1 Trace Analysis of Anionic and Nonionic Surfactants from Oil Dispersants in Gulf of 2 Mexico Seawater Using Large Volume Injection Liquid Chromatography with Tandem 3 **Mass Spectrometry** 4 Benjamin J. Place¹, Matt J. Perkins², Adam Barsamian¹, Paul Blakemore¹, Jennifer A. Field² 5 ¹ Department of Chemistry, Oregon State University, Corvallis, OR 6 7 ² Department of Environmental and Molecular Toxicology, Oregon State University, Corvallis, 8 OR 9 10 11 **Table of Contents** 12 Page.....Contents 13 1.....Cover Page 14 2.....Experimental Methods 15 2.....Results and Discussion 16 17 Figures 18 3.....Figure S1: DOSS hydrolysis transformation pathway 19 4.....Figure S2: Salt elution timeline 20 5.....Figure S3: Stability of analytes in parent stock standards 21 6.....Figure S4: Stability of analytes in analytical standards. 22 7.....Figure S5: Matrix effects of nonionic analytes 8.....Figure S6: Effect of filtration on analyte recovery 23 9.....Figure S7: Chromatographic separation of DOSS contamination 24 11.....Figure S8: Timeline of LC/MS parameters 25 12.....Figure S9: Schematic of LC/MS flow 26 27 13.....Figure S10: Chromatographic separation and MS detection of polyethoxylates. 28 29 Tables 30 10.....Table S1: MS parameters for all analytes 31

32 **EXPERIMENTAL METHODS**

33 LVI-LC-MS/MS Method Details

34 A single MRM transition was identified and scanned for α -/ β -EHSS (m/z 309 > 81), 35 while two MRM transitions were scanned for DOSS (quantitative: m/z 421 > 81, qualitative: m/z36 421 > 227). The negatively ionized analytes were detected during the first 17 min. Span 80 was 37 detected in positive mode with two MRM transitions (quantitative: m/z 429 > 411, qualitative: 38 m/z 446 > 429). The qualitative transitions for DOSS and Span 80 were monitored to verify 39 presence, although there were no ratio requirements due to poor sensitivity of the qualitative 40 transitions. The positively ionized analytes were detected for the latter 16 min of the analytical 41 run. A timeline is illustrated in shown in Figure S8 and the MS Parameters is shown in Table 42 **S1**. 43 44 Salt Elution Timeline.

45 In order to determine the required time for the complete elution of the salts in the seawater samples, an eluent precipitation experiment was developed. A sample of 1,800 µL 46 (25% Isopropanol, 75% Instant Ocean sample) was injected using the developed analytical 47 HPLC parameters except with the post-column eluent collected instead of injected into the MS. 48 After the injection, 30 s (0.25 mL) fractions of eluent were collected over the first 10 min of the 49 50 analytical run. 51 A solution of 1 M AgNO₃/0.6 M HNO₃ was made in deionized water for the precipitation

52 of solid AgCl (Cl⁻ as a broad indicator of seawater). Preliminary experiments found samples 53 containing > 0.1% seawater showed visible AgCl precipitate. A 100-µL aliquot of the AgNO₃ solution was added to each eluent and the samples were briefly shaken.

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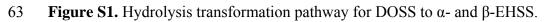
56 **RESULTS AND DISCUSSION**

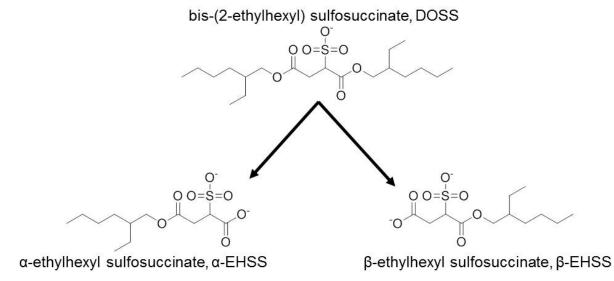
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58 Salt Elution Timeline.

59 AgCl precipitate was observed for samples from 1 min to 7.5 min, suggesting majority of 60 the salts eluted during this time. The post-column divert valve was set to divert the eluent flow to the mass spectrometer only after 9.5 min, which verified that no high concentrations of salt 61

62 would be directed to the MS (Figure S1).





- **Figure S2.** Visual demonstration of salt elution from the analytical column using AgNO₃ as a
- 66 precipitation indicator of Cl^{-} ions.

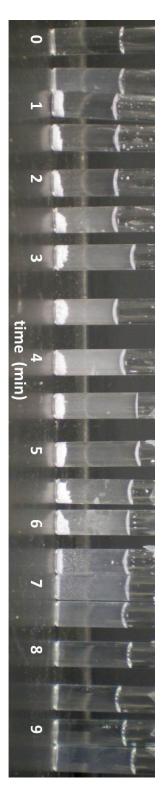
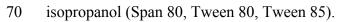
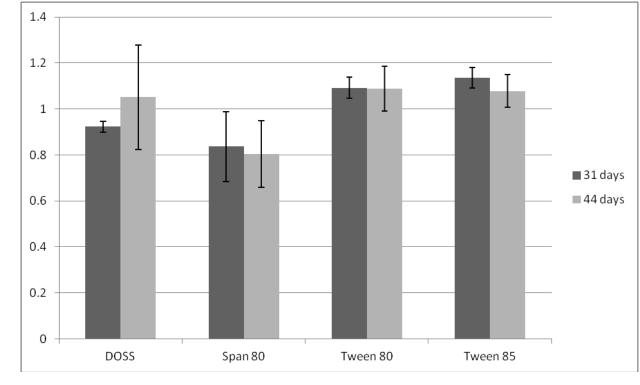
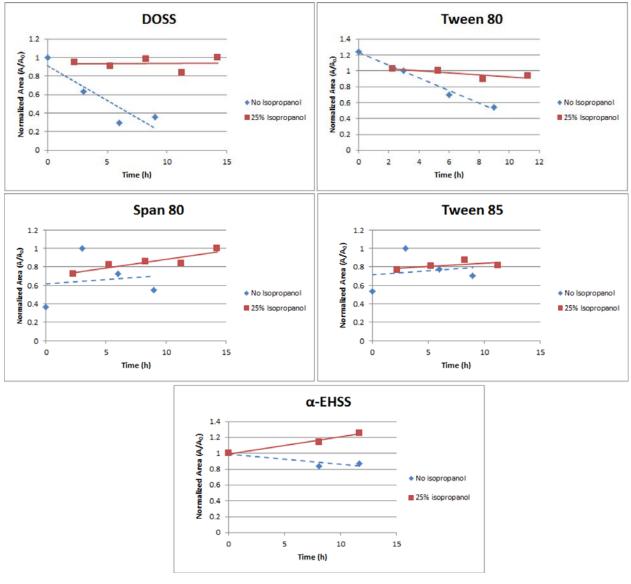


Figure S3. Stability of analytes in parent stock standards prepared in methanol (DOSS) and





- 73 **Figure S4**. Stability of analytical standards in HPLC autosampler vials with/without 25%
- 74 isopropanol. Normalized area was calculated by dividing the resultant area counts by the largest
- 75 area counts for that compound.



- 78 **Figure S5.** Matrix effects, as indicated by normalized peak area counts, of nonionic analytes
- 79 prepared in buffered water (0.5 mM ammonium acetate), Instant Ocean, and Oregon Coast
- 80 water.

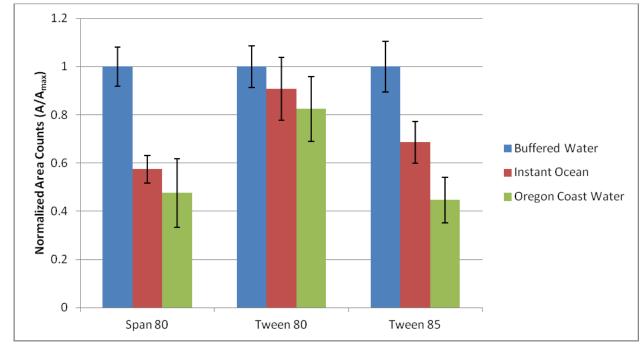
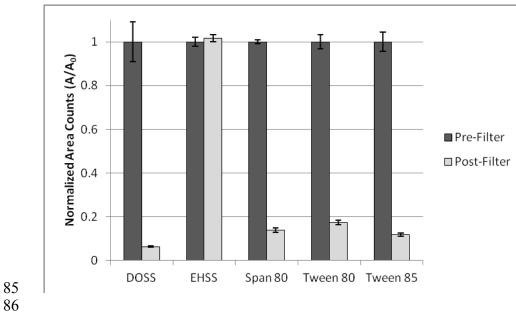
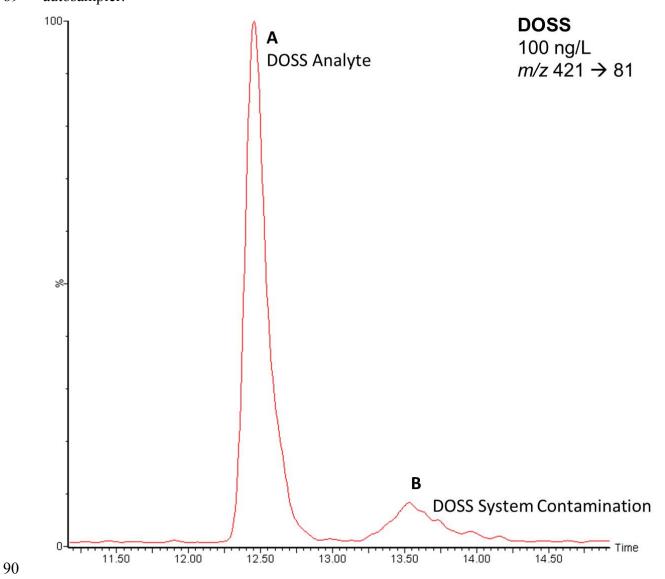


Figure S6. Recovery of all analytes before and after filtration through a Sterivex 0.2-μm



84 sterilization filter.

- 87 **Figure S7**. Chromatographic separation of DOSS analyte in sample (A) and the system
- background contamination (B) using a guard column placed after the pump mixer, but before theautosampler.



	Retention						
	Time	MS	Scan	Parent Ion	Daughter	Cone	Collision
Analyte	(min)	Ionization	Mode	(m/z)	Ion (m/z)	Voltage (V)	Energy (V)
DOSS	12.7	Negative	MRM	421	81	44	26
		Negative	MRM	421	227*	42	18
³ C ₄ -DOSS	12.7	Negative	MRM	425	81	44	26
α-/β-EHSS	10.1	Negative	MRM	309	81	44	26

446

411

429*

309

309

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91 **Table S1**. MS Parameters for the detection of all analytes. * indicates secondary MRM

MRM

MRM

Positive Parent Ion 400 - 1300

Positive Parent Ion 400 - 1300

Positive

Positive

92 transitions used for analyte verification.

26.8

26.0

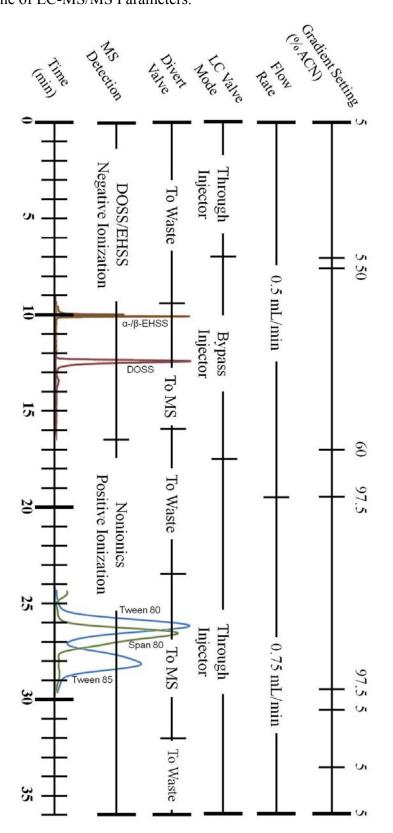
28.2

93

Span 80

Tween 80

Tween 85



- 97 **Figure S9**. Schematic of LC/MS design. Top figure is the LC/MS in main pass LC mode and
- 98 post-column solvent flow is diverted to the waste (column wash step). Bottom figure is the
- 99 LC/MS in bypass LC mode with post-column solvent flow diverted to the MS (analyte elution
- 100 step)
- 101

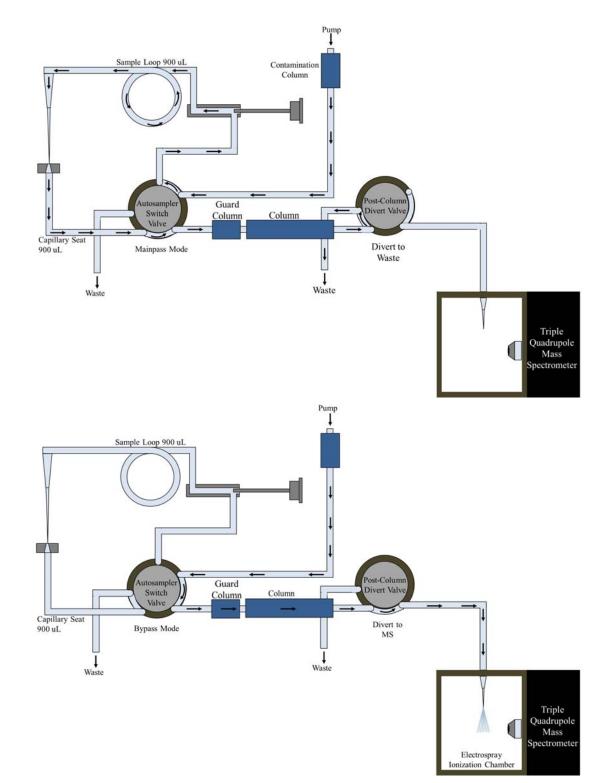


Figure S10. Top: Chromatogram of the parent ion scanning for m/z 309, Tween 80 and Tween 85 are designated as peaks A and B, respectively. Middle: Parent ions of product ion m/z 309 for peak A. Bottom: Parent ions of product ion m/z 309 for peak B.

