 71%. *Ariomma indica* dominated

the catch but not as much as *Trichiurus haumela* during the previous season. A total of 16 families was represented in the top 20 species/taxa list. There were six exclusive species/taxa during this season, i.e., *Sphyraena obtusata* (obtuse barracuda, family: Sphyraenidae), *Seriolina nigrofasciata* (black-banded trevally, family: Carangidae), *Gymnocaesio gymnoptera* (snappers, family: Lutjanidae), *Acanthurus* spp. (unicornfishes, family: Balistidae), *Leiognathus leuciscus* and Labridae (wrasses).

e. *Summary of Abundance Patterns.* The overall sequence of the seasonal variation in the species composition of RABUTINOS shows a number of recurring patterns. For example, the total number of families represented in the top 20 species/taxa list is constant at about 16 families. This is primarily brought about by two factors: (1) the total number of exclusive members added to the top 20 list in each season varied between 5 to 6 new entrant species and (2) the commonly occurring species (i.e., present in all seasons) were also stable at 9 to 11 species. These fishes include: Dasyatidae (ray), *Ariomma indica* (Indian driftfish, family: Ariommidae), *Leiognathus bindus* (orange ponyfish, family: Leiognathidae), *Saurida undusquamis* (brushtooth lizardfish, family: Synodidae), *Upeneus sulphureus* (yellow goatfish, family: Mullidae), *Decapterus muruadsi* (round scad, family: Carangidae), *Trichiurus haumela* (hairtail, family: Trichiuridae), Diodontidae (pufferfish), *Carangoides speciosus* (jack, family: Carangidae), *Abalistes stellaris* (starry triggerfish, family: Balistidae) and *Priacanthus macracanthus* (red bigeye, family: Priacanthidae). Many of these common species/taxa were also included in the top 5 most dominant species list, compared to the new entrants that usually ranked 10th or lower in abundance.

In summary, the majority (2/3) of the species included in the top 20 fishes remained common throughout the year, including during the ENSO period. About 1/3 of the top 20 most abundant species/taxa occurred during one season only. This seasonal species/taxa turn-over resulted in compositionally distinct species assemblages. The geographic areas most affected by this seasonal fluctuation will be covered in the next section.

C. Results of Ordination and Classification:

This section presents the results of the ordination and classification of the trawl samples aimed at unraveling the species/taxa assemblage and station groupings that comprise the demersal fish communities of RABUTINOS. The seven major, seasonally standardized SU and species groups (see methods section for details of the standardization procedures) are presented and described first in order to provide the necessary biological context to the succeeding discussions. The seasonal species/taxa composition, geographical distribution and environmental correlation of the different groups are presented thereafter.

The TWINSpan dendrogram, DECORANA ordination diagram, and table of correlation between the different environmental variables and the ordination axes were combined in one figure to show their interrelationships better. In the TWINSpan dendrogram, the small letter-number combinations inside the parentheses refer to the temporary seasonal group designations assigned to each SU cluster prior to standardization. The large numbers/letters in bold font refer to the final group designations after standardization. Table 3 summarizes the relationships among fishing stations, temporary seasonal clusters and the final standard cluster designation for each season (or group of sampling cruises). Appendix II lists some of the habitat characteristics of the fishing stations.

In the case of DECORANA ordination diagrams, (e.g., Figure 12), numbers refer to the fishing stations while diamonds mark their positions in the ordination space. The relationship between environmental variables and species ordination scores are shown as radiating lines coming from the ordination centroid of the "joint plot." The angle and length of the line indicate the direction and relative strength of the relationship. Only environmental variables having an $r^2 \geq 0.2$ with the scores on either axis were plotted.

Axis scaling using PC-ORD's 'minimum to maximum' option was used to maximize the spread of points on the graph and provide better visual resolution of the gradients. The second axis is stretched more relative to the first due to differences in their eigenvalues.

1. Standardized SU Groups and Their Fish Assemblages.

a. *Standardization of Classes.* Seven major standard groups and two sub-groups were identified by TWINSpan based on the classification of all seasonal by derived clusters (Figure 12, upper diagram). Table 3 contains more detailed information on the temporary seasonal cluster designations. The first dichotomy separated the deep station (i.e., Groups 1 to 3) from the shallow fishing stations. The shallow fishing stations were further subdivided into the coralline station clusters (i.e., Group 7) near the Sorsogon Bay entrance and those further north (i.e., Group 5), including the riverine stations of Ragay Gulf and Northern Samar Sea (Groups 4A and 4B). The seasonal shifting of member species/taxa at the boundaries of Group 5 and Group 7 were captured by the transition Group 6. The major TWINSpan classes formed relatively distinct and easily separable groupings either on the first or second DCA axis (Figure 12, lower diagram). The eigenvalues of the first three DCA axes were 0.75, 0.33 and 0.23 respectively.

b. *Characteristic Species/Taxa of the Major Site Groups.* A description of the different species assemblages comprising the seven major SU groupings are presented next. Figure 13 graphically illustrates and summarizes the corresponding habitat characteristics and principal families and species that make up the major SU groups. Appendix I lists the fish species/taxa arranged by families while Appendix III gives a complete list of all the fish species/taxa included in each major site group. The fishes shown in the upper section of the figure represent the most dominant species/taxa and/or the indicator species for the given community. The assemblage provides a visual approximation of what the trawl catch looks like on the deck of the trawler ship prior to sorting, boxing and storage.

Table 3. Standardized Group Designation and Equivalent Preliminary Seasonal Group Designation of Sampling Units with their Component Fishing Stations.

TWINSpan GROUPING		CRUISE	FISHING STATIONS
Standard	Preliminary		
• <u>DEEP FISHING AREAS:</u>			
1	a3	3, 4 & 5	FS6
2	b3	6, 8 & 9	FS6, FS7, FS17, FS18
	c4	10 & 12	FS17, FS18
3	a4	3, 4 & 5	FS7, FS8, FS9, FS16, FS17, FS18, FS19
	b4	6, 8 & 9	FS8, FS9, FS16, FS19
	c3	10 & 12	FS6, FS7, FS9, FS16, FS19
	d2	13	FS6, FS7, FS8, FS9, FS16, FS17, FS18, FS19
• <u>SHALLOW RIVERINE/ESTUARINE FISHING AREAS:</u>			
4A	a2	3, 4 & 5	FS1, FS2, FS3, FS4, FS21, FS22, FS23
	b2	6, 8 & 9	FS1, FS2, FS3, FS4, FS21, FS22, FS23
	c2	10 & 12	FS1, FS2, FS3, FS4, FS5, FS21, FS22, FS23
	d1	13	FS1, FS2, FS3, FS4, FS5, FS20, FS21, FS22, FS23
4B	a1	3, 4 & 5	FS5, FS20
	b1	6, 8 & 9	FS5, FS20
	c1	10 & 12	FS8, FS20
5	b6	6, 8 & 9	FS10, FS11
	c6	10 & 12	FS10, FS11
• <u>SHALLOW CORALLINE FISHING AREAS:</u>			
6	a6	3, 4 & 5	FS10, FS11, FS12
	b5	6, 8 & 9	FS12, FS13, FS14, FS15
	d3	13	FS11, FS13, FS14, FS15
7	a5	3, 4 & 5	FS13, FS14, FS15
	c5	10 & 12	FS12, FS13, FS14, FS15

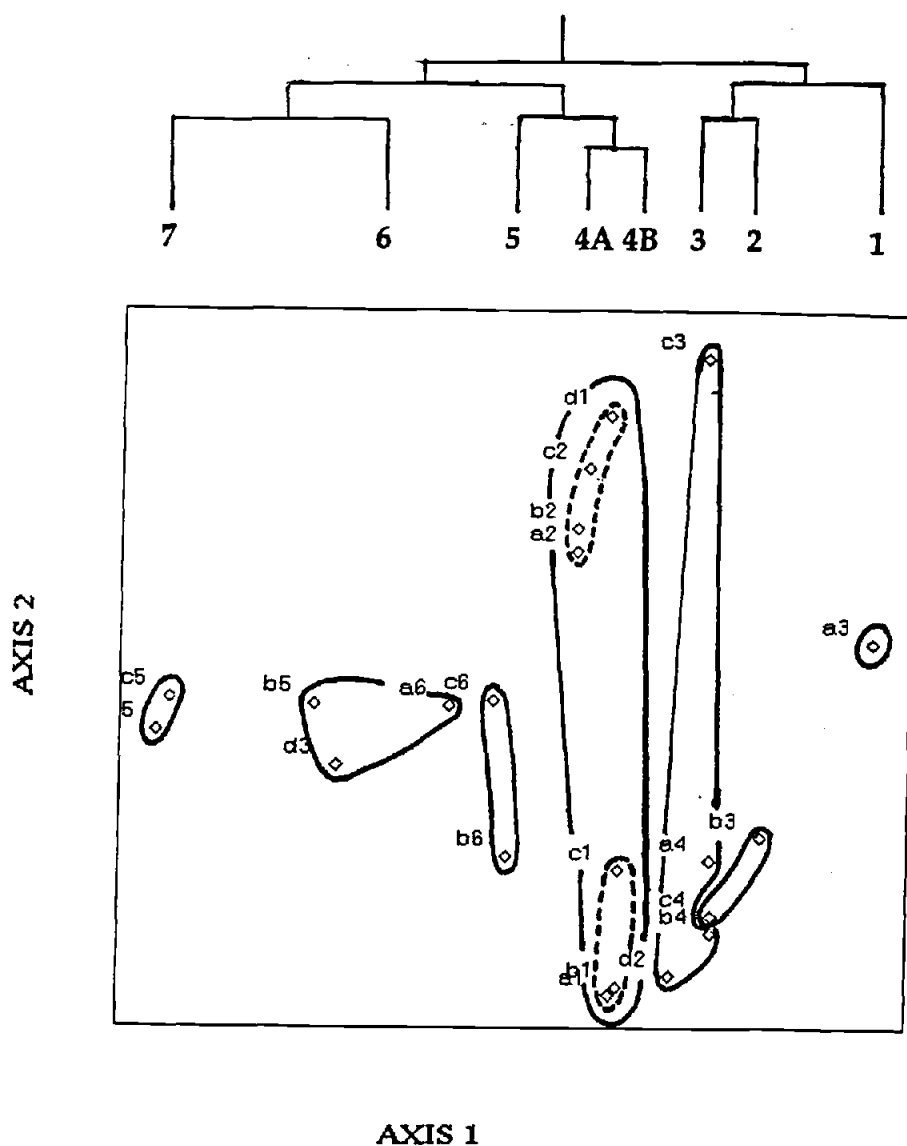


Figure 12. Results of the Q-Type Classification and Ordination Analyses on the Preliminary Seasonal Clusters. Upper diagram: TWINSpan dendrogram showing the "Standardized Group Designation." Lower diagram: DECORANA ordination plot showing the clustering of preliminary seasonal groups into standardized groups.

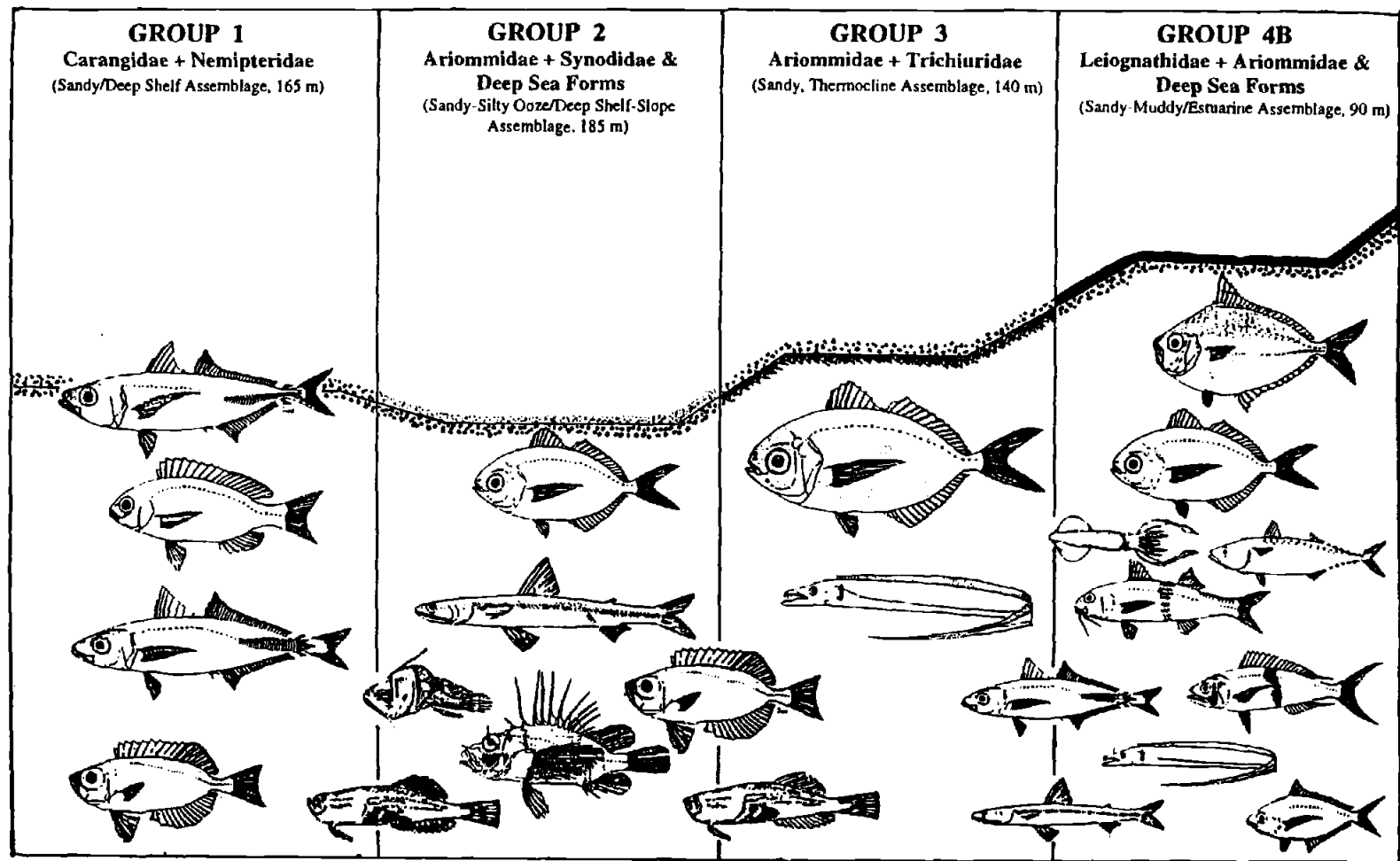


Figure 13. Schematic Representation of the Typical Species Composition and Habitat Characteristics of the Different Fish Assemblages found in RABUTINOS.

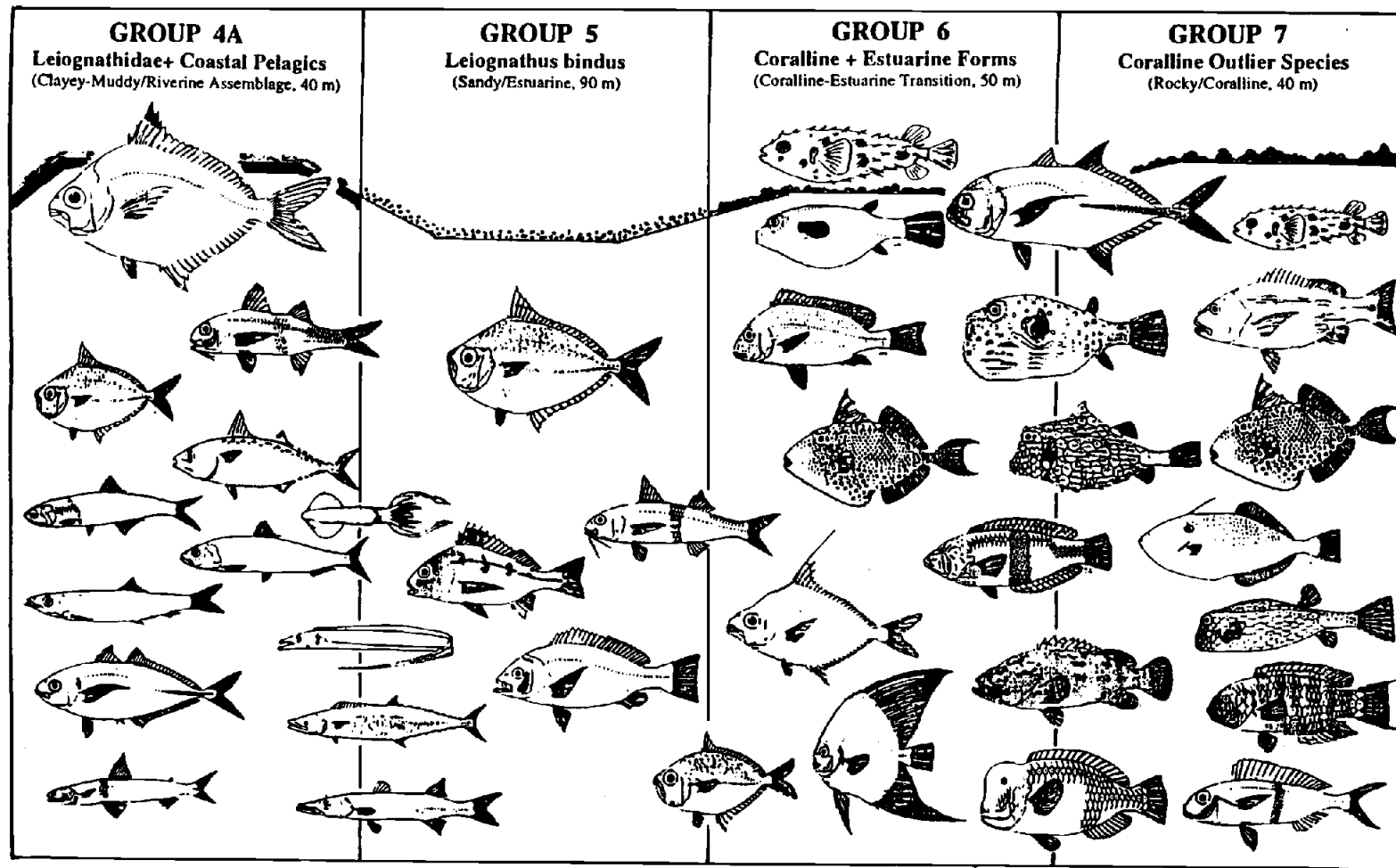


Figure 13 continued. Schematic Representation of the Typical Species Composition and Habitat Characteristics of the Different Fish Assemblages found in RABUTINOS.

Figure 13. (continued). Index to Species and Habitat Characteristics of the Fish Assemblage Groups.

GROUP 1	GROUP 2	GROUP 3	GROUP 4B
Primary Species: <i>Decapterus kurroides</i> Nemipteridae <i>Decapterus muruadsi</i>	Primary Species: <i>Arioma indica</i> <i>Saurida undosquamis</i>	Primary Species: <i>Arioma indica</i> <i>Trichiurus haumela</i>	Primary Species: <i>Leiognathus bindus</i> <i>Arioma indica</i> <i>Loligo sp.</i> Indicator Species: <i>Rastrelliger kanagurta</i>
Associated Species: <i>Priacanthus macracanthus</i> Lophiidae Uranoscopidae	Associated Species: Lophiidae <i>Priacanthus macracanthus</i> Scorpaenidae Uranoscopidae	Associated Species: <i>Priacanthus macracanthus</i> <i>Decapterus muruadsi</i> Uranoscopidae <i>Saurida undusquamis</i>	Associated Species: <i>Upeneus mollucensis</i> <i>Decapterus muruadsi</i> <i>Aphareus rutilans</i> <i>Trichiurus haumela</i> <i>Saurida undusquamis</i> <i>Pentaprion longimanus</i>
Habitat Characteristics: Substrate: Sandy Mean Temp.: 18.17°C Mean Salinity: 34.42‰ Mean DO: 1.80 ml/L Mean Depth: 166 m Others: Below Thermocline; Ragay Basin Sill; Wide Shelf N = 18 sp. ; CPUE = 101 kg	Habitat Characteristics: Substrate: Sandy-Silty Ooze Mean Temp.: 19.31°C Mean Salinity: 34.46‰ Mean DO: 2.39 ml/L Mean Depth: 185 m Others: Below Thermocline; Sill or Ridge; Wide Shelf N = 57 sp. ; CPUE = 236 kg	Habitat Characteristics: Substrate: Sandy-Hard Surface Mean Temp.: 21.98°C Mean Salinity: 34.46‰ Mean DO: 3.68 ml/L Mean Depth: 139 m Others: Within Thermocline; Sill or Ridge; Wide Shelf N = 87 sp. ; CPUE = 229 kg	Habitat Characteristics: Substrate: Sandy-Muddy Mean Temp.: 26.07°C Mean Salinity: 34.32 Mean DO: 5.18 ml/L Mean Depth: 88 m Others: Close to River; Near Shelf Break N 104 sp. ; CPUE = 148 kg

Figure 13. (continued). Index to Species and Habitat Characteristics of the Fish Assemblage Groups.

GROUP 4A	GROUP 5	GROUP 6	GROUP 7
Primary Species: <i>Leiognathus splendens</i> <i>Upeneus sulphureus</i> <i>Leiognathus bindus</i>	Primary Species: <i>Leiognathus bindus</i>	Primary Species: Diodontidae <i>Caranx speciosus</i> Triodontidae <i>Plectorhincus pictus</i> Triodontidae <i>Abalistes stellaris</i> Ostraciidae <i>Upeneus mollucensis</i> Labridae <i>Leiognathus fasciatus</i> Serranidae <i>Platax orbicularis</i> Leiognathidae (<i>Gazza minuta</i>) Scaridae	Primary Species: <i>Caranx speciosus</i> Diodontidae Triodontidae <i>Lutjanus bohar</i> Triodontidae <i>Abalistes stellaris</i> Ostraciidae Labridae <i>Alutera monoceros</i> Ostraciidae Serranidae Labridae Scaridae <i>Nemipterus peronii</i>
Indicator Species: <i>Rastrelliger brachysoma</i>			
Associated Species: Engraulidae (<i>Stolephorus indica</i>) Clupeidae (<i>Sardinella longiceps</i>) <i>Loligo sp.</i> Clupeidae (<i>Dussumiera acuta</i>) <i>Trichiurus haumela</i> <i>Scomberomorus commersonii</i> <i>Saurida tumbil</i> Sphyraenidae (<i>S. barracuda</i>)	Associated Species: <i>Upeneus mollucensis</i> <i>Loligo sp.</i> <i>Pomadasys maculatus</i> <i>Trichiurus haumela</i> <i>Lutjanus malabaricus</i> <i>Scomberomorus commersonii</i> <i>Sphyraena barracuda</i> <i>Gazza minuta</i>		
Habitat Characteristics: Substrate: Clayish-Muddy Mean Temp.: 25.94°C Mean Salinity: 34.26‰ Mean DO: 5.58 ml/L Mean Depth: 46 m Others: Riverine/Estuarine N = 120 sp. ; CPUE = 142 kg	Habitat Characteristics: Substrate: Sandy Mean Temp.: 25.74°C Mean Salinity: 34.39‰ Mean DO: 5.18 ml/L Mean Depth: 80 m Others: Intermittent River Influence; Narrow shelf N = 105 sp. ; CPUE = 355 kg	Habitat Characteristics: Substrate: Rocky-Coralline Mean Temp.: 25.42°C Mean Salinity: 34.32‰ Mean DO: 5.47 ml/L Mean Depth: 47 m Others: Near Entrance to Sorsogon Bay/Coral Reefs; N = 183 sp. ; CPUE = 339 kg	Habitat Characteristics: Substrate: Rocky-Coralline Mean Temp.: 26.35°C Mean Salinity: 34.27‰ Mean DO: 4.98 ml/L Mean Depth: 40 m Others: Near Entrance to Sorsogon Bay/Coral Reefs; Narrow Shelf N = 120 sp. ; CPUE = 489 kg

- **DEEP SEA STATIONS:**

GROUP 1

Key Species/Taxa: Carangidae and Nemipteridae

The main distinguishing features of this group are its very low species richness and the dominance of two fish families that account for 77% of the group's catch: Carangidae (*Decapterus kurroides*, *D. muruadsi*) and Nemipteridae (*Scolopsis inermis*, *Nemipterus marginatus*, *N. bathybius*). This may be a highly seasonal group.

Type of Substrate: Sandy to Silty Sand.

Average Depth: 165 meters

Average CPUE; Biomass: 101 kg/h; 8 kg/ha (0.76 mt/km²)

This group contains only 18 species/taxa (Table 4), the least among the seven groups. The five most abundant species (by wet weight) accounted for almost all of the catch (87%). These were: *Decapterus kurroides*, family: Carangidae (28%); *Scolopsis inermis*, family: Nemipteridae (24%); *Decapterus muruadsi*, family: Carangidae (18%); *Priacanthus macracanthus*, family: Priacanthidae (10%) and *Nemipterus marginatus*, family: Carangidae (8%). The exclusive species among this assemblage are the greeneyes (*Chlorophthalmus albatros*, family: Chlorophthalmidae), deepwater shrimps and crabs, grouper (*Epinephelus* sp., family: Serranidae), and threadfin bream (*Nemipterus marginatus*, family: Nemipteridae).

GROUP 2

Key Species/Taxa: Ariommidae + Mixture of Deep Sea Forms

The main distinguishing characteristic of this group is the dominance of *Ariomma indica*, family: Ariommidae, comprising 19% of the group's catch, stingrays (13%) and a mix of

Table 4. Top 20 Most Abundant (Net Weight) Species/Taxa Characteristic of Standardized Group 1.

Rank	SPECIES	Species Code	Mean	%	Cumulative Frequency
1	<i>Decapterus kurroides</i>	DECAKU	28.18	27.81%	27.81%
2	<i>Scolopsis inermis</i>	SCOLIN	23.83	23.51%	51.32%
3	<i>Decapterus muruadsi</i>	DECAMU	18.19	17.95%	69.27%
4	<i>Priacanthus macracanthus</i>	PRIAMA	9.77	9.64%	78.91%
5	<i>Nemipterus marginatus</i>	NEMIMA	8.50	8.39%	87.30%
6	<i>Saurida undosquamis</i>	SAURUN	4.27	4.21%	91.51%
7	<i>Epinephelus</i> spp.	EPISPP	4.00	3.95%	95.46%
8	<i>Chlorophthalmus albatros</i>	CHLORO	1.87	1.85%	97.31%
9	<i>Saurida tumbil</i>	SAURTU	1.67	1.65%	98.95%
10	Champsodontidae	CHAMPO	0.37	0.37%	99.32%
11	Shrimps	SHRIMP	0.19	0.19%	99.51%
12	Crabs	CRABSS	0.14	0.14%	99.64%
13	<i>Nemipterus bathybius</i>	NEMIBA	0.11	0.11%	99.75%
14	Rhinobatidae	RHINO	0.07	0.07%	99.82%
15	<i>Bothus</i> spp.	BOTHUS	0.06	0.06%	99.88%
16	<i>Rexea solandri</i>	REXEAS	0.06	0.06%	99.94%
17	Uranoscopidae	URANOS	0.04	0.04%	99.98%
18	<i>Fistularia petimba</i>	FISTUL	0.02	0.02%	100.00%

deep sea demersal forms, i.e., big-eyes (family: Priacanthidae), rockfishes (family: Triglidae), stargazers (family: Champsodontidae), anglerfishes (Lophiidae), Scorpaenidae (lionfishes), and lizardfishes (Synodidae) (47%) (Table 5). Together, they comprised 79% of this group.

Type of Substrate: Sandy/Silty Ooze (at Central Ragay Gulf) to Hard-Sandy Bottom (west of San Bernardino Strait).

Average Depth: 185 meters.

Average CPUE; Biomass: 236 kg/h; 18 kg/ha (1.78 mt/km²)

This group was represented by a total of 57 species/taxa and was second lowest in species richness among the seven groups (Appendix III). Group 2 is dominated by deep water fauna composed of *Ariomma indica*, family: Ariommidae (20% of the group's catch), *Saurida undusquamis*, family: Synodidae (12%), *Priacanthus macracanthus*, family: Priacanthidae (10%), Lophiidae (8%) and Dasyatidae (7%). These taxa comprise more than half (57%) of the fish landed in this group. The fish fauna that were exclusive to this group included members of the family Lophiidae (goosefish akin to anglerfish), Scorpaenidae (lionfish), *Decapterus russelli* (round scad, family: Carangidae), Triglidae (searobin related to scorpionfish) and Chimaeridae (ratfish).

GROUP 3

Key Species/Taxa: *Ariomma indica* + *Trichiurus haumela*

The key distinguishing feature of this group is that half of the catch is composed mainly of hairtails and Indian driftfishes (Table 6).

Type of Substrate: Sandy/Silty Ooze

Average Depth: 140 meters.

Average CPUE; Biomass: 229 kg/h; 17 kg/ha (1.73 mt/km²)

This group is dominated by only two species: *Ariomma indica*, family: Ariommidae (27%) and *Trichiurus haumela*, Trichiuridae (23%). Each of the remaining

Table 5. Top 20 Most Abundant (Net Weight) Species/Taxa Characteristic of Standardized Group 2.

Rank	SPECIES	Species Code	Mean	%	Cumulative Frequency
1	<i>Arioma indica</i>	ARIOMA	46.00	19.51%	19.51%
2	<i>Saurida undosquamis</i>	SAURUN	29.51	12.51%	32.02%
3	<i>Priacanthus macracanthus</i>	PRIAMA	23.43	9.94%	41.96%
4	Lophiidae	LOPHII	18.70	7.93%	49.89%
5	Dasyatidae	DASYAT	16.80	7.12%	57.01%
6	<i>Decapterus muruadsi</i>	DECAMU	15.66	6.64%	63.65%
7	Rhinobatidae	RHINO	14.25	6.04%	69.69%
8	<i>Trichiurus haumela</i>	TRICHI	9.14	3.88%	73.57%
9	Uranoscopidae	URANOS	6.93	2.94%	76.50%
10	<i>Nemipterus bathybius</i>	NEMIBA	5.66	2.40%	78.91%
11	<i>Champsodontidae</i>	CHAMPO	5.48	2.32%	81.23%
12	Scorpaenidae	SCORPI	4.90	2.08%	83.31%
13	<i>Decapterus russeli</i>	DECARU	4.81	2.04%	85.35%
14	Bothus spp.	BOTHUS	4.06	1.72%	87.07%
15	Triglidae	TRIGLI	3.88	1.64%	88.71%
16	Sharks	SHARKS	3.42	1.45%	90.16%
17	<i>Decapterus kurroides</i>	DECAKU	1.85	0.78%	90.94%
18	<i>Scolopsis inermis</i>	SCOLIN	1.84	0.78%	91.72%
19	<i>Rexea solandri</i>	REXEAS	1.76	0.75%	92.47%
20	Chimaeridae	CHIMAE	1.64	0.69%	93.17%

Table 6. Top 20 Most Abundant (Net Weight) Species/Taxa Characteristic of Standardized Group 3.

Rank	SPECIES	Species Code	Mean	%	Cumulative Frequency
1	<i>Arioma indica</i>	ARIOMA	61.36	26.81%	26.81%
2	<i>Trichiurus haumela</i>	TRICHI	53.32	23.29%	50.10%
3	<i>Priacanthus macracanthus</i>	PRIAMA	14.91	6.51%	56.61%
4	Dasyatidae	DASYAT	13.28	5.80%	62.42%
5	<i>Decapterus muruadsi</i>	DECAMU	10.40	4.55%	66.96%
6	<i>Saurida undosquamis</i>	SAURUN	9.04	3.95%	70.91%
7	Epinephelus spp.	EPISPP	8.15	3.56%	74.47%
8	<i>Seriola nigrofasciata</i>	SERION	6.88	3.00%	77.48%
9	Sharks	SHARKS	6.62	2.89%	80.37%
10	Loligo spp.	LOLIGO	5.49	2.40%	82.77%
11	Peristidiinae	PERIST	3.73	1.63%	84.40%
12	Lophiidae	LOPHII	3.15	1.38%	85.78%
13	Uranoscopidae	URANOS	2.63	1.15%	86.92%
14	<i>Upeneus vittatus</i>	UPEVIT	2.00	0.87%	87.80%
15	<i>Decapterus macrosoma</i>	DECAMA	1.95	0.85%	88.65%
16	<i>Nemipterus bathybius</i>	NEMIBA	1.85	0.81%	89.46%
17	Champsodontidae	CHAMPO	1.83	0.80%	90.26%
18	Tetraodontidae	TETRAO	1.53	0.67%	90.92%
19	<i>Selar crumenophthalmus</i>	SELACR	1.49	0.65%	91.57%
20	<i>Saurida tumbil</i>	SAURTU	1.27	0.56%	92.13%

85 species/taxa (Appendix III) comprise only small fractions of the total catch (Table 6). The top 20 most abundant species accounts for 92% of the group's total catch in this category. Exclusive to this list are *Seriolina nigrofasciata* (black-barred amberjack, family: Carangidae), Peristidiinae (lizardfish, family: Synodidae), Lophiidae (anglerfish), *Upeneus vittatus* (goatfish, family: Mullidae) and *Decapterus macrosoma* (roundscad, family: Carangidae).

- **SHALLOW RIVERINE/ESTUARINE STATIONS:**

GROUP 4A

Key Species/Taxa: Leiognathidae + Coastal Pelagics; neritic scombrid, *Rastrelliger brachysoma* (family: Scombridae) as indicator species.

The main distinguishing feature of this group is the dominance of Leiognathidae that accounts for 27% of the group's catch, Mullidae (10%) and coastal pelagics composed of Carangidae, Scombridae, Engraulidae and Clupeidae (25%) (Table 7). Together, they comprised 62% of the group's catch. The neritic species, *Rastrelliger brachysoma* are known to prefer shallow waters with muddy or clayish bottom (Jones and Rosa, 1965) and can be used as an indicator species for this group.

Type of Substrate: River Mud to Sandy Muddy.

Average Depth: 45 meters.

Average CPUE; Biomass: 142 kg/h; 11 kg/ha (1.07 mt/km²)

This shallow, riverine sub-group contains 120 species/taxa (Appendix III) in which the top 20 most abundant species account for 81% of the group's catch (Table 7). The predominant species in this category include: *Leiognathus splendens* and *L. bindus* (ponyfish, family: Leiognathidae), *Upeneus sulphureus* (goatfish, family: Mullidae), *Trichiurus haumela* (hairtails, family: Trichiuridae), and *Selaroides leptolepis* (yellow-

Table 7. Top 20 Most Abundant (Net Weight) Species/Taxa Characteristic of Standardized Group 4a.

Rank	SPECIES	Species Code	Mean	%	Cumulative Frequency
1	<i>Leiognathus splendens</i>	LEOSPL	18.25	12.85%	12.85%
2	<i>Upeneus sulphureus</i>	UPESUL	15.60	10.99%	23.84%
3	<i>Leiognathus bindus</i>	LEOBIN	14.43	10.16%	34.00%
4	<i>Trichiurus haumela</i>	TRICHI	11.86	8.35%	42.36%
5	<i>Selaroides leptolepis</i>	SELARO	5.96	4.20%	46.55%
6	<i>Rastrelliger brachysoma</i>	RASTBR	5.38	3.79%	50.34%
7	<i>Sardinella longiceps</i>	SARDLO	4.70	3.31%	53.65%
8	<i>Leiognathus leuciscus</i>	LEOLEU	4.57	3.22%	56.87%
9	<i>Loligo spp.</i>	LOLIGO	4.41	3.10%	59.97%
10	<i>Dussumiera acuta</i>	DUSSUM	4.24	2.98%	62.96%
11	<i>Sphyræna obtusata</i>	SPHYOB	3.86	2.72%	65.68%
12	<i>Decapterus muruadsi</i>	DECAMU	3.39	2.38%	68.06%
13	<i>Selar crumenophthalmus</i>	SELACR	3.19	2.25%	70.31%
14	<i>Scomberomorus commersoni</i>	SCOMME	3.14	2.21%	72.52%
15	<i>Saurida tumbil</i>	SAURTU	2.65	1.87%	74.39%
16	<i>Stolephorus tri</i>	STOLTR	2.13	1.50%	75.89%
17	<i>Uraspis helvolus</i>	URASPI	1.90	1.34%	77.22%
18	<i>Leiognathus equulus</i>	LEOEQU	1.81	1.27%	78.50%
19	<i>Saurida undosquamis</i>	SAURUN	1.77	1.25%	79.74%
20	Tetraodontidae	TETRAO	1.72	1.21%	80.95%

striped crevalle, family: Carangidae). Together, they represented almost half (46%) of this group's total catch.

Exclusive to this group are *Leiognathus splendens* (black-tipped ponyfish), *L. leuciscus* (whipfin ponyfish) and *L. equulus* (common ponyfish), family: Leiognathidae; *Selaroides leptolepis*, (yellow-striped trevally, family: Carangidae); *Rastrelliger brachysoma* (short-bodied mackerel, family: Scombridae); *Sardinella longiceps* (Indian oil sardine, family: Clupeidae); *Sphyraena obtusata* (obtuse barracuda, family: Sphyracidae); *Stolephorus tri* (anchovy, family: Engraulidae) and *Uraspis helvolus* (black ulua, family: Carangidae).

GROUP 4B

Key Species/Taxa: Leiognathidae + Ariommidae and other Deep Shelf Forms; oceanic scombrid, *Rastrelliger kanagurta* (family: Scombridae) as indicator species.

The main distinguishing feature of this group is the dominance of Leiognathidae that comprise 25% of this group's catch and deep sea forms like Ariommidae (11%), Loliginidae (8%), Mullidae (9%). The oceanic species, *Rastrelliger kanagurta*, known to prefer clear waters with salinities $\geq 34\text{‰}$ (Jones and Rosa, 1965) and can be used as an indicator species to separate this group from Group 4A.

Type of Substrate: Muddy to Sandy Muddy.

Average Depth: 88 meters.

Average CPUE; Biomass: 148 kg/h; 11 kg/ha (1.12 mt/km²)

This sub-group, composed of 104 species/taxa (Appendix III), represents the transition between the deep shelf and the shallow, riverine assemblages. The top 20 most abundant species/taxa account for 90% of the group's catch, while the top 5 species comprise more than half (54%) (Table 8). The predominant species include: *Leiognathus bindus* (slipmouth/ponyfish, family: Leiognathidae), *Ariomma indica* (Indian driftfish,

Table 8. Top 20 Most Abundant (Net Weight) Species/Taxa Characteristic of Standardized Group 4b.

Rank	SPECIES	Species Code	Mean	%	Cumulative Frequency
1	<i>Leiognathus bindus</i>	LEOBIN	34.69	23.36%	23.36%
2	<i>Arioma indica</i>	ARIOMA	16.36	11.02%	34.39%
3	<i>Loligo</i> spp.	LOLIGO	11.88	8.00%	42.39%
4	<i>Upeneus mollucensis</i>	UPEMOL	9.32	6.28%	48.66%
5	<i>Aphareus rutilans</i>	APHARE	8.26	5.56%	54.22%
6	<i>Decapterus muruadsi</i>	DECAMU	7.99	5.38%	59.60%
7	<i>Eupleurogrammus nuticus</i>	EUPLEU	7.09	4.78%	64.38%
8	<i>Saurida undosquamis</i>	SAURUN	6.57	4.43%	68.81%
9	<i>Fistularia petimba</i>	FISTUL	4.85	3.27%	72.08%
10	<i>Saurida tumbil</i>	SAURTU	4.83	3.25%	75.33%
11	<i>Dussumiera acuta</i>	DUSSUM	3.29	2.22%	77.55%
12	<i>Pentaprion longimanus</i>	PENTAP	3.26	2.20%	79.75%
13	<i>Rastrelliger kanagurta</i>	RASTKA	2.62	1.76%	81.51%
14	<i>Trichiurus haumela</i>	TRICHI	2.21	1.49%	83.00%
15	<i>Gymnocaesio gymnoptera</i>	GYMNOS	2.15	1.45%	84.45%
16	<i>Upeneus sulphureus</i>	UPESUL	2.08	1.40%	85.85%
17	<i>Leiognathus elongatus</i>	LEOELO	1.56	1.05%	86.90%
18	<i>Upeneus</i> spp.	UPESPP	1.37	0.93%	87.82%
19	Tetraodontidae	TETRAO	1.35	0.91%	88.73%
20	Rhinobatidae	RHINO	1.33	0.90%	89.63%

family: Ariommidae), *Loligo* spp. (squid, family: Loliginidae), *Upeneus mollucensis* (goatfish, family: Mullidae) and *Aphareus rutilans* (small-tooth jobfish, family: Lutjanidae).

Exclusive members of this sub-group were: *Aphareus rutilans* (small-tooth jobfish, family: Lutjanidae), *Eupleurogrammus nuticus* (Malayan hairtail, family: Trichiuridae), *Pentaprion longimanus* (longfin mojarra, family: Gobiidae), *Rastrelliger kanagurta* (Indian mackerel, family: Scombridae), *Gymnocaesio gymnoptera* (family: Lutjanidae) and *Upeneus* spp. (goatfish, family: Mullidae).

GROUP 5

Key Species/Taxa: Leiognathidae, primarily *Leiognathus bindus*.

This group represents the main fishing ground for Leiognathidae where it comprises about 40% of the catch.

Type of Substrate: Sandy.

Average Depth: 80 meters.

Average CPUE; Biomass: 355 kg/h; 27 kg/ha (2.68 mt/km²)

This group contains 105 species (Appendix III), with the top 20 most abundant species/taxa accounting for 80% of the group's catch (Table 9). It is dominated largely by the family Leiognathidae with the orange pony, *Leiognathus bindus*, comprising the bulk of the catch (34%). Other associated species/taxa include: *Upeneus sulphureus* (yellow goatfish) and *Upeneus mollucensis* (gold band goatfish, family: Mullidae); *Pomadasys maculatus* (blotched grunt, family: Pomadasyidae = Haemulidae); and Dasyatidae (stingray), together comprising 53% of the group's catch.

The exclusive species comprising this group include: *Pomadasys maculatus* (blotched grunt) family: Haemulidae; *Alectis ciliaris* (pennantfish), *Carangoides dinema* (shadow kingfish) and *C. ciliaris* (longfin cavalla), family: Carangidae; *Formio niger*

Table 9. Top 20 Most Abundant (Net Weight) Species/Taxa Characteristic of Standardized Group 5.

Rank	SPECIES	Species Code	Mean	%	Cumulative Frequency
1	<i>Leiognathus bindus</i>	LEOBIN	119.19	33.60%	33.60%
2	<i>Upeneus sulphureus</i>	UPESUL	22.43	6.32%	39.93%
3	<i>Pomadasys maculatus</i>	POMAMA	19.70	5.55%	45.48%
4	Dasyatidae	DASYAT	14.16	3.99%	49.47%
5	<i>Upeneus mollucensis</i>	UPEMOL	11.28	3.18%	52.65%
6	<i>Lutjanus malabaricus</i>	LUTMAL	10.57	2.98%	55.63%
7	<i>Trichiurus haumela</i>	TRICHI	10.36	2.92%	58.55%
8	<i>Fistularia petimba</i>	FISTUL	8.83	2.49%	61.04%
9	<i>Gazza minuta</i>	GAZAMI	7.63	2.15%	63.19%
10	<i>Caranx ciliaris</i>	CRXCIL	7.07	1.99%	65.19%
11	<i>Alectis ciliaris</i>	ALECIL	6.95	1.96%	67.14%
12	<i>Leiognathus fasciatus</i>	LEOFAS	6.57	1.85%	69.00%
13	<i>Formio niger</i>	FORMIO	6.31	1.78%	70.77%
14	<i>Sphyraena forsteri</i>	SPHYFO	6.21	1.75%	72.53%
15	<i>Scomberomorus commersoni</i>	SCOMME	5.78	1.63%	74.15%
16	<i>Leiognathus elongatus</i>	LEOELO	4.67	1.32%	75.47%
17	<i>Lutjanus bojar</i>	LUTBOJ	4.35	1.23%	76.70%
18	Diodontidae	DIODON	4.24	1.19%	77.89%
19	<i>Carangoides dinema</i>	CRXDIN	3.66	1.03%	78.92%
20	<i>Epinephelus tauvina</i>	EPITAV	3.63	1.02%	79.95%

(black pomfret, family: Formionidae); *Sphyraena forsterii* (Forster's barracuda, family: Sphyraenidae); and *Epinephelus tauvina* (greasy grouper, family: Serranidae).

- **SHALLOW CORALLINE STATIONS:**

GROUP 6

Key Species/Taxa: Coralline Outliers + Leiognathidae and Haemulidae.

This group is characterized by a large number of coralline and estuarine species that are evenly distributed in the catch.

Type of Substrate: Sandy to Rocky/Coralline Rubble Mix.

Average Depth: 50 meters.

Average CPUE; Biomass: 339 kg/h; 69 kg/ha (2.56 mt/km²)

A total of 183 species/taxa comprised this group (Appendix III). The top 20 most dominant species amounts to 62% of the group's catch, while the top 5 species/taxa, consisting mainly of Dasyatidae (stingray), Diodontidae (porcupinefish), *Carangoides speciosus* (crevalla, family: Carangidae), Tetraodontidae (pufferfish), and *Aeliscus strigatus* (shrimpfish, family: Centriscidae), account for 36% (Table 10).

Two members were exclusive to this group, i.e., *Plectorhincus pictus* (painted sweetlip, family: Haemulidae) and *Platax orbicularis* (batfish, family: Platacidae). Basically, Group 6 is a transition group containing a mixture of coralline and estuarine (Group 5) assemblages.

GROUP 7

Key Species/Taxa: Coralline Outliers; < 1% Leiognathidae, Pomadasidae, Mullidae.

Table 10. Top 20 Most Abundant (Net Weight) Species/Taxa Characteristic of Standardized Group 6.

Rank	SPECIES	Species Code	Mean	%	Cumulative Frequency
1	Diodontidae	DIODON	40.64	11.99%	11.99%
2	Dasyatidae	DASYAT	25.76	7.60%	19.59%
3	<i>Carangoides speciosus</i>	CRXSPE	22.93	6.76%	26.35%
4	<i>Aeliscus strigatus</i>	AELISC	16.35	4.82%	31.17%
5	Tetraodontidae	TETRAO	15.87	4.68%	35.86%
6	<i>Leiognathus bindus</i>	LEOBIN	8.91	2.63%	38.48%
7	<i>Plectorhincus pictus</i>	PLECIN	8.50	2.51%	40.99%
8	<i>Abalistes stellaris</i>	ABALIS	7.93	2.34%	43.33%
9	<i>Lutjanus lineolatus</i>	LUTLIN	7.64	2.26%	45.59%
10	<i>Lethrinus opercularis</i>	LTRIOP	7.00	2.07%	47.65%
11	<i>Gazza minuta</i>	GAZAMI	6.28	1.85%	49.50%
12	Ostraciidae	OSTRAC	6.21	1.83%	51.34%
13	<i>Alutera monoceros</i>	ALUTMO	6.00	1.77%	53.11%
14	<i>Upeneus mollucensis</i>	UPEMOL	5.50	1.62%	54.73%
15	Labridae	LABRID	4.73	1.39%	56.12%
16	Centriscidae	CENTRI	4.60	1.36%	57.48%
17	<i>Platax orbicularis</i>	PLATAX	4.58	1.35%	58.83%
18	<i>Leiognathus fasciatus</i>	LEOFAS	4.53	1.34%	60.17%
19	<i>Acanthurus</i> spp.	ACANSP	4.25	1.25%	61.42%
20	<i>Acanthurus bleekeri</i>	ACANBL	4.08	1.20%	62.62%

This group is distinguishable by the absence or rarity of Leiognathidae (slipmouths or ponyfishes), Haemulidae (grunts) and Mullidae (goatfishes).

Type of Substrate: Rocky/Coralline Mix.

Average Depth: 40 meters.

Average CPUE; Biomass: 489 kg/h; 37 kg/ha (3.69 mt/km²)

Group 7 consisted of 120 coralline outlier species (Appendix III). The top 20 most abundant species/taxa accounted for 76% of the group's catch, while the top 5 percent in the list contributed 40% of the catch (Table 11). There is no dominant family. Exclusive members of this group included: *Lethrinus lentjan* (red spot emperor, family: Lethrinidae), *Naso* spp. (surgeonfish/unicornfish, family: Acanthuridae), *Epinephelus guttatus* (grouper, family: Serranidae), *Macolor macolor* (snapper, family: Lutjanidae), Scaridae (parrotfish), and *Nemipterus peronii* (Peron's butterfly bream, family: Nemipteridae).

In general, the above assemblage groupings were well separated by the two numerical techniques. What is more important is that the resulting group also made biological sense, i.e., using the categories described and illustrated in this section, visual discrimination and classification of trawl catches can easily be made based only on their distinct faunal composition and relative abundance (see Appendix V).

2. Species Clusters.

The resemblance among species based on the similarity of their station/habitat characteristics (r-type analysis) is shown as a TWINSpan two-way ordered classification table (Figure 14). Due to the graphics limitation of the PC-ORD program, this diagram was abbreviated and shows only the 100 most important species/taxa. The site classification portion, i.e., q-type analysis, basically shows the seven major cluster groupings described in Figure 12 that represent the three basic assemblage/habitat types, i.e., deep marine, shallow riverine/estuarine, and shallow coralline SU groups.

Table 11. Top 20 Most Abundant (Net Weight) Species/Taxa Characteristic of Standardized Group 7.

Rank	SPECIES	Species Code	Mean	%	Cumulative Frequency
1	<i>Carangoides speciosus</i>	CRXSPE	47.55	9.73%	9.73%
2	Diodontidae	DIODON	47.25	9.67%	19.40%
3	<i>Lutjanus bojar</i>	LUTBOJ	37.29	7.63%	27.04%
4	Dasyatidae	DASYAT	34.94	7.15%	34.19%
5	<i>Abalistes stellaris</i>	ABALIS	25.98	5.32%	39.51%
6	Tetraodontidae	TETRAO	22.02	4.51%	44.02%
7	<i>Alutera monoceros</i>	ALUTMO	18.58	3.80%	47.82%
8	<i>Lethrinus lentjan</i>	LTRILE	14.82	3.03%	50.85%
9	<i>Lethrinus opercularis</i>	LTRIOP	14.69	3.01%	53.86%
10	Ostraciidae	OSTRAC	14.46	2.96%	56.82%
11	<i>Epinephelus guttatus</i>	EPIGUT	14.00	2.87%	59.69%
12	Labridae	LABRID	12.38	2.53%	62.22%
13	Naso spp.	NASOSP	10.36	2.12%	64.34%
14	Acanthurus spp.	ACANSP	9.45	1.94%	66.28%
15	<i>Macolor macolor</i>	MACOLO	9.33	1.91%	68.19%
16	<i>Lutjanus lineolatus</i>	LUTLIN	8.89	1.82%	70.01%
17	Centriscidae	CENTRI	8.76	1.79%	71.80%
18	Scaridae	SCARID	7.76	1.59%	73.39%
19	<i>Aeliscus strigatus</i>	AELISC	6.88	1.41%	74.79%
20	<i>Nemipterus peronii</i>	NEMIPE	6.74	1.38%	76.17%

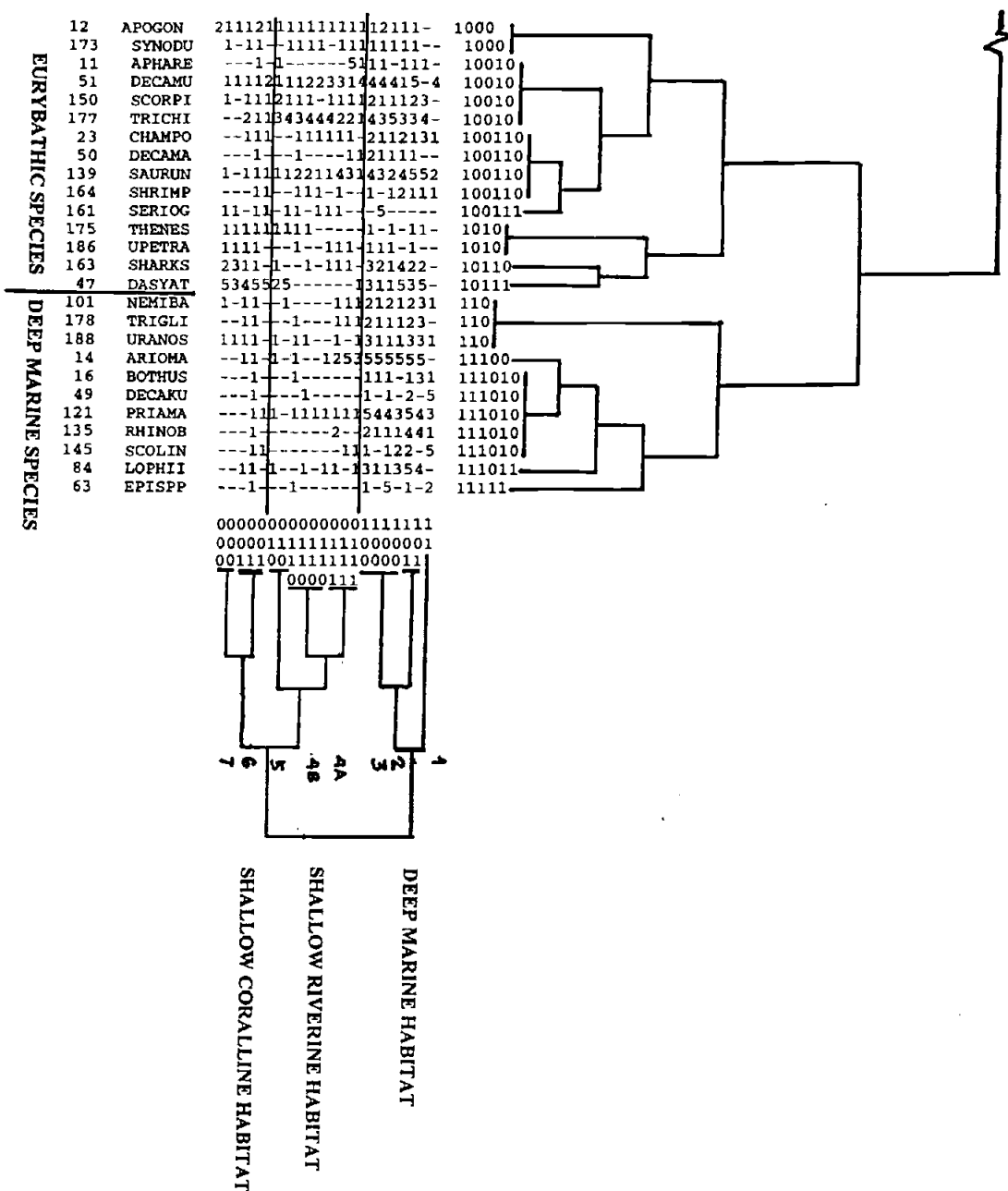


Figure 14. TWINSpan 2-Way Indicator Species Table for RABUTINOS (continued).

Based on the species characteristics and location among the different fishing stations, the first dichotomy separated the deep sea from the shallow water fish assemblage. The deep sea group were further subdivided into the truly deep marine assemblage (i.e., species/taxa whose primary habitat is the deep sea, e.g., Myctophidae, lanternfish) and the eurybathic assemblage (i.e., deep preferential species/taxa found mostly in deeper regions but may also be present in shallower areas).

Based on the substrate characteristics of the fishing stations, the shallow water assemblages were subdivided into three general habitat categories: (1) rocky/coralline assemblage; (2) shallow soft bottom (muddy-sandy) assemblages of the inner shelf; and (3) ubiquitous assemblage that live primarily in shallow regions but also has the ability to traverse a wide range of depths. The coralline assemblages were separated further into two categories, i.e., the ubiquitous coralline species assemblage (belonging mainly to Group 6 or 7 SU assemblage) and those confined primarily to the shallow coralline areas (mostly of Group 7 SU assemblage). The vertical range of distribution of the shallow coralline and soft bottom fish assemblages was more defined compared to the deep sea species. The ubiquitous species belonging to this category are spread over a wide range of depths although concentrated more on shallow water areas.

The key points derived from the TWINSpan species grouping are: (1) the range of species distribution varies from eurybathic/ubiquitous to narrow habitat zones and (2) the distribution of species shows gradual overlaps, not sharp boundaries.

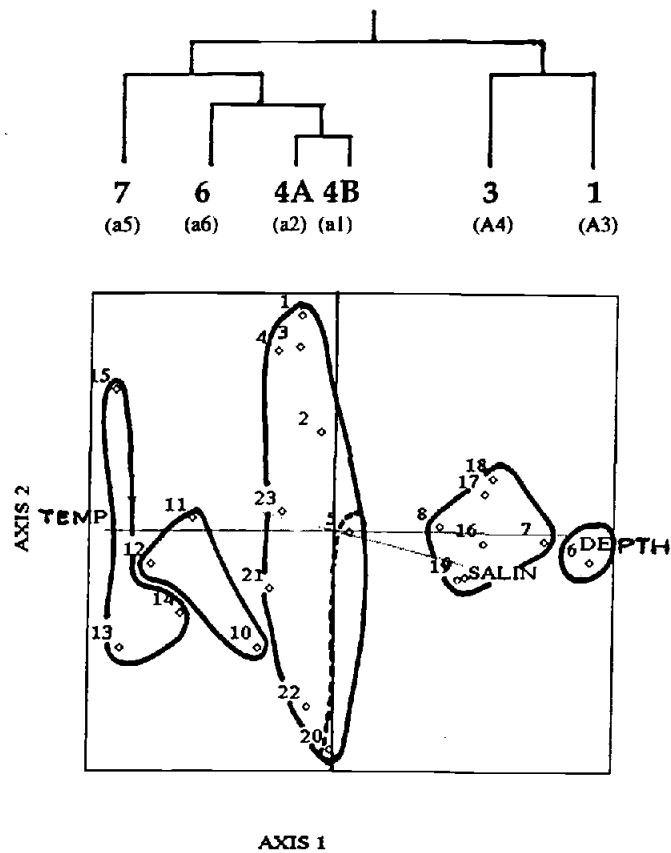
3. Seasonality and Geographic Distribution of the SU Groups.

The SU groups that comprised the demersal fish communities of RABUTINOS during the different seasons, including their species composition and geographic distribution, are presented in this section. Some of the possible environmental gradients

that correlate with this distribution were also explored. The seasonal as well as equivalent standard group designations are included in the dendograms. Refer to Table 3 for a complete list of the standard group designations, their seasonal equivalent, inclusive fishing stations and the specific cruises where they occurred.

a. *Northeast Monsoon Season.* The upper diagram in Figure 15 shows the combined results of TWINSpan classification and DECORANA ordination of the northeast monsoon samples based on the similarities between each station's fish occurrences and abundances. The first dichotomy in the dendogram separated the shallow (Groups 4A, 4B, 6 and 7) from the deep (Groups 1 and 3) fishing station clusters. During this season, the deep fishing stations were further subdivided into Group 1 (composed of FS6) and Group 3 (composed of FS7, FS8, FS9, FS16, FS17, FS18 and FS19). FS6 is the fishing station at the Ragay Gulf Basin sill nearest to Viñas River while FS7, FS8 and FS9 are those further away. FS16 to FS19 are fishing stations located at the deep sea ridge west of San Bernardino Strait.

The shallow fishing station clusters were also subdivided into the coralline stations, i.e., Group 7 (composed of FS13 to FS15), and the coastal riverine stations, i.e., Group 4B (composed of FS5, FS20), Group 4A (composed of FS1 to FS4, FS21 to FS23), and Group 6 (composed of FS10 to FS12). Since demersal trawls cannot operate on coral reefs, the term "coralline" refers to the similarity of the characteristic fauna to coral reef fish assemblages. "Estuarine" or "Riverine" refers to stations located near major river systems with soft, muddy to sandy-muddy substrates of terrigenous origins shallow depths and low salinities during the rainy season. The riverine stations were further subdivided into fishing stations near major sources of freshwater and over wide insular shelves, i.e., Group 4B (composed of FS1 to FS 5) and Group 4A (composed of FS20 to FS23), versus those with narrow shelves and further away from major rivers, i.e., Group 6 (composed of FS10 and FS11). Finer but minor distinctions were made between Group 4B (relatively deeper) and Group 4A (relatively shallower) fishing station groups.



PEARSON AND KENDALL CORRELATIONS WITH ORDINATION AXES N= 23									
AXIS:	1			2			3		
	r	r-sq	tau	r	r-sq	tau	r	r-sq	tau
DEPTH	.879	.772	.715	-.100	.010	-.008	.087	.008	.231
DOXY	-.861	.741	-.766	.037	.001	.004	-.154	.024	-.200
SALIN	.644	.415	.490	-.339	.115	-.164	.108	.012	.196
TEMP	-.808	.653	-.684	.038	.001	.040	-.111	.012	-.119

Figure 15. Results of the Q-Type Classification and Ordination Analyses on Fish Abundance Data During the Northeast Monsoon Season. Upper diagram: TWINSpan dendrogram of the seasonal fish assemblage grouping. Middle diagram: DECORANA ordination plots of the seasonal fish assemblage grouping. Bottom table: correlation between environmental variables and SU scores on the first three DCA axes.

The clusters derived from TWINSpan were used to show the groups of fishing stations in the DCA ordination diagram (Figure 15, middle diagram). The eigenvalues for the first three DCA axes were 0.82, 0.42 and 0.25 respectively. The first DCA axis was sufficient enough to differentiate the TWINSpan derived clusters. The seeming disparity between the possible group membership of FS12 and FS14 is just the result of the axis distortion mentioned above (see Methods section). The separation between the deep and shallow waters stations was very distinct as indicated by their cluster separation and distance.

Figure 15 (bottom table) shows the correlation between the first three DCA Axes and the four measured environmental variables. The highest correlations achieved were between the first DCA Axis and the environmental variables. Depth and salinity increases while DO and temperature decreased as one moves from left to right along the first DCA Axis, thereby separating the shallow water site clusters (i.e., Groups 7, 6, 4A and 4B) from the deep water site clusters (i.e., Groups 3 and 1).

The second DCA Axis hints of salinity influence, however, the generally low correlation between the second axis and the measured environmental variables indicate that some other important factors may be important in structuring the community. Given the ordination results (Figure 15; middle diagram), Groups 4A and 4B showed the widest ranges of salinity variations during the wet northeast monsoon season, followed by Group 7. The fishing stations near Viñas River (i.e., FS1, FS2, FS3, FS4, FS5) had the lowest salinity compared to the shallow fishing stations along the coast of northern Samar Sea (i.e., FS20, FS21, FS22, FS23). For Group 7, FS15 had the lowest salinity values compared to the rest. The fishing stations least affected by the northeast monsoon rains and corresponding drops in salinity were those from the deep water group (i.e., Groups 1 and 3) located within and below the halocline.

Although the correlation coefficients with the third DCA Axis were relatively higher than the second, the eigenvalues of the third DCA Axis was too low and basically

gave the same information as the first DCA Axis. Hence, it was not included in the analysis.

The geographic boundaries of the six different site groups identified by TWINSpan and DECORANA during this season are shown in Figure 16. Appendix II gives a list of the geographic characteristics of each Fishing Station. Starting at the mid-section of RABUTINOS, Group 5 composed of FS10 to FS12, hugs the narrow shelf of north of Sorsogon Bay and basically defined the main fishing area for *Leiognathus bindus*. Just outside the bay entrance and extending to the south along the narrow Ticao Pass shelf are the coralline stations (FS13 to FS15) belonging to Group 5.

The shallow, muddy shelf areas extending seawards from the coasts of Northern Samar Sea and south of Viñas River to about 50 m isobath defined the boundaries of sampling Group 4A. These were the locations of FS1 to FS4 and FS21 to FS23, and denoted the boundaries of Leiognathidae + coastal pelagic fish assemblages. Adjacent to this group and farther offshore were the areas that comprised Group 4B. The boundaries of this group were defined approximately by the 50 m and 100 m isobaths. This was the transition zone between the shallow and deep water stations and contained a fish assemblage dominated by Leiognathidae + Ariommidae and a variety of deep sea forms.

The deep water stations from the 100 m down to the 200 m isobaths defined the boundaries of Group 3 or the deep shelf stations dominated by Ariommidae + Trichiuridae. The stations included in this group were FS7 to FS9 for Ragay Gulf Basin and FS16 to FS19 for Northern Samar Sea. During this cruise, FS6 defined Group 1, the deep shelf/slope assemblage dominated by Nemipteridae and Carangidae.

b. *Dry Intermonsoon Period.* The deep fishing stations were separated from the shallow fishing stations by TWINSpan in the first dichotomy (Figure 17, upper diagram), similar to the previous season's dendrogram. The deep fishing stations were further subdivided into Group 2 (composed of FS6, FS7, FS17 and FS18) from Group 3

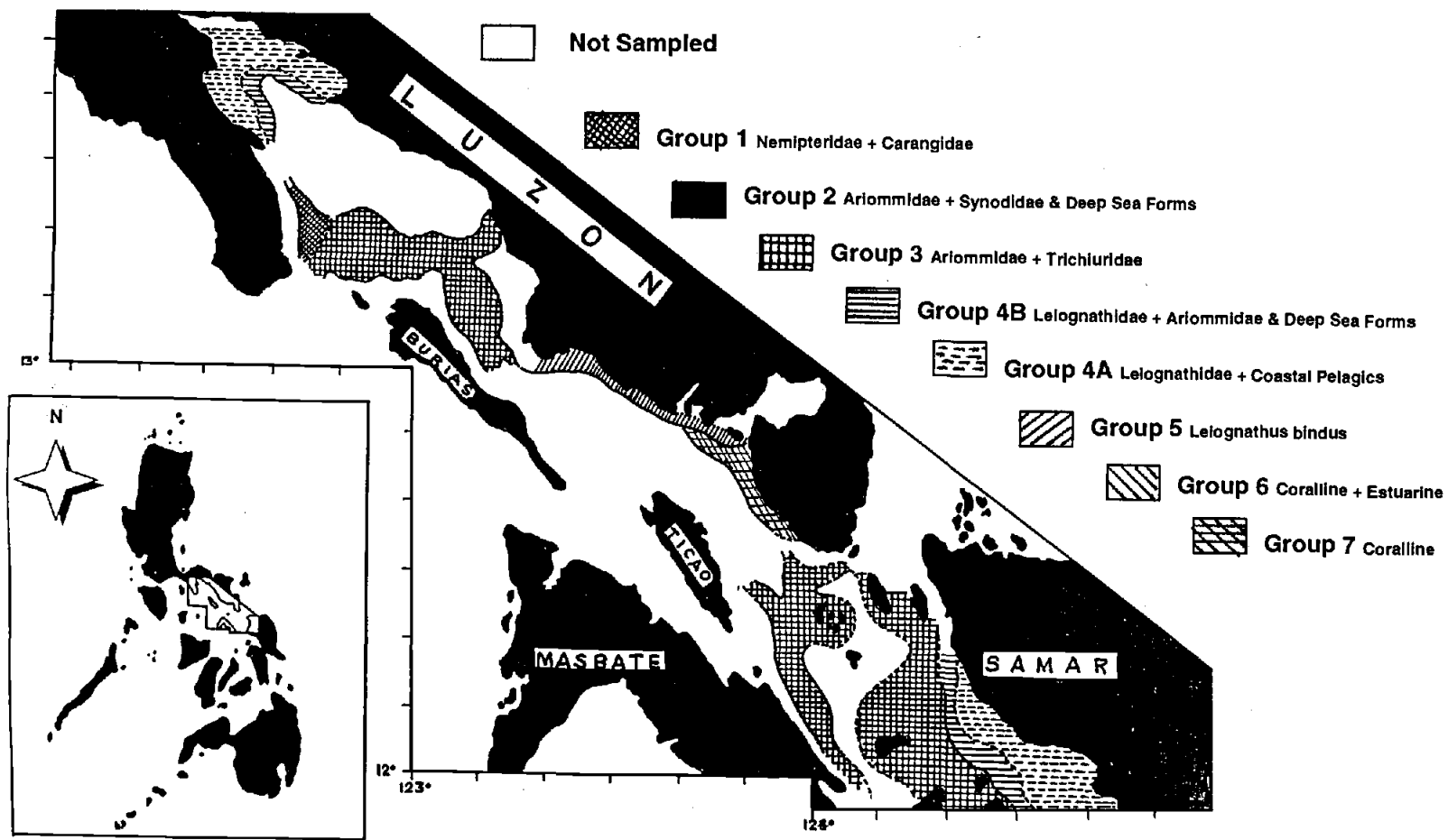


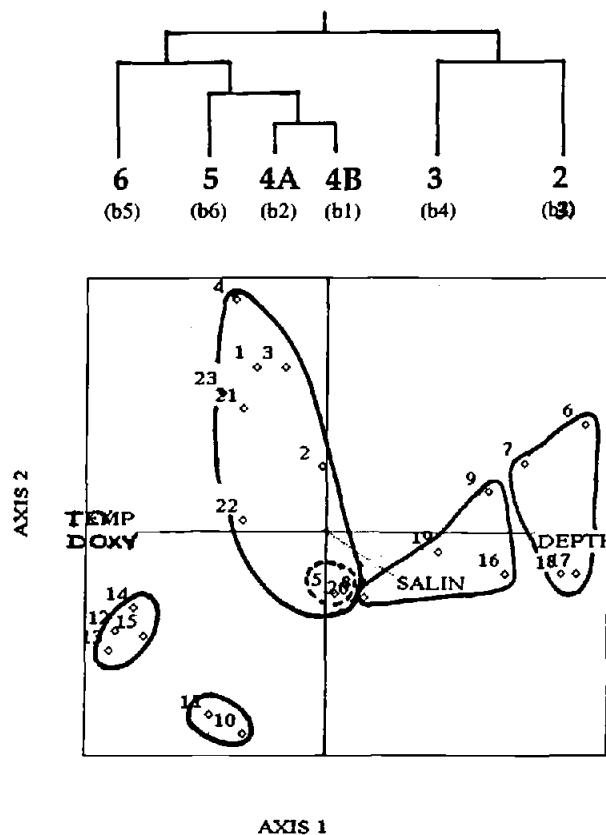
Figure 16. Geographic Location of the Different Fish Assemblage Groups at RABUTINOS During the Northeast Monsoon Season.

(composed of FS8, FS9, FS16 and FS19). The shallow fishing stations were also subdivided into the coralline station group, i.e., Group 6 (composed of FS12 to FS15) and the coastal riverine stations, i.e., Group 5 (composed of FS10, FS11), Group 4A (composed of FS1 to FS4 and FS21 to FS23) and Group 4B (composed of FS5, FS20). Further subdivisions on the coastal riverine group separated stations located on narrow insular shelves and relatively far from major freshwater influence, i.e., Group 5 from stations near major river systems emptying into wider insular shelves, i.e., Groups 4A and 4B.

The DECORANA ordination and TWINSpan classification were both able to distinguish the different cluster groupings based on the first and second axis (Figure 17, middle diagram). The eigenvalues for the first three axes were 0.86, 0.43 and 0.36 respectively. Groups 4B and 4A were clearly separated from Group 5 by the second DCA axis while the rest were highly discernible from the first DCA axis alone. The compact clustering of stations within Group 6 and Group 5 and their distance of separation from the other clusters indicate stronger similarities among its members relative to the others.

The correlation between the DCA axes and environmental variables during the dry intermonsoon period is shown in Figure 17 (bottom table). The correlations were basically the same as the ones derived for the northeast monsoon season, except for the lower temperature correlation on the first DCA Axis. The distinct and compact clusters formed by Groups 5 and 6 (Figure 17; middle diagram) indicate strong similarities among their members and the conditions of their environment. Both groups were also in distinctively more saline habitats than the rest.

During this period, salinity differences among fishing stations located at similar depths were not discernible for stations located in northern Samar Sea and in the vicinity of Viñas River. However, the salinity difference was very pronounced between Group 4A and Group 4B. Also, the fishing stations comprising Group 4B and Group 3 were not as distant as the previous cruise.



AXIS:	1			2			3		
	r	r-sq	tau	r	r-sq	tau	r	r-sq	tau
DEPTH	.935	.875	.787	-.077	.006	.012	.249	.062	.197
DOXY	-.890	.793	-.679	.060	.004	.044	-.307	.094	-.165
SALIN	.523	.273	.585	-.448	.201	-.129	.156	.024	.126
TEMP	-.833	.694	-.521	.003	.000	.185	-.371	.138	-.167

Figure 17. Results of the Q-Type Classification and Ordination Analyses on Fish Abundance Data During the Dry Inter-Monsoon Season. Upper diagram: TWINSpan dendrogram of the seasonal fish assemblage grouping. Middle diagram: DECORANA ordination plots of the seasonal fish assemblage grouping. Bottom table: correlation between environmental variables and SU scores on the first three DCA axes.

During the dry months, the areal extent of Group 5 that defined the *Leiognathus bindus* fishing grounds, shrank to include just FS10 to FS11 located along the narrow shelf north of Sorsogon Bay shown in Figure 18. The southern half of this narrow shelf (FS12 to FS15) changed from Group 7, i.e., coralline fish assemblage, into transition Group 6 assemblage characterized by coralline + Leiognathidae and other estuarine species/taxa. The shift from Group 7 into Group 6 was brought about by substantial migration of slipmouths and grunts into areas occupied by the coralline fish assemblage during the dry inter-monsoon period.

For the riverine stations, the locations of Groups 4A (i.e., where Leiognathidae + coastal pelagic fishes abound) and 4B (i.e., where Leiognathidae + Ariommidae and other deep sea forms dominate), remained similar to the previous season. Some major changes occurred in the species assemblage of the deep sea fishing stations. In the Ragay Gulf Basin, areas formerly designated as Group 1 (Nemipteridae and Carangidae assemblage), were now occupied by Group 2 assemblage (characterized by Ariommidae + a wide variety of deep sea forms), which included FS6 and FS7. The deep fishing stations FS17 and FS18 in Northern Samar Sea were now also occupied by the Group 2 fish assemblage. This was brought about by outmigration of Trichiuridae and an immigration of deep sea forms into the area. The rest of the deep water stations remained as Group 3 where Ariommidae + Trichiuridae abound.

c. *Southwest Monsoon Season.* On the first dichotomy, the TWINSPAN dendrogram separated the stations into the deep water group and the shallow water group (Figure 19, upper diagram). The deep water stations were further subdivided into Group 2 (composed of FS17, FS18) and Group 3 (composed of FS6, FS7, FS9, FS16, FS19). The shallow water group were subdivided into the coralline stations, i.e., Group 7 (composed of F12 to F15), and the riverine stations composed of two minor sub-groups, i.e., Group 4A (composed of FS1 to FS5 and FS21 to FS23) and Group 4B (composed of FS8 and FS20).

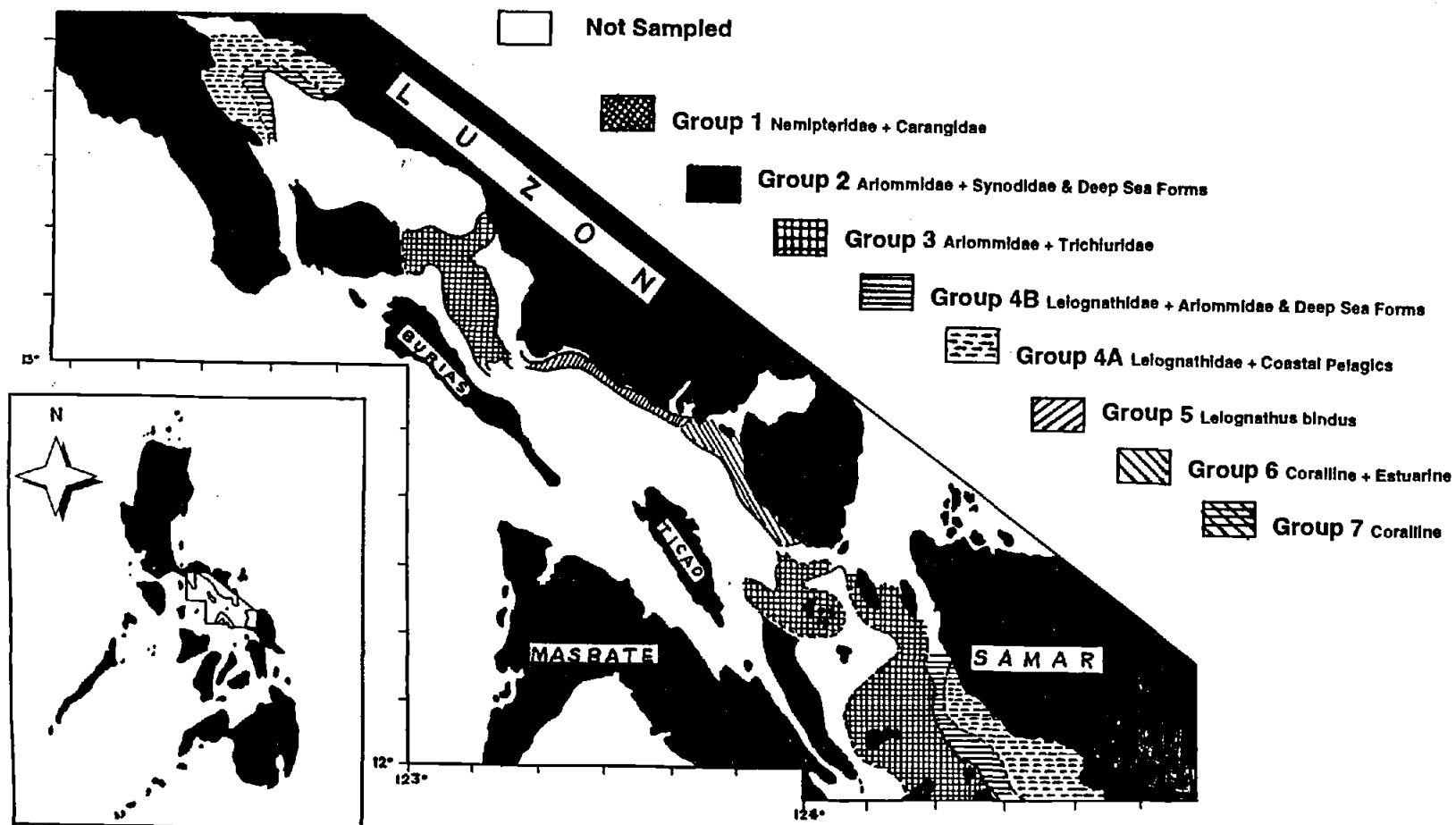
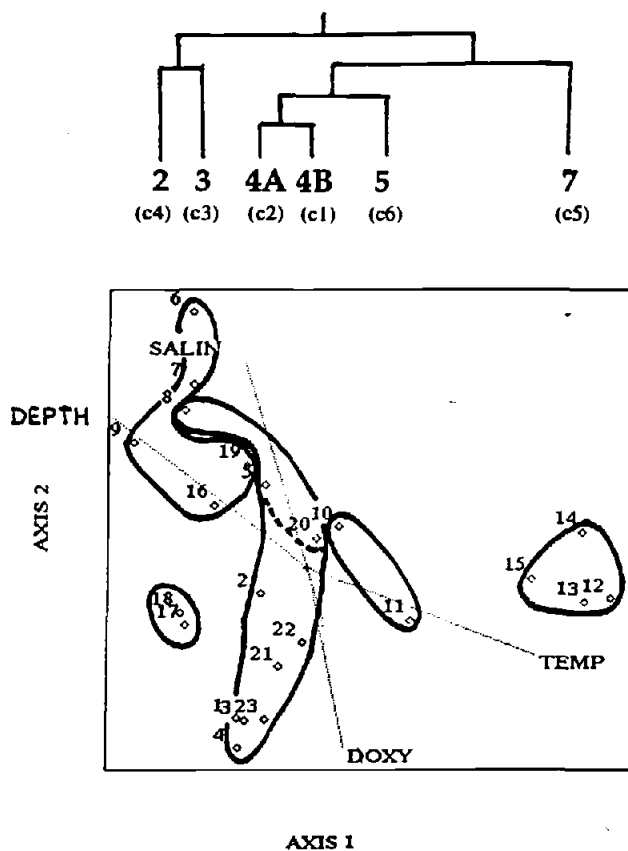


Figure 18. Geographic Location of the Different Fish Assemblage Groups at RABUTINOS During the Dry Inter-Monsoon Period.



PEARSON AND KENDALL CORRELATIONS WITH ORDINATION AXES N= 23									
AXIS:	1			2			3		
	r	r-sq	tau	r	r-sq	tau	r	r-sq	tau
DEPTH	-.561	.314	-.449	.525	.275	.512	-.365	.133	-.116
DOXY	.228	.052	.246	-.524	.274	-.404	.502	.252	.298
SALIN	-.294	.087	-.261	.559	.313	.348	-.330	.109	-.181
TEMP	.572	.327	.600	-.366	.134	-.319	.489	.239	.024

Figure 19. Results of the Q-Type Classification and Ordination Analyses on Fish Abundance Data During the Southwest Monsoon Season. Upper diagram: TWINSpan dendrogram of the seasonal fish assemblage grouping. Middle diagram: DECORANA ordination plots of the seasonal fish assemblage grouping. Bottom table: correlation between environmental variables and SU scores on the first three DCA axes.

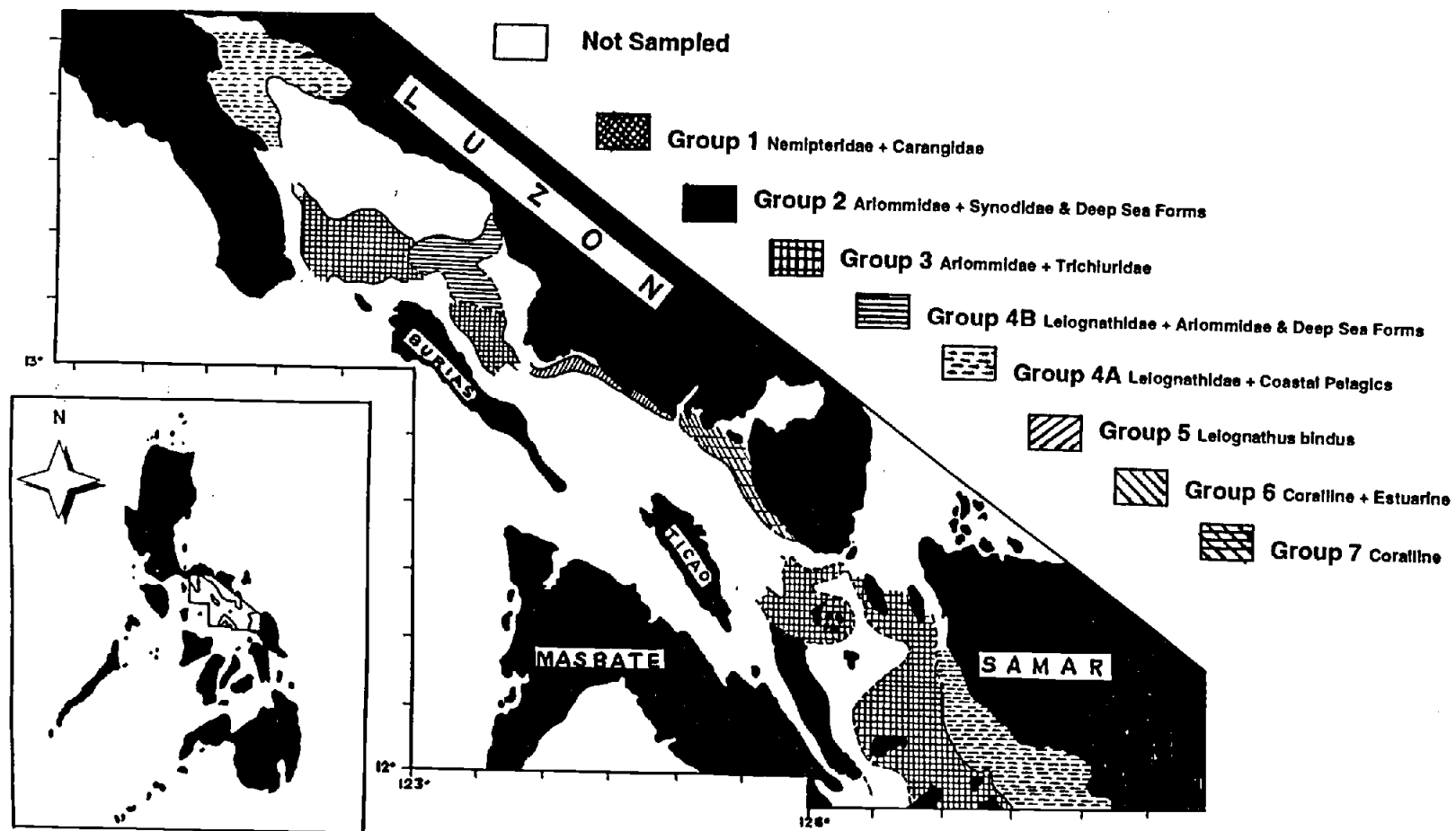


Figure 20. Geographic Location of the Different Fish Assemblage Groups at RABUTINOS During the Southwest Monsoon Season.

DCA ordination of the southwest monsoon data (Figure 19, middle diagram) distinctly separated the coralline Group 7 from the rest. During this season, Group 5 showed much closer affinity with Groups 4B and 4A compared to the previous season. The second DCA axis clearly separated the deepest stations, (i.e., Group 2), from the less deep adjacent stations, (i.e., Group 3). Apart from FS8, the coastal riverine stations formed a relatively distinct cluster. FS8 may have been misclassified as indicated by its greater affinity with Group 3 in the ordination diagram. The eigenvalues of the first three DCA axes were 0.86, 0.43 and 0.31 respectively.

The rainy southwest monsoon season reduced the correlation between the first DCA Axis and the environmental variables (Figure 19; bottom table). The first DCA Axis is adequately represented by both temperature and depth gradients, i.e., depth decreases and temperature increased as one moved from left to right along the first DCA Axis (Figure 19; middle diagram). The second DCA Axis represented a complex gradient involving all of the environmental variables, i.e., salinity and depth decrease while temperature and DO concentration increase as one moved from top to bottom of the diagram. The fishing stations comprising Group 7 were distinctly separated from the rest due to its relatively high water temperature and medium salinity and DO values. The shallow water stations (Group 4A) were easily distinguishable from the deeper stations, e.g., Group 4B and Group 3 due to its high DO concentration. However, the separation between Groups 4B and 3 were not as distinct.

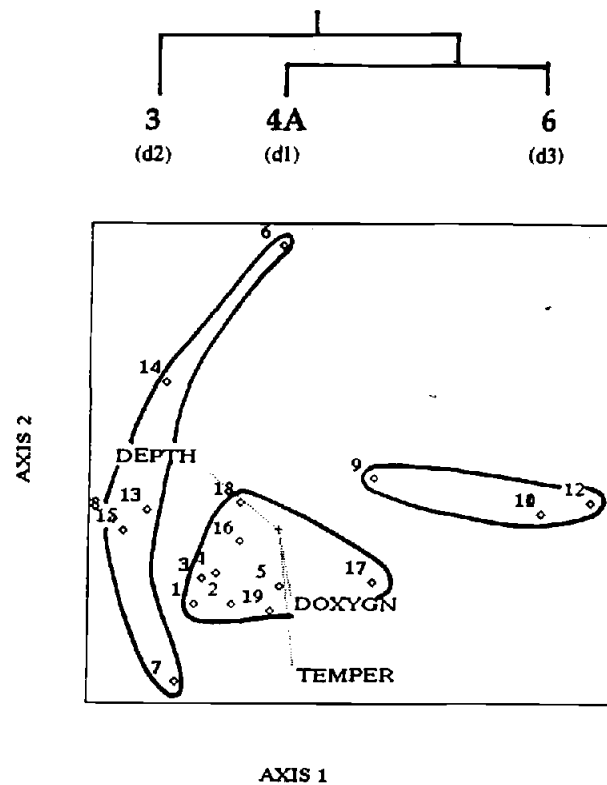
The rainy season brought some changes in the station groupings at RABUTINOS (Figure 20). On the narrow shelf off Sorsogon Bay and vicinity, the areas formerly occupied by transition Group 6 assemblage (i.e., composed of a mixed bag of coralline + Leiognathidae and other estuarine species/taxa) reverted back to Group 7 (composed mainly of coralline fish assemblage), resulting from the outmigration of Leiognathidae and associated species/taxa from the coralline fish assemblage boundaries and a return to

their own main habitats to the north. Once again, a clear separation existed between Group 5 (*Leiognathus bindus* assemblage) and Group 7 (coralline assemblage).

On the shallow inner shelves of northern Samar Sea and Ragay Gulf, Group 4A (composed of Leiognathidae + other coastal pelagics) now extended all the way offshore into areas formerly occupied by Group 4B fish assemblage (i.e., Leiognathidae + Ariommidae and other deep sea forms). This was due to the outmigration of deep sea forms and the extension of the range of coastal/riverine pelagic species into these areas.

Except for FS8, most of the deep shelf areas in the central Ragay Gulf Basin were re-occupied by Group 3 assemblages composed of Ariommidae + Trichiuridae. FS8 was classified by TWINSpan as belonging to Group 4B but was closer to Group 3 in the DECORANA ordination. Group 4B represented the transition zone between the shallow, riverine group (Group 4A) and the deep preferential species (Group 3) and was differentiated mainly by the relative abundances of Trichiuridae, Ariommidae and Leiognathidae. Since this period corresponds to the rainy southwest monsoon season, FS8 was assigned to Group 4B.

d. *Dry ENSO Early Maturity Phase.* The ENSO period showed radical structural and compositional departures from the previous seasonal dendograms (Figure 21, upper diagram). Only three major clusters were discerned by TWINSpan, i.e., the deep water stations within the Southwestern region (Group 3), and the shallow water stations composed of coralline-estuarine mixture (Group 6) and riverine (Group 4A) clusters. The DCA ordination pattern (Figure 21, middle diagram) also indicated the same grouping. The eigenvalues of the first three DCA axes were 0.86, 0.49 and 0.40 respectively. (Note: Since four fishing stations were not sampled during this cruise, the sequential arrangement of numbers was not the same as the fishing station codes; see Table 3 for details).



PEARSON AND KENDALL CORRELATIONS WITH ORDINATION AXES N= 19									
AXIS:	1			2			3		
	r	r-sq	tau	r	r-sq	tau	r	r-sq	tau
TEMPER	.251	.063	.144	-.793	.630	-.406	.099	.010	.154
SALINI	-.433	.187	-.314	.062	.004	.264	-.481	.231	-.165
DOXYGN	.224	.050	.135	-.553	.305	-.306	.256	.065	.207
DEPTH	-.553	.306	-.429	.519	.269	.261	-.311	.097	-.295

Figure 21. Results of the Q-Type Classification and Ordination Analyses on Fish Abundance Data During the ENSO Early Maturity Stage. Upper diagram: TWINSpan dendrogram of the seasonal fish assemblage grouping. Middle diagram: DECORANA ordination plots of the seasonal fish assemblage grouping. Bottom table: correlation between environmental variables and SU scores on the first three DCA axes.

The correlation between the first DCA Axis and the environmental variables were weakest during the Early Maturity Stage of ENSO (Figure 21; bottom table). Depth was the primary gradient along the first DCA Axis while temperature and DO concentration were the gradients along the second DCA Axis (Figure 21; middle diagram). Instead of producing noisy ordination and classification output, this group produced three very distinct clusters representing shallow-riverine (Group 4A), coralline/estuarine transition (Group 6), and deep thermocline water transition (Group 3) assemblages.

During this period (Figure 22), there was no change in the species assemblages of the estuarine areas as indicated by the same geographic extent of Group 4A. However, both the *Leiognathus bindus* (Family: Leiognathidae) assemblage north of Sorsogon Bay (Group 5) and the coralline assemblage (Group 7) were now replaced by Group 6 (coralline + Leiognathidae and other estuarine assemblage) indicating cross-mixing among species belonging to both groups along the whole length of the narrow Ticao Pass shelf. All the deep sea stations, i.e., from approximately 50 m to 200 m depth, exhibited a homogenous fish assemblage composed mainly of Ariommidae + Trichiuridae (Group 3).

e. *Summary of Seasonal Patterns.* In general, the primary dichotomy clearly identified by the ordination and classification analyses were along the bathymetric gradient. The depth of separation between the shallow and deep water communities was around the 90 m isobath. Secondary gradients were most pronounced in the shallow areas (<90 m depth). These were related mainly to geographical features, i.e., (1) proximity to riverine sources and associated physical attributes like varying levels of salinity; or (2) proximity to rocky or coral reefs characterized by high habitat complexity and rugosity.

Table 12 outlines the seasonal and spatial variation in the cluster membership of the different SU's. Contrary to expectations, the shallow coastal stations near major rivers and freshwater run-offs did not show any change in group membership with the season, even with ENSO-induced drought conditions. Instead of changing from Group 4A to Group 4B as freshwater discharge from rivers declined, the shallow water stations stayed the same during the dry ENSO early maturity period. This resulted in the enlargement of

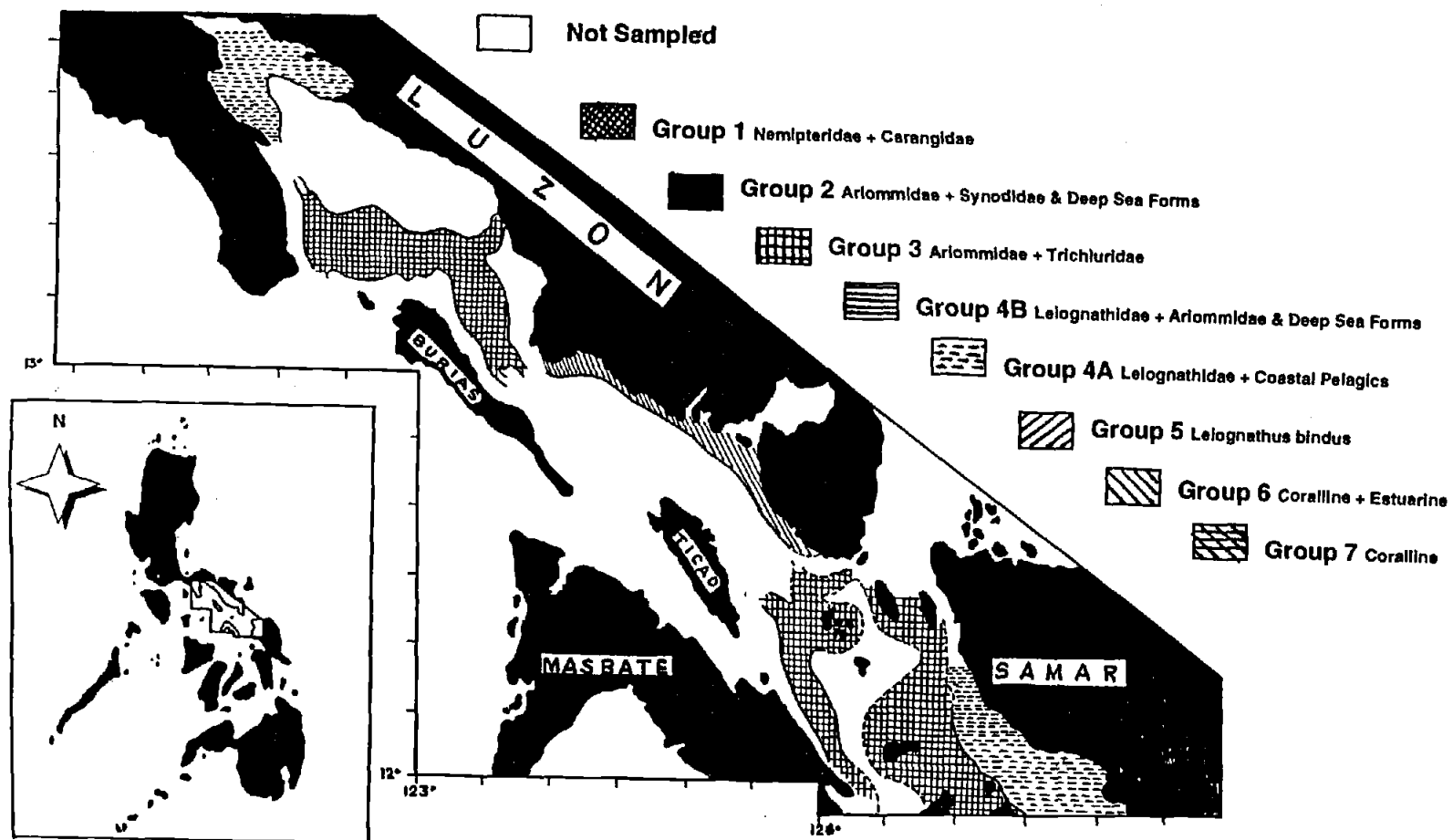


Figure 22. Geographic Location of the Different Fish Assemblage Groups at RABUTINOS During the ENSO Early Maturity Stage.

Table 12. Seasonal and Spatial Variation in the Station Cluster Membership. Standard group designations were derived from ordination and classification. Shaded cells indicate stations that show ENSO variations in group membership.

SAMPLING UNITS (FISHING STATIONS)																							
SEASON	Riverine					Deep				Ticao Pass/Coralline						Deep			Riverine				
	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23
Wet NE Monsoon	4A	4A	4A	4A	4B	1	3	3	3	6	6	6	7	7	7	3	3	3	3	4B	4A	4A	4A
Dry Intermonsoon	4A	4A	4A	4A	4B	2	2	3	3	5	5	6	6	6	6	3	2	3	3	4B	4A	4A	4A
Wet SW Monsoon	4A	4A	4A	4A	4A	3	3	4B	3	5	5	7	7	7	7	3	2	3	3	4B	4A	4A	4A
ENSO Maturity	4A	4A	4A	4A	4A	3	3	3	3	6	6	(6)	6	6	6	3	3	3	3	4A	4A	4A	4A
Normal NE Group	4A	4A	4A	4A	4B	1	3	3	3	6	6	6	7	7	7	3	3	3	3	4B	4A	4A	4A
Enso Effect?	No	No	No	No	Yes	Yes	No	No	No	No	No	No	Yes	Yes	Yes	No	No	No	No	Yes	No	No	No
Seasonality?	No	No	No	No	Yes	Yes	Yes	Yes	No	Yes	Yes	Yes	Yes	Yes	Yes	No	Yes	No	No	No	No	No	No

Legend:  Possible ENSO-Induced Change

 Possible Regular Seasonal Variation Only

the area occupied by Group 4A. Most of the seasonal variations and ENSO impacts were confined to the locations of the coralline station clusters and the riverine-deep sea transition groups (i.e., FS5 and FS20; FS10 to FS15), including some deep sea stations (FS 6 to FS8 and FS17).

The coralline stations alternate from Group 7 (purely coralline) during the wet season and into Group 6 (coralline-estuarine transition) during the dry season. The northerly stations (FS10 and FS11) alternate from Group 6 during the wet season, to Group 5 (mainly *Leiognathus bindus* assembly) during the dry season indicating stronger horizontal rather than vertical migration and species interchange between these two station clusters.

The deep water stations (FS 17, FS6 and FS7) exhibited greater vertical interchange of species, involving mainly the thermocline associated fauna and the deeper zones during the dry season. This was indicated by the change from Group3 to Group 2 and *vice-versa* among those stations. The fish assemblage in FS8 was primarily of transition Group 3 except during the wet rainy season when it became the shallower Group 4B transition assemblage. It is not definite from the data whether ENSO conditions were responsible for the non-appearance of Group 1 during the averted 1982 northeast monsoon season.

f. *Seasonal Assemblage Contribution to Overall Fish Abundance.* The seasonal variation in the mean catches of each fish assemblage is shown in Figure 23. Significant decline in the total abundance and number of fish assemblages occurred during the ENSO period. The mean catch dropped from approximately 1.5 metric tons during the northeast, intermonsoon and southwest monsoon seasons to about one half of this value (0.7 metric tons) by the early maturity stages of ENSO. Furthermore, the number of fish assemblages contributing to the observed abundances decreased from six to just three groups during ENSO. The three remaining assemblages all came from relatively less stable or highly variable marine environments, i.e., Group 3 (thermocline SU assemblage), Group 4A

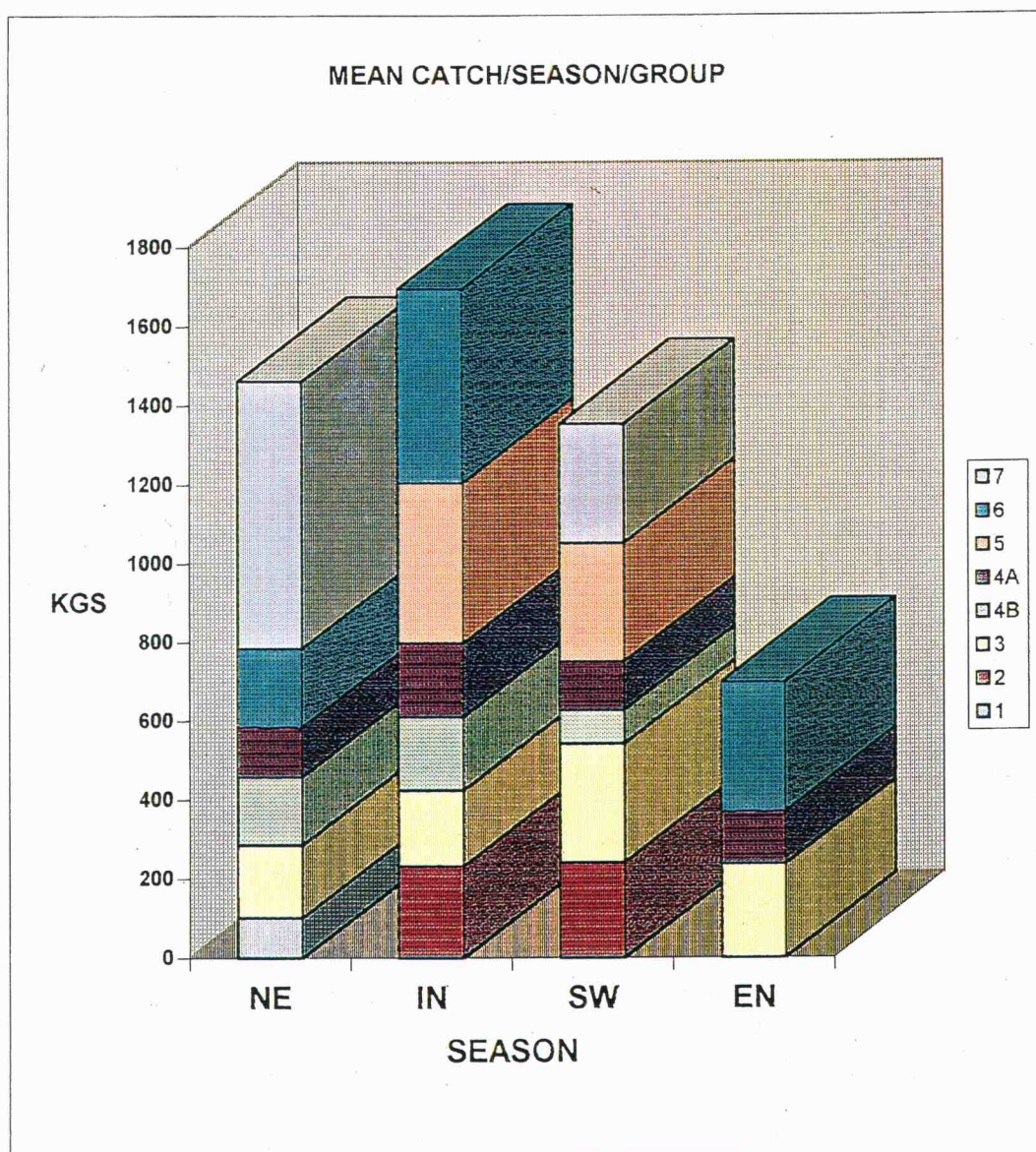


Figure 23. Seasonal Variation in Abundance of Demersal Trawl Fauna in Rabutinos

(shallow, riverine SU assemblage) and Group 6 (coralline-estuarine transition SU assemblage).

D. Environmental Association:

This section presents the results of the analyses done on the environmental characteristics of the different SU groups with the objective of identifying the principal gradient(s) that influence the observed fish assemblage structure at RABUTINOS. There were two basic environmental association methods employed and were presented as follows: (1) a comparative analysis of habitat characteristics aimed at statistically isolating significant differences between each SU groups and (2) multiple linear regression analysis for the simultaneous analysis of the direction and magnitude of each environmental variable's impact on the individual species comprising the community.

1. Seasonal Variation of Environmental Factors.

a. *Salinity.* The time series pattern of salinity distribution in the upper 100 m layer during the survey period is shown as a Tukey box plot in Figure 24. In general, the average salinity in the upper 100 m layer of water was low during the rainy months, (i.e., Cruise 3/ November '81, Cruise 4/December '81, Cruise 9/July '82 and Cruise 10/September '82), and high during the dry months, (i.e., Cruise 5/January '82, Cruise 6/March '82, Cruise 8/May '82, Cruise 12/November '82 and Cruise 13/January '83). Also, the average salinity during January '83 (Cruise 13), two months after the start of the ENSO-induced drought, was significantly higher than the previous January '82 (Cruise 5) based on the results of Fisher's Protected LSD test (Table 13). The dry months of December '81 (Cruise 4), July '82 (Cruise 9) and September '82 (Cruise 10) were also significantly more saline than the wet months of March '82 (Cruise 6), November '82 (Cruise 12) and January '83 (Cruise 13).

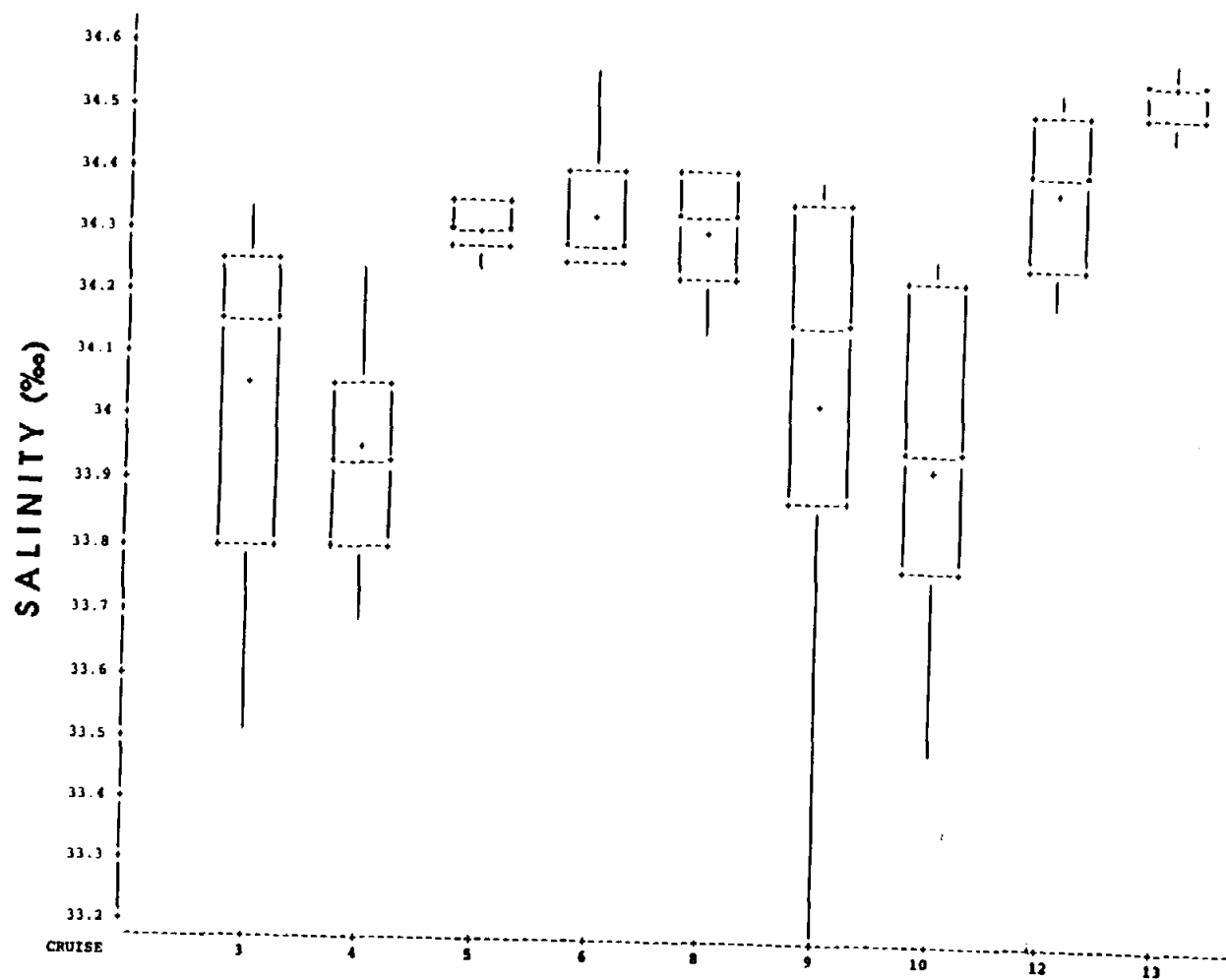


Figure 24. Time Series Plot of the Mean Salinity (Upper 100 m Layer) Variation at Ragay Gulf and Vicinity.

Table 13. Fisher's Protected Least Significant Difference (FPLSD) Test Comparing the Mean Salinity/Cruise in the Upper 100 m Layer of RABUTINOS.

T Grouping		Mean	Cruise
B	A	34.54	13
	A	34.39	12
	A	34.33	6
		34.31	5
		34.31	8
	C	34.04	3
	C	34.03	9
	C	33.94	4
	C	33.93	10
Alpha = 0.05		LSD=0.22	

Table 14. Fisher's Protected Least Significant Difference (FPLSD) Test Comparing the Mean Temperature/Cruise in the Upper 100 m Layer of RABUTINOS.

T Grouping			Mean	Cruise
B	A		27.69	9
	A		27.22	10
	A		27.11	8
B	A	C	26.63	3
B	D	C	26.38	12
E	D	C	25.62	4
E	D		25.41	13
E			24.85	6
E			24.81	5
Alpha = 0.05			LSD=1.16	

The temporal variation in salinity, from the sea surface down to 300 m depth, is shown in Figure 25. The cross-hatched area in the diagram indicates seawater salinities $\leq 34.3\text{‰}$ used as arbitrary reference point in the discussion. Low salinity waters extended down to 120 m depths during the 1981 and 1982 rainy southwest monsoon seasons, indicating the degree in which heavy rainfall affects the "estuarization" of the marine environment in tropics. Salinity stratification was accentuated during the rainy season as indicated by the number of salinity isopleths stacked on top of each other. The reverse happened during the dry season when most of the low salinity isopleths disappeared and the water column approached isohaline conditions. While it took more time to lower the salinity of the water column, once the heavy rains stopped, e.g., January - February '82 and November '82 onwards, it only took a short period of time for salinities at RABUTINOS to increase and for the water column to attain near isohaline conditions again.

b. *Water Temperature.* The temporal variation in seawater temperature during the survey period is given in Figure 26. Except for some phase delay, the diagram showed basic congruence in the pattern of variation between air temperature and the temperature of the water column. The local marine environment was coolest during the northeast monsoon months and warmest during the southwest monsoon season. Peak seawater temperature was reached around July, approximately one month after air temperature reached its maximum values. Fisher's Protected LSD procedure (Table 14), indicated that the mean seawater temperature in the upper 100 m water column during January '83 (Cruise 13) was not significantly higher than the average temperature of the previous year (January '82/Cruise 5).

The temporal variation in seawater temperature with depth is shown in Figure 27. The compactly spaced isotherms, i.e., the band of water between 18°C and 24°C , located at depths between 100 m to 150 m defined the typical location and vertical extent of the thermocline on this side of the equatorial Pacific Ocean. The cross-hatched portion in the

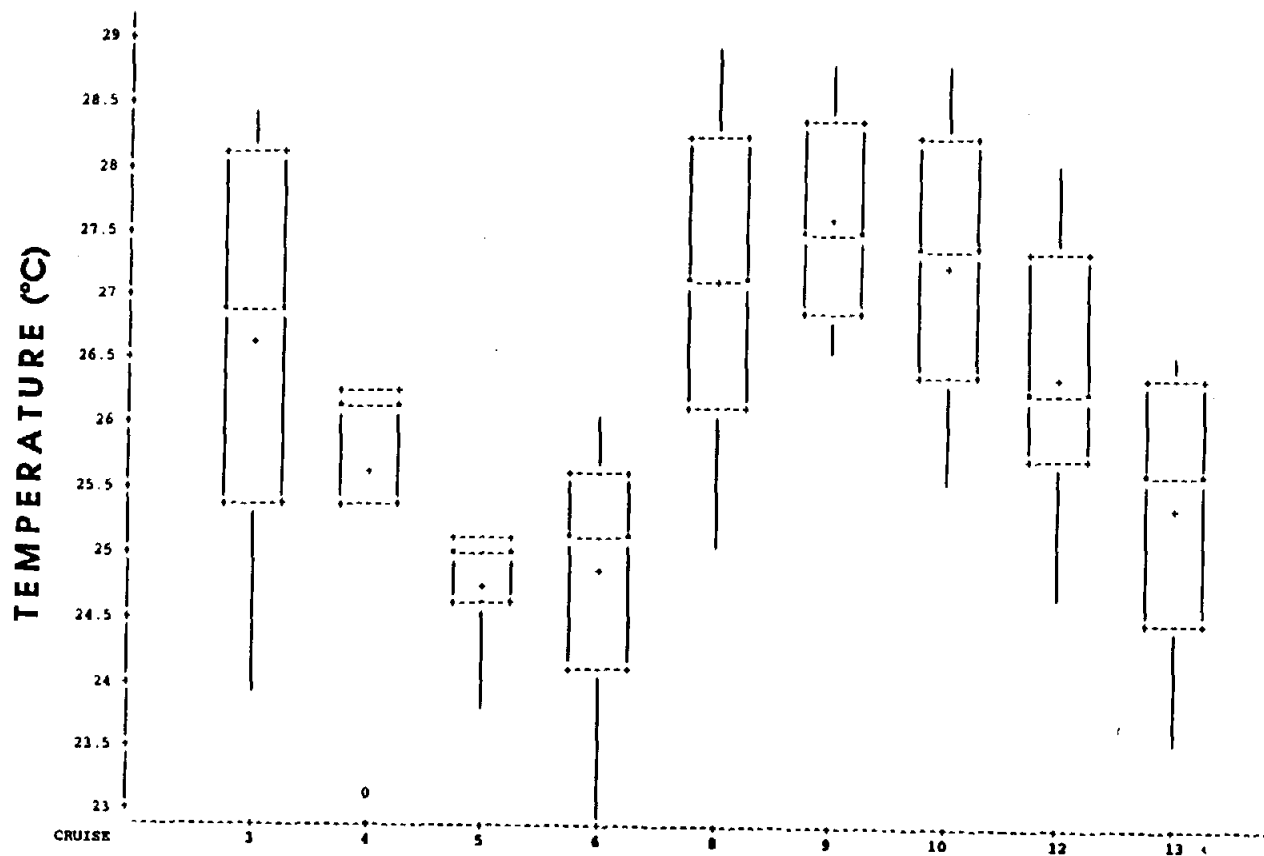


Figure 26. Time Series Plot of the Mean Sea Temperature (Upper 100 m Layer) Variation at Ragay Gulf and Vicinity.

diagram indicated waters warmer than 26°C. Much of the seasonal fluctuations in water temperature occurred above the thermocline (i.e., <100 m depth). This was manifested in the position of the 26°C isotherm as the seasonal warming and cooling process went on. Accentuated thermal stratification of the water column was also evident during the warm southwest monsoon season (Figure 27). The reverse was true during the cool northeast monsoon season as shown by water temperatures in the upper 100 m water column approaching the upper limits of the thermocline.

c. *Dissolved Oxygen.* The temporal variation in the dissolved oxygen (DO) content of sea water within the upper 100 m depth is shown in Figure 28. The time series pattern showed DO concentrations following a regular seasonal cycle. Low DO values were observed during the northeast monsoon while high values were typical during the southwest monsoon season. Comparing the pre-ENSO and ENSO cruises, Fisher's Protected LSD test (Table 15) indicated that the average dissolved oxygen concentration during January '82 (Cruise 5) was significantly higher compared to those observed in January '83 (Cruise 13).

The seasonal variation in DO concentration with depth is illustrated in Figure 29. The cross-hatched area delineated water layers with DO values > 6.0 ml/L and showed the extent in which the water column was oxygenated. High DO concentrations were prevalent throughout the surface layers but rapidly decreased with depth. An oxycline was present at around 75 m to 150 m depth during the dry intermonsoon period (February to May). This marked the boundary between the highly oxygenated surface layers from the oxygen deficient bottom waters. This layer mimicked the thermocline but exhibited wider vertical fluctuations. Tracing the 5.5 ml/L DO isopleth, the low oxygen boundary layer extended upwards near the sea surface during the northeast monsoon season and down to 40 m depth during the peak of the rainy southwest monsoon season.

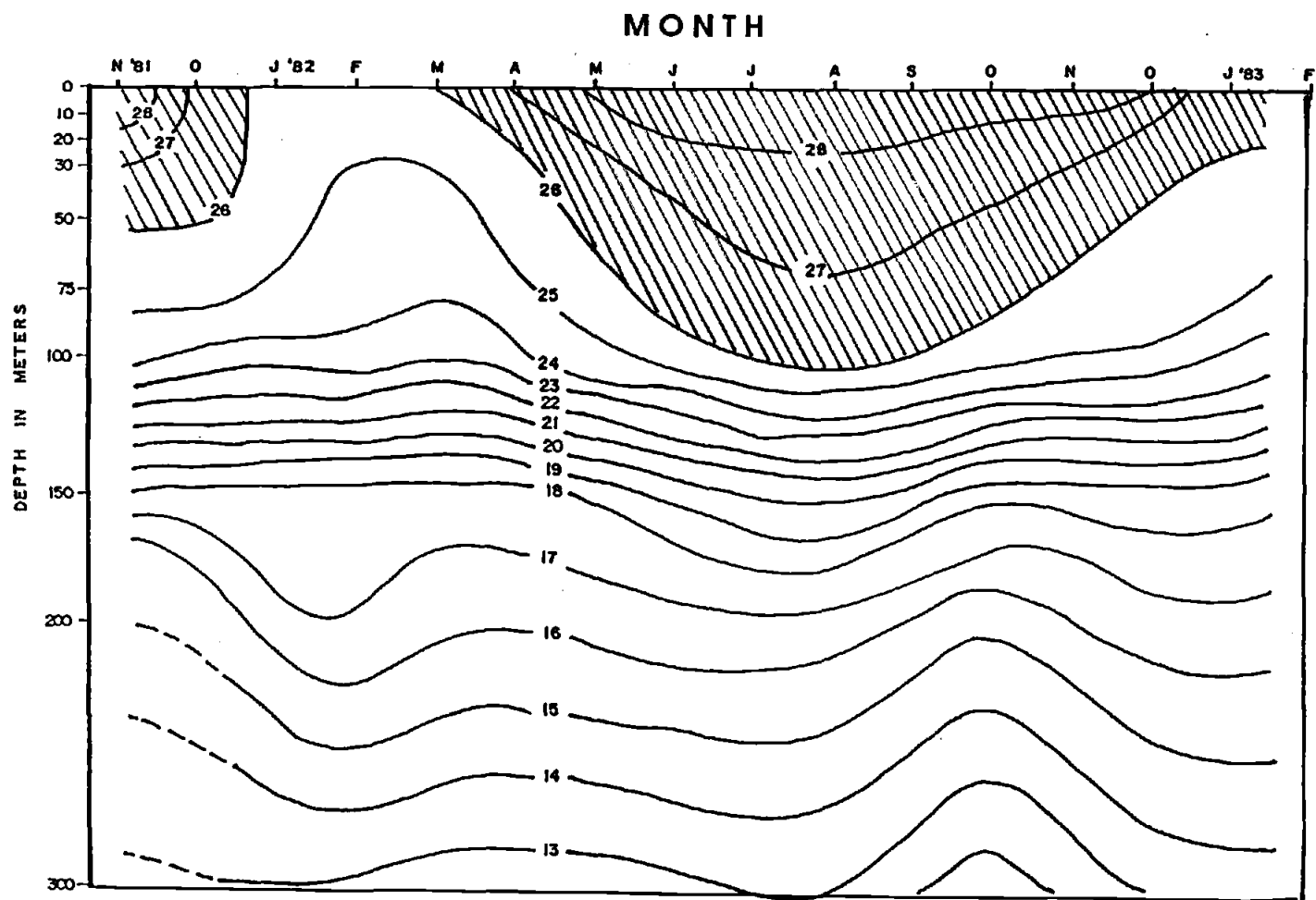


Figure 27. Isopleth Map of the Seasonal Variation in the Mean Temperature of Ragay Gulf and Vicinity.

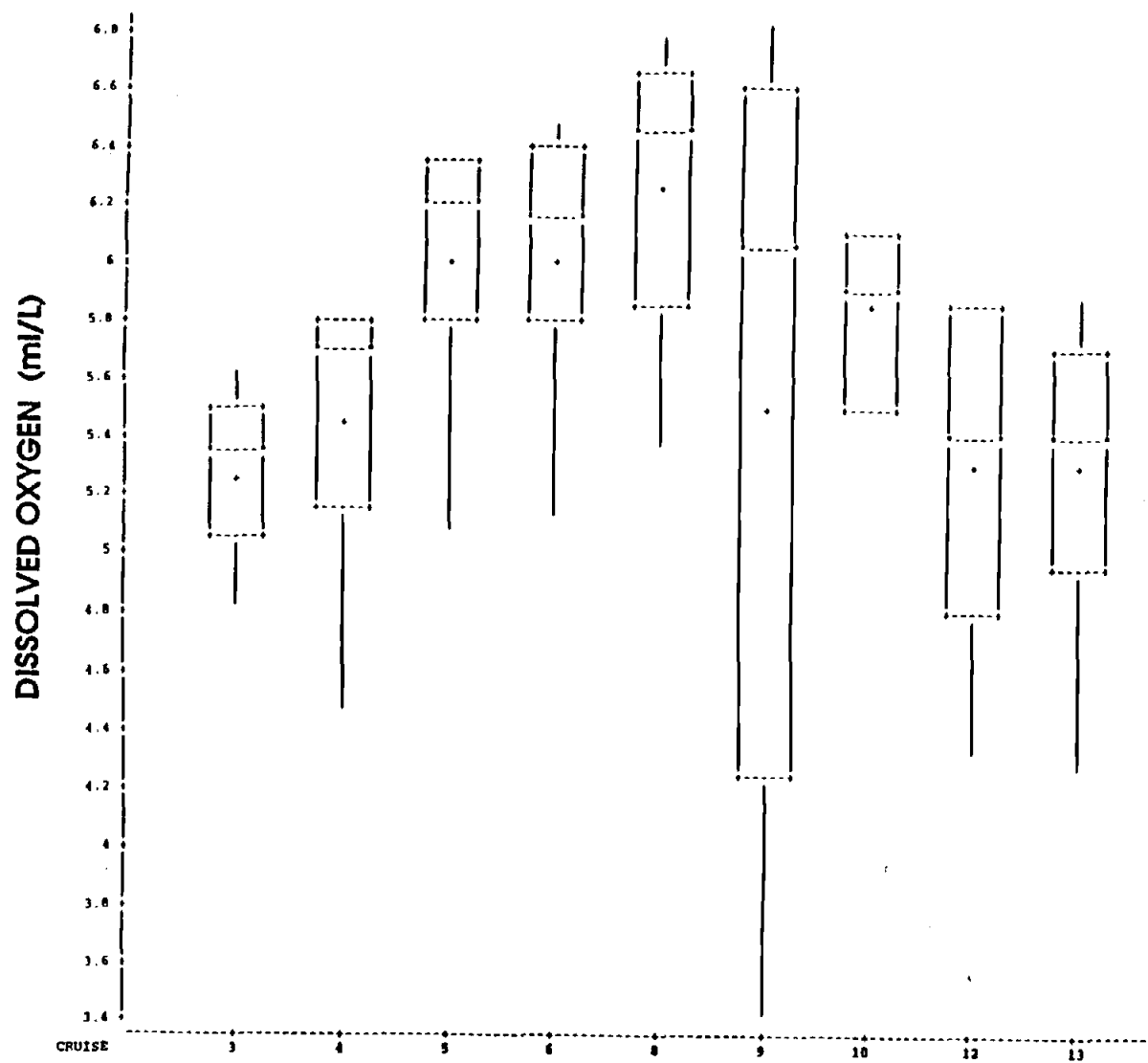


Figure 28. Time Series Plot of the Mean Dissolved Oxygen Concentration in the Upper 100 m Layers of Ragay Gulf and Vicinity.

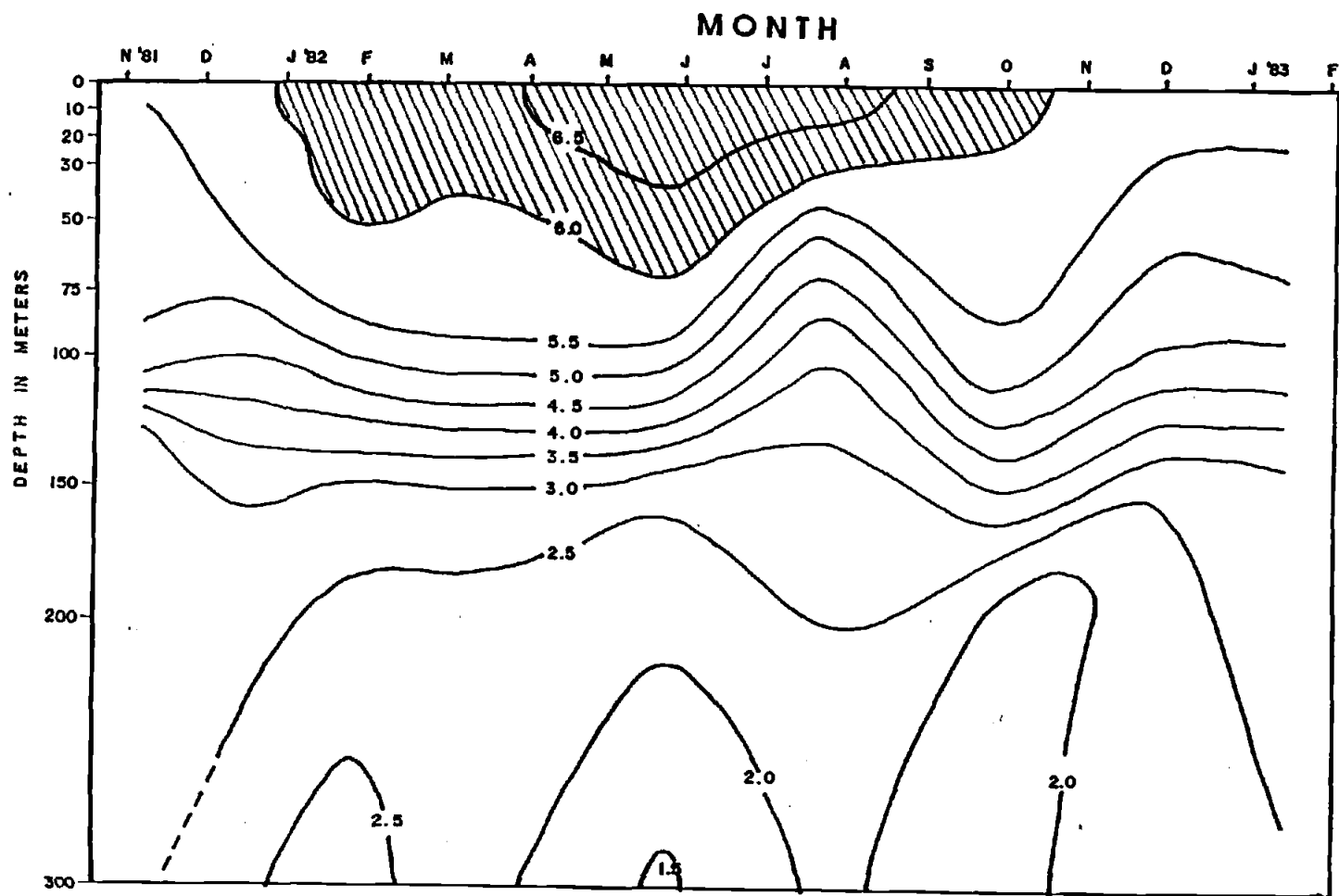


Figure 29. Isopleth Map of the Seasonal Variation in the Mean Dissolved Oxygen Concentration in Ragay Gulf and Vicinity.

Table 15. Fisher's Protected Least Significant Difference (FPLSD) Test Comparing the Mean Dissolved Oxygen/Cruise in the Upper 100 m Layer of RABUTINOS.

T Grouping			Mean	Cruise
	A		6.27	8
B	A		6.01	5
B	A		5.99	6
B	A	C	5.83	10
B		C	5.51	9
B		C	5.46	4
		C	5.29	13
		C	5.28	12
		C	5.26	3
Alpha = 0.05			LSD=0.64	

2. Relationship Between Biotic Groups and Environmental Factors.

a. *Environmental Characteristics of the Biotic Groups.* The results of Fisher's Protected LSD test among environmental variables are listed in Table 16. The shallow water group (i.e., Groups 4A, 4B, 5, 6 and 7) were significantly different from the deep water group (i.e., Groups 1, 2 and 3) in most of the environmental categories listed. Basically, the shallow water fishing stations were all above the thermocline while the deep water stations were either situated within or below the thermocline.

Within the deep water category, salinity was not a separating factor as most of the variability in salinity occurred near the sea surface and not at the bottom (Figure 23). What distinguished Group 3 (Ariommidae and Trichiuridae Assemblage) from Group 1 (Nemipteridae and Carangidae Assemblage) and Group 2 (Ariommidae + Deep Sea Forms) were temperature and DO concentration, i.e., Group 3 was significantly warmer and more oxygenated. At an average depth of 140 m, fishing stations belonging to Group 3 were located right at the thermocline and oxycline while fishing stations belonging to Group 1 (166 m) and Group 2 (185 m) were all located deeper. The vertical variations in temperature (Figure 27) and dissolved oxygen (Figure 29) below the thermocline were more gradual as shown by the widely spread isopleths below the thermocline/oxycline. None of the environmental factors measured in Group 1 and Group 2 were significantly different which implied that some other factors may have caused their separation.

Among the shallow water groups, the environmental conditions observed between fishing stations located above (i.e., Groups 4A, 6 and 7) and below (i.e., Groups 4B and 5) the 35 m isobath were significantly different from each other. Group 7 (Coralline Assemblage) was distinguishable from Group 6 (Coralline + Leiognathidae and Associated Estuarine Assemblage) and 4A (Leiognathidae + Coastal Pelagics) based on significantly lower DO concentration. Group 3 was warmer and had higher dissolved oxygen concentration than Groups 1 and 2 although it was not significantly shallower than Group 2.

Table 16. Fisher's Protected Least Significant Difference (FPLSD) Test Comparing the Mean Environmental Characteristics/Standardized Groups at RABUTINOS.

GROUP	TEMPERATURE*	SALINITY	DISSOLVED OXYGEN*	DEPTH*
	°C	(ppt)	(ml/liter)	(meter)
1	18.17 ^c	34.42 ^a	1.80 ^d	165.67 ^{ab}
2	19.31 ^c	34.46 ^a	2.39 ^d	185.25 ^a
3	21.98 ^b	34.42 ^a	3.68 ^c	139.26 ^b
4A	25.94 ^a	34.26 ^b	5.58 ^a	45.75 ^d
4B	26.07 ^a	34.32 ^b	5.18 ^{ab}	88.20 ^c
5	25.74 ^a	34.39 ^{ab}	5.18 ^{ab}	79.90 ^c
6	25.42 ^a	34.32 ^b	5.47 ^a	47.24 ^d
7	26.35 ^a	34.27 ^b	4.98 ^b	40.00 ^d

*Means in the same column with different letters are significantly different ($p < 0.05$)

^{ns}Not significant ($p > 0.05$)

Table 17. Fisher's Protected Least Significant Difference (FPLSD) Test Comparing the Mean Environmental Characteristics/Northeast Monsoon Seasonal Clusters at RABUTINOS.

GROUP	TEMPERATURE*	SALINITY ^{NS}	DISSOLVED OXYGEN*	DEPTH*
	°C	(ppt)	(ml/liter)	(meter)
NE1	18.17 ^c	34.42	1.80 ^b	165.67 ^a
NE3	21.73 ^b	34.37	3.75 ^c	147.40 ^a
NE4A	25.62 ^a	34.16	5.36 ^a	44.76 ^c
NE4B	27.14 ^a	34.21	5.00 ^a	85.00 ^b
NE6	25.61 ^a	34.22	5.46 ^a	56.75 ^c
NE7	25.88 ^a	34.23	5.51 ^a	42.78 ^c

*Means in the same column with different letters are significantly different ($p < 0.05$)

^{ns}Not significant ($p > 0.05$)

b. *Seasonal Differences in the Environmental Characteristics of Biotic Groups.*

The environmental characteristics of the different fishing stations corresponding to the SU fish assemblages are presented below according to the monsoon seasons:

(1) Northeast Monsoon Season. Six major biotic groups and sub-groups were present during the northeast monsoon season (Table 17), i.e., Groups 1, 3, 4A, 4B, 6 and 7. The "NE" prefix denotes that the groups being compared were from the northeast monsoon season. Salinity was not a significant environmental variable for discriminating the different groups. Fisher's Protected LSD test showed that the shallow water fishing stations (i.e., Groups NE4A, NE4B, NE6 and NE7) were significantly different from the deep water fishing stations (i.e., Groups NE1 and NE3) in all of the other environmental categories. While the depths of NE1 (Carangidae and Nemipteridae Assemblage) and NE3 (Ariommidae + Trichiuridae Assemblage) were not considerably different, NE3 stations located within the thermocline were substantially more oxygenated and warmer.

Among the shallow water groups, depth (i.e., above and below the 35 m isobath) was the only measured environmental variable that differentiated N4B (Leiognathidae + Ariommidae and Deep Sea Forms) from NE4A (Leiognathidae + Coastal Pelagics), NE6 (Coralline + Leiognathidae and Estuarine Assemblage) and NE7 (Coralline Assemblage).

(2) Dry Intermonsoon Period. Six major SU fish assemblage groups and sub-groups were represented during this period (Table 18). The prefix "IN" identifies the group to be compared as belonging to the intermonsoon period. Salinity was not very helpful in separating the different groups, but the rest of the measured environmental variables significantly discriminated the shallow water fishing stations (i.e., Groups IN4A, IN4b, IN5 and IN6) from the deep water fishing stations (i.e., Groups IN2 and IN3). Within the deep water group, all of the remaining environmental factors were also able to separate Group IN3 (stations within the thermocline) from Group IN2 (stations below the thermocline). For the shallow water stations, only depth was significantly better at separating the different biotic groups.

Table 18. Fisher's Protected Least Significant Difference (FPLSD) Test Comparing the Mean Environmental Characteristics/Dry Inter-Monsoon Period Clusters at RABUTINOS.

GROUP	TEMPERATURE*	SALINITY*	DISSOLVED OXYGEN*	DEPTH*
	°C	(ppt)	(ml/liter)	(meter)
IN2	20.18 ^c	34.44 ^a	2.48 ^d	176.58 ^a
IN3	23.67 ^b	34.40 ^a	4.12 ^c	120.27 ^b
IN4a	26.54 ^a	34.22 ^b	5.99 ^a	48.52 ^e
IN4b	25.68 ^a	34.35 ^{ab}	5.35 ^{ab}	86.33 ^c
IN5	25.58 ^{ab}	34.40 ^a	5.25 ^b	74.67 ^d
IN6	25.58 ^{ab}	34.32 ^{ab}	5.69 ^a	42.38 ^e

*Means in the same column with different letters are significantly different ($p < 0.05$)

^{ns} Not significant ($p > 0.05$)

Table 19. Fisher's Protected Least Significant Difference (FPLSD) Test Comparing the Mean Environmental Characteristics/Southwest Monsoon Seasonal Clusters at RABUTINOS.

GROUP	TEMPERATURE*	SALINITY ^{ns}	DISSOLVED OXYGEN*	DEPTH*
	°C	(ppt)	(ml/liter)	(meter)
SW2	16.70 ^c	34.52	2.15 ^d	211.25 ^a
SW3	20.92 ^b	34.46	3.12 ^c	143.60 ^b
SW4a	26.16 ^a	34.28	5.50 ^a	47.94 ^d
SW4b	25.33 ^a	34.42	5.15 ^{ab}	95.00 ^c
SW5	25.98 ^a	34.37	5.08 ^{ab}	87.75 ^c
SW7	26.88 ^a	34.31	4.38 ^b	36.88 ^d

*Means in the same column with different letters are significantly different ($p < 0.05$)

^{ns} Not significant ($p > 0.05$)

(3) Southwest Monsoon Season. There was no discernible difference in the salinities of the six biotic groups during the Southwest monsoon season (Table 19). Apart from these, all other environmental variables were considerably different between the deep fishing stations (Groups SW2 and SW3) and the shallow ones (Groups SW4A, SW4B, SW5 and SW7). The "SW" prefix refers to southwest monsoon season. Among the deep water fishing stations temperature, DO concentration and depth were significantly different between Group SW2 (Ariommidae + Deep Sea Forms) and Group SW3 (Ariommidae + Trichiuridae Assemblage).

Temperature and salinity were not substantially different among the shallow water groups. The DO concentration of the fishing stations belonging to Group SW4B (Leiognathidae + Arioma and Deep Sea Forms) were significantly higher than those of Group SW7 (Coralline Assemblage). Depth was also comparably different between Groups SW4B and SW5.

(4) ENSO Early Maturity Phase. Only three biotic groups were discernible during this period (Table 20). Except salinity, all other environmental factors measured for group EN3 (i.e., temperature, DO concentration and depth), were considerably different from the rest based on Fisher's Protected LSD test. Only dissolved oxygen concentration significantly differentiated Group EN4A and Group EN6.

c. Correlations Among Environmental Variables. The degree of interdependence or simple correlation among the measured environmental variables is listed in Table 21. All of the indicated correlations were significant at the $\alpha \leq 0.05$ level. Depth was positively correlated with salinity and negatively correlated with dissolved oxygen and temperature. The strength and direction of the correlation were as expected and reflected the normal variation of the temperature, salinity and dissolved oxygen concentration with depth. Salinity had the lowest and most variable correlation with the other environmental

Table 20. Fisher's Protected Least Significant Difference (FPLSD) Test Comparing the Mean Environmental Characteristics of the 1982-'83 El Niño-Southern Oscillation's Early Maturity Stage Clusters at RABUTINOS.

GROUP	TEMPERATURE*	SALINITY ^{NS}	DISSOLVED OXYGEN*	DEPTH*
	°C	(ppt)	(mL/liter)	(meter)
EN3	21.48 ^b	34.57	3.57 ^c	139.67 ^a
EN4a	24.91 ^a	34.53	5.30 ^a	49.33 ^b
EN6	24.50 ^a	34.53	4.75 ^b	44.00 ^b

*Means in the same column with different letters are significantly different ($p < 0.05$)

^{NS} Not significant ($p > 0.05$)

factors due to the greater influence of exogenous factors, i.e., precipitation levels, in its distribution and concentration.

The correlations between salinity and the rest of the measured environmental variables were strongest during periods of heavy precipitation (i.e., during the “estuarization” of RABUTINOS) and weakest during drought situations (i.e., when the vertical salinity structure broke down and the water column approached isohaline conditions).

e. *Multiple Regression on Fish Abundance and Environmental Variables.* Tables 22 to 25 list the seasonal multiple regression coefficients of the models for the different species that met the acceptable significance level and minimum r^2 contribution criteria. A total of 45, 33, 25 and 31 species have significant regression models (i.e., $\alpha \leq 0.05$) in the northeast monsoon (Cruises 3, 4 and 5), intermonsoon period (Cruises 6, 8 and 9), southwest monsoon (Cruises 10 and 12) and ENSO early maturity phase (Cruise 13), respectively. This accounted for 13% to 23% of the total number of species/taxa that were affected and/or responded to changes in environmental conditions.

For the northeast monsoon season, depth and salinity were the variables that figured prominently in the models with 23 and 22 species associations respectively. During the intermonsoon period, depth was associated with 20 species followed by salinity with 10. Depth was associated with 13 species during the southwest monsoon season followed by salinity and temperature with 7 and 6 species associations each. None of the species included DO concentration level as a significant variable in the model. Salinity and temperature were the environmental variables included in most of the models during the ENSO early maturity phase with 14 and 13 species associations, respectively. Multicollinearity was evident in models with more than one independent variable as expected from the table of correlations among environmental factors. Most of the models

Table 22. Multiple Linear Regression Coefficients of the Environmental Variables Included in the Species/Taxa Models for the NE Monsoon Season.

SPECIES NAME	CODE	Intercept	TEMP	SALIN	DOXY	DEPTH	R ²	Cp
<i>Ariomma indica</i>	ARIOMA	-9.39				0.2200	0.28 ²	3.3
Brotulinae	BROTUL	-0.03				0.0006	0.26 ²	0.8
Carapidae	CARAPI	-295.73		8.645			0.27 ²	-0.4
Champsodontidae	CHAMPO	-357.28		10.461			0.19 ¹	1.5
Chimaeridae	CHIMAE	-109.53		3.202			0.26 ¹	-0.4
<i>Chlorophthalmus albat.</i>	CHLORO	123.49		-3.497	-0.732		0.62 ¹	2.5
<i>Chirocentrus dorab</i>	CIROCE	27.97		-0.814			0.34 ²	2.8
Congridae	CONGRI	-140.14		4.096			0.27 ¹	-0.3
<i>Caranx chrysophrys</i>	CRXCRY	-277.88		8.196		-0.0260	0.36 ¹	1.1
<i>Caranx dinema</i>	CRXDIN	1.82				-0.0130	0.17 ¹	1.3
<i>Caranx malabaricus</i>	CRXMAL	2.08				-0.0150	0.17 ¹	0.4
Cynoglossidae	CYNOGL	-522.85		15.284			0.27 ¹	-0.3
<i>Decapterus kurroides</i>	DECAKU	57.10			-9.329	-0.1350	0.65 ²	1.4
<i>Decapterus macrosoma</i>	DECAMA	16.90	-0.655				0.18 ¹	1.5
<i>Dussumiera acuta</i>	DUSSUM	-19.14	1.326		-2.549		0.39 ¹	5.9
<i>Epinephelus spp.</i>	EPISPP	6.75			-1.114	-0.0150	0.49 ¹	1.2
<i>Formio niger</i>	FORMIO	110.49		-3.214			0.24 ¹	0
<i>Leiognathus splendens</i>	LEOSPL	849.85		-24.738			0.21 ¹	0.6
<i>Loligo spp.</i>	LOLIGO	-205.73	11.602		-17.867	0.1610	0.81 ¹	3.9
Lophiidae	LOPHII	-530.57		15.538			0.28 ²	5.7
Myctophidae	MYCTOP	-23.71		0.693			0.26 ¹	-0.4
<i>Nemipterus bathybius</i>	NEMIBA	-0.60				0.0180	0.30 ²	1.7
<i>Nemipterus hexodon</i>	NEMIHE	17.42		-0.507			0.18 ¹	0.1
<i>Nemipterus marginatus</i>	NEMIMA	16.70			-2.717	-0.0410	0.58 ²	1.1
<i>Priacanthus macracan.</i>	PRIAMA	-9.55				0.2130	0.63 ²	3.1
<i>Promethichthys promet.</i>	PROMET	-0.11				0.0020	0.35 ²	0.3
<i>Pseudorhombus eleva.</i>	PSEUEL	-472.52		13.814			0.25 ¹	-0.5
<i>Pseudorhombus spp.</i>	PSEUSP	-908.21		26.549			0.26 ¹	-0.4
<i>Rastrelliger brachysoma</i>	RASTBR	241.11		-7.012			0.26 ¹	0.5
<i>Sardinella longiceps</i>	SARDLO	295.59		-8.605			0.22 ¹	0.7
<i>Saurida undosquamis</i>	SAURUN	-3.20				0.1100	0.67 ¹	5.1
<i>Scolopsis inermis</i>	SCOLIN	52.85				-0.1240	0.69 ²	1.9
<i>Scomberomorus comm.</i>	SCOMME	4.79				-0.0300	0.24 ²	-0.3
<i>Secutor ruconius</i>	SECURU	275.83	-0.232	-7.882			0.48 ¹	3
<i>Selar boops</i>	SELABO	39.61	-0.032	-1.133			0.65 ²	3.9
<i>Selar crumenophthalmus</i>	SELACR	39.22		-1.140			0.27 ²	0.1
<i>Selar melanoptera</i>	SELAME	85.37	-0.063	-2.444			0.50 ¹	1.5
<i>Selaroides leptolepis</i>	SELARO	343.15		-9.984			0.34 ²	2.6
<i>Seiola nigrofasciata</i>	SERION	-6.66	0.456		-0.086		0.50 ²	1.5
Sharks	SHARKS	-1.90				0.0580	0.27 ²	2.4
<i>Sphyaena langsar</i>	SPHYLA	1.11				-0.0070	0.17 ¹	-0.8
Tetraodontidae	TETRAO	-12.88			2.022	0.0510	0.42 ²	1.5
<i>Trichiurus haumela</i>	TRICHI	-2.73				0.0820	0.23 ¹	2
<i>Upeneus mollucensis</i>	UPEMOL	-61.01	2.255			0.0930	0.38 ²	1.6
Uranoscopidae	URANOS	-5.20				0.0880	0.40 ²	0

Legend: 1 = $p < 0.05$ 2 = $p < 0.01$ 3 = $p < 0.001$

Table 23. Multiple Linear Regression Coefficients of the Environmental Variables Included in the Species/Taxa Models for the Dry Inter-Monsoon Period.

SPECIES NAME	CODE	Intercept	TEMP	SALIN	DOXY	DEPTH	R ²	Cp
<i>Aluterus scriptus</i>	ALUTSC	120.22		-3.493			0.23 ¹	0.4
<i>Aphareus rutilans</i>	APHARE	-0.02				0.000	0.20 ¹	1
<i>Ariomma indica</i>	ARIOMA	-15.61				0.333	0.24 ¹	0.3
<i>Bothus spp.</i>	BOTHUS	-1.25				0.022	0.22 ¹	0.4
Brotulinae	BROTUL	-0.48				0.009	0.24 ¹	0.2
<i>Canthigaster compressus</i>	CANTIG	-0.37				0.006	0.18 ¹	0.8
Chimaeridae	CHIMAE	-0.51				0.001	0.18 ¹	0.9
<i>Chirocentrus dorab</i>	CIROCE	-1.19			0.349		0.20 ¹	1
Congridae	CONGRI	-0.07				0.001	0.20 ¹	0.8
Cynoglossidae	CYNOGL	-0.06				0.001	0.18 ¹	0.8
Diodontidae	DIODON	261.62			-33.865	-1.008	0.37 ¹	3.7
Labridae	LABRID	34.85			-4.524	-0.134	0.35 ¹	3
Lactariidae	LACTAR	-0.03				0.001	0.18 ¹	0.8
<i>Leiognathus splendens</i>	LEOSPL	930.39		-27.017			0.17 ¹	0.1
<i>Loligo spp.</i>	LOLIGO	480.39		-13.944			0.57 ³	0.5
Lophiidae	LOPHII	-1.86				0.045	0.19 ¹	2.2
Myctophidae	MYCTOP	-0.35				0.007	0.29 ²	-0.2
<i>Nemipterus bathybius</i>	NEMIBA	6.58			-1.131		0.30 ²	5.6
<i>Priacanthus macracanth.</i>	PRIAMA	80.42	-3.072				0.70 ³	-0.2
<i>Promethichthys promet.</i>	PROMET	-0.17				0.003	0.19 ¹	0.9
<i>Rastrelliger kanagurta</i>	RASTKA	168.28		-4.881			0.21 ¹	-0.2
<i>Rexea solandri</i>	REXEAS	-14.44			2.052	0.060	0.29 ¹	1
<i>Sardinella fimbriata</i>	SARDFI	878.03		-25.628		0.029	0.84 ³	4.6
<i>Saurida undosquamis</i>	SAURUN	-3.25				0.081	0.22 ¹	1.9
<i>Scolopsis inermis</i>	SCOLIN	1.04	-0.040				0.21 ¹	3.3
<i>Scomberomorus comm.</i>	SCOMME	421.11		-12.224			0.29 ²	-0.7
<i>Secutor insidiator</i>	SECUIN	26.51		-0.770			0.19 ¹	4.9
<i>Selar malam</i>	SELAMA	47.22		-1.372			0.34 ²	3.7
<i>Selar mate</i>	SELAMT	1.22				-0.008	0.18 ¹	-0.3
<i>Selaroides leptolepis</i>	SELARO	909.17		-26.432			0.60 ³	1
Shrimp	SHRIMP	-0.60				0.011	0.26 ¹	0.2
<i>Stolephorus tri</i>	STOLTR	234.66		-6.815			0.44 ³	-0.5
<i>Upeneus vittatus</i>	UPEVIT	15.72			-2.307	-0.048	0.32 ¹	2.1

Legend: 1 = $p < 0.05$ 2 = $p < 0.01$ 3 = $p < 0.001$

Table 24. Multiple Linear Regression Coefficients of the Environmental Variables Included in the Species/Taxa Models for the Southwest Monsoon Season.

SPECIES NAME	CODE	Intercept	TEMP	SALIN	DOXY	DEPTH	R ²	Cp
<i>Ariomma indica</i>	ARIOMA	183.98	-6.537				0.18 ¹	0.9
Balistidae	BALIST	47.10		-1.370			1.00 ³	11.5
<i>Bothus spp.</i>	BOTHUS	2.75	-0.102				0.30 ²	2
Carapidae	CARAPI	-0.24				0.0040	0.39 ³	0.5
Champsodontidae	CHAMPO	2.87	-0.106				0.26 ¹	1
<i>Chelmon rostratum</i>	CHELMO	0.02		-0.001			1.00 ³	28
Chimaeridae	CHIMAE	-0.24				0.0040	0.35 ²	0.3
<i>Chlorophthalmus albatros</i>	CHLORO	-0.20				0.0030	0.54 ³	2.7
Cynoglossidae	CYNOGL	-0.01				0.0001	0.39 ²	0.5
Dactylopteridae	DACTYL	1.15	-0.041				0.28 ²	3.2
<i>Decapterus kurroides</i>	DECAKU	-1.79				0.0298	0.43 ³	0.8
<i>Decapterus muruadsi</i>	DECAMU	-12.82				0.2650	0.41 ³	-0.4
<i>Decapterus russeli</i>	DECARU	8.19		-0.342		0.0540	0.45 ¹	1.8
<i>Formio niger</i>	FORMIO	-3.54	0.171				0.23 ¹	1
<i>Histioporus indicus</i>	HISTIN	4.49	-0.167				0.35 ²	3.4
Lophiidae	LOPHII	-0.49				0.0310	0.19 ¹	2.4
<i>Nemipterus hexodon</i>	NEMIHE	3.05		-0.088			0.95 ³	3.2
<i>Priacanthus macracanthus</i>	PRIAMA	-2.66				0.1020	0.36 ²	0.5
<i>Promethichthys prometheus</i>	PROMET	-0.54				0.0090	0.41 ³	0.3
Rhinobatidae	RHINOB	-6.95				0.1130	0.39 ²	0.5
<i>Saurida undosquamis</i>	SAURUN	-1.43				0.0740	0.30 ²	0.1
Shrimp	SHRIMP	13.13		-0.351			0.37 ²	1.6
Triglidae	TRIGLI	30.34		-0.816			0.42 ³	1.6
<i>Upeneus vittatus</i>	UPEVIT	-0.70				0.0120	0.44 ³	0.9
Uranoscopidae	URANOS	27.73		-0.796			0.95 ³	24

Legend: 1 = p<0.05 2 = p<0.01 3 = p<0.001

Table 25. Multiple Linear Regression Coefficients of the Environmental Variables Included in the Species/Taxa Models for the ENSO Early Maturity Stage

SPECIES NAME	CODE	Intercept	TEMP	SALIN	DOXY	DEPTH	R ²	Cp
<i>Apogonidae</i>	APOGON	1004.06		-29.048			0.27 ¹	-0.8
<i>Brotulidae</i>	BROTUL	0.86	-.034				0.25 ¹	3.7
<i>Canthigaster compressus</i>	CANTIG	658.70		-19.063			0.46 ²	-0.1
<i>Centricus scutatus</i>	CENTRI	7325.39		-212.004			0.47 ²	0.3
<i>Champsodontidae</i>	CHAMPO	38.00	-1.544				0.31 ²	4.0
<i>Chimaeridae</i>	CHIMAE	7.57	-.031				0.31 ²	4.0
<i>Chlorophthalmus albatro.</i>	CHLORO	0.76	-.031				0.53 ³	2.5
<i>Congridae</i>	CONGRI	0.31	-.013				0.31 ²	4.0
<i>Coradion altiveles</i>	CORADI	31.37		-.907			0.22 ¹	3.9
<i>Caranx chrysophrys</i>	CRXCRY	2247.56		-65.047			0.47 ²	0.3
<i>Caranx malabaricus</i>	CRXMAL	2501.73		-72.391			0.54 ³	-0.7
<i>Cynoglossidae</i>	CYNOGL	0.62	-.025				0.31 ²	4.0
<i>Dasyatidae</i>	DASYAT	344.68	-13.684				0.35 ²	3.7
<i>Diodontidae</i>	DIODON	108.73			-16.006	-.362	0.41 ¹	4.5
<i>Diplopriion bifasciatus</i>	DIPLOP	3.30		-.096			0.47 ²	0.3
<i>Labridae</i>	LABRID	36.01			-5.335	-.121	0.42 ¹	3.0
<i>Lactariidae</i>	LACTAR	27.59	-1.121				0.31 ¹	3.9
<i>Leiognathus elongatus</i>	LEOELO	71.47		-2.068			0.29 ¹	1.9
<i>Lutjanus lineolatus</i>	LUTLIN	3394.14		-98.228			0.47 ²	0.1
<i>Myctophidae</i>	MYCTOP	4.90	-.308		.569		0.37 ¹	2.1
<i>Nemipterus nematophorus</i>	NEMINE	-2.35	.251		-.732		0.62 ³	17.7
<i>Parupeneus indicus</i>	PARUIN	298.83		-8.645			0.27 ¹	3.7
<i>Plectorhincus pictus</i>	PLECIN	94.85			-13.987	-.321	0.41 ¹	2.9
<i>Priacanthus macracanthus</i>	PRIAMA	-2.10				.046	0.22 ¹	0.3
<i>Pseudorhombus oligodon</i>	PSEUOL	3.38	-.184		.236		0.55 ¹	5.0
<i>Saurida undosquamis</i>	SAURUN	-32.73			5.252	.141	0.40 ¹	1.7
<i>Scolopsis inermis</i>	SCOLIN	1791.70	-1.524	-50.792			0.54 ¹	9.3
<i>Scolopsis taeniopterus</i>	SCOLTA	170.32		-4.926			0.22 ¹	3.5
<i>Scorpaenidae</i>	SCORPI	9.49	-.374				0.24 ¹	3.4
<i>Secutor insidiator</i>	SECUIN	2.66		-.077			0.47 ²	0.3
<i>Selar mate</i>	SELAMT	579.12		-16.751			0.24 ¹	1.0

Legend: 1 = $p < 0.05$ 2 = $p < 0.01$ 3 = $p < 0.001$

with $C_p < p$ were the result of the trade-off between the high minimum r^2 incremental contribution required of a variable *versus* a better C_p index.

The seasonal regression coefficients of the environmental variables and their signs are summarized in Table 26. Within their normal range of distribution, the shallow (deep) water species generally had negative (positive) coefficients for depth. The signs were also consistent with the seasons except for *Decapterus kurroides* and *Upeneus vittatus*. The abundance of *D. kurroides* increased with decreasing depth during the northeast monsoon and the reversed during the southwest monsoon season. *U. vittatus* had a similar but reverse response.

The abundance of coastal species increased with decreasing salinity (negative salinity coefficients) while for the deep sea species, it increased with increasing depth. The relationship between salinity and abundance was more prevalent and strong among shallow, coastal dwellers during the dry season as indicated by the presence of strictly negative salinity coefficients in the models involving the intermonsoon period and ENSO early maturity phase. A reversal in sign from negative (northeast monsoon) to positive (southwest monsoon) was seen only in *Formio niger*, a riverine/estuarine species found in Group 4A/B and Group 6.

Most of the temperature coefficients were negative and associated with species that inhabited the deep seas. Temperature was important in predicting the abundance of only a few coastal species, e.g., *Dussumiera acuta*, *Loligo* spp., *Nemipterus nematophorus*, *Seriolina nigrofasciata* and *Upeneus mollucensis*. Except for two species, DO concentration levels factor in the regression model only in conjunction with another primary variable, mostly depth and temperature.

Table 26. Signs of the Multiple Linear Regression Coefficients Included Respective Species/Taxa Models for the Different Seasons at RABUTINOS.

SPECIES NAME	CODE	NE Monso				Intermonso				SW Monsoon				ENSO			
		T	S	O	D	T	S	O	D	T	S	O	D	T	S	O	D
<i>Aluteres scriptus</i>	ALUTSC					-											
<i>Aphareus rutilans</i>	APHARE							+									
Apogonidae	APOGON														-		
<i>Arioma indica</i>	ARIOMA			+				+	-								
Balistidae	BALIST									-							
<i>Bothus spp.</i>	BOTHUS							+	-								
Brotulinae	BROTUL			+				+						-			
<i>Canthigaster compressus</i>	CANTIG							+						-			
Carapidae	CARAPI	+											+				
<i>Centriscus scutatus</i>	CENTRI													-			
Champsodontidae	CHAMPO	+							-					-			
<i>Chelmon rostratum</i>	CHELMO								-								
Chimaeridae	CHIMAE	+						+				+	-				
<i>Chlorophthalmus albatros</i>	CHLORO	-	-									+	-				
<i>Chirocentrus dorab</i>	CIROCE	-				+											
Congridae	CONGRI	+						+					-				
<i>Coradion altiveles</i>	CORADI													-			
<i>Caranx chrysophrys</i>	CRXCRY	+	-											-			
<i>Caranx dinema</i>	CRXDIN		-														
<i>Caranx malabaricus</i>	CRXMAL		-											-			
Cynoglossidae	CYNOGL	+						+				+	-				
Dactylopteridae	DACTYL								-								
Dasyatidae	DASYAT													-			
<i>Decapterus kurroides</i>	DECAKU		-	-								+					
<i>Decapterus macrosoma</i>	DECAMA	-															
<i>Decapterus muruadsi</i>	DECAMU											+					
<i>Decapterus russeli</i>	DECARU								-			+					
Diodontidae	DIODON							-	-						-	-	
<i>Diploprion bifasciatus</i>	DIPLOP													-			
<i>Dussumiera acuta</i>	DUSSUM	+	-														
<i>Epinephelus spp.</i>	EPISPP		-	-													
<i>Formio niger</i>	FORMIO	-								+							
<i>Histioporus indicus</i>	HISTIN									-							
Labridae	LABRID							-	-						-	-	
Lactariidae	LACTAR							+						-			
<i>Leiognathus elongatus</i>	LEOELO													-			
<i>Leiognathus splendens</i>	LEOSPL	-				-											
<i>Loligo spp.</i>	LOLIGO	+	-	+		-											
Lophiidae	LOPHII	+						+				+					
<i>Lutjanus lineolatus</i>	LUTLIN													-			
Myctophidae	MYCTOP	+						+					-		+		
<i>Nemipterus bathybius</i>	NEMIBA			+		-											
<i>Nemipterus hexodon</i>	NEMIHE	-								-							
<i>Nemipterus marginatus</i>	NEMIMA		-	-													
<i>Nemipterus nematophorus</i>	NEMINE												+	-			
<i>Parupeneus indicus</i>	PARUTN													-			
<i>Plectorhincus indicus</i>	PLECIN													-	-		
<i>Priacanthus maculatus</i>	PRIAMA			+	-							+				+	
<i>Promethichthys prometheus</i>	PROMET			+				+				+					
<i>Pseudorhombus elevatus</i>	PSEUEL	+															
<i>Pseudorhombus oligodon</i>	PSEUOL													-	+		

DISCUSSION

A. Community Structure:

This study demonstrated that the demersal fish assemblages of RABUTINOS display distinctive patterns of distribution and abundance. The sharpest changes in species composition occurred along the depth gradient corresponding to the three major oceanic zones similar to those established in the eastern tropical Pacific Ocean (Bianchi, 1991). These were the upper zone (down to about 90 - 100 m depth), the intermediate zone (between 100 - 150 m depth) and the deeper zone (>150 m). Each of these zones has their own distinct physico-chemical characteristics. The upper zone corresponds to the warm, low salinity, highly oxygenated wind-mixed layer. The intermediate zone is highly influenced by the thermocline and displays rapid changes in physico-chemical characteristics of the water masses while the deeper zone corresponds to the cold, poorly oxygenated and high salinity waters.

Fager and Longhurst (1968) found that separation boundary between different fish assemblages in the Gulf of Guinea was related to the thermocline as well as to sediment type, which also co-varies with depth (Bianchi, 1991). McManus (1985) found depth-dependent faunal distinction between 30 and 40 m in Samar Sea, independent of the season and substrate type. Federizon (1992) also found two important gradients in his ordination: the depth gradient and the coralline gradient. The dividing depth for Federizon's demersal fish assemblages was the 90 m isobath. The discrepancy between the separation depths of these two adjacent bodies of water and their resolution is discussed in the next section.

Hutchinson's (1957) view of a niche as an N-dimensional hypervolume combined with Fry's autecological paradigms on the responses of fishes to their environment can be used to relate depth to fish distribution (Magnuson et al., 1979; Kerr, 1980; Crowder and

Magnuson, 1983). For example, the thermal niche of a fish can be described as a range of preferred temperatures 4°C to 10°C wide. Using this concept, Magnuson *et al.* (1979) classified fishes into three thermal preference categories, i.e., coldwater fishes (10°C to 15°C), coolwater fishes (20°C to 25°C) and warmwater fishes (25°C to 30°C). This corresponds to the temperature of water within the deeper, intermediate and upper zones of the oceans, respectively, and provides a physiological basis for the way fish communities are structured.

Although depth, acting as a proxy variable, may be the principal determinant of fish distribution in RABUTINOS, the results of the correlation and linear regression analysis suggest other factors may also be important. This may include sediment type, turbidity and water mass characteristics. Turbidity is a primary factor affecting the distribution of estuarine and inshore fishes in the tropics (Cyrus and Blaber, 1987; Blaber *et al.*, 1994) but has not been studied well with respect to tropical marine shelf assemblages. While temperature, salinity and DO concentration of water were adequately sampled, sediment type and turbidity were not. Refinements and quantification of these variables need to be carried out in the future. The approach followed in the northern Australian shelf (see Sainsbury, 1988) of taking continuous photographs of the sediment characteristics along the path of the trawl is a novel idea that may be worth trying.

Within each depth zone, secondary environmental gradients become more relevant in structuring demersal fish communities. At RABUTINOS these range from substrate characteristics (e.g., presence of rocky-coraline habitat or sandy to muddy bottom), geographic location (e.g., proximity to river mouths), and characteristics of the water masses (e.g., temperature, salinity, and DO concentration level) either alone or in tandem (Ramm *et al.*, 1990; Bianchi, 1991; Blaber *et al.*, 1994). Quinn (1980) postulated that salinity is the common feature between temperate and tropical ecosystem in the maintenance of community cycles.

My community analysis suggested that the fish fauna of RABUTINOS could be split into seven major groups. The primary separation found in the analysis is based on the 100 m isobath which separates the deep (cold water, sub-tropical fauna) from the shallow (warm, tropical faunal) assemblages. The deep assemblages can be subdivided further into Group 3 assemblage typical of the intermediate zone (or thermocline), Group 2 assemblage of the deep sea zone, and Group 1 assemblage composed primarily of scombrids and nemipterids characteristic of the cold, subtropical habitats (Longhurst and Pauly, 1987). Group 1's singular appearance, i.e., during the northeast monsoon season may reflect its seasonal nature, inadequacy of samples from the deeper regions, or both. Group 2 and Group 3 are essentially *Ariomma indica* and *Trichiurus haumela* dominated assemblages. Group 3 represented a community clearly dominated by these two species while Group 2 contained a mixture of deep sea forms like stargazers, angler fishes, bigeyes, and goosefishes.

The shallow water assemblages can be subdivided into two sub-groups: the coralline group or the Leiognathidae group. The coralline group includes Group 7 with its characteristic even distribution of species/taxa abundance and high similarity with coral reef species, and Group 6 -- a transition group composed of coralline fish assemblages mixed with the *Leiognathus bindus* assemblage. The Leiognathidae assemblage can be subdivided into those in close proximity of river mouths (Group 4A) and those relatively deeper (>50 m) and farther away in clear waters (Group 4B). Group 4A is basically composed of Leiognathidae family + coastal pelagic species while Group 4B is a combination of Leiognathidae family + *Ariomma indica* and other deep sea forms. Group 5 defined the *Leiognathus bindus* dominated assemblage inhabiting the relatively deeper (>50 m), sandy bottom areas intermittently flushed by distant rivers.

Aside from abiotic factors, biotic factors also contribute to the way communities are structured. For example, the spatial distributions of some congeneric species were partitioned along geographic or bathymetric gradients, or both (Ramm et al., 1990). Based on the analysis of each group's species composition, *Rastrelliger brachysoma* -- a

neritic scombrid form, occurred predominantly in Group 4A assemblage while *R. kanagurta*, the oceanic form of the scombrid family, was associated more with Group 4B assemblage. These two scombrid forms are separated based on salinity preference, food habits, and avoidance or tolerance of highly turbid waters (Jones and Rosa, 1965; Collete and Nauen, 1983; Chullasorn and Martosubroto, 1986). The members of the family Leiognathidae were also separated according to depth (Villoso and Aprieto, 1983) as well as food (Jones, 1985; Chullasorn and Martosubroto, 1986).

Classic predator-prey interactions were also evident in the way Group 4A assemblage composition was structured. The principal species comprising Group 4A are prey species which include the Leiognathidae family (*Leiognathus splendens*, *L. bindus*, *L. leuciscus*, *L. equulus*), the coastal pelagics represented by the sardines (*Sardinella longiceps*, *Dussumiera acuta*, family: Clupeidae), anchovy (*Stolephorus tri*, family: Engraulididae), round scad (*Decapterus muruadsi*, family: Carangidae) and mackerel (*Rastrelliger brachysoma*, family: Scombridae). These species are attracted to shallow areas, especially near estuaries due to the abundance of their main food items (Hardenberg, 1955; Jones and Rosa, 1965; Tiews *et al.*, 1968; Tiews *et al.*, 1970a; Tiews *et al.*, 1970b; Weng, 1990), i.e., mainly phytoplankton dominated microplankton, fish eggs and fish larvae (Manacop, 1956). In turn, the abundance of these prey species in a particular area attracts their main predators, e.g., mackerel (*Scomberomorus commerson*, family: Scombridae), hairtail (*Trichiurus haumela*, family: Scombridae), squid (*Loligo* spp., family: Loliginidae) and lizardfish (*Saurida tumbil*, family: Synodidae) to aggregate into the same area (Jones and Rosa, 1965; Tiews *et al.*, 1972; Chullasorn and Martosubroto, 1986; Robertson and Duke, 1990).

Some evidence of competition as a mechanism structuring tropical communities is evident from the abundances of *Rastrelliger kanagurta* and *Sardinella longiceps*. Both species have similar food and habitat preferences (mainly Group 4A and Group 4B fishing station locations) such that a high abundance of one group in a particular area, e.g., *S. longiceps* in Group 4A stations, corresponds with a low abundance of the other,

i.e., *R. kanagurta*. This was similar to the descriptions of Jones and Rosa (1965), Chullasorn and Martosubroto (1986), and Longhurst and Pauly (1987).

B. Zoogeographic Affinities:

A number of authors (e.g., Rainer and Munro, 1982; McManus, 1985; Ramm *et al.*, 1990; Bianchi, 1991; Federizon, 1992; this study) have already established depth as a major gradient associated with the zonation of demersal fish communities (i.e., species composition changes with increasing depth). However, Ramm *et al.* (1990) and Federizon (1992) expressed caution when demarcating survey areas based on depth gradation alone as problems of spatial scales and seasonal factors (i.e., multiple gradients) may also be important in structuring fish communities. In most cases, depth is only a proxy variable representing the vertical distribution of some other environmental gradient that is physiologically of consequence (Magnuson, 1991; Jamir *et al.*, 1994).

The demarcation depth separating “shallow” and “deep” communities in Ragay Gulf and the adjacent Samar Sea highlights the limitations on the use of depth as the sole explanatory variable without searching for its environmental correlates. In partitioning the ordinated station points in Ragay Gulf, Federizon (1992) distinguished two subcommunities (i.e., “shallow” and “deep”) separated by the 100 m isobath. In Samar Sea, McManus (1985) also came up with “shallow” and “deep” subcommunity designations separated by the 35 m isobath. In the absence of environmental data and detailed analysis of the ecological characteristics of the component species, Federizon attributed this discrepancy to possible bias induced by differences in the sample depth-frequency distribution. As a result, Federizon was not so sure whether either the 35 m or 100 m isobath really marked the boundaries of natural communities in Samar Sea and Ragay Gulf.

Similar disagreements existed in the findings of Rainer and Munro (1982) and Rainer (1984), vis-à-vis, Harris and Poiner (1990) and Ramm *et al.* (1990). The former asserted that seasonal variations in temperature and salinity were the main determinants of faunal distribution in the shallow, northern Australian continental shelves while the latter group believed that depth and spatial factors were more important. Ramm *et al.* (1990) suggested the possibility that sampling scale (both spatial and temporal) may be behind the differences in the observed results.

Ramm *et al.* (1990) were right in bringing sampling scale into the question. The maximum depth of Samar Sea is only 90 m, similar to the Gulf of Carpentaria (<80 m), with predominantly sandy and muddy substrates. RABUTINOS has a maximum sampling depth of 210 m covering a wide range of habitat types (e.g., riverine to deep marine, sandy to muddy substrates, soft bottom to rocky-coralline surface). Temporal scale is also important. In the case of Ramm *et al.* (1990), their sampling methodology prevented stratification by depth, season, fishing ground and time of day while Rainer and Munro's (1982) survey was limited to the post wet monsoon season months of March to May only. Although the data set used by Federizon and McManus spanned a period of 15 months, seasonal information was lost due to annual pooling and averaging the data.

Figure 30 compares the fish assemblage groupings derived by McManus (1985), Federizon (1992) and this study. In general, this study agrees with Federizon on the three major subcommunities, i.e., Coralline, Shallow and Deep. However, this study goes further by refining these subdivisions to include the possible effects of seasonal and other environmental factors in structuring the fish communities of RABUTINOS. The value of cluster standardization when doing comparative analysis of different author's work was also highlighted in this figure. It was clear that the two subcommunities identified by McManus were not the same as Federizon's "shallow" and "deep" categories. They were actually subsets of the latter's shallow water (<100 m) community. Combined with an analysis of the principal species comprising McManus's two subgroups, the shallow (<40 m, muddy substrate, dominated by Leiognathidae + coastal pelagics + *Rastrelliger*

COMPARISON OF SITE GROUP HABITAT & ASSEMBLAGE CHARACTERISTICS

FEDERIZON JAMIR MCMANUS

Shallow/Estuarine Assemblage:
 "Shallow" Group 4A (muddy deposit; riverine) "Shallow"
 Group 4B (muddy deposit; clear water)
 Group 5 (sandy deposit; clear water) "Deep"

Deep/Cold Water Assemblage:
 "Deep" Lower Thermocline/Halocline:
 Group 3 (sandy)
 Deep Shelf:
 Group 1 (sandy)
 Deep Shelf/Slope:
 Group 2 (sandy/silty ooze)

OTHERS:

Shallow Coralline Assemblage:
 "Coralline" Group 6 (Coralline-Sandy/Estuarine Transition)
 Group 7 (Coralline)

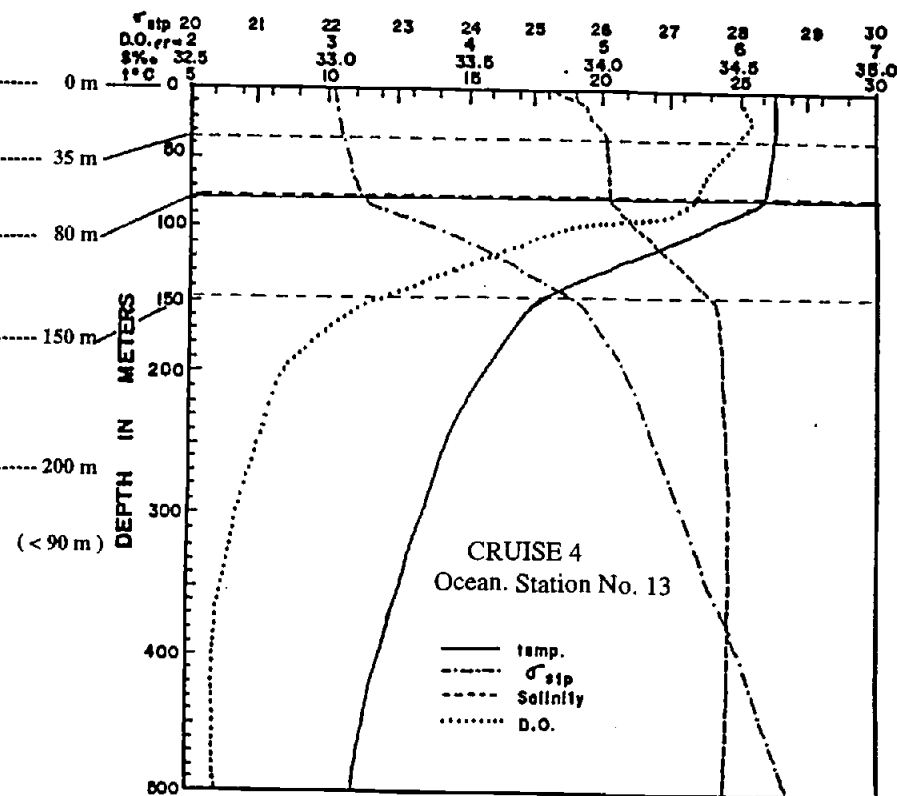


Figure 30. Comparison of McManus', Federizon's and Jamir's Site Group Categories In Relation to Habitat Characteristics.

brachysoma) community corresponded to Group 4A of this series, while the deep (>40 m, sandy substrate, dominated by *Leiognathus bindus*) community corresponded to Group 5.

The 35 - 40 m demarcation between the two communities most probably reflect the effects of riverine conditions on substrate characteristics, depth, salinity and productivity (Caddy and Bakun, 1995; Crivelli *et al.*, 1995; Halim *et al.*, 1995). This depth range roughly corresponds to water bodies subjected to freshwater dilution by rivers and other freshwater sources (Caddy and Bakun, 1995). For example, at the end of the rainy season, low salinity layers extending down to 30 m depth were commonly observed along the east coast of Sumatra (Wyrski, 1961) and in Lingayen Gulf (Sebastian *et al.*, 1959), San Miguel Bay (Pauly and Mines, 1982), Samar Sea (Labao, 1980), Manila Bay, Philippines (Megia *et al.*, 1953), Central Ragay Gulf (Jamir, 1986) and this study. Surnarayana *et al.* (1992) showed that Indian monsoon rains mixes within the upper 50 m of the water column and spreads offshore to distances of up to 150 km as a lens of low-salinity water. Seasonal estuarization of the shallow Gulf of Carpentaria during the rainy season has also been reported (Longhurst and Pauly, 1987).

For the coastal and estuarine fish assemblages studied by Sheaves (1992, 1993, 1996, 1998), the physical environment (mainly related to salinity) and biological processes (supply of recruits) interact to control the distribution of individual species and determine locality-specific assemblage patterns. Besides these, Weng (1990) attributed the interaction between food availability, habitat preference and hydrological characteristics for the observed distribution of fish assemblages. The principal species in the shallow regions of Samar Sea included *Stolephorus indicus* (Family: Engraulidae) and *Rastrelliger brachysoma* (Family: Scombridae) while in waters deeper than 40 m, the dominant species were *Saurida undosquamis* (Family: Synodidae), *Decapterus macrosoma* (Family: Carangidae) and *Rastrelliger kanagurta* (Family: Scombridae) (McManus, 1985). The evolution of food partitioning among scombrids as a means of reducing competition (Collette and Nauen, 1983) limited the distribution of *Rastrelliger brachysoma* primarily within shallow, low salinity (<32‰), high turbidity coastal waters

and estuaries rich in microplankton mix dominated by phytoplankton (Jones and Rosa, 1965). *R. kanagurta*, however, prefer salinities $>32\text{‰}$ and microplankton mix dominated by foraminiferans (Jones and Rosa, 1965; Chullasorn and Martosubroto, 1986). In the Southeast Asian waters, anchovies are usually confined within the nearshore and estuarine areas (Chullasorn and Martosubroto).

While all marine and most estuarine organisms can withstand full sea water, some cannot withstand lowered salinities (Gunter, 1961). *Decapterus macrosoma* (Family: Carangidae) and *Rastrelliger kanagurta* (Family: Scombridae) both avoid waters $<30\text{‰}$ and prefer depths greater than 40 m (Jones and Rosa, 1965; Chullasorn and Martosubroto, 1986). *D. macrosoma* (Family: Carangidae) does not enter Manila Bay (Tiews *et al.*, 1970) as well as the bays of Jakarta (Soemarto, 1960) where they avoid zones of lower salinities. *Saurida undosquamis* (Family: Synodidae) can be found in depths ranging from 30 - 90 m but prefer deeper regions (Chullasorn and Martosubroto, 1986).

Environmental association also indicated that Federizon's "shallow" and "deep" fish assemblage categories actually correspond to natural ecological boundaries, i.e., corresponding to Magnuson *et al.*'s (1979) fishes living above the thermocline, within the thermocline and below the thermocline (Bianchi, 1990). The underlying physiological basis for such separation may be related to the characteristics of Magnuson *et al.*'s (1990) warm water, cool water and cold water fish. Understanding this principle facilitates regional comparison of research results. For example, given the oceanographic characteristics of western and eastern boundary regions (Pickard and Emery, 1982), it is easy to see that the 90m to 100 m demarcation depth separating the warm from the cold/cool water fishes in the western tropical Pacific is essentially the same as the 50 m demarcation depth found by Bianchi (1991) in the eastern tropical Pacific Ocean and by Longhurst and Pauly (1987) in the eastern tropical Atlantic Ocean.

The significance of the thermocline as a natural zone of separation between two physiologically distinct fish communities were recognized by Longhurst and Pauly

(1987) in their analysis of the Guinean Trawling Survey (GTS). The equatorial submergence through the Gulf of Guinea of the sparid-dominated fauna from the cooler regions to the north and south is a good example to illustrate this point. The western Pacific counterpart, composed mainly of a subtropical fauna dominated heavily by sea bream (Sparidae, Nemipteridae), large croaker (Sciaenidae) and lizardfish (Synodidae), replaced the shallow-water tropical fauna not far to the south of Hong Kong, and near Carnarvon in western Australia and Cooktown in Queensland (Longhurst and Pauly, 1987). These subtropical species corresponded to Group 2 of this study, but were caught mainly in the cold, deep waters below the thermocline at RABUTINOS.

Reasonable comparison of different demersal fish communities cannot be made without reference to the region's geographic and oceanographic characteristics. The Southeast Asian continental shelf consists of the Mainland, Sunda, Arafura and Sahul shelves (Morgan and Fryer, 1985). These areas are relatively shallow with maximum depths around the South China Sea ranging only from 40 - 100 m. Depths within the wide and gently sloping Gulf of Thailand are no greater than 80 m. The Arafura Shelf connecting Australia and New Guinea has depths ranging only from 30 m to 90 m while the Sahul Shelf extending along the northwest coast of Australia has depths of only 80 m to 100 m. It is not surprising that most of the demersal fishing activities in Southeast Asia are confined to depths <50 m (Pauly, 1988).

From an oceanographic point of view, this depth ranges basically define the extent of shallow, warm water habitats above the thermocline. Hence, the assemblage species expected from these areas would fall under Group 4A for predominantly shallow, muddy and riverine-influenced regions and Group 4B for similar but deeper areas (>30 m) and clear waters; Group 5 for coastal areas dominated by sandy substrates; Group 7 for shallow, rocky-coraline substrates; and Group 6 for the Group 7/Group 5 transition areas. In regions characterized by a complex array of marine environments, the resulting aggregate species assemblage will depend on the relative areal extent of each habitat/depth category.

These predictions parallel Okera's (1982) recurrent species groups derived from the shallow continental shelf of northern Australia. Okera's "Inner Shelf Assemblage" (muddy sands, 10m - 50 m) is similar to Group 4A of this study. The "Shelf Break Assemblage" (mixed deposits, 12-220 m) compares with Group 2 but not as clear due to the wide depth range included by Okera. The "Midshelf Assemblage" (mixed deposits, 60-110 m) as well as the "Offshore Sand Assemblage" (sand, 80-90 m) compared with Group 3 and Group 4B or Group 5 assemblage minus the leiognathids. The Leiognathidae family is most abundant in the catch of Southeast Asian trawlers but is only a minor component of the fishery in the northern Australia (Pauly, 1988) which explains the relative absence of ponyfishes in Australia's Group 4B counterpart. Finally, Okera's "Hard Bottom Assemblage" (boulders and reefs, various depths) shows similarity to Group 7 assemblage.

The fit between Okera (1982) and this study's community groups were not as tight mainly because Okera used substrate type as the primary classification variable instead of depth. This was unfortunate since depth has been documented as the primary gradient structuring the demersal fish communities in this region (e.g., Rainer and Munro, 1982; Ramm, *et al.*, 1990; Watson *et al.*, 1990; Blaber *et al.*, 1994). As a result, there was an unnecessary agglomeration of species coming from different communities.

The average catch composition of the Gulf of Thailand prior to the squid population explosion (Pauly, 1988) was primarily dominated by Leiognathidae (24%); Carangidae (7%); *Nemipterus* spp., Family: Nemipteridae (6%); Sciaenidae (6%); Mullidae (5%); Rajidae (5%); Gerridae (4%); *Saurida* spp., Family: Synodidae (4%); *Scolopsis* spp., Family: Nemipteridae (3%); and Ariidae (3%). This corresponded well with Group 4A and Group 5 of this study, mixed with a sciaenid assemblage. This assemblage was as expected from the Gulf of Thailand composed primarily of muddy inshore and sandy offshore deposits. The sciaenid assemblage found in shallow (<15 m),

muddy, riverine areas were not included in this study due to the depth limitation of the research vessel and existing fisheries management regulations in the survey areas.

Twenty years later, overfishing has significantly altered the catch composition of the Gulf of Thailand into a predominantly *Loligo* spp., Family: Loliginidae (20%); *Priacanthus* spp., Family: Priacanthidae (9%); *Nemipterus* spp., Family: Nemipteridae (9%); Leionathidae (8%); *Saurida* spp., family: Synodidae (6%); *Sepia* spp., family: Loliginidae (3%); Carangidae (3%); Lutjanidae (2%); Ariidae (2%) and *Scolopsis* spp., family: Nemipteridae (1%) assemblage. These species are typical of the sciaenid assemblage mixed with Group 4B and Group 3 assemblages (excluding the leognathids) found in waters deeper than 50 m. This implies that as one community is decimated by overharvesting, the niche they vacate is immediately taken over by some species from the nearest assemblage capable of utilizing the available resources (Sainsbury, 1988). As mentioned earlier, demersal trawling in Southeast Asia is confined primarily to depths less than 50 m deep. This suggests that commercial fishing pressure in these waters have little influence on many of the species inhabiting the deeper communities (Pauly, 1988).

Within the normal depth range of most Southeast Asian trawl fishery, most of the demersal fish communities of the region were adequately represented in this study. Exceptions to this include the sciaenid community found in waters less than 15 m deep (Longhurst and Pauly, 1987) and the lutjanid community that prefers the rocky-coraline areas 60 - 120 m deep (Okera, 1982; Longhurst and Pauly, 1987; Sainsbury, 1988). Most of the lutjanids in this study were caught in the vicinity of Sorsogon Bay (FS 10 to FS15). The absence of deeper fishing stations in these locations may explain the relative absence of a distinct lutjanid assemblage in this study.

Consistent with the findings of Marten and Polovina (1982) and Pauly (1982), the highest biomass at RABUTINOS were from the shallow coralline fish assemblages (3.7 metric tons/km²) and the lowest (0.8 metric tons/ km²) were from the deep, sandy areas. The shallow, muddy/riverine areas gave a much lower biomass at about 1.1 tons/ km²

while the deeper regions (i.e., >100 m) had a much higher biomass (1.7 tons/ km²). This may be a reflection of the effects of intensive fishing operation within the shallow (<50 m depths) coastal shelves off Viñas River and northern Samar Sea.

In terms of magnitude, the biomass of the deep areas (>150 m) were comparable with the 1.8 tons/km² estimates of Yutuc and Trono (1977) for the Philippine shelves. The shallow, soft-bottom communities of RABUTINOS were much less productive compared to the estimated biomasses at the Gulf of Thailand (3.9 tons/ km²) for depths of 0 - 50 m (SCS, 1978), north coast of Java (2.6 tons/ km²) for 0 - 50 m (SCS, 1979), Sunda Shelf - South (2.3 tons/ km²) for 0 - 50 m (SCS 1978), South China Sea (2.0 tons/ km²) continental shelf (Aoyama, 1973), Sunda Shelf - NW Borneo (1.7 tons/ km²) for 0 - 50 m (SCS, 1978). The limited extent of the shelf area (Marten and Polovina, 1982) and the intensity of fishing activities (Warfel and Manacop, 1950; Pauly, 1988) may explain the depauperate conditions of Ragay Gulf's fish stocks.

C. The Effects of El Niño-Southern Oscillation.

The main impact of the 1982-'83 ENSO event is best grasped by looking at how the normal seasonal pattern has been altered in the region. The normal sequence of the monsoon season at RABUTINOS is as follows:

(WET)NE ⇒ (DRY)INT ⇒ (WET)SW ⇒ (WET)NE ⇒ (DRY)INT ⇒ (WET)SW...
 (Cool) (Warm) (Warm) (Cool) (Warm) (Warm)

During the strong 1982-1983 El Niño-Southern Oscillation event, this normal seasonal sequence was altered as follows:

(WET)NE \Rightarrow (DRY)INT \Rightarrow (WET)SW \Rightarrow (DRY)NE \Rightarrow (DRY)INT \Rightarrow (WET)SW...
 (Cool) (Warm) (Warm) (Warm) (Warm) (Warm)

Main ENSO effects felt here

This pattern resulted in an early warm and dry season starting in November 1982 which basically mimicked the environmental conditions during the intermonsoon period (i.e., similar to spring conditions in the upper latitudes). Since the rains started to pick up only in July 1983 with the passage of a typhoon, the marine environment of RABUTINOS essentially experienced an extended “spring” condition which lasted for seven months instead of just three. The six months delay in the anomalous tradewind reversals associated with the 1982-1983 ENSO event tempered its impact on the fishery of the western Pacific Ocean by shortening the drought period from one full year to just half.

This anomalous condition was believed to be the cause of the extensive *Pyrodinium bahamense* red tide bloom that hit Samar Sea and Carigara Bay (Hermes *et al.*, 1985) in mid-1983. The above conditions may have also tricked some marine organisms to basically speed up their biological clocks. For example, in November and December of 1982, an anomalously high abundance of fish eggs were collected in the vicinity of Viñas River (i.e., located within the areas bounded by Groups 4A and 4B fish assemblages) which rivaled the regular peak egg-laying months of April to May recorded by Villosio *et al.* (1984) and Jamir (1986) for Ragay Gulf.

As mentioned in the introduction section, the ENSO-induced environmental perturbation offered an empirical test on the validity of Longhurst and Pauly's (1987) community stability hypothesis (“LPCS Hypothesis”). The performance of LPCS Hypothesis based on the stability criteria of Krebs' (1978) is as follows:

The constant 1:3 ratio in the number of seasonal entrants *versus* recurrent species appearing in the top 20 most abundant specie/taxa list attests to the relative stability of

this community with respect to species composition. In terms dominance stability, if the LPCS Hypothesis is judged according to the continued dominance of a single species or family group, then it fails to satisfy this criterion since the demersal fish communities of RABUTINOS show distinct seasonal patterns of abundance and shifting species dominance. Taken as a group, however, the ability of the recurring species' membership to maintain the overall species dominance within their group regardless of the season attests to the dominance stability of the system. Also, the fact that most of the recurring group members never ranked lower than 10th place in relative abundance attests to the RABUTINOS community's stability with respect to this criterion.

The performance of the LPCS Hypothesis with respect to the last criteria, i.e., stability of spatial distribution, needs some qualification. There is no question that most of the geographic boundaries between the different demersal fish assemblages readily shift with the season. The changes are even more dramatic in the context of the early maturity phase of the ENSO event. Here, not only were the physical boundaries of the different assemblage groupings radically altered -- even the basic structure of the whole community was modified. The six assemblage groups typical of the previous seasons were reduced to just three assemblage groups by the time ENSO reached its early maturity stage.

While it is possible that the above may just be a sampling artifact, the distinctness of the groupings derived by the DCA ordination and TWINSPAN classification *vis-à-vis* other seasonal groupings indicate otherwise. What was more revealing and interesting from an ecological point of view were the characteristics of the three remaining groups. Group 6 is the transition group between the coralline assemblage (Group 7) and the estuarine/riverine assemblage (Group 5). Group 6 contains a mixture of fishes coming from both groups. Group 4A represents the estuarine/riverine assemblage -- the transition zone between the sea and the rivers. Finally, Group 3 represents the intermediate zone or thermocline assemblage which is the transition zone between the warm upper ocean layers and the deep, cold oceanic zone below. All three groups are located in relatively

less stable or inhospitable environments and may be the reason why their component species persisted despite the environmental perturbations brought about by ENSO.

Theoretically, the more stable the environment, the more species will be present and the more stable is the resulting community (Krebs, 1978). According to this idea, areas with stable environments allow the evolution of finer specializations and adaptations than do areas with erratic, unpredictable environments. Therefore, species are expected to be more flexible in less stable habitat and more specialized in more stable environments. In the face of adverse environmental perturbations, communities coming from less stable habitat may be expected to survive and persist better than those coming from more stable environments. This was clearly demonstrated in Ragay Gulf during the early stages of ENSO.

Some of the hard, nagging questions that need at least partial resolution include the possible mistaken membership of FS8 during the rainy southwest monsoon season, the reasons why Group 4A remained stable despite ENSO drought conditions, and whether Group 1 was a true, seasonal group or just a sampling aberration (Table 12). Spikes in rainfall (see Figure 4) are common at RABUTINOS due to its location along the "typhoon belt." Typhoons or hurricanes passing right through RABUTINOS could easily dump several tens of inches of rain in just a few days (Hardjawinata, 1980) within the survey area. The semi-enclosed nature of the Ragay Gulf Basin makes it susceptible to extensive freshwater dilution from various point and non-point sources (Jamir, 1986) similar to the Gulf of Carpentaria (Longhurst and Pauly, 1987) or Bay of Bengal (Ansari et al., 1995). This may be the reason for the observed change in the group membership of FS 8 from Group 3 to Group 4B conditions.

Longhurst and Pauly (1987) have recorded carangids, scombrids and nemipterids as part of the regular catches coming from the deep tropical regions. These species are also known for their long migrations and seasonal occurrences in different fishing and spawning grounds (Jones and Rosa, 1965; Tiews *et al.*, 1970a; Ronquillo, 1975). In the

Philippines, for example, *Decapterus muruadsi*, family: Carangidae, are known to spawn and dominate the catch in Palawan during the months of December - March before migrating outside of Manila Bay two months later (Chullasorn and Martosubroto, 1986). The adults of the Scombridae family (e.g., the oceanic form, *Rastrelliger kanagurta*) are caught inside Ragay Gulf during the rainy months of May - December (Jones and Rosa, 1965; Chullasorn and Martosubroto, 1986). These months also correspond to one of their intense feeding periods (Manacop, 1961) associated with rapid gonadal development (Jones and Rosa, 1965) and the November - December spawning period inside the Ragay Gulf Basin (Mines, 1984). Therefore, it is highly likely that Group 1 assemblage is a regular, seasonal member of the Ragay Gulf demersal fish community and that the dry ENSO conditions may have aborted their November - December, 1982 spawning migration, much like the disappearance of *Sardinella longiceps* along the coast of India following an ENSO-induced recruitment failure (Raja, 1972, 1973; Longhurst and Pauly, 1987).

The most intense effects of nutrient runoff under open marine conditions have involved semi-enclosed systems (Caddy and Bakun, 1995), similar to the Ragay Gulf Basin. Concentration of floating debris, larval stages and food resources results from the convergence processes at the boundary of the river plume (Pauly, 1982; Pickard and Emery, 1982). This explains the year-round, high abundance of fish eggs and larvae in the vicinity of Viñas River (Villoso *et al.*, 1984; Jamir, 1986) and the corresponding accumulation of coastal pelagic fishes and associated predators which basically comprise the Group 4A assemblage. The spring-like conditions and high productivity associated with the dry intermonsoon period and also during the early maturity stages of ENSO explain the prevalence of Group 4A during the rest of the year.

As Gunter (1961) pointed out, most marine and estuarine organisms can withstand full sea water but some marine fishes cannot tolerate lowered salinities. The estuarization of the coastal areas explains the expansion of Group 4A habitats further offshore during the rainy southwest and northeast monsoon seasons as low salinity intolerant species

migrate offshore. The relatively warm and dry ENSO conditions may have deterred some species from the deep, cold waters from venturing above the thermocline. Despite the shrinking low salinity lenses, the high productivity brought about by ENSO-induced "spring transition" conditions near major river systems act as oases that draw planktivores like leiognathids and coastal pelagics, as well as their associated predator assemblage, i.e., Group 4A, to agglomerate in the area. This may be the mechanism that enabled Group A assemblages to remain geographically stable or even expand their boundaries offshore during ENSO.

D. Implications for Fisheries Management and Research.

This study further confirms the existence of relatively stable demersal fish communities in such diverse habitats as Ragay Gulf, Burias Pass, Ticao Pass and northern Samar Sea. The primary separation of each demersal fish communities by depth (i.e., <35 m shallow/riverine assemblage, 35 - 90 m shallow/further offshore assemblage, 90 - 100 m thermocline assemblage, and >150 m cold, deep water assemblage) and further subdivisions according to substrate types have lots of implications for better fisheries management policies and practices in the Philippines and the Southeast Asian region.

Most of the commercial demersal trawl operations in the region are confined to waters less than 50 m deep. In the Philippines, the national government banned commercial trawling and purse seining operations in waters less than 7 fathoms (13 m) deep and/or within 3 nautical miles from the coast (see Presidential Decree No. 704, Republic of the Philippines, 1975). Based on the findings of this research as well as by McManus (1985) and Federizon (1992), legislations aimed at reducing the impact of the growing commercial trawl fisheries on the municipal or sustenance fisheries by a physical separation should use 35 m (or 15 fathoms) as the dividing line rather than the current 7 fathom limit (Pauly, 1988).

Adoption of a deeper depth limit will practically close most of the country's overexploited traditional fishing grounds from commercial trawl operations. These include the shallow Lingayen Gulf, Manila Bay, San Miguel Bay, Samar Sea, Maqueda Bay, Villareal Bay, Visayan Sea and Panguil Bay (Smith et al., 1980; Ganaden, 1990). While this may be good for the fishes and the municipal fishery sector, the economic displacement that this will entail may not warrant the political risks involved unless alternate fishing grounds are explored and developed ahead of time. Again, the results of this research can be used to classify, map and project the catch composition and biomass potentials of new fishing grounds with more precision and less cost than the techniques currently available, e.g., Kvrn (1971); Aoyama (1973); Menasveta *et al.* (1973); Marten and Polovina (1982); Silvestre (1986); Silvestre and Pauly (1986); and Silvestre *et al.* (1986).

The ENSO period covered in this study is limited to just the early maturity stages and drought conditions, however, a number of insights can be gained for more targeted studies in the future. The following are some recommendations on how to conduct this monitoring program. First, a re-examination of the present status of the fishery needs to be done to determine if significant alterations in the fish communities were evident. This will also serve as a benchmark for judging the short and long-term impacts of ENSO on the demersal fisheries of the area.

Second, knowing the location of fishing grounds that show signs of seasonal and ENSO related fish assemblage adjustments, the number of stations can be reduced by half and limited to just the Ragay Gulf Basin (close to Viñas River) and Ticao Pass. Given the progress in the early detection of an ENSO event, adequate preparation time is available between the commencement of an ENSO event, i.e., Fall (year -1), the time its impacts are felt throughout the Philippines, i.e., northeast monsoon (year 0), and the ocean's return to normal conditions sometime between February to June (year +1). Instead of monthly cruises, representative seasonal samples would significantly cut the cost of research and justify extension over a longer period of time, possibly at least one to two

years after an ENSO event. The monitoring program should include plankton studies, physical oceanography and hydrology besides demersal trawl sampling.

REFERENCES

- Allan, R. 1988. El Niño-Southern Oscillation influences in the Australasian region. *Progress in Physical Geography*, 12:4-40.
- _____. 1991. Australasia. In *Teleconnections: linking worldwide climate anomalies*, eds. M. Glantz, R. Katz, and N. Nicholls, 13-42. Cambridge: Cambridge University Press.
- Ansari, Z., A. Chatterji, B. Ingole, R. Sreepada, C. Rivonkar, and A. Parulekar. 1995. Community structure and seasonal variation of an inshore demersal fish community at Goa, West Coast of India. *Estuarine, Coastal and Shelf Science* 41:593-610.
- Aoyama, T. 1973. The demersal fish stocks and fisheries of the South China Sea. SCS/DEV/73/3. Rome. UN Food and Agricultural Organization/UN Development Programme.
- Aprieto, V. 1978. Hydrobiological studies of the Visayan Sea demersal fisheries. A report on trawl fishing investigations of traditional and non-traditional fishing grounds in the Philippines. *Technical Report*. Quezon City: Institute of Fisheries Development and Research, University of the Philippines. (mimeographed).
- Barnett, T. 1991. The interaction of multiple time scales in the tropical climate system. *Journal of Climate* 4:269-285.
- Beals, E. 1984. Bray-Curtis ordination: an effective strategy for analysis of multivariate ecological data. *Advances in Ecological Research* 14:1-55.
- Belbin, L. 1991. *PATN technical reference*. Canberra: CSIRO Division of Wildlife and Ecology.
- Bialek, E. 1966. *Handbook of Oceanographic Tables*. Special Publications, US Navy Hydrographic Office. Washington, D.C.: US Government Printing Office.
- Bianchi, G. 1991. Demersal assemblages of the continental shelf and slope edge between the Gulf of Tehuantepec (Mexico) and the Gulf of Papagayo (Costa Rica). *Marine Ecology Progress Series* 73:121-140.
- Binet, D., L. Reste, and P. Diouf. 1995. The influence of runoff and fluvial outflow on the ecosystems and living resources of west African coastal waters. In *Effects of riverine inputs on coastal ecosystems and fisheries resources*, 89-118. FAO Fisheries Technical Paper No. 349. Rome: UN Food and Agricultural Organization.

- Blaber, S., J. Young, and M. Dunning. 1985. Community structure and zoogeographic affinities of the coastal fishes of the Dampier region of north-western Australia. *Australian Journal of Marine and Freshwater Research* 36:247-66.
- Blaber, S., D. Brewer, and J. Salini. 1989. Species composition and biomasses of fishes in different habitats of a tropical northern Australian estuary: their occurrence in the adjoining sea and estuarine dependence. *Estuarine, Coastal and Shelf Science* 29:509-531.
- Blaber, S., D. Brewer, and A. Harris. 1994. Distribution, biomass and community structure of demersal fishes of the Gulf of Carpentaria, Australia. *Australian Journal of Marine and Freshwater Research* 45:375-96.
- Blaber, S., D. Brewer, and J. Salini. 1995. Fish communities and the nursery role of the shallow inshore waters of a tropical bay in the Gulf of Carpentaria, Australia. *Estuarine, Coastal and Shelf Science* 40:177-193.
- Brander, K. 1988. Multispecies fisheries of the Irish Sea. In *Fish population dynamics* (2 ed.), ed. J. Gulland, 303-328. New York: John Wiley and Sons.
- Caddy, J. and A. Bakun. 1995. Marine catchment basins and anthropogenic effects on coastal fishery ecosystems. In *Effects of riverine inputs on coastal ecosystems and fisheries resources*, 119-133. FAO Fisheries Technical Paper No. 349. Rome: UN Food and Agricultural Organization.
- Caddy, J. and G. Sharp. 1986. An ecological framework for marine fishery investigations, 1-152. FAO Fisheries Technical Paper No. 283. Rome: UN Food and Agricultural Organization.
- Cane, M. 1983. Oceanographic events during El Niño. *Science*, 222:1189-95.
- Chan, E. and H. Liew. 1986. Characteristics of an exploited tropical shallow-water demersal fish community in Malaysia. In *The first Asian fisheries forum. Proceedings of the first Asian fisheries forum*, eds. J. MacLean, L. Dizon and L. Hosillos, 349-352. Manila: Asian Fisheries Society.
- Chessman, B. and D. Robinson. 1987. Some effects of the 1982-83 drought on water quality and macroinvertebrate fauna in the Lower LaTrobe River, Victoria. *Australian Journal of Marine and Freshwater Research* 38:289-99.
- Chong, V., A. Sasekumar, M. Leh, and R. D'Cruz. 1990. The fish and prawn communities of a Malaysian coastal mangrove system with comparisons to adjacent mud flats and inshore waters. *Estuarine, Coastal and Shelf Science* 31:703-722.

- Chullasorn, S. and P. Martosubroto. 1986. *Distribution and important biological features of coastal fish resources in Southeast Asia*. FAO Fisheries Technical Report No. 278. Rome: UN Food and Agricultural Organization.
- Cody, R. and J. Smith. 1991. *Applied statistics and the SAS programming language*, 3ed. New Jersey: Prentice-Hall.
- Collette, B. and C. Nauen. 1983. *Species catalogue, vol. 2, scombrids of the world: an annotated and illustrated catalogue of tunas, mackerels, bonitos and related species known to date*. FAO Fisheries Synopsis No. 125. Rome: UN Food and Agricultural Organization.
- Connoly, R. 1994. A comparison of fish assemblages from seagrass and unvegetated areas of a southern Australian estuary. *Australian Journal of Marine and Freshwater Research* 45:1033-44.
- Corpuz, A., J. Saeger and V. Sambilay Jr. 1985. Population parameters of commercially important fishes in Philippine waters. *Technical Report No. 6*. Quezon City: Department of Marine Fisheries, University of the Philippines.
- Courtney, A., J. Masel, and D. Die. 1995. Temporal and spatial patterns in recruitment of three penaeid prawns in Moreton Bay, Queensland, Australia. *Estuarine, Coastal and Shelf Science* 41:377-392.
- Crivelli, A., M. Ximenes, G. Gout, G. Lassere, P. Freon and T. Do Chi. 1995. Causes and effects of terrestrial runoff and riverine outflow on brackish/coastal marine fisheries ecosystems in the northern Mediterranean region. In *Effects of riverine inputs on coastal ecosystems and fisheries resources*, 59-88. Rome: FAO Fisheries Technical Paper 349.
- Crowder, L. and J. Magnuson. 1983. Cost-benefit analysis of temperature and food resource use: a synthesis with examples from the fishes. In *Behavioral energetics*, eds. W. Aspey and S. Lustick, 189-221. Ohio: Ohio State University Press.
- Cunningham, K. and J. Ogilvie. 1972. Evaluation of hierarchical grouping techniques: a preliminary study. *Computer Journal* 15:209-213.
- Cushing, D. 1982. *Climate and Fisheries*. London: Academic Press.
- Cyrus, D. and S. Blaber. 1992. Turbidity and salinity in a tropical northern Australian estuary and their influence on fish distribution. *Estuarine, Coastal and Shelf Science* 35:545-563.

- Day, J., J. Field and M. Montgomery. 1971. The use of numerical methods to determine the distribution of the benthic fauna across the continental shelf of North Carolina. *Journal of Animal Ecology* 40:93-126.
- Day, R. and G. Quinn. 1989. Comparison of treatments after an analysis of variance in ecology. *Ecological Monographs* 59:433-63.
- Dennis, G. and T. Bright. 1988. Reef fish assemblages on hard banks in the northwestern Gulf of Mexico. *Bulletin of Marine Science* 43(2):280-307.
- Diaz, H. and V. Markgraf. 1992. Introduction. In *El Niño: Historical and Paleoclimatic Aspects of the Southern Oscillation*, eds., H. Diaz and V. Markgraf, 1-4. Cambridge: Cambridge University Press.
- Diaz, H. and G. Kiladis. 1992. Atmospheric teleconnections associated with the extreme phases of the Southern Oscillation. In *El Niño: Historical and Paleoclimatic Aspects of the Southern Oscillation*, eds., H. Diaz and V. Markgraf, 7-28. Cambridge: Cambridge University Press.
- Digby, P. and R. Kempton. 1987. *Multivariate analysis of ecological communities*. New York: Chapman and Hall.
- Enfield, D. 1992. Historical and prehistorical overview of El Niño/Southern Oscillation. In *El Niño: Historical and Paleoclimatic Aspects of the Southern Oscillation*, eds., H. Diaz and V. Markgraf, 95-117. Cambridge: Cambridge University Press.
- Fager, E. and A. Longhurst. 1968. Recurrent group analysis of species assemblages of demersal fish in the Gulf of Guinea. *Journal of the Fisheries Research Board of Canada* 25:1405-1421.
- FAO. 1974. *FAO species identification sheets for fishery purposes - eastern Indian Ocean (fishing area 57) and western central Pacific (fishing area 71)*, vol. I - IV. Rome: UN Food and Agricultural Organization.
- Federizon, R. 1992. Description of the subareas of Ragay Gulf, Philippines, and their fish assemblages by exploratory data analysis. *Australian Journal of Marine and Freshwater Research* 43:379-91.
- Ferrel, D., S. McNeill, D. Worthington, and J. Bell. 1993. Temporal and spatial variation in the abundance of fish associated with the seagrass *Posidonia australis* in south-eastern Australia. *Australian Journal of Marine and Freshwater Research* 44:881-98.

- Field, J. 1971. A numerical analysis of changes in the soft-bottom fauna along a transect across False Bay, South Africa. *Journal of Experimental Marine Biology and Ecology* 7:215-253.
- Field, J., K. Clarke, and R. Warwick. 1982. A practical strategy for analysing multispecies distribution patterns. *Marine Ecology Progress Series* 8:37-52.
- Fishelson, L. 1980. Partitioning and sharing of space and food resources by fishes. In *Fish behavior and its use in the culture and capture of fishes*, eds. J. Bardach, J. Magnuson, R. May, and J. Reinhart, 415-445. Manila: ICLARM Conference Proceedings 5, International Center for Living Aquatic Resources Management.
- Ganaden, S. 1992. Multi-species demersal resources and multi-gear fisheries in the Philippines. *FAO Fisheries Report* 463. Rome: UN Food and Agricultural Organization.
- Gauch, H. 1982. *Multivariate analysis in community ecology*. London: Cambridge University Press.
- Gjosaeter, J., and K. Kawaguchi. 1980. *A review of the world resources of mesopelagic fish*. FAO Technical Paper No. 193. UN Food and Agricultural Organization.
- Glantz, M. 1991. Introduction. In *Teleconnections: linking worldwide climate anomalies*, eds. M. Glantz, R. Katz, and N. Nicholls, 13-42. Cambridge: Cambridge University Press.
- Glynn, P. (ed.) 1990. *Global Ecological Consequences of the 1982-1983 El Niño-Southern Oscillation*. New York: Elsevier .
- Graham, H. 1952. A contribution to the oceanography of the Sulu Sea. *Proceedings of the Seventh Pacific Science Congress* 3:1-42.
- _____. 1953. The topography of the sea surface in the region of the Philippines. *Proceedings of the Eight Pacific Science Congress* 673-678.
- Grashoff, J. (ed.). 1977. *Methods of seawater analysis*. Weinheim: Verlag Chemie.
- Grieg-Smith, P. 1983. *Quantitative Plant Ecology* (3 ed.). Los Angeles: University of California Press.
- Gunter, G. 1961. Some relations of estuarine organisms to salinity. *Limnology and Oceanography* 6:182-90.
- Halim, Y., S. Morcos, S. Rizkalla and M. El-Sayed. 1995. The impact of the Nile and the Suez canal on the living marine resources of the Egyptian Mediterranean waters

- (1958-1986). In *Effects of riverine inputs on coastal ecosystems and fisheries resources*, 19-57. Rome: FAO Fisheries Technical Paper 349.
- Hardenberg, J. 1955. A review of current knowledge of *Rastrelliger*. Paper presented to IPFC 6th Session, Tokyo: IPFC/C55/WK25):1-10.
- Hardjawanata, S. 1980. Macroclimatic aspects of rice production in Southeast Asia. In *Proceedings of the symposium on the agrometeorology of the rice crop*, 57-68. Los Baños, Laguna, Philippines: World Meteorological Organization and The International Rice Research Institute.
- Harris, A. and I. Poiner. 1990. By-catch of the prawn fishery of Torres Strait: composition and partitioning of the discards into components that float or sink. *Australian Journal of Marine and Freshwater Research* 41:37-52.
- Hayek, L. 1994. Analysis of amphibian biodiversity data. In *Measuring and monitoring biological diversity: Standard methods for amphibians*, eds. R. Heyer, M. Donnelly, R. McDiarmid, L. Hayek, and M. Foster, 207-269. Washington, D.C.: Smithsonian Institution Press.
- Helvey, M. and R. Smith. 1985. Influence of habitat structure on the fish assemblages associated with two cooling-water intake structures in southern California. *Bulletin of Marine Science* 37(1):189-199.
- Hermes, R., T. V. Jamir and E. Villosio. 1985. Spatial distribution of *Pyrodinium bahamense* var. *compressa* in the Samar Sea and associated oceanographic parameters. *UPV Fisheries Journal* 1(1):1-12.
- Hill, M. 1973. Reciprocal averaging: an eigenvector method of ordination. *Journal of Ecology* 61:237-249.
- _____. 1979a. *DECORANA -- A FORTRAN Program for Detrended Correspondence Analysis and Reciprocal Averaging*. New York: Cornell University Press.
- _____. 1979b. *TWINSPAN -- A FORTRAN Program for Arranging Multivariate Data in an Ordered Two-Way Table by Classification of the Individuals and Attributes*. New York: Cornell University Press.
- Hill, M. and H. Gauch. 1980. Detrended correspondence analysis: an improved ordination technique. *Vegetatio* 42:47-58.
- Hoese, D. 1960. Biotic changes in a bay associated with the end of a drought. *Limnology and Oceanography* 5:326-36.

- Hutchinson, G. 1957. Concluding remarks. *Cold Spring Harbor Symposium on Quantitative Biology* 22: 415-427.
- Ingles, J. and D. Pauly. 1984. An atlas of the growth, mortality and recruitment of Philippine fishes. *ICLARM Technical Report No. 13*. Manila: International Center for Living Aquatic Resource Management.
- Isdale, P. 1984. Fluorescent bands in massive corals record centuries of coastal rainfall. *Nature* 310:578-9.
- Isdale, P. and V. Kotwicki. 1987. Lake Eyre and the Great Barrier Reef: A paleohydrological ENSO connection. *South Australian Geographical Journal* 87:48-55.
- Jacinto, G. 1983. Flow characteristics and distribution of nutrients, temperature and salinity in Calatagan lagoon, Batangas. *Unpublished M.S. Thesis*. Quezon City: University of the Philippines.
- Jain, A. and R. Dubes. 1988. *Algorithms for clustering data*. New Jersey: Prentice-Hall.
- Jamir, T.V. 1986. The oceanographic condition of Ragay Gulf and Burias Pass, Philippines and its relationship with the fisheries of the area. *Unpublished M.S. Thesis*. Quezon City: University of the Philippines.
- Jamir, T.V., A. Huyer, W. Pearcy and J. Fisher. 1994. The influence of environmental factors on the marine survival of Oregon hatchery coho (*Oncorhynchus kisutch*). In *Salmon ecosystem restoration: myth and reality*, ed. M. Keefe, 115-138. Eugene, Oregon: American Fisheries Society.
- Johannes, R. 1980. Using knowledge of the reproductive behavior of reef and lagoon fishes to improve fishing yield. In *Fish behavior and its use in the culture and capture of fishes*, eds. J. Bardach, J. Magnuson, R. May, and J. Reinhart, 247-270. Manila: ICLARM Conference Proceedings 5, International Center for Living Aquatic Resources Management.
- Jones, G. 1985. Revision of the Australian species of the fish family Leiognathidae. *Australian Journal of Marine and Freshwater Research* 36:559-613.
- Jones, G., J. Baker, K. Edyvane, and G. Wright. 1996. Nearshore fish community of the Port River-Barker inlet estuary, South Australia. I. Effect of thermal effluent on the fish community structure, and distribution and growth of economically important fish species. *Marine and Freshwater Research* 47:785-99.
- Jones, S. and H. Rosa Jr. 1965. *Synopsis of biological data on Indian mackerel Rastrelliger kanagurta (Cuvier) 1817 and short bodied mackerel Rastrelliger*

- brachysoma* (Bleeker) 1851. FAO Fisheries Synopsis No. 29. Rome: UN Food and Agricultural Organization.
- Kerr, S. 1980. Niche theory in fisheries ecology. *Transactions of the American Fisheries Society* 109:254-260.
- Krebs, C. 1978. *Ecology: the experimental analysis of distribution and abundance* (2ed.). New York: Harper and Row Publishers.
- _____. 1989. *Ecological Methodology*. New York: Harper-Collins Publishers.
- Kumagai, S. and T. Bagarinao. 1979. Result of drift card experiments and considerations on the movement of milkfish eggs and larvae in the Northern Sulu Sea. *Fisheries Research Journal of the Philippines* 4:64-81.
- Kvran, E. 1971. Marine fisheries potential in the Philippines and Southeast Asia. *Fisheries newsletter* July - December, 8-17. Manila: Philippine Fisheries Commission.
- Labao, E. 1980. Oceanographic survey of Samar Sea. *Technical Report*. Quezon City: Department of Marine Fisheries, University of the Philippines. (mimeographed).
- La Fond, E. 1951. *Processing Oceanographic Data. H.O. Publication 614*. Washington, D.C.: US Navy Hydrographic Office.
- Lasiak, T. 1984. Structural aspects of the surf-zone fish assemblage at King's Beach, Algoa Bay, South Africa: Long-term fluctuations. *Estuarine, Coastal and Shelf Science* 18:459-483.
- Lau, K. and P. Sheu. 1991. Teleconnections in global rainfall anomalies: Seasonal to inter-decadal time scales. In *Teleconnections: linking worldwide climate anomalies*, eds. M. Glantz, R. Katz, and N. Nicholls, 13-42. Cambridge: Cambridge University Press.
- Legasto, R. C. del Mundo and K. Carpenter. 1975. On the hydro-biological and socio-economic surveys of San Miguel Bay for the proposed fish nurseries/reservation. *Philippine Journal of Fisheries* 13:205-246.
- Limpus, C. and N. Nicholls. 1988. The Southern Oscillation regulates the annual numbers of green turtles (*Chelonia mydas*) breeding around northern Australia. *Australian Journal of Wildlife Research* 15:157-61.
- Long, B. and I. Poinet. 1994. Infaunal benthic community structure and function in the Gulf of Carpentaria, northern Australia. *Australian Journal of Marine and Freshwater Research* 45:293-316.

- Longhurst, A. and D. Pauly. 1987. *Ecology of tropical oceans*. New York: Academic Press.
- Lopez, R. 1964. "Problemas de la distribucion geografica de los peces marinos suramericanos," pp. 57-62. Bolivian Institute of Marine Biology, Argentina.
- Love, G. 1987. Banana prawns and the Southern Oscillation Index. *Australian Meteorological Magazine* 35:47-9.
- Ludwig, J. and J. Reynolds. 1988. *Statistical Ecology*. New York: John Wiley and Sons.
- Magnuson, J. 1991. Fish and fisheries ecology. *Ecological Applications* 1(1):13-26.
- Magnuson, J., L. Crowder and P. Medvick. 1979. Temperature as an ecological resource. *American Zoologist* 19:331-343.
- Manacop, P. 1955a. A preliminary systematic study of the Philippine chub mackerels, family Scombridae, genera *Pneumatophorus* and *Rastrelliger*. Paper presented to IPFC 6th session. Tokyo: IPFC/C55/Tech.50 (Sea): 1-18.
- Manacop P. 1955b. Commercial trawling in the Philippines. *Philippine Journal of Fisheries* 3:117-188.
- Marchant, R., L. Barmuta, and B. Chessman. 1994. Preliminary study of the ordination and classification of macroinvertebrate communities from running waters in Victoria, Australia. *Australian Journal of Marine and Freshwater Research* 45:945-62.
- Marten, G. and J. Polovina. 1982. A comparative study of fish yields from various tropical ecosystems. In: *Theory and management of tropical fisheries*, eds. D. Pauly and G. Murphy, 255-289. Manila: ICLARM Conference Proceedings, 9. International Center for Living Aquatic Resource Management, Manila, Philippines and Commonwealth Scientific and Industrial Research Organization, Cronulla, Australia.
- Masuda, K., S. Nakane, S. Saito and T. Fujii. 1964. Survey of trawl grounds off the north-west coast of Australia with species reference to hydrographic conditions of the ground. *Bulletin of the Faculty of Fisheries of Hokkaido University* 15(2):77-88.
- May, R. (ed.). 1984. *Exploitation of Marine Communities*, Dahlem Konferenzen. Berlin: Springer-Verlag.
- McCune, B. 1988. Ecological diversity in north American pines. *American Journal of Botany* 75(3):353-368.

- McCune, B. and M. Mefford. 1995. *PC-ORD. Multivariate analysis of ecological data, version 2.0*. Oregon: MjM Software Design.
- McGregor, G. 1989. An assessment of the annual variability of rainfall: Port Moresby, Papua New Guinea. *Singapore Journal of Tropical Geography*, 10(1):43-54.
- McManus, J. 1985. Descriptive community dynamics: background and an application to tropical fisheries management. *Unpublished Ph.D. Dissertation*. Kingston: University of Rhode Island.
- McManus, J., C. Nañola, A. del Norte, R. Reyes Jr., J. Pasamonte, N. Armada, E. Gomez and P. Aliño. 1996. Coral reef fishery sampling methods. In: *Stock Assessment: Quantitative Methods and Applications for Small-Scale Fisheries*, eds. V. Galluci, S. Saila, D. Gustafson, and B. Rothschild, 226-270. New York: CRC Press.
- Megia, T. 1953. A contribution to the oceanography of Manila Bay. *Proceedings of the Eight Pacific Science Congress* 18-64.
- Megia, T. and A. Sebastian. 1951. The thermal structure of the surface waters off western Philippines based on BT observations. *Philippine Journal of Fisheries* 1(2):131-143.
- Megia, T. and D. Villadolid. 1953. A contribution to the knowledge of the structure and composition of the water masses off Eastern Philippines. *Philippine Journal of Fisheries* 2(1):59-67.
- Menasveta, D., S. Shindo and S. Chullasorn. 1973. *Pelagic fishery resources of the South China Sea and prospects for their development*. SCS/DEV/73/6. Rome: UN Food and Agricultural Organization.
- Metzeling, L. 1993. Benthic macroinvertebrate community structure in streams of different salinities. *Australian Journal of Marine and Freshwater Research* 44:335-51.
- Milligan, G. and P. Isaac. 1980. The validation of four ultrametric clustering algorithms. *Pattern Recognition* 12:41-51.
- Mines, A. (ed.). 1984. Assessment of trawlable areas in the Philippines II: Ragay Gulf, Burias Pass, Ticao Pass and waters north of Samar Sea. *Technical Report No. 5*. Quezon City: Department of Marine Fisheries, University of the Philippines.
- Miller, R. and R. Death. 1997. Seasonal and spatial dynamics in the phytomacrofaunal community of Lake Henley, New Zealand. *New Zealand Journal of Marine and Freshwater Research* 31:423-434.

- Morgan, J. 1964. *A preliminary fishery survey of bottom fishing on the north Kenya Banks, No. 21*. London: Colonial Office Fisheries Publication.
- Morgan, J. and D. Fryer. 1985. The marine geography of Southeast Asia. In *Marine Policy in Southeast Asia*, eds. G. Kent and M. Valencia, 9-32. Los Angeles: University of California Press.
- Morin, B., C. Hudon, and F. Whoriskey. 1992. Environmental influences on seasonal distribution of coastal and estuarine fish assemblages at Wemindji, eastern James Bay. *Environmental Biology of Fishes* 35:219-229.
- Nair, R. and R. Subrahmanyam. 1955. The diatom, *Fragillaria oceanica*, an indicator of abundance of the Indian Oil sardine. *Current Science* 24(2):41-42.
- Newman, S., D. Williams, and G. Russ. 1997. Patterns of zonation of assemblages of Lutjanidae, Lethrinidae and Serranidae (Epinephelinae) within and among mid-shelf and outer-shelf reefs in the central Great Barrier Reef. *Marine and Freshwater Research* 48:119-28.
- Nicholls, N. 1988. El Niño-Southern Oscillation impact prediction. *Bulletin of the American Meteorological Society* 69:173-6.
- _____. 1992. Historical El Niño-Southern Oscillation variability in the Australasian region. In: *El Niño: Historical and Paleoclimatic Aspects of the Southern Oscillation*, eds., H. Diaz and V. Markgraf, 151-173. Cambridge: Cambridge University Press.
- Ohman, M., A. Rajasuriya, and E. Olafsson. 1997. Reef fish assemblages in north-western Sri Lanka: distribution patterns and influences of fishing practices. *Environmental Biology of Fishes* 49:45-61.
- Okera, W. 1982. Organization of Fish Assemblages on the Northern Australian Continental Shelf. Cronula: CSIRO (unpublished).
- Ommaney, F. 1961. "Malayan Offshore Trawling Grounds -- The Experimental and Exploratory Fishing of the FRV Manahine in Malayan and Bornean Waters, 1955-56," No. 18, 1-95. London: Colonial Office Fisheries Publication.
- Ordoñez, J., F. Arce, R. Ganaden, and N. Metrillo Jr. 1976. On the hydrobiological and fisheries survey of Sorsogon Bay, Luzon Island. *Philippine Journal of Fisheries* 14(1):178-203.

- Pauly, D. 1979. *Theory and management of tropical multispecies stocks: a review with emphasis on the southeast Asian demersal fisheries*. Manila: ICLARM Studies and Reviews 1, International Center for Living Aquatic Resource Management.
- _____. 1980. *A Selection of Simple Methods for the Assessment of Tropical Fish Stocks*. Rome: UN Food and Agricultural Organization Fisheries Circular No. 729. FIRM/c 729.
- _____. 1983. *Some simple methods for the assessment of tropical fish stocks*. FAO Fisheries Technical Paper No. 234. Rome: UN Food and Agricultural Organization.
- _____. 1988. Fisheries research and the demersal fisheries of Southeast Asia. In *Fish population dynamics (2ed.)*, ed. J. Gulland, 329-348. New York: John Wiley and Sons.
- Pauly, D. and A. Mines (eds.). 1982. Small-scale fisheries of San Miguel Bay, Philippines: biology and stock assessment. *ICLARM Technical Report No. 7*. Manila: International Center for Living Aquatic Resources Management.
- Pauly, D. and G. Murphy. 1982. *Theory and Management of Tropical Fisheries*. Manila: ICLARM. International Center for Aquatic Resource Management.
- Pauly, D. and N. Navaluna. 1983. Monsoon-induced seasonality in the recruitment of Philippine fishes. In *Proceedings of the Expert Consultation of Neritic Fish Resources*, eds. G. Sharp and J. Csirke, 834-843. Costa Rica: FAO Fisheries Report No. 291(3).
- Peterson, M. and S. Ross. 1991. Dynamics of littoral fishes and decapods along a coastal river-estuarine gradient. *Estuarine, Coastal and Shelf Science* 33:467-483.
- Philippi, T. 1993. Multiple regression: herbivory. In *Design and Analysis of Ecological Experiments*, eds. S. Scheiner and J. Gurevich, 183-210. New York: Chapman and Hall.
- Philippines, Republic. Office of the President. 1975. Presidential decree no. 704. Fishery decree of 1975 - revising and consolidating all laws affecting fishing and fisheries. Malacañang, Manila.
- Pickard, G. and W. Emery. 1982. *Descriptive physical oceanography (4ed)*. New York: Pergamon Press.
- Polonsky, A. 1994. Comparative study of the Pacific ENSO event of 1991-92 and the Atlantic ENSO-like event of 1991. *Australian Journal of Marine and Freshwater Research* 45:705-25.

- Ponton, D. and G. Copp. 1997. Early dry-season community structure and habitat use of young fish in tributaries of the river Sinnamary (French Guiana, South America) before and after hydrodam operation. *Environmental Biology of Fishes* 50:235-256.
- Prochazka, K. 1998. Spatial and trophic partitioning in cryptic fish communities of shallow subtidal reefs in False Bay, South Africa. *Environmental Biology of Fishes* 51:201-220.
- Pusey, B. and D. Edward. 1990. Structure of fish assemblages in waters of the Southern Acid Flats, South-western Australia. *Australian Journal of Marine and Freshwater Research* 41:721-34.
- Quinn, W., D. Zopf, K. Short, and R. Kuo Yang. 1978. Historical trends and statistics of the Southern Oscillation, El Niño and Indonesian droughts. *Fishery Bulletin*, 76:663-78.
- Quinn, N. 1980. Analysis of temporal changes in fish assemblages in Serpentine Creek, Queensland. *Environmental Biology of Fishes* 5(2):117-133.
- Rainer, S. 1984. Temporal changes in a demersal fish and cephalopod communities of an unexploited coastal environment in northern Australia. *Australian Journal of Marine and Freshwater Research* 33:1039-1055.
- Raja, B. 1972. A forecast for the ensuing oil-sardine fishery. *Seafood Export Journal* 4(10):27-33.
- _____. 1973. Possible explanation for the fluctuations in abundance of the Indian oil-sardine. *Proceedings of the Indo-Pacific Fish Commission* 15:241-252.
- Ramm, D., P. Pender, R. Willing, and R. Buckworth. 1990. Large-scale spatial patterns of abundance within the assemblage of fish caught by prawn trawlers in Northern Australian waters. *Australian Journal of Marine and Freshwater Research* 41:79-95.
- Rainer, S. and I. Munro. 1982. Demersal fish and cephalopod communities of an unexploited coastal environment in northern Australia. *Australian Journal of Marine and Freshwater Research* 33:1039-55.
- Rasmusson, E. and J. Wallace. 1983. Meteorological aspects of the El Niño/Southern Oscillation. *Science* 222:1195-1202.
- Rau, N. and A. Rau. 1980. *Commercial marine fishes of the central Philippines (bony fishes)*. Eschborn, Germany: German Agency for Technical Cooperation.

- Ray, A. (ed.) 1982. *SAS user's guide: Statistics*. Cary, North Carolina: SAS Institute.
- Richards, A. 1955. Trawling in the Southeast Caribbean. *A Report Prepared for the Government of Trinidad and Tobago and the Caribbean Commission*. Port-of-Spain, Trinidad: Caribbean Commission.
- Robertson, A. and N. Duke. 1990. Recruitment, growth and residence time of fishes in a tropical Australian mangrove system. *Estuarine, Coastal and Shelf Science* 31:723-743.
- Rochford, D. 1991. 'Upwelling': Does it need a stricter definition? *Australian Journal of Marine and Freshwater Research* 42:45-6.
- Roel, B. 1987. Demersal communities off the west coast of South Africa. In *The Benguela and Comparable Ecosystems*, 575-84, eds. A. Payne, J. Gulland and K. Brink. Cape Town: Sea Fisheries Institute.
- Ronquillo, I. 1975. A review of the roundscad fishery in the Philippines. *Philippine Journal of Fisheries* 3(1 & 2):86-126.
- Ronquillo, I., P. Caces-Borja, and A. Mines. 1960. Preliminary observations on the otter trawl fishery of Manila Bay. *Philippine Journal of Fisheries* 8(1):47-56.
- Ross, S. and T. Doherty. 1994. Short-term persistence and stability of Barrier Island fish assemblages. *Estuarine, Coastal and Shelf Science* 38:49-67.
- Rotschild, B. 1983. Achievement of fisheries management goals in the 1980s. In *Global fisheries*, ed. B. Rothschild, 151-77. New York: Springer.
- Russ, G. 1991. Coral reef fisheries: effects and yields. In *The ecology of fishes on coral reefs*, ed. P. Sale, 601-635. San Diego: Academic Press.
- Sainsbury, K. 1982. The ecological basis of tropical fisheries management. In *Theory and management of tropical fisheries*, eds. D. Pauly and J. Murphy, 167-188. Manila: ICLARM Conference Proceedings, 9.
- _____. 1988. The ecological basis of multispecies fisheries, and management of a demersal fishery in tropical Australia. In *Fish population dynamics (2ed.)*, ed. J. Gulland, 349-382. New York: John Wiley and Sons.
- Sambilay, V. Jr., A. Corpuz, and L. Reyes. (1990). Catch and length frequency data sets of the RV Albacore and the RV Sardinella bottom trawl surveys in Central

- Philippine waters, 1979 to 1982. *Technical Report No. 11*, Iloilo: Department of Marine Fisheries, University of the Philippines in the Visayas.
- San Diego, M. 1985. Measurements of physical variables and nutrient concentration of Tambac Bay. *Unpublished M.S. Thesis*. Quezon City: University of the Philippines.
- Santos, R. and R. Nash. 1995. Seasonal changes in a sandy beach fish assemblage at Porto Pim, Faial, Azores. *Estuarine, Coastal and Shelf Science* 41:579-591.
- SCSP. 1978a. Pelagic resource evaluation. Report on the workshop on the biology and resources of mackerels (*Rastreliger* spp.) and round scads (*Decapterus* spp.) in the South China Sea - Part I. SCS/GEN/78/17. Manila: South China Sea Development and Coordinating Programme.
- _____. 1978b. Report on the workshop on the demersal resources of the Sunda Shelf - Part I. SCS/GEN/78/19. Manila: South China Sea Development and Coordinating Programme.
- Sebastian, A., M. Llorca and V. Encina. 1959. Oceanography of Lingayen Gulf. *Philippine Journal of Fisheries* 7:13-33.
- Sheaves, M. 1992. Patterns of distribution and abundance of fishes in different habitats of a mangrove-lined tropical estuary, as determined by fish trapping. *Australian Journal of Marine and Freshwater Research* 43:1461-79.
- _____. 1993. Patterns of movement of some fishes within an estuary in tropical Australia. *Australian Journal of Marine and Freshwater Research* 44:867-80.
- _____. 1996a. Do spatial difference in the abundance of two serranid fishes in estuaries of tropical Australia reflect long-term salinity patterns? *Marine Ecology Progress Series* 137:39-49.
- _____. 1996b. Habitat specific distributions of some fishes in a tropical estuary. *Marine and Freshwater Research* 47:827-30.
- _____. 1998. Spatial patterns in estuarine fish faunas in tropical Queensland: a reflection of interaction between long-term physical and biological processes? *Marine and Freshwater Research* 49:31-40.
- Sherman, K. and L. Alexander (eds.). 1986. *Variability and Management of Large Marine Ecosystems*. American Society for the Advancement of Science selected symposium series, No. 99.

- Silvestre, G. 1986. Assessment of the multispecies demersal stock of the Samar Sea, Philippines. *Technical report no. 7*. Quezon City: Department of Marine Fisheries, University of the Philippines.
- Silvestre, G. and D. Pauly. 1986. Estimate of yield and economic rent from Philippine demersal stocks 1946-1985. *ICLARM contribution no. 332*, 1-6. Townsville: WESTPAC Symposium on Marine Science in the Western Pacific: The Indo-Pacific Convergence.
- Silvestre, G., R. Regalado and D. Pauly. 1986. Status of Philippine demersal stocks: inferences from under-utilized catch rate data. In *Resources, management and socio-economics of Philippine marine fisheries, Technical report no. 10*, eds. D. Pauly, J. Saeger and G. Silvestre, 47-96. Quezon City: Department of Marine Fisheries, University of the Philippines.
- Smith, I., M. Puzon, and C. Vidal-Libunao. 1980. *Philippine municipal fisheries: a review of resources, technology and socioeconomics*. ICLARM Studies and Reviews 4. Manila: International Center for Living Aquatic Resources Management and Fishery Industry Development Council.
- Soemarto. 1960. Fish behaviour with special reference to pelagic shoaling species: *Lajang (Decapтерus spp.)*. *Indo-Pacific Fisheries Commission Proceedings* 8(3):89-95.
- Sparre, P., E. Ursin and S. Venema. 1989. *Introduction to tropical fish stock assessment. Part I. Manual*. FAO Fisheries Technical Paper No. 306. Rome: UN Food and Agricultural Organization.
- Staples, D. 1983. Gulf of Carpentaria and rainfall. *Proceedings of the Colloquium on the Significance of the Southern Oscillation-El Niño Phenomena and the Need for a Comprehensive Ocean Monitoring System in Australia*. Melbourne: AMSTAC, Department of Science and Technology.
- Studenmund A. and H. Cassidy. 1987. *Using Econometrics: A Practical Guide*. Boston: Little, Brown and Company.
- Suryanarayana, A., C. Murty, and D. Rao. 1992. Characteristics of coastal waters of the western Bay of Bengal during different monsoon seasons. *Australian Journal of Marine and Freshwater Research* 43:1517-33.
- Tabachnick, B. and L. Fidell. 1989. *Using Multivariate Statistics* (2 ed.). New York: Harper and Row, Publishers.

- Tejerina-Garro, F., R. Fortin, and M. Rodriguez. 1998. Fish community structure in relation to environmental variation in floodplain lakes of the Araguaia River, Amazon Basin. *Environmental Biology of Fishes* 51:399-410.
- Ter Braak, C. 1986. Canonical correspondence analysis: a new eigenvector technique for multivariate direct gradient analysis. *Ecology* 67:1167-1179.
- Ter Braak, C. and J. Prentice. 1988. A theory of gradient analysis. *Advances in Ecological Research* 18:271-317.
- Thresher, R. 1991. Geographic variability in the ecology of coral reef fishes: evidence, evolution, and possible implications. In: *The ecology of fishes on coral reefs*, ed. P. Sale, 401-436. San Diego: Academic Press.
- Tiews, K., P. Sucondharman and A. Isarankura. 1967. On the changes in the abundance of demersal fish stocks in the Gulf of Thailand from 1963/1964 to 1966 as a consequence of the trawl fisheries development. *Bangkok Marine Fisheries Laboratory Contribution* 8:1-39.
- Tiews, K., A. Mines, and I. Ronquillo. 1972. On the biology of *Saurida tumbil* (Bloch) 1801 Family Synodontidae in Philippine waters. *Philippine Journal of Fisheries* 10(1 & 2):1-29.
- Tiews, K., P. Divino, I. Ronquillo, and J. Marquez. 1968. On the food and feeding habits of eight species of *Leiognathus* found in Manila Bay and San Miguel Bay. *Proceedings of the Indo-Pacific Fisheries Council* 13(3):93-99.
- Tiews, K., I. Ronquillo, and P. Caces-Borja. 1970a. On the biology of roundscads (*Decapterus bleeker*) in Philippine waters. *Proceedings of the Indo-Pacific Fisheries Council* 13(2):82-106.
- Tiews, K., I. Ronquillo, and L. Santos. 1970b. On the biology of anchovies (*Stolephorus lacepede*) in Philippine waters. *Proceedings of the Indo-Pacific Fisheries Council* 13(2):20-48.
- Trenberth, K. 1991. General characteristics of El Niño-Southern Oscillation. In *Teleconnections: linking worldwide climate anomalies*, eds. M. Glantz, R. Katz, and N. Nicholls, 13-42. Cambridge: Cambridge University Press.
- Vance, D., D. Staples, and J. Kerr. 1985. Factors affecting year-to-year variation in the catch of banana prawns (*Penaeus merguensis*) in the Gulf of Carpentaria. *Journal du Conseil International pour l'Exploration de la Mer*, 42:83-97.

- Villoso, E. and V. Aprieto. 1983. On the relative abundance and distribution of slipmouths (Pisces: Leiognathidae) in Lingayen Gulf, Philippines. *Fisheries Research Journal of the Philippines* 8(1):26-43.
- Villoso, E. E. Cinco, M. Tenmatay, E. Manlapig and M. Ferrer. 1984. Ichthyoplankton survey of Ragay Gulf, Burias Pass, Ticao Pass and northwestern Samar Sea. In *Assessment of trawlable areas in the Philippines II: Ragay Gulf, Burias Pass, Ticao Pass and Waters North of Samar Sea*, ed. A. Mines, 54-66. Quezon City: Technical Report of the Department of Marine Fisheries, University of the Philippines 5.
- Warfel, H. and P. Manacop. 1950. *Otter trawl explorations in Philippine waters*. Research Report 25. Washington, D.C.: Fish and Wildlife Service. U.S. Department of the Interior.
- Watson, R., M. Dredge, and D. Mayer. 1990. Spatial and seasonal variation in demersal trawl fauna associated with a prawn fishery on the Central Great Barrier Reef, Australia. *Australian Journal of Marine and Freshwater Research* 41:65-77.
- Weng, H. 1990. Fish in shallow areas in Moreton Bay, Queensland and factors affecting their distribution. *Estuarine, Coastal and Shelf Science* 30:569-578.
- Wyrski, K. 1961. *Physical oceanography of Southeast Asian waters*. Naga Report No. 2. San Diego: Scripps Institute of Oceanography.
- Yutuc, S. and G. Trono. 1977. A review of the Philippine fisheries resources and their productivity. In *PREPF, Volume II-3B*, 1138-1200. Manila: Development Academy of the Philippines.

APPENDICES

Appendix I. List of Species and Families.

Acanthuridae:

- Acanthurus* spp. (Surgeonfish, Unicornfish)
Naso spp. (Surgeonfish, Unicornfish)
Acanthurus bleekeri (Bleeker's Surgeonfish)

Apogonidae:

(Cardinalfish)

Ariommidae:

- Ariomma indica* (Indian Driftfish)

Balistidae:

- Abalistes stellaris* (Starry Triggerfish)

Bothidae:

- Bothus* spp. (Left-eyed Flounder)

Carangidae:

- Alectis ciliaris* (Pennantfish, Threadfin Mirrorfish)
Carangoides chrysophrys (Longnose Cavalla, Longnose Kingfish)
Carangoides ciliaris (Longfin Cavalla)
Carangoides fulvoguttatus (Goldspotted Trevally)
Carangoides dinema (Shadow Kingfish)
Carangoides malabaricus (Malabar Cavalla, Nakedshield Kingfish)
Caranx melampygus (Bluefin Jack, Blue Kingfish, Blue Trevally)
Carangoides speciosus (Jack)
Decapterus kurroides (Scad)
Decapterus macrosoma (Layang Scad, Cherooffish)
Decapterus muruadsi (Round Scad)
Decapterus russelli (Russel's Scad)
Selar crumenophthalmus (Big-eye Scad, Purse-eye Scad)
Selaroides leptolepis (Yellowstripe Trevally, Smooth-tailed Trevally)
Seriolina nigrofasciata (Blackbanded Trevally, Butter Yellowtail)
Seriola grandis (Trevally)
Uraspis helvolus (Black Ulua)

Champsodontidae:

(Stargazer)

Chlorophthalmidae:

- Chlorophthalmus albatros* (Greeneyes)

Chimaeridae:

(Ratfish)

Centriscidae:

- Aeliscus strigatus* (Shrimpfish, Razorfish)
Centriscus scutatus (Shrimpfish, Razorfish)

Clupeidae:

- Dussumiera acuta* (Rainbow Sardine)
Sardinella longiceps (Indian Oil Sardine)

Dasyatidae:

(Ray)

Diodontidae:	(Two-Toothed Pufferfish)
Engraulididae: <i>Stolephorus tri</i>	(Anchovy)
Fistulariidae: <i>Fistularia petimba</i>	(Cornetfishes)
Formionidae: <i>Formio niger</i>	(Black Pomfret)
Gobiidae: <i>Pentaprion longimanus</i>	(Long-fin Mojarra)
Haemulidae (=Pomadasydae): <i>Plectorhinchus pictus</i> <i>Pomadasys maculatus</i>	(Painted Sweetlip, Grunt) (Blotched Grunt)
Labridae:	(Rainbowfish, Wrass, Tuskfish)
Leiognathidae: <i>Gazza minuta</i> <i>Leiognathus bindus</i> <i>Leiognathus elongatus</i> <i>Leiognathus equulus</i> <i>Leiognathus fasciatus</i> <i>Leiognathus leuciscus</i> <i>Leiognathus splendens</i>	(Toothed Ponyfish, Toothed Soapy) (Orange Ponyfish/Slipmouth) (Slender Ponyfish, Elongate Slimy) (Slimy, Common Ponyfish) (Striped Ponyfish) (Whipfin Ponyfish/Slipmouth) (Splendid Ponyfish, Black-tipped Ponyfish)
Lethrinidae: <i>Lethrinus opercularis</i> <i>Lethrinus lentjan</i>	(Emperor) (Redspot Emperor)
Loliginidae: <i>Loligo</i> spp.	(Squid)
Lophiidae:	(Goosefish, Anglerfish)
Lutjanidae: <i>Aphareus rutilans</i> <i>Lutjanus bojar</i> <i>Lutjanus lineolatus</i> <i>Lutjanus malabaricus</i> <i>Lutjanus vitta</i> <i>Gymnoaesio gymnoptera</i> <i>Macolor macolor</i>	(Small-tooth Jobfish) (Two Spot Red Snapper) (Bigeye Snapper, Yellow Snapper) (Malabar Red Snapper) (Brownstripe Red Snapper) (Snapper) (Snapper)
Mobulidae:	(Ray)
Monacanthidae: <i>Alutera monoceros</i>	(Unicorn Filefish, Unicorn Leatherjacket)
Mullidae: <i>Upeneus mollucensis</i> <i>Upeneus</i> spp.	(Goldband Goatfish) (Goatfish)

<i>Upeneus sulphureus</i>	(Yellow Goatfish)
<i>Upeneus vittatus</i>	(Yellowstriped Goatfish)
Nemipteridae:	
<i>Nemipterus marginatus</i>	(Threadfin Bream)
<i>Nemipterus bathybius</i>	(Yellowbelly Threadfin Bream)
<i>Nemipterus peronii</i>	(Peron's Butterfly Bream)
<i>Scolopsis inermis</i>	(Monocle Bream)
Ostraciidae:	
	(Boxfishes, Coffinfishes, Trunkfishes)
Platacidae:	
<i>Platax orbicularis</i>	(Batfish, Leafish)
Priacanthidae:	
<i>Priacanthus macracanthus</i>	(Red Bigeye)
Rhinobatidae:	
	(Ray, Guitarfish)
Scaridae:	
	(Parrotfish)
Serranidae:	
<i>Epinephelus fuscoguttatus</i>	(Brown-marbled Grouper, Blotched Rockcod)
<i>Epinephelus guttatus</i>	(Grouper)
<i>Epinephelus</i> spp.	(Grouper)
<i>Epinephelus tauvina</i>	(Greasy Grouper)
Scombridae:	
<i>Rastrelliger brachysoma</i>	(Short-bodied Mackerel)
<i>Rastrelliger kanagurta</i>	(Long-jaw Mackerel, Indian Mackerel)
<i>Scomberomorus commerson</i>	(Narrow-barred Spanish Mackerel)
Scorpaenidae:	
	(Lionfish, Scorpionfish)
Sphyraenidae:	
<i>Sphyraena obtusata</i>	(Obtuse Barracuda)
<i>Sphyraena forsterii</i>	(Forster's Barracuda)
Synodidae (=Synodontidae):	
<i>Saurida tumbil</i>	(Common Saury)
<i>Saurida undusquamis</i>	(Brushtooth Lizardfish)
Peristidiinae	
Tetraodontidae:	
	(Pufferfish)
Trichiuridae:	
<i>Trichiurus haumela</i>	(Hairtail, Cutlass)
<i>Eupleurogrammus muticus</i>	(Malayan Hairtail)
Triglidae:	
	(Sea Robins)
Uranoscopidae:	
	(Stargazer)

Appendix II. Site Characteristics of the Different Sampling Units (Fishing Stations).

FISH. STN.	GEOGRAPHIC ATTRIBUTES						OTHER GEOGRAPHIC FEATURES
	SUBSTRA. TYPE	DEPTH (meters)	NO. RIVER TRIBUTARY	SLOPE (X1000)	SHR. DIST. (nau. mi.)	SHLF.WIDTH (nau. mi.)	
1	Clayey-Muddy	46	4	5	3	8	Near entrance of Vinas River
2	Clayey-Muddy	63	4	7	5	8	Near entrance of Vinas River
3	Clayey-Muddy	45	4	10	1	8	Near Vinas River
4	Clayey-Muddy	32	4	10	2	8	Near Vinas River
5	Clayey-Muddy	88	0	27	66	8	South of Vinas River/Deep Section of Shelf
6	Sandy	173	1	27	4	5	Near Coral Island & Intermittent River
7	Sandy	168	0	0	8	20	Basin Site
8	Sandy-Silty Ooze	119	0	0	8	20	Atop Basin Site
9	Sandy-Silty Ooze	132	0	0	4	20	Atop Basin Site
10	Sandy	85	0	13	2	4	
11	Sandy	72	1	27	1	2	Near Donsol River/Sorsogon Bay
12	Coralline	28	0	9	1	5	Off Sorsogon Bay
13	Coralline	29	0	13	1	3	At South Entrance of Sorsogon Bay
14	Coralline	59	0	9	3	4	Off Sorsogon Bay
15	Coralline	32	1	27	0.5	4	Off Sorsogon Bay
16	Hard-Sandy	141	0	0	7	4	Atop Ridge/Sn. Bernard Current Converg. Zone
17	Hard-Sandy	160	0	0	5	5	Atop Ridge/Sn. Bernard Current Converg. Zone
18	Hard-Sandy	216	0	0	3	4	Atop Ridge/Sn. Bernard Current Converg. Zone
19	Hard-Sandy	109	0	12	8	16	Atop Ridge/Sn. Bernard Current Converg. Zone
20	Muddy	77	1	9	4	20	Near Calbayog River
21	Muddy	39	2	5	4	20	Near Calbayog River
22	Muddy	57	1	5	7	20	Near Calbayog River
23	Muddy	25	2	4	6	20	Near Calbayog River

Appendix III. Complete List of All the Species/Taxa Belonging to the Different Fish Assemblages or Groups Found in RABUTINOS.

GROUP	Grp 1	Grp 2	Grp 3	Grp 4a	Grp 4b	Grp 5	Grp 6	Grp 7
# SPECIES	n=18	n=57	n=57	n=120	n=104	n=105	n=183	n=120
TOTAL WT.	wt=101.34	wt=235.80	wt=228.80	wt=141.97	wt=148.46	wt=354.87	wt=339.01	wt=468.51
1	DECAKU 28.180	AFIOMA 46.003	AFIOMA 61.356	LEOSPL 18.248	LEOBIN 34.687	LEOBIN 119.185	DIODON 40.840	CRUXPE 47.548
2	SCOLIN 23.830	SAURUN 29.505	TRICH 53.316	UPEBUL 16.803	AFIOMA 16.382	UPEBUL 22.428	DASYAT 25.757	DIODON 47.245
3	DECAMU 18.180	PRAMA 23.428	PRAMA 14.910	LEOBIN 14.425	LOUGO 11.877	POMAMA 19.700	CRUXPE 22.933	LUTBOJ 37.282
4	PRAMA 9.770	LOPHI 18.685	DASYAT 13.283	TRICH 11.855	UPEMOL 9.317	DASYAT 14.168	AELISC 16.348	DASYAT 34.943
5	NEMBA 8.500	DASYAT 16.795	DECAMU 10.405	SELARO 5.956	APHARE 8.258	UPEMOL 11.275	TETRAO 15.874	ABALIS 25.886
6	SAURUN 4.270	DECAMU 16.658	SAURUN 9.042	RASTBR 5.380	DECAMU 7.988	LUTMAL 10.585	LEOBIN 8.914	TETRAO 22.020
7	EPISPP 4.000	RHINOB 14.245	EPISPP 8.154	SARDLO 4.888	EUPLEU 7.083	TRICH 10.383	PLECN 8.500	ALUTMO 18.581
8	CHLORO 1.870	TRICH 9.144	SERION 6.875	LEOLEU 4.572	SAURUN 6.573	FISTUL 8.825	ABALIS 7.928	LTRILE 14.814
9	SAURTU 1.670	URANOS 6.928	SHARKS 6.822	LOUGO 4.405	FISTUL 4.853	GAZAMI 7.830	LUTLIN 7.845	LTRIP 14.887
10	CHAMPO 0.370	NEMBA 5.864	LOUGO 5.488	DUSSUM 4.235	SAURTU 4.830	CRUXIL 7.088	LTRIP 7.004	OSTRAC 14.480
11	SHRIMP 0.180	CHAMPO 5.479	PERIST 3.731	SPHYOS 3.880	DUSSUM 3.293	ALECIL 6.948	GAZAMI 6.290	EPGUT 14.000
12	CRABSS 0.140	SCORPI 4.898	LOPHI 3.151	DECAMU 3.385	PENTAP 3.282	LEOFAS 6.588	OSTRAC 6.290	LABRID 12.575
13	NEMBA 0.110	DECARU 4.810	URANOS 2.828	SELARU 3.189	RASTKA 2.820	FORMIO 6.305	ALUTMO 6.000	NASOSP 10.358
14	RHINOB 0.070	BOTHUS 4.058	UPEVIT 1.995	SCOMME 3.142	TRICH 2.210	SPHYFO 6.210	UPEMOL 5.497	ACANSP 9.453
15	BOTHUS 0.080	TRIGLI 3.478	DECAMU 1.850	STOLTR 2.648	GYMNOS 2.150	SCOMME 6.775	LABRID 5.728	MACOLO 9.333
16	REKEAS 0.080	SHARKS 3.416	NEMBA 1.830	URASPI 1.898	UPEBUL 2.077	LEOLEU 4.673	CENTRI 4.601	LUTLIN 8.888
17	URANOS 0.040	DECAMU 1.850	CHAMPO 1.830	LEOEQU 1.808	UPEBUL 2.077	LUTBOJ 4.350	PLATAX 4.583	CENTRI 8.758
18	FISTUL 0.020	SCOLIN 1.838	TETRAO 1.527	LEOEQU 1.808	UPEBUL 2.077	DIODON 4.235	LEOFAS 4.528	SCARD 7.700
19		REKEAS 1.763	SELARU 1.487	SAURUN 1.788	TETRAO 1.350	CRUXIN 3.880	ACANSP 4.250	AELISC 6.875
20		CHAMPO 1.638	SAURTU 1.271	TETRAO 1.720	RHINOB 1.333	EPITAV 3.825	ACANBL 4.080	NEMPE 6.737
21		BROTUL 1.534	SCOLIN 1.190	RASTKA 1.388	NEMBA 1.152	CRUXEL 3.558	CRUXIN 4.008	CRUXIL 6.585
22		HISTIN 1.483	PSEULS 1.090	SEPIAT 1.170	SEPIAS 1.140	CRUXPL 3.418	SCARD 3.738	SCOMME 6.088
23		PROMET 1.433	RHINOB 1.087	UPEMOL 1.089	URASPI 1.082	PSETOD 3.325	LTRILE 3.556	LUTVIT 4.985
24		MYCTOP 1.208	SCORPI 0.983	MEGALA 0.943	ALUTMO 1.085	ABALIS 3.315	CRUXIL 3.178	SEPIAS 4.946
25		CARAPI 1.205	UPEMOL 0.958	ALEPDI 0.918	SCOMME 0.833	LTRIM 3.170	PARUHE 3.134	EPISPP 4.272
26		TRIGLI 0.728	SCORPI 0.867	SARDLO 0.917	ABALIS 0.817	LEOEQU 2.905	SCOLBI 3.133	SCOLPE 4.227
27		THEMES 0.688	APOGON 0.774	CRUXARM 0.861	LEOEQU 0.568	SAURTU 2.813	RACHYS 3.108	PRABO 4.188
28		NEMBA 0.683	LACTAR 0.758	SECUN 0.875	DECAMU 0.525	CRUXMAL 2.700	ALECN 2.984	ALECN 4.024
29		EPISPP 0.675	SHRIMP 0.698	DIPLOP 0.857	SERION 0.525	MEGALA 2.418	CRUXRY 2.850	SHARKS 3.888
30		APHARE 0.580	CYNMOGL 0.661	FORMIO 0.834	PRAMA 0.510	SEPIAS 2.415	DREPAN 2.849	CRUXIN 3.443
31		SERION 0.481	PSEULS 0.583	SELAMT 0.824	SERION 0.445	EPGUT 2.375	SPHYBA 2.811	CRUXIL 3.402
32		SHRIMP 0.480	URASPI 0.574	GAZAMI 0.794	PRAMA 0.418	TETRAO 2.308	GYMNOS 2.774	SYMPOR 3.022
33		TETRAO 0.451	SCOMME 0.504	CRUXMAL 0.757	APOGON 0.382	ALUTMO 2.208	MYLJOB 2.687	CHAECC 2.828
34		HISTYP 0.444	APHARE 0.410	CRUCE 0.738	ARGYRO 0.380	LOUGO 1.970	MEGALA 2.831	LEOLEU 2.588
35		SEPIAS 0.408	CARAPI 0.381	FISTUL 0.734	UPETRA 0.342	SELABO 1.953	LEOEQU 2.448	PARASP 2.512
36		CHLORO 0.375	DACTYL 0.359	SPHYLA 0.719	PSETOD 0.270	ALECN 1.858	CRUXIN 2.370	LOUGO 2.428
37		LOUGO 0.368	CHAMPO 0.352	GYMNOS 0.678	SARDLO 0.280	URASPI 1.858	LTRIM 2.250	CRUXIN 2.305
38		DECAMU 0.341	SYNOOU 0.352	RASTFA 0.688	RASTFA 0.233	SCORPI 1.758	SAURTU 2.217	HEMOC 2.253
39		SCOMBE 0.298	FISTUL 0.344	SELAMA 0.659	SELACR 0.232	SELACR 1.585	SCOMME 2.205	ACANBL 2.035
40		APOGON 0.291	PRAMA 0.296	ALECIL 0.610	LOPHI 0.215	LUTLIN 1.398	CANTIG 2.188	SPHYBA 2.030
41		CYNMOGL 0.238	HISTIN 0.288	SEPIAS 0.591	SEPIAT 0.212	SPHYBA 1.398	ALECN 1.901	CRUXRY 1.977
42		CRUXIN 0.233	ALUTMO 0.270	SERION 0.588	SCORPI 0.205	DREPAN 1.275	CRUXCOM 1.785	PARUHE 1.867
43		TRWOOD 0.183	THEMES 0.251	SECUN 0.585	EPITAV 0.200	ARGYRO 1.265	UPEBUL 1.869	DREPAN 1.838
44		LACTAR 0.179	PRASP 0.250	SHARKS 0.545	TRIGLI 0.187	UPEBEN 1.250	SEPIAS 1.707	CRUXMAL 1.835
45		URASPI 0.144	PSEULS 0.215	SPHYBA 0.541	DASYAT 0.175	MEGALA 1.208	PSETOD 1.699	ARGYRO 1.880
46		DIODON 0.139	HISTYP 0.208	SPHYE 0.471	FORMIO 0.165	SELAMT 1.085	CRUXIN 1.688	CRUXCOM 1.728
47		MYLJOB 0.138	DECARU 0.201	STOLIN 0.429	ALECIL 0.150	SEPIAT 0.985	SPHYOS 1.637	SPHYLA 1.686
48		DACTYL 0.135	CONGRI 0.188	PENTAP 0.388	CRUXMAL 0.150	CRUCE 0.805	APOGON 1.512	LEOBIN 1.543
49		SAURTU 0.123	UPEBEN 0.198	SCOLTA 0.350	SCOLTA 0.138	CRUXCOM 0.725	SCOLPE 1.489	APOGON 1.487
50		PARPER 0.103	LEOBIN 0.195	PRAMA 0.338	SHARKS 0.125	SCARD 0.725	CRUXMAL 1.438	BAUST 1.333
51		CRABSS 0.100	RASTKA 0.180	SELAME 0.330	NEMBA 0.123	SAURUN 0.823	CRUXARM 1.427	CRUXIN 1.290
52		PSETOD 0.071	SEPIAS 0.177	APOGON 0.303	EPILU 0.117	PENTAP 0.578	POMAMA 1.417	EPITAV 1.250
53		TRACA 0.063	PARPER 0.172	LEODAU 0.277	CHAMPO 0.108	SHARKS 0.578	FISTUL 1.370	CRUXEL 1.148
54		UPETRA 0.041	SCOMBE 0.171	MEGALA 0.284	SPHYLA 0.108	CRUXEQU 0.575	PENTAP 1.343	SELAMT 1.134
55		TODARO 0.041	NEMBA 0.154	CRUXIN 0.243	CRABSS 0.105	CRUXIN 0.583	LUTGB 1.333	CRUXPL 1.100
56		ABALIS 0.018	EUPLEU 0.144	EUPLEU 0.238	PRABO 0.103	PRAMA 0.505	LUTMAC 1.333	RACHYS 1.098
57		CHLORO 0.010	ALUTMO 0.142	ALUTMO 0.217	EPISPP 0.100	LTRILE 0.500	LOUGO 1.303	ALUTSC 1.078
58			UPETRA 0.139	SERION 0.207	STOLIN 0.085	PRAMA 0.420	CHAECC 1.273	FISTUL 1.083
59			CRABSS 0.138	NEMBA 0.203	TRACA 0.080	AFIOMA 0.415	DECAMU 1.130	SCOLDU 0.982
60			MYCTOP 0.132	CRUXIL 0.183	DACTYL 0.082	APOGON 0.405	PARUN 1.082	SCOLBI 0.880
61			TRWOOD 0.118	SCENA 0.181	TRWOOD 0.082	EPIMOR 0.400	CRUXIL 0.989	CORADI 0.880
62			NEMBA 0.102	ALEPDI 0.153	URANOS 0.082	DECAMU 0.388	SECUN 0.991	LTRIM 0.955
63			PROMET 0.099	ALUTSC 0.148	LUTVIT 0.078	CRUXIL 0.375	TRICH 0.971	NEMTO 0.832
64			EPITAV 0.088	UPEBUL 0.142	DECARU 0.088	PRABO 0.370	SELAMT 0.958	TRWOOD 0.813
65			BAUST 0.075	ALECN 0.134	SYNOOU 0.088	ACANBL 0.333	LUTVIT 0.875	CHANEL 0.810
66			NEMBA 0.068	NEMBA 0.128	EPIMOR 0.080	LEOSPL 0.303	PRABO 0.874	ALECN 0.794
67			OSTRAC 0.055	EPISPP 0.120	SPHYE 0.055	CRUXIL 0.288	ARGYRO 0.828	RASTKA 0.791
68			PRABO 0.051	PRAMA 0.118	CRUXIL 0.057	CRUXARM 0.283	SPHYLA 0.829	CRUXARM 0.708
69			BROTUL 0.042	ABALIS 0.118	LEOFAS 0.055	PARUHE 0.280	NASOSP 0.803	THEMES 0.685
70			BOTHUS 0.042	CRABSS 0.104	RASTBR 0.048	PRASP 0.280	CHAEU 0.790	SCOLTA 0.627
71			TODARO 0.041	NEMBA 0.103	LEOLEU 0.047	OSTRAC 0.270	BAUST 0.783	POMAMA 0.550
72			TRACA 0.038	EPIMOR 0.088	UPEBEN 0.047	CRUXPE 0.250	SELACR 0.771	CANTIG 0.540
73			SEPIAT 0.027	SELABO 0.087	LUTGB 0.042	SECUN 0.198	SPHYFO 0.758	EPILU 0.533
74			PSEULS 0.025	AFIOMA 0.081	SCOLIN 0.038	GERESK 0.185	LEOLEO 0.680	PSEULS 0.532
75			EPIMOR 0.022	ARGYRO 0.088	MEGALA 0.037	APHARE 0.180	MACOLO 0.687	PARUN 0.505
76			LEOLEO 0.020	SCORPI 0.088	NEMBA 0.037	SERION 0.163	PRAMA 0.659	UPEBUN 0.487
77			NEMTO 0.020	EPISPP 0.058	STOLTR 0.033	EUPLEU 0.153	NEMTO 0.667	PARUCR 0.478
78			PENTAP 0.014	LOPHI 0.062	SELAMT 0.030	LOPHI 0.150	SCOLTA 0.808	UPETRA 0.467
79			SARDLO 0.013	PRASP 0.048	DIODON 0.025	LEOLEU 0.120	THEMES 0.551	SERION 0.480
80			ABALIS 0.013	CHAMPO 0.048	PSEULS 0.023	SCOLTA 0.118	GERESK 0.540	LTRICO 0.425
81			DIODON 0.012	TRWOOD 0.038	SCOMBE 0.023	PARUN 0.113	SAURUN 0.538	EQHNE 0.358
82			UPESUN 0.007	RACHYS 0.038	NEMTO 0.020	STOLIN 0.103	PRAMA 0.524	PSEULS 0.345
83			DECARU 0.007	PSEULS 0.035	OSTRAC 0.020	CRUXIL 0.080	SCOLIN 0.508	URASPI 0.315
84			SPHYLA 0.004	TRACA 0.032	NEMPE 0.018	THEMES 0.078	LEOLEU 0.483	SELACR 0.247
85			REKEAS 0.002	PSETOD 0.032	SELARO 0.018	EPISPP 0.078	CRUXEL 0.472	PENTAP 0.233
86			UPEBEN 0.001	CONGRI 0.031	URANOS 0.016	URANOS 0.080	DAYAJE 0.462	SEPIAT 0.228
87			GAZAMI 0.001	GERESK 0.031	LABRID 0.015	SPHYOS 0.083	URASPI 0.452	UPEBEN 0.205
88				CRUXPL 0.028	SCOLBI 0.016	EQHNE 0.046	SCORPI 0.448	DAYAJE 0.200
89				CRUXIN 0.028	CRUXPL 0.013	RASTKA 0.043	NEMPE 0.443	SPHYE 0.200
90				LUTLIN 0.028	UPESUN 0.013	UPEVIT 0.043	TRACA 0.432	SPHYFO 0.191
91				LUTMAL 0.025	ALECN 0.012	TRACA 0.038	PRIAL 0.417	DACTYL 0.188
92				CHAECC 0.024	CRUXARM 0.012	SPHYE 0.035	SCOLDU 0.408	DIPLOP 0.183
93				URANOS 0.023	NEMBA 0.012	CRABSS 0.020	PSEULS 0.405	LEOLEO 0.174
94				SYNOOU 0.022	CRUXRY 0.010	DIPLOP 0.020	UPEBEN 0.387	RASTFA 0.159
95				LABRID 0.019	PARUCR 0.010	NEMBA 0.015	CRUXPL 0.363	TRACA 0.141
96				OSTRAC 0.019	MEGALA 0.005	RACHYS 0.015	HEMOC 0.348	URANOS 0.088
97				LEOFAS 0.018	SELAME 0.005	RASTFA 0.013	EPITAV 0.343	DECAMU 0.087
98				LUTVIT 0.017	SCENA 0.003	SARDLO 0.008	NEMBA 0.343	SAURTU 0.080
99				EPILU 0.016	BROTUL 0.002	CANTIG 0.003	SPHYE 0.328	MEGALA 0.078
100				DACTYL 0.014	CRUXIN 0.002	SYNOOU 0.003	SELARO 0.325	UPEVIT 0.063
101				DECAMU 0.009	GAZAMI 0.002		STOLIN 0.323	CRUXEQU 0.080
102				THEMES 0.009	PARPER 0.002		FORMIO 0.322	CRABSS 0.052

Appendix III. Complete List of All the Species/Taxa Belonging to the Different Fish Assemblages or Groups Found in RABUTINOS.

	Grp 4a	Grp 4b	Grp 5	Grp 6	Grp 7
103	UPETRA 0.008	SHRIMP 0.002		CORADI 0.308	GERESK 0.044
104	SPHYFO 0.008	ACANBL 0.000		SEPION 0.308	SELARO 0.035
105	OREPAN 0.008			URANOS 0.282	PSEUOL 0.033
106	SHRIMP 0.007			NEMIBA 0.287	SELABO 0.030
107	TRIGLI 0.007			LOPHI 0.284	SCORPI 0.025
108	CRXGL 0.008			ECHINE 0.273	PRIATA 0.020
109	BOTHUS 0.005			RASTKA 0.268	STOUN 0.018
110	CHAELO 0.003			SEPION 0.264	SAURUN 0.012
111	SCOMBE 0.003			EPIGUT 0.263	STOLTR 0.012
112	DECAKU 0.003			UPESPP 0.262	SYNODU 0.012
113	LTRILE 0.003			TRIGLI 0.256	SECUN 0.010
114	DAYAJE 0.002			SYNODU 0.251	NEMIBA 0.006
115	NEMITO 0.002			SYMPOR 0.250	SARDLO 0.003
116	EPISPP 0.002			SHRIMP 0.248	
117	LEOLO 0.001			PSEUOL 0.242	
118	CRKEQU 0.001			RHINOB 0.238	
119	PARLUP 0.001			UPETRA 0.236	
120	PARLIN 0.001			DACTYL 0.234	
121				DIFLOP 0.233	
122				CRKEQU 0.213	
123				UPESUN 0.210	
124				SELAMA 0.206	
125				CHELMO 0.187	
126				SEPIAT 0.187	
127				STOLTR 0.183	
128				PARLUP 0.181	
129				CHAMPO 0.181	
130				NEMHE 0.180	
131				CRABSS 0.180	
132				SHARKS 0.180	
133				BROTUL 0.180	
134				TRICOO 0.182	
135				NEMAJA 0.178	
136				EPISIX 0.176	
137				ALUTSC 0.171	
138				AROMA 0.169	
139				CONGRI 0.168	
140				ALEPOJ 0.167	
141				ALEPKA 0.167	
142				APHARE 0.167	
143				BOTHUS 0.167	
144				CARAPI 0.167	
145				CHIMAE 0.167	
146				CHLORO 0.167	
147				CRIOCE 0.167	
148				CRITIL 0.167	
149				CYNOCOL 0.167	
150				DECAKU 0.167	
151				DECAMA 0.167	
152				DECARI 0.167	
153				OUSSUM 0.167	
154				EPISLI 0.167	
155				EPISUS 0.167	
156				EPISPP 0.167	
157				EPISLEU 0.167	
158				HISTIN 0.167	
159				HISTYP 0.167	
160				LACTAR 0.167	
161				LEODAU 0.167	
162				LEOSPL 0.167	
163				LTRICO 0.167	
164				LUTMAL 0.167	
165				MENEMA 0.167	
166				MYCTOP 0.167	
167				PARPER 0.167	
168				PRIASP 0.167	
169				PROMET 0.167	
170				RASTBR 0.167	
171				RASTFA 0.167	
172				RENEAS 0.167	
173				SARDPI 0.167	
174				SARDLO 0.167	
175				SOENA 0.167	
176				SCOMBE 0.167	
177				SECURU 0.167	
178				SELABO 0.167	
179				SELAME 0.167	
180				UPEVIT 0.167	
181				PERIST 0.072	
182				PSEUSP 0.023	
183				PARUCR 0.017	

Appendix IV. Complete List of the Names and Code Designations of All the Species or Taxa Included in the RABUTINOS Demersal Trawl Samples.

No.	CODE	SPECIES	No.	CODE	SPECIES	No.	CODE	SPECIES
1	ABALIS	Abalistes stellaris	76	LABRID	Labridae	152	SCOLBI	Scolopsis bimaculatus
2	ACANBL	Acanthurus bleekeri	77	LACTAR	Lactaridae	153	SCOLDU	Scolopsis dubiosus
3	ACANSP	Acanthurus sp.	78	LEOBIN	Leiognathus bindus	154	SCOLIN	Scolopsis inermis
4	AELISC	Alelucis strigatus	79	LEODAU	Leiognathus daura	155	SCOLPE	Scolopsis personatus
5	ALECIL	Alectis ciliaris	80	LEOELO	Leiognathus elongatus	156	SCOLTA	Scolopsis taeniopterus
6	ALEGIN	Alectis indicus	81	LEOEQU	Leiognathus equulus	157	SCOMBE	Scomber australicus
7	ALEPDJ	Alepes djedaba	82	LEOFAS	Leiognathus fasciatus	159	SCOMME	Scomberomorus commersonii
8	ALEPKA	Alepes kalla	83	LEOLEU	Leiognathus leuciscus	160	SCORPI	Scorpaenidae
9	ALUTMO	Alutera monoceros	84	LEOSPL	Leiognathus splendens	161	SECUIN	Secutor insidiator
10	ALUTSC	Aluteres scriptus	85	LTRICO	Lethrinus choerorhincus	162	SECURU	Secutor ruconius
11	APHARE	Aphareus rutilans	86	LTRILE	Lethrinus lentjan	163	SELABO	Selar boops
12	APOGON	Apogonidae	87	LTRIMI	Lethrinus miniatus	164	SELACR	Selar crumenophthalmus
13	ARGYRO	Argyrops spinifer	88	LTRIOP	Lethrinus opercularis	165	SELAMA	Selar malam
15	ARIOMA	Arioma indica	89	LOLIGO	Loligo sp.	166	SELAMT	Selar mate
16	BALIST	Balistes sp.	90	LOPHII	Lophidae	167	SELAME	Selar melanoptera
17	BOTHUS	Bothus sp.	91	LUTBOJ	Lutjanus bojar	168	SELARO	Selaroides leptolepis
18	BROTUL	Brotulidae	92	LUTGIB	Lutjanus gibbus	169	SEPIAS	Sepia sp.
19	CANTIG	Canthigaster compressus	93	LUTLIN	Lutjanus lineolatus	170	SEPIAT	Sepiathetus
20	CRXARM	Caranx armatus	94	LUTMAC	Lutjanus macolor	171	SERIOG	Seriola grandis
21	CRXCRY	Caranx chrysophrys	95	LUTMAL	Lutjanus malabaricus	172	SERION	Seriola nigrofasciata
22	CRXCIL	Caranx ciliaris	98	LUTVIT	Lutjanus vitta	173	SHARKS	Sharks
23	CRXCOM	Caranx compressus	99	MACOLO	Macolor macolor	174	SHRIMP	Shrimps
24	CRXDIN	Caranx dinema	100	MEGALA	Megalaspis cordyla	175	SPHYBA	Sphyræna barracuda
25	CRXEQU	Caranx equula	101	MENEMA	Mene maculata	176	SPHYFO	Sphyræna forsteri
26	CRXFUL	Caranx fulvoguttatus	102	MOBULI	Mobulidae	177	SPHYJE	Sphyræna jello
27	CRXIGN	Caranx ignobilis	103	MYCTOP	Myctophidae	178	SPHYLA	Sphyræna langar
28	CRXLIN	Caranx linear-fulvoguttatus	104	MYLIOB	Myliobatidae	179	SPHYOB	Sphyræna obtusata
29	CRXMAL	Caranx malabaricus	105	NASOSP	Naso sp.	180	STOLIN	Stolephorus indicus
30	CRXMEL	Caranx melampygus	106	NEMIBA	Nemipterus bathybius	182	STOLTR	Stolephorus tri
31	CRXPPL	Caranx plumbeus	107	NEMIHE	Nemipterus hexodon	183	SYMPOR	Symphorus nematophorus
33	CRXSPE	Caranx speciosus	108	NEMIJA	Nemipterus japonicus	184	SYNODU	Synodus variegatus
34	CRXTIL	Caranx tile	109	NEMIMA	Nemipterus marginatus	185	TETRAO	Tetraodontidae
35	CARAPI	Carapidae	110	NEMINE	Nemipterus nematophorus	186	THENES	Thunnus orientalis
36	CENTRI	Centricus scutatus	111	NEMIME	Nemipterus peronii	187	TODARO	Todarodes pacificus
37	CHAEUL	Chaetodon lunula	112	NEMITO	Nemipterus tolu	188	TRIACA	Triacanthidae
38	CHAEOC	Chaetodon ocellatus	113	OSTRAC	Ostraciidae	189	TRICHI	Trichiurus haumela
39	CHAMPO	Champsodontidae	114	PARPER	Paraperidae	190	TRIGLI	Trigidae
40	CHELMO	Chelmon rostratum	115	PARUCR	Parupeneus chryserydros	191	TRIODO	Triodontidae
41	CHIMAE	Chimaeridae	116	PARUHE	Parupeneus heptacanthus	192	UPEBEN	Upeneus bensasi
42	CIROCE	Chirocentrus dorab	117	PARUIN	Parupeneus indicus	193	UPEMOL	Upeneus molluscensis
43	CHLORO	Chlorophthalmus abrotosus	118	PARUSP	Parupeneus sp.	194	UPESPP	Upeneus sp.
44	CONGRI	Congridae	119	PENTAP	Pentapton longimanus	195	UPESUL	Upeneus sulphureus
45	CORADI	Coradion altiveles	120	PERIST	Peristidae	196	UPESUN	Upeneus undulatus
46	CRABSS	Crabs	121	PINJAL	Pinjalo pinjalo	197	UPETRA	Upeneus tragula
47	CYNOGL	Cynoglossidae	122	PLATAX	Platax orbicularis	198	UPEVIT	Upeneus vittatus
48	DACTYL	Dactyloptendae	123	PLATYC	Platycephalus indicus	199	URANOS	Uranoscopidae
49	DASYAT	Dasyatidae	124	PLECIN	Plectorniscus pictus	200	URASPI	Uraspis helvolus
50	DAYAJE	Daya jerdoni	125	PLECLE	Plectropomus leopardus			
51	DECAKU	Decapterus kurroides	126	PLECMA	Plectropomus maculatus			
52	DECAMA	Decapterus macrosoma	128	POMAHA	Pomadourys hasta			
53	DECAMU	Decapterus muradsi	129	POMAMA	Pomadourys maculatus			
54	DECARU	Decapterus russeli	130	PRIABO	Priacanthus boops			
55	DIODON	Diodontidae	131	PRIAMA	Priacanthus macracanthus			
56	DIPLOP	Diplopterus bifasciatus	132	PRIASP	Priacanthus sp.			
57	DREPAN	Drepane punctata	133	PRIATA	Priacanthus tayenus			
58	DUSSUM	Dussumiera acuta	134	PROMET	Promethichthys prometheus			
59	ECHINE	Echineis naucrates	135	PSETOD	Psettodes erumei			
60	EPIBLI	Epinephelus bleekeri	136	PSEUDU	Pseudorhombus duplici-cellatus			
61	EPIFUS	Epinephelus fuscoguttatus	137	PSEUEL	Pseudorhombus elevatus			
62	EPIGUT	Epinephelus guttatus	138	PSEUOL	Pseudorhombus oligodon			
63	EPIMOR	Epinephelus morhua	139	PSEUSP	Pseudorhombus sp.			
64	EPISEX	Epinephelus sexfasciatus	140	RACHYS	Rachycentron canadus			
65	EPISTP	Epinephelus sp.	141	RASTBR	Rastrelliger brachysoma			
66	EPIYAV	Epinephelus tauvina	142	RASTFA	Rastrelliger faughni			
67	EUPLEU	Eupleurogrammus noticus	143	RASTKA	Rastrelliger kanagurta			
68	FISTUL	Fistularia petimba	144	REXNAS	Rexia solandri			
69	FORMIO	Formio niger	145	RHINOB	Rhinobatidae			
70	GAZAMI	Gazza minuta	146	SARDFI	Sardinella fimbriata			
71	GERESK	Gerrus kapas	147	SARDLO	Sardinella longiceps			
72	GYMNOS	Gymnoaesio gymnoptera	148	SAURTU	Saurida tumbil			
73	HENIOC	Heniochus acuminatus	149	SAURUN	Saurida undosquams			
74	HISTIN	Histiogaster indicus	150	SCARID	Scaridae			
75	HISTYP	Histiogaster typus	151	SCIENA	Sciæna dussumieri			


Appendix V. Species/Taxa Composition of Standard SU Groups at RABUTINOS.


SPECIES NAME	G-1	G-2	G-3	G-4B	G-4A	G-5	G-6	G-7
<i>Decapterus kurroides</i>	28.2	1.9						
<i>Scolopsis inermis</i>	23.8	1.8						
<i>Decapterus muruadsi</i>	18.2	15.7	10.4	8.0	3.4			
<i>Priacanthus macracanthus</i>	9.8	23.4	14.9					
<i>Nemipterus marginatus</i>	8.5							
<i>Saurida undosquamis</i>	4.3	29.5	9.0	6.6	1.8			
<i>Epinephelus</i> spp.	4.0		8.2					
<i>Chlorophthalmus albatros</i>	1.9							
<i>Saurida tumbil</i>	1.7			4.8				
Champsodontidae	0.4	5.5	1.8					
Shrimps	0.2							
Crabs	0.1							
<i>Nemipterus bathybius</i>	0.1	5.7	1.9					
Rhinobatidae	0.1	14.3		1.3				
<i>Bothus</i> spp.	0.1	4.1						
<i>Rexea solandrii</i>	0.1	1.8						
Uranoscopidae	0.0	6.9	2.6					
<i>Fistularia petimba</i>	0.0			4.9		8.8		
<i>Ariomma indica</i>		46.0	61.4	16.4				
Lophiidae		18.7	3.2					
Dasyatidae		16.8	13.3			14.2	25.8	34.9
<i>Trichiurus haumela</i>		9.1	53.3	2.2	11.9	10.4		
Scorpaenidae		4.9						
<i>Decapterus russelli</i>		4.8						
Triglidae		3.9						
Sharks		3.4	6.6					
Chimaeridae		1.6						
<i>Seriola nigrofasciata</i>			6.9					
<i>Loligo</i> spp.			5.5	11.9	4.4			
Peristidiinae			3.7					
<i>Upeneus vittatus</i>			2.0					
<i>Decapterus macrosoma</i>			2.0					
Tetraodontidae			1.5	1.4	1.7		15.9	22.0
<i>Selar crumenophthalmus</i>			1.5		3.2			
<i>Saurida tumbil</i>			1.3		2.7			
<i>Leiognathus bindus</i>				34.7	14.4	119.2	8.9	
<i>Upeneus mollucensis</i>				9.3		11.3	5.5	
<i>Aphareus rutilans</i>				8.3				
<i>Eupleurorammus nuticus</i>				7.1				
<i>Dussumiera acuta</i>				3.3	4.2			
<i>Pentaprion longimanus</i>				3.3				
<i>Rastrelliger kanagurta</i>				2.6				
<i>Gymnoaesio gymnoptera</i>				2.2				
<i>Upeneus sulphureus</i>				2.1	15.6	22.4		
<i>Leiognathus elongatus</i>				1.6		4.7		
<i>Upeneus</i> spp.				1.4				
<i>Leiognathus splendens</i>					18.3			
<i>Selaroides leptolepis</i>					6.0			

Appendix V. Species/Taxa Composition of Standard SU Groups at
RABUTINOS (continued).

SPECIES NAME	G-1	G-2	G-3	G-4B	G-4A	G-5	G-6	G-7
<i>Rastrelliger brachysoma</i>					5.4			
<i>Sardinella longiceps</i>					4.7			
<i>Leiognathus leuciscus</i>					4.6			
<i>Sphyraena obtusata</i>					3.9			
<i>Scomberomorus commerson</i>					3.1	5.8		
<i>Stolephorus tri</i>					2.1			
<i>Uraspis helvolus</i>					1.9			
<i>Leiognathus equulus</i>					1.8			
<i>Pomadasys maculatus</i>						19.7		
<i>Lutjanus malabaricus</i>						10.6		
<i>Gazza minuta</i>						7.6	6.3	
<i>Carangoides ciliaris</i>						7.1		
<i>Alectis ciliaris</i>						7.0		
<i>Leiognathus fasciatus</i>						6.6	4.5	
<i>Formio niger</i>						6.3		
<i>Sphyraena forsteri</i>						6.2		
<i>Lutjanus bohar</i>						4.4		37.3
Diodontidae						4.2	40.6	47.3
<i>Carangoides dinema</i>						3.7		
<i>Epinephelus tauvina</i>						3.6		
<i>Carangoides speciosus</i>							22.9	47.6
<i>Aeliscus strigatus</i>							16.4	6.9
<i>Plectorhincus pictus</i>							8.5	
<i>Abalistes stellaris</i>							7.9	26.0
<i>Lutjanus lineolatus</i>							7.6	8.9
<i>Lethrinus opercularis</i>							7.0	14.7
Ostraciidae							6.2	14.5
<i>Alutera monoceros</i>							6.0	18.6
Labridae							4.7	
<i>Centriscus scutatus</i>							4.6	8.8
<i>Platax orbicularis</i>							4.6	
<i>Acanthurus spp.</i>							4.3	9.5
<i>Acanthurus bleekeri</i>							4.1	
<i>Lethrinus lentjan</i>								14.8
<i>Epinephelus guttatus</i>								14.0
Labridae								12.4
<i>Naso spp.</i>								10.4
<i>Macolor macolor</i>								9.3
Scaridae								7.8
<i>Nemipterus peroni</i>								6.7

Legend:

 = major distribution mode

 = minor distribution mode