

# Development of a Continuous Flow Analysis System for Trace Iron

-Data Report-



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## 2 Introduction

Iron levels in the surface ocean are generally very low—in the picomolar to nanomolar range—reaching levels as low as 0.1 nM as a result of biological uptake [Bruland and Rue 2001; Measures 1995]. Iron's role as an essential nutrient for phytoplankton suggests that it may play a major role in determining total biomass and community structure [Bruland 2001, Johnson, 2001]. In much of the ocean, iron is assumed to be the limiting nutrient [Bruland, Lohan 2003]. While this is especially true for the open ocean, coastal oceans may also experience iron limitation [Bruland, 2001, Hutchins and Bruland, 1998, Johnson, 2001].

To better understand iron's role in determining the rate of phytoplankton growth, and by extension, the rate of carbon uptake in the coastal ocean the measurement of iron was included as part of the SUCCES (Seasonal Upwelling Coastal Carbon Export and Sequestration) cruises in summer 2009. We needed to develop a system to measure iron concentrations using a continuous sample stream from our towed pump vehicles, SuperSucker and SeaSoar.

Iron's relative scarcity combined with its ubiquity in the human environment makes it a challenge to measure. Methods must be very sensitive but also minimize the potential for contamination by minimizing the exposure of samples to the open air. For our purposes, the system must also be able to quickly measure iron in a continuous sample stream to resolve iron gradients in the shelf waters.

Iron has been measured to a high degree of accuracy by several methods. Inductively coupled plasma mass spectrometry (ICPMS), a highly sensitive method, is very useful in the laboratory [Wu and Boyle, 1998] but is not a feasible method of shipboard analysis. Electrochemical techniques, including cathodic stripping voltammetry, can be used at sea and can give speciation data. Electrochemistry can only be used on discrete samples.

Two other analysis methods can be used to measure iron. Flow injection analysis (FIA) is the most common measurement of iron done at sea. FIA relies on discrete injections of sample into a constant carrier fluid that then mixes with a reagent stream and flows to the detector. FIA has been used for continuous analysis of surface waters [Measures, 1995; Johnson, 2001; Chase, 2005] because of its ease of use, efficient sampling, and high precision [Zhang 2001]. The total analysis time of approximately 5 minutes per sample (Measures 1995) leads to both smearing (due to load time) and lack of the desired temporal resolution. In classical continuous flow analysis (CFA), the sample stream continuously flows into the stream of reagents and this reaction mixture is continuously delivered to the detector. When used in this way, CFA generates results that may be "smeared."

To achieve our goal of very high frequency measurements, we have adapted the CFA method by segmenting the sample with injected air bubbles in a process called gas segmented continuous flow analysis (GSCFA). It would allow for high temporal resolution of our sample stream with minimal smearing. GSCFA has been used successfully by Hales, et. al. [2004] for high resolution nitrate measurements in similar shipboard settings. GSCFA allows for higher frequency measurements than FIA as there is no pre-concentration column, the absence of a column also reduces smearing. In addition, this method, like FIA, minimizes the sample handling which can lead to contamination [Weeks 2002].

## Introduction

We have developed a continuous flow analysis (CFA) system capable of high frequency measurements to profile the bottom boundary layer with chemistry based on previous flow injection analysis systems (FIA) utilizing N,N-dimethyl-p-phenylenediamine dihydrochloride (DPD) [Measures et al. 1995; Sedwick et al. 1997; Weeks and Bruland 2002; Chase 2005].

### 3 Methods

#### 3.1 Equipment set up

*Refer to figure 1*

The CFA set up is contained in a plastic box that has been modified specifically for this purpose (see Figure 1). The input ports for DPD, buffer, and H<sub>2</sub>O<sub>2</sub> all have check valves to prevent reagent contamination. There is also an input port for the sample and seven ports connected to the VCI ChemInert (06U-0396L) valve for the sample and standards.

Reagents were kept in a AirClean 600 PCR workstation clean air hood and pumped into the CFA box. PTFE 1/16" OD x 0.5mm ID plastic tubing is used to draw the reagents from the hood to the Ismatec (C.P. 7800-40) peristaltic pump. Various sizes of Cole Parmer peristaltic pump tubing are used to deliver the appropriate reagent mixture—DPD, acid, and air are delivered in tygon 0.38mm ID, buffer and sample in Pharmed BPT 0.89 mm ID, and H<sub>2</sub>O<sub>2</sub> in Pharmed 0.51 mm ID.

The buffer and DPD are then mixed and run through a GE Healthcare HiTrap 1mL (17-0408-01) chelating column. H<sub>2</sub>O<sub>2</sub> is then added to the reaction mixture. The sample is combined with 0.10N HCl and flowed through three 25-turn glass coils (approx ID 1mm). After acidification is complete, the reaction mixture and acidified sample are combined and flowed through one 25-turn glass coil before flowing to the photodiode detector and out of the box to a waste bottle. The photodiode is connected to a Macintosh computer via a National Instruments USP 6009 card and data is collected using a LabView program written by Burke Hales.

The peristaltic pump is set at 35 which generates reagent flow rates of 0.92 mL/min for DPD (5.25 mM/min), 0.315 mL/min for buffer, 0.10 mL/min for H<sub>2</sub>O<sub>2</sub>, 0.328 mL/min for sample, and 0.092 mL/min for acid. This gives a total flow of 0.84 mL/min. The temperature of the acidification coil was controlled to 40°C while the reaction coil was not temperature controlled.

#### 3.2 Reagents

DPD is ordered in a powdered form (both Sigma and Fluka were used during this research) and stored in the refrigerator to prevent oxidation. The reagent is prepared daily as a mixture of 0.6 g DPD in 60 mL of MilliQ H<sub>2</sub>O plus 4uL/mL HCl. Hydrogen peroxide is mixed as needed. The reagent is mixed as 125 mL of a 5% solution made from J.T. Baker Ultrex II ultrapure 30% H<sub>2</sub>O<sub>2</sub>.

1L of buffer is made from 170 mL isoplastically distilled NH<sub>4</sub>OH (~4M) and 70 mL J.T. Baker Ultrex II ultrapure glacial acetic acid and filled to 1L with MilliQH<sub>2</sub>O. The pH is then measured and adjusted to 6.3 with NH<sub>4</sub>OH. The acidification solution is 0.1 N HCl mixed in 500 mL batches from J.T. Baker Ultrex II ultrapure hydrochloric acid.

Two stock iron solutions (14.46 µM and 189 nM) were made in MilliQ (with 1 mL 6N HCl/125mL) from Ultra Scientific Analytical Solutions commercial iron standard (10,000 µg/mL). The standards used for running the instrument were made from these two standards with seawater (at pH 6) chelated in a GE Healthcare HiTrap 5mL chelating column. These standards were acidified at 4uL/mL. Standards varied from 0.4 nM to as high as 100 nM and still remained in a linear range.

## 3.3 Shipboard setup

Refer to Figure 2

On board R.V. Wecoma, water was pumped from the underwater towed vehicle Supersucker and then routed into the wet lab using nylon tubing. The sample was collected via a tangential flow filter (TFF) built and designed by Chris Holm. The TFF was fitted with a 0.2 micron 47 mm filter that had been rinsed in acid and stored in 10% HCl. A bypass loop was constructed around the TFF to prevent flow disruptions downstream. A Polyethylene line from the surface towed iron fish was also plumbed to the GCFA so it could run either supersucker or surface fish sample.

## 3.4 Sample collection

Discrete samples were collected on the cruises in several different ways. Some samples were collected at the sink outlet of the supersucker. These were collected in a syringe rinsed with the flow three times and then filtered with an acid rinsed 0.45 micron Supor membrane PALL Life Sciences IC Acrodisk 25mm syringe filter and acidified. Other samples were collected at the input to the GCFA system. These samples were collected with a syringe in two ways: via the tangential flow filter or from the sample flow and filtered as with the samples collected at the sink. All samples were treated with 240  $\mu$ L 6N HCl per 60 mL of sample.

CTD samples were collected in a similar manner. We were second to sample the CTD bottles (after methane sampling). A PALL SUPOR AcroPak 200 0.2 micron cartridge filter was affixed to the spout and allowed to flush for about a minute before collecting 60 mL samples. These samples were also acidified with 240  $\mu$ L 6N HCl per 60 mL of sample.

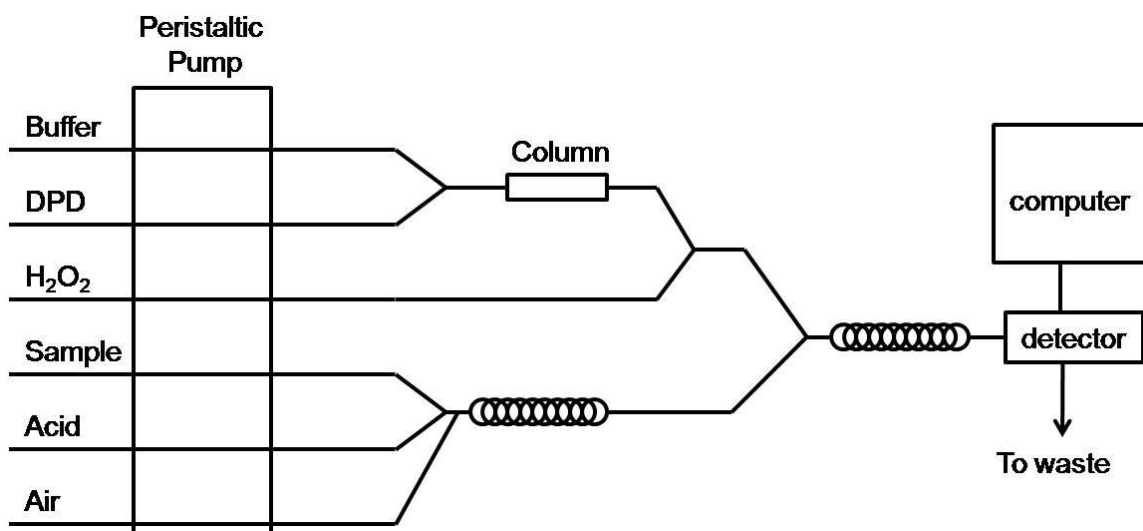


Figure 1: GCFA Schematic

## Methods

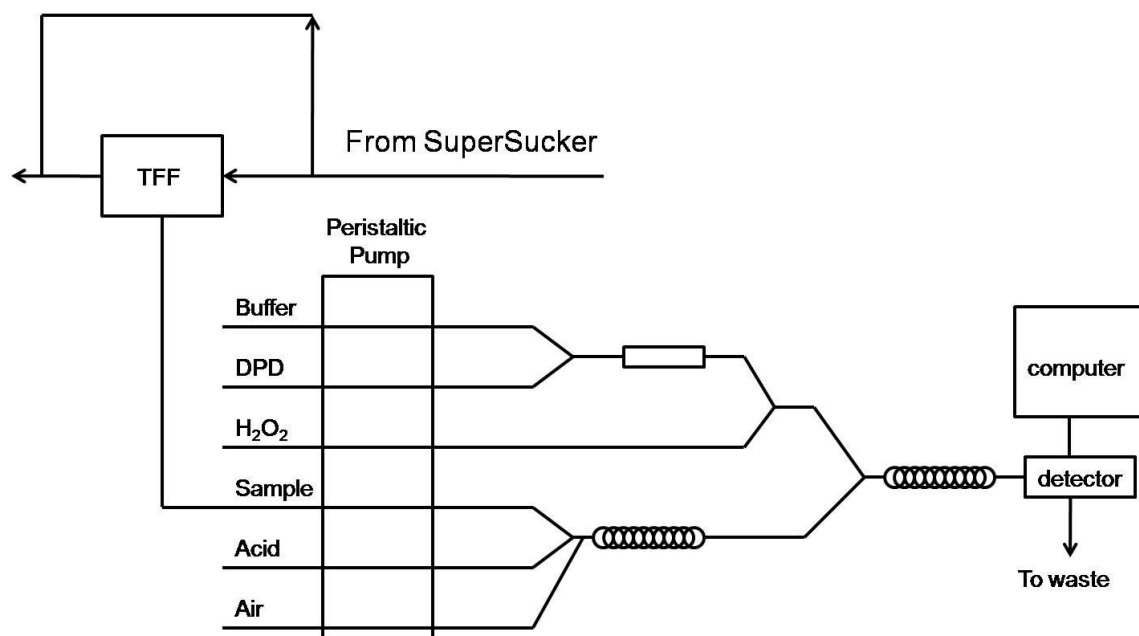


Figure 2: GCFA as on SXS Cruises

## Results

### 4 Results

#### 4.1 Lab tests

The system was tested in the lab to prepare it for deployment on the SUCCES cruises throughout 2008 and through the early summer of 2009. After deployment on SXS2, the system was tested further, especially with respect to the disagreement between discrete and inline samples (see May 26 and final discussion), before it was deployed on SXS3

##### 4.1.1 High-low comparisons and response time

Several tests of a high-low response were conducted to determine the response time of the system. These tests were rather inconclusive as the response time seemed to vary in an undermined fashion.

The run conducted in February 2009 (see chart below) illustrated the typical response of an initial fast response followed by a longer and slower secondary tapering. This test was done with two standards alternating back and forth. There does appear to be a drift problem evident in this sample—this result was not consistently reproducible. The high standard in this case was beyond the linear range of the system at 225.5 nM which may have had other effects on the system.

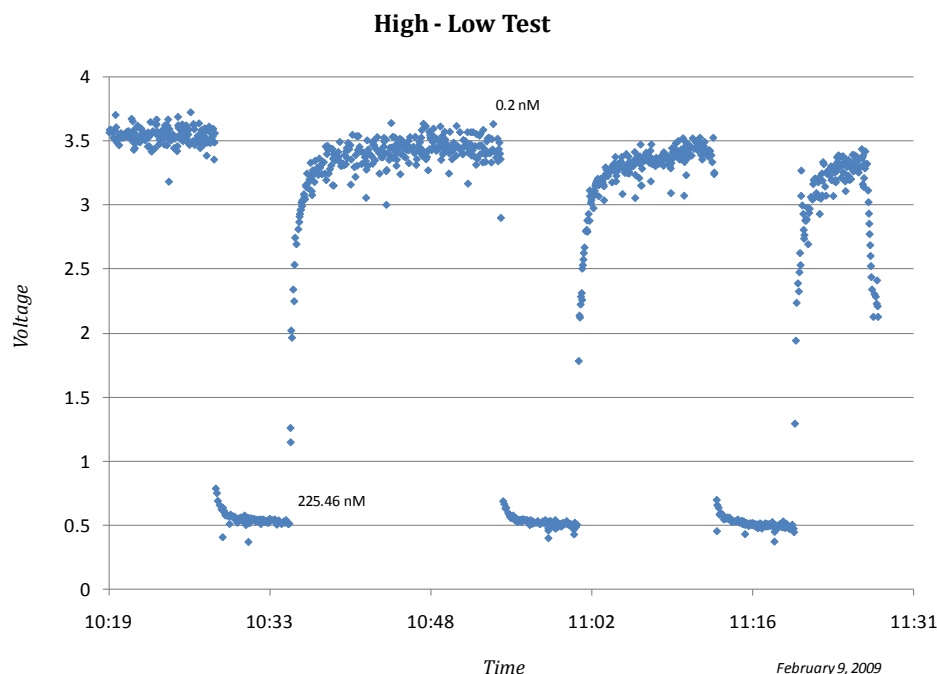


Figure 3: Laboratory high-low testing (1)

A test using two different standards (mixed at the same time as the example above) showed very little drift in the lower standard but still maintains the fast initial response followed by a slower magnitude tapering.

## Results

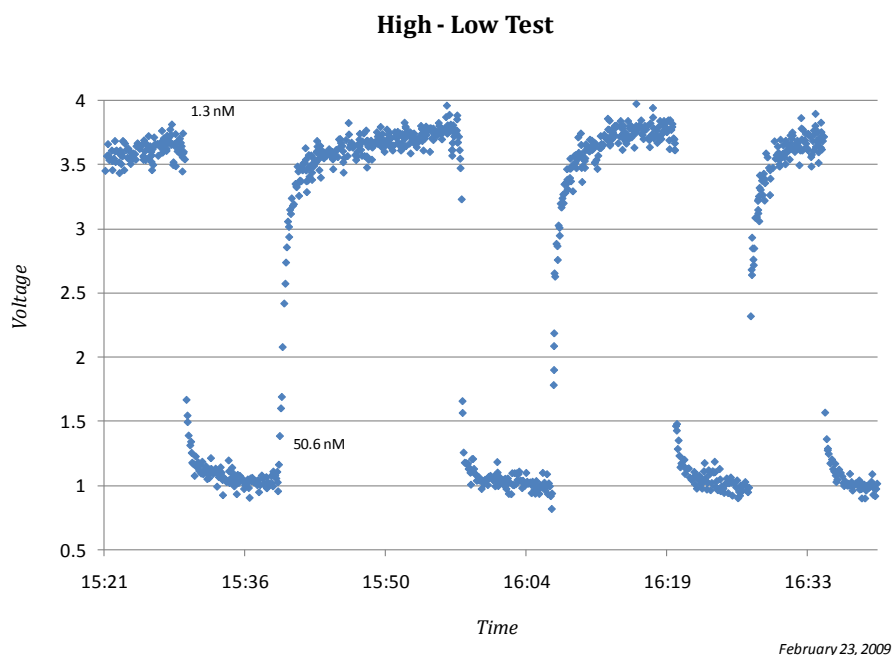


Figure 4: Laboratory high-low testing (2)

The system has shown the ability to give repeatable fast results. In the test (chart below) conducted on August 7, 2008 a one meter plastic reaction coil was used. There was no inline acidification. This test shows little to no drift, consistency in the curve shape, and relatively fast responses.

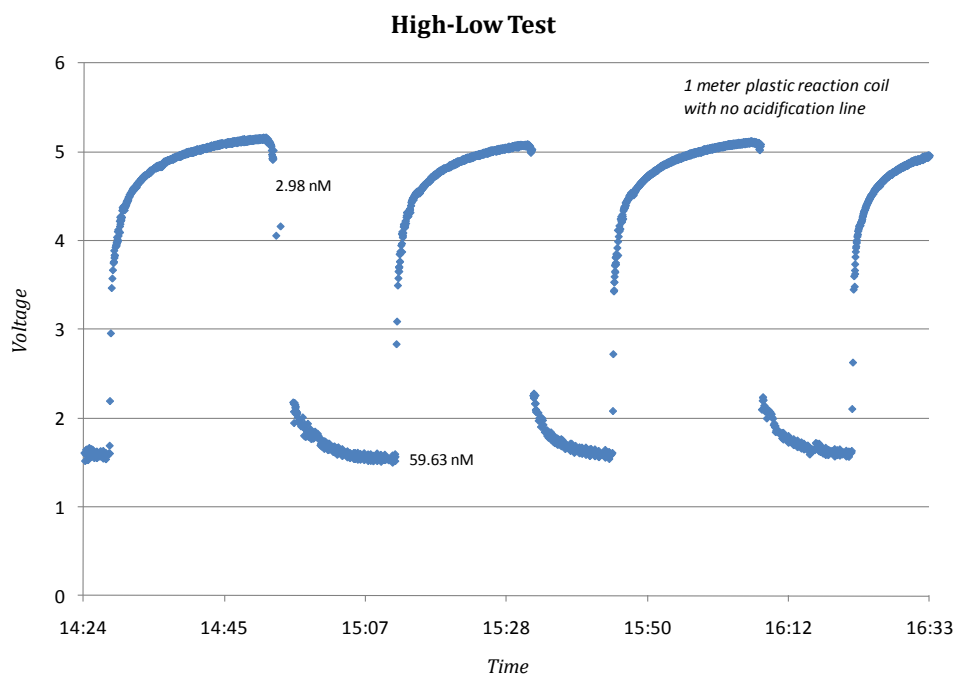


Figure 5: Laboratory high-low testing (3)

An overlaid look at the data shows the February dates had very similar response times. This is to be expected because very little changed in the system

## Results

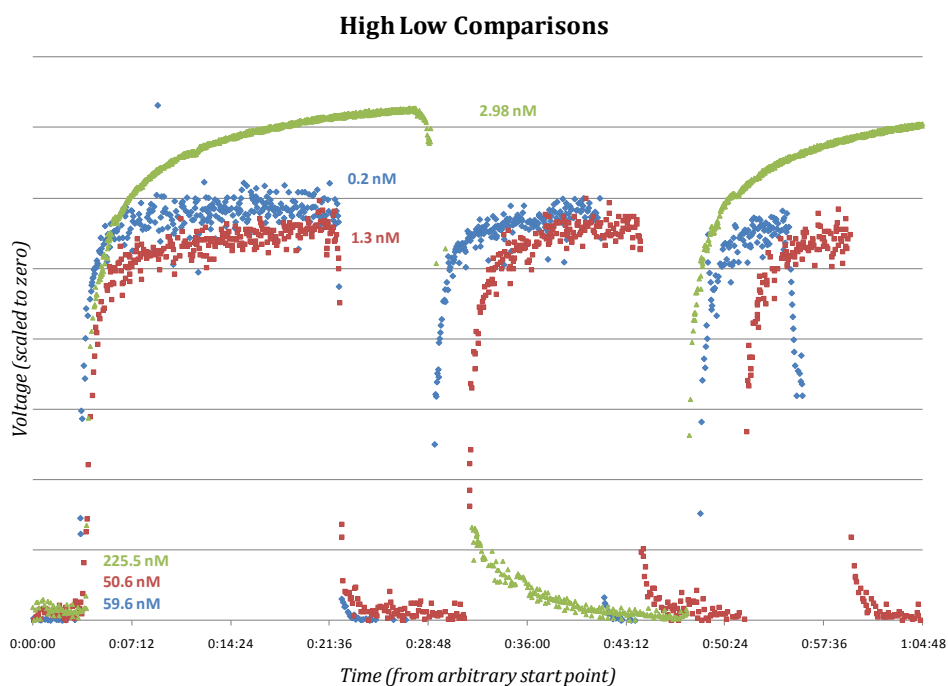


Figure 6: High-Low Test Comparison

### 4.1.2 Acidification and pH

Acidification was ruled out as a possible cause of the discrete and underway mismatches through a series of laboratory tests.

First, a set of five 60 mL standards (0.19, 0.91, 2.81, 9.26, 51.19), plus a blank, was mixed and acidified to the usual level of acidification used for standards (240  $\mu$ L 6N HCl/60 mL sample). These samples were run in the GCFA with the inline acidification line removed. After each sample was run, they were acidified with an additional 240  $\mu$ L 6N HCl and run again as “double acidified” samples. The standards and blank were acidified one more time with 240  $\mu$ L 6N HCl and run as “triple acidified” samples. There were no significant differences between the absorbancies of single, double, and triple acidified standards as seen in the chart below.

## Results

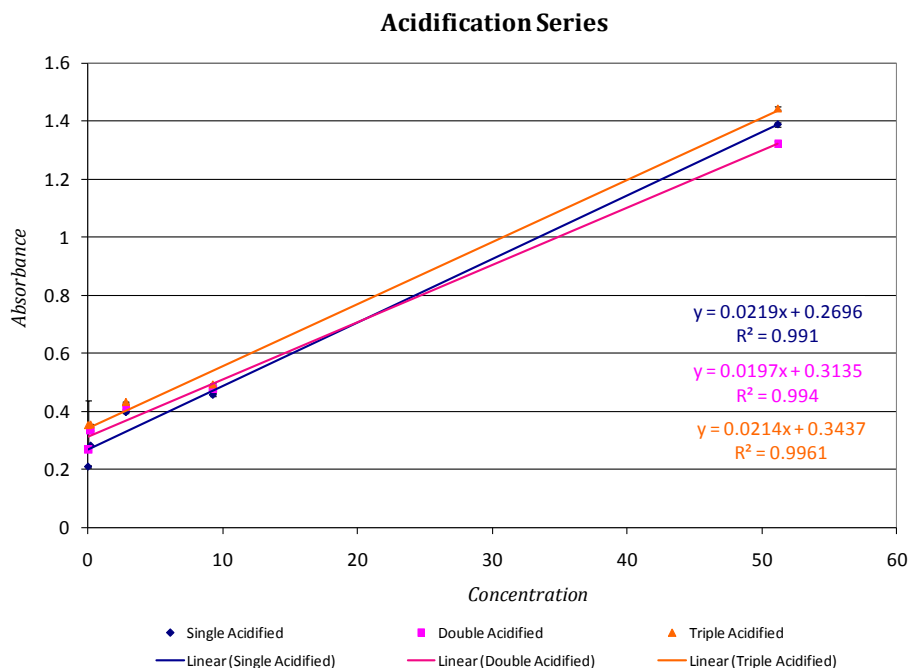


Figure 7: Acidification Series

A second test of acidification was conducted to be sure that the acidification line itself was not the cause of the offsets. For this test, a sample of approximately 20 nM was made up in unchelated seawater and allowed to sit for approximately 1 hour to allow for any iron to stick to the sides of the bottle. This sample was then run and acidified via the inline acidification system with 0.25, 0.10, and 0.05 N HCl. This sample was then transferred to a new bottle and acidified with 240  $\mu$ L 6N HCl and run with the same range of acids. There is still some variation in the absorbances however the magnitude of this variation is not large enough to completely explain the difference between discrete and underway measurements.

## Results

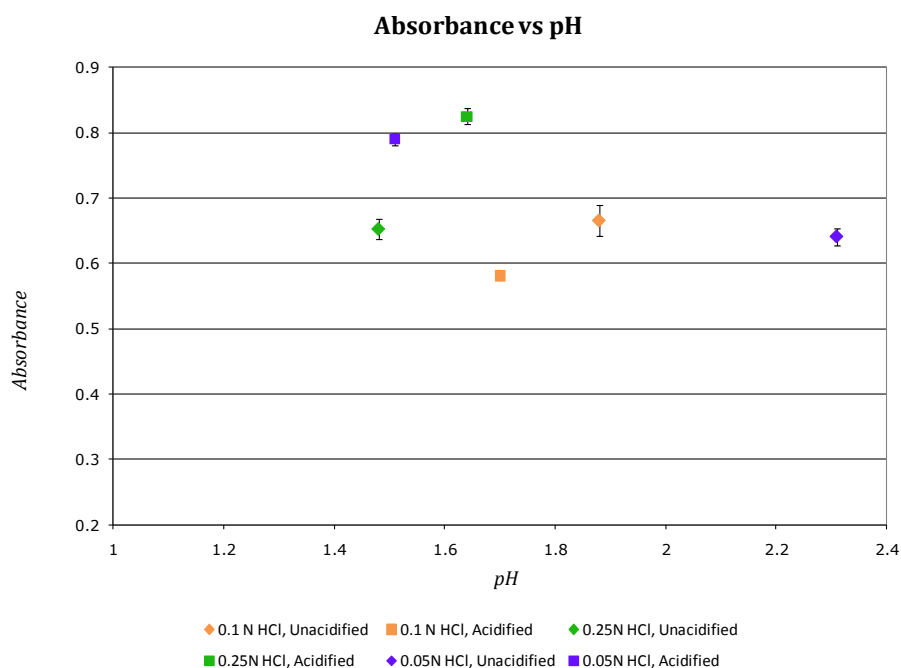


Figure 8: pH effects

### 4.1.3 Temperature

The effect of temperature was also tested as a possible source of the discrete and underway sample mismatch. This test was conducted by making up a 125 mL sample of approximately 20 nM. The sample was allowed to sit for approximately one hour for iron to stick to the sides of the bottle. The sample was then divided into four vials. The two of the vials were acidified and two were left unacidified. One of each sample was then placed into a 10°C chiller. The samples were then run with both 0.05 and 0.10 N HCl to control for the effects of pH.

The results of this test are shown in the figure below. There is a large variation between the samples acidified to the higher pH levels (pH 2.3, and pH 1.9) and the lower pH levels (pH 1.7 and pH 1.5) however in practice the sample is always acidified to at least pH 1.7. There is no significant trend between the sample run at 10°C and the sample run at room temperature. This is probably partially attributable to the acidification tubing being heated to 40°C which may minimize the temperature differences during the reaction. Within the pH 1.7 and pH 1.5 samples the variation is similar to that seen during the acidification tests pointing towards a yet undetermined source of variation.

## Results

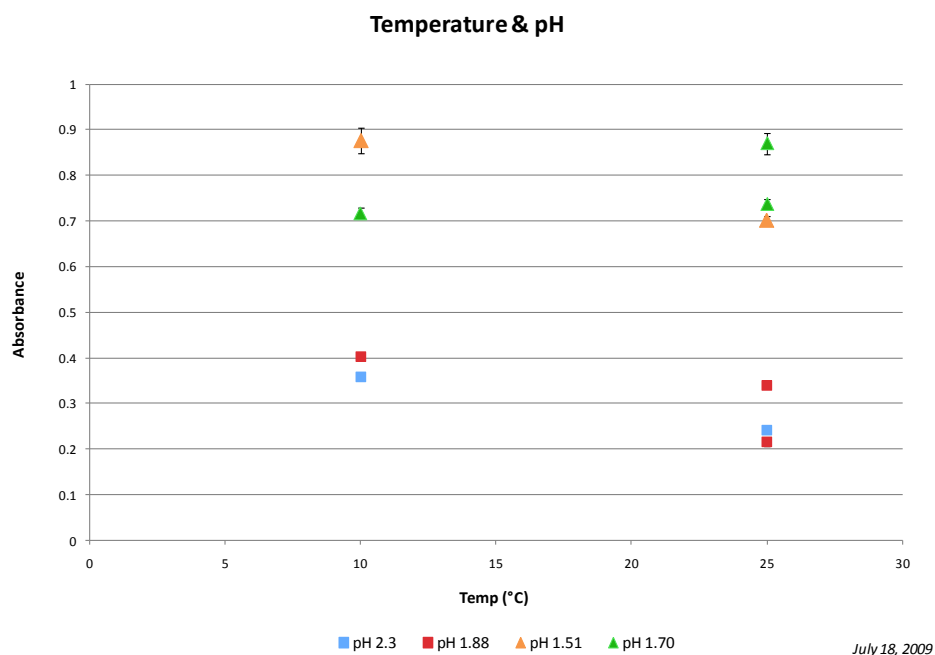


Figure 9: Temperature effects

### 4.2 Lab results

The discrete samples collected during SXS2 did not match the values measured during the underway analyses. *See section 4.3.1.1.3 (May 26).*

The above tests were conducted during June and July 2009 to determine the origin of the mismatch and to resolve the problem before the beginning of SXS3. The problem still appeared to be an issue (see cruise discussion in section 5).

## Results

### 4.2.1 Lab results files

**Table 1: Laboratory data files**

Date	File	Contents
6/13/2008	6132008.xlsx	Standard Curve, 5m plastic
6/16/2008	6162008.xlsx	Standard Curve, 5m plastic
6/24/2008	6242008.xlsx	Standard Curve, 5m plastic
6/25/2008	6252008.xlsx	Standard Curve, 5m plastic
6/30/2008	6302008.xlsx	Standard Curve, 4m plastic
7/1/2008	7012008.xlsx	Standard Curve, 4m plastic
7/7/2008	7072008.xlsx	Standard Curve, 3m plastic
7/10/2008	7102008.xlsx	Standard Curve, 4m plastic, 40°C
7/21/2008	7212008.xlsx	Standard Curve, 3m plastic
7/23/2008	7232008.xlsx	High-Low drift test
7/25/2008	7252008.xlsx	Standard Curve, 3m plastic
7/28/2008	7282008.xlsx	High-Low drift test
8/1/2008	8012008.xlsx	Standard Curve, 3m plastic
8/5/2008	8052008.xlsx	1m plastic coil tau estimation
8/6/2008	8062008.xlsx	2m plastic coil tau estimation
8/8/2008	8082008.xlsx	2m plastic high-low
8/11/2008	8112008.xlsx	3m plastic high-low
8/12/2008	8122008a.xlsx	4m plastic high-low
	8122008b.xlsx	5m plastic high-low
8/27/2008	8272008.xlsx	PreSXS1 acidification testing
11/2/2008	11022008.xlsx	Standard Curve
11/21/2008	11212008.xlsx	Glass tubing
1/5/2009	1052009.xlsx	Pump samples
2/9/2009	2092009.xlsx	High-low drift test
2/23/2009	2232009.xlsx	High-low drift test
3/2/2009	3022009.xlsx	Pump samples
4/13/2009	4132009.xlsx	Standard curve, 5m plastic
4/14/2009	4142009.xlsx	Standard curve, 3m plastic
4/23/2009	4232009.xlsx	No-bubble run
4/27/2009	4272009.xlsx	Manual bubble run
6/18/2009	6182009.xlsx	1x, 2x, 3x acidification test
6/22/2009	6222009.xlsx	SXS2 Samples
7/16/2009	7162009.xlsx	pH effect test
7/18/2009	7182009.xlsx	Temperature effect test
9/14/2009	9142009.xlsx	SXS3 samples
9/15/2009	9152009.xlsx	SXS3 samples
9/16/2009	9162009.xlsx	SXS3 samples
9/23/2009	9232009.xlsx	New vs cruise standards
10/7/2009	10072009.xlsx	Holm/Lakin standard comparison
10/8/2009	10082009.xlsx	Fluka vs Sigma DPD test
	HighLowComparisons.xlsx	Figure 6 and data

## Results

### 4.3 Cruise Results

*Internal lag times are estimated from the recorded time for changing to a new standard to the beginning of the fast initial change in voltage.*

#### 4.3.1 SXS2 (May) Cruise

##### 4.3.1.1 Daily Results

\*Times given in the notes section reflect the time the information was recorded.

##### 4.3.1.1.1 May 23—Transect 1, 45°N (SXS2\_xsct1)

A transect of 45°N was run on May 23. The system was started several hours before supersucker flow in order to improve bubble quality over results in port call. Some improvement were made during the standards run but most of the bubble progress was made during the early part of the days supersucker flow. The improvements included replacing the bpt links between the glass tubing sections in the acidification coil with Tygon which seemed to improve the bubbles as well as giving us a better view of what happened to the bubbles inside the tubing. It was also determined that a close fit of the glass tubing inside the Tygon connections is imperative to maintaining the bubbles inside the acidification coil so the connections were also changed to Tygon and adjusted in the reaction coil. The reaction coil was also shortened from 4 25-turn coils to 3 coils.

The run was interrupted by some valve switching problems related to the writing of new data files (the system was being run on valve position 2 but when a new file would start it would revert to position 1 and draw air). The system pulled large amounts of air at the end of the run and the second standards run was indecipherable.

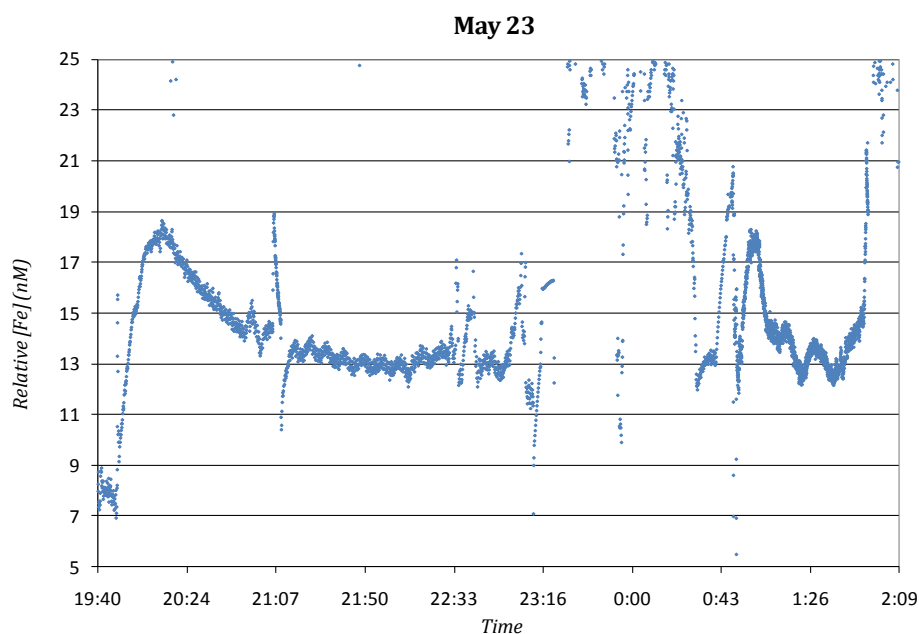


Figure 10: Relative Fe (SXS2\_xsct1)

## Results

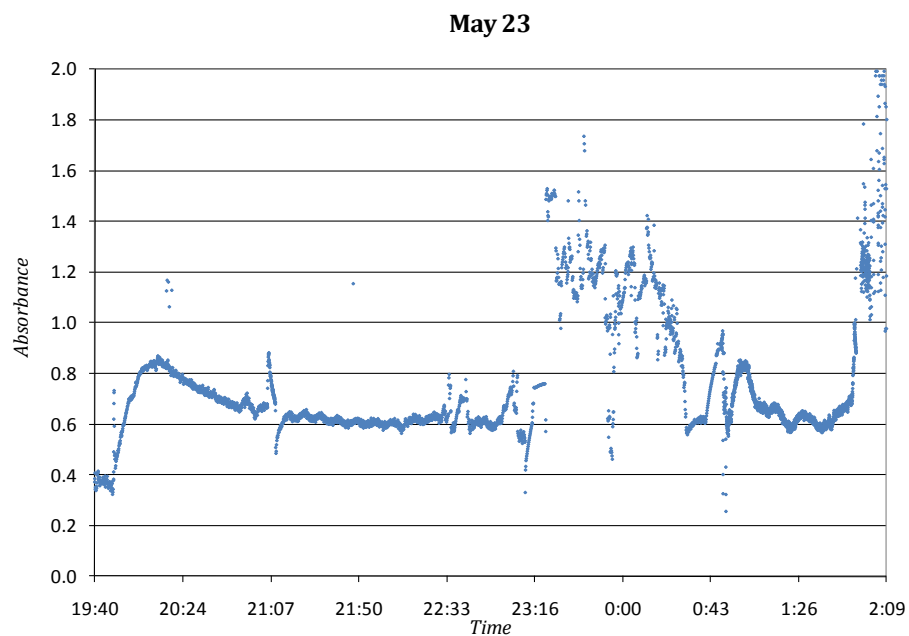


Figure 11: Absorbance (SXS2\_xsct1)

**Table 2: SXS2\_xsct1 summary**

	<i>Absorbance</i>	<i>Stdev</i>	<i>Drift (Abs 2-Abs1)</i>		<i>Detection limit</i>		<i>Slope</i>
MilliQ	0.5826	0.0186	n/a		0.92		0.0466
Std 1	0.6780	0.0143	n/a		(Std1)		
Std 2	0.7651	0.0196	n/a				
Std 3	1.6402	0.0461	n/a				
Std 4	3.0571	0.3232	n/a				
Average		0.0843					

**Internal Lag time:** ≈15 minutes

## Results

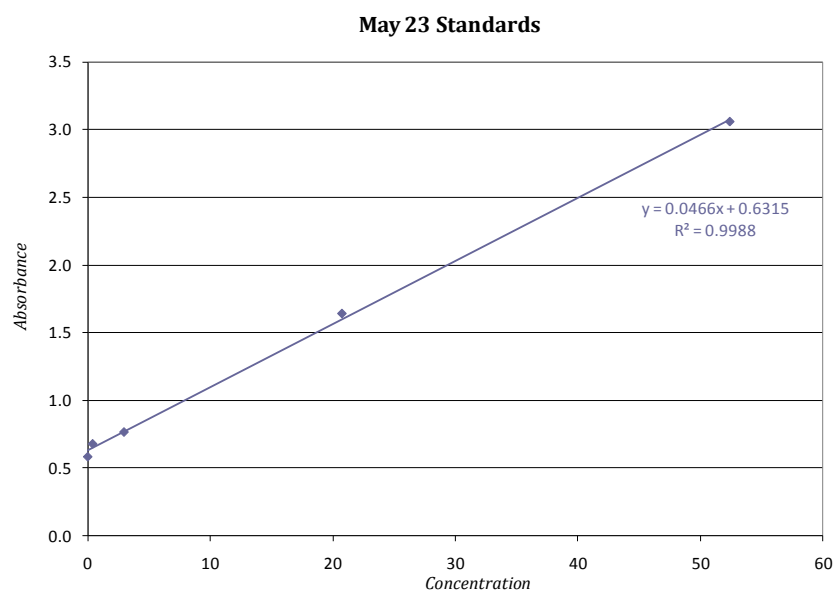


Figure 12: SXS2\_xsct1 standards

**Table 3: SXS2\_xsct1 standards**

<i>Standard</i>	<i>Concentration</i>	<i>Date</i>	<i>Time</i>	<i>Absorbance</i>	<i>Stdev</i>
MilliQ	0	5/23/09	17:44	0.5826	0.01856
Std 1	0.4	5/23/09	17:55	0.6778	0.01427
Std 2	2.96	5/23/09	18:05	0.7651	0.01963
Std 3	20.75	5/23/09	18:20	1.6402	0.04610
Std 4	52.41	5/23/09	18:41	3.0571	0.32318

## Results

**Table 4: SXS2\_xsct1 notes**

5/23/09	15:50	reagents running
5/23/09	17:10	replacing glass tubing links, improving bubbles
5/23/09	17:33	begin standards
5/23/09	19:08	air stuck in the line
5/23/09	19:26	begin supersucker transect (45°N); allow lines to flush ~10 minutes before running
5/23/09	19:35	begin supersucker flow to GCFA
5/23/09	20:50	accidental valve switch
5/23/09	22:13	noticed slow upward trend of voltage; there are small oscillations within the trend
5/23/09	22:39	acidification/reagents junction made more direct
5/23/09	23:34	changed to larger pump tubing for bubbles with inconclusive results; also changed to larger ID link between bubble injection and acidification line which seemed to make things worse so returned to smaller ID; perfected fits between glass tubing which seems to make the flow more even
5/24/09	0:52	changed to tygon links in the reaction coil; switched from 4 coils to 3
5/24/09	1:56	drawing air during standards; abandon run

### 4.3.1.1.2 May 25—Patch day 1 (SXS2\_srvy1)

May 25, 2009 was the first day of surveying the tracer patch during SXS2. The system was run with only three 25-turn glass coils in the acidification line and one 25-turn coil as the reaction line. Some experimentation was done with the temperature but increasing the temperature to 50°C only seemed to boost noise so it was returned to 40°C. Two discrete samples were run (#128 and #111) during the run. A vertical profile was conducted at the end of the day (5/26, 6:04) that included some surface pumping before shutting down.

Overall, the days run was very noisy. This is most likely related to bubble issues but could also be caused by contamination from the supersucker line or opening the system lines for adjustment.

## Results

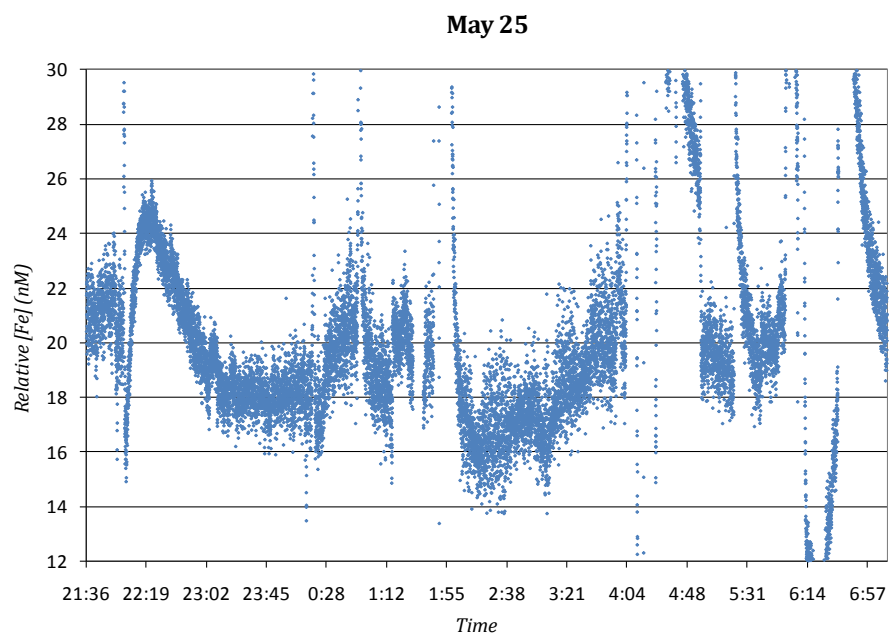


Figure 13: Relative [Fe] (SXS2\_srvy1)

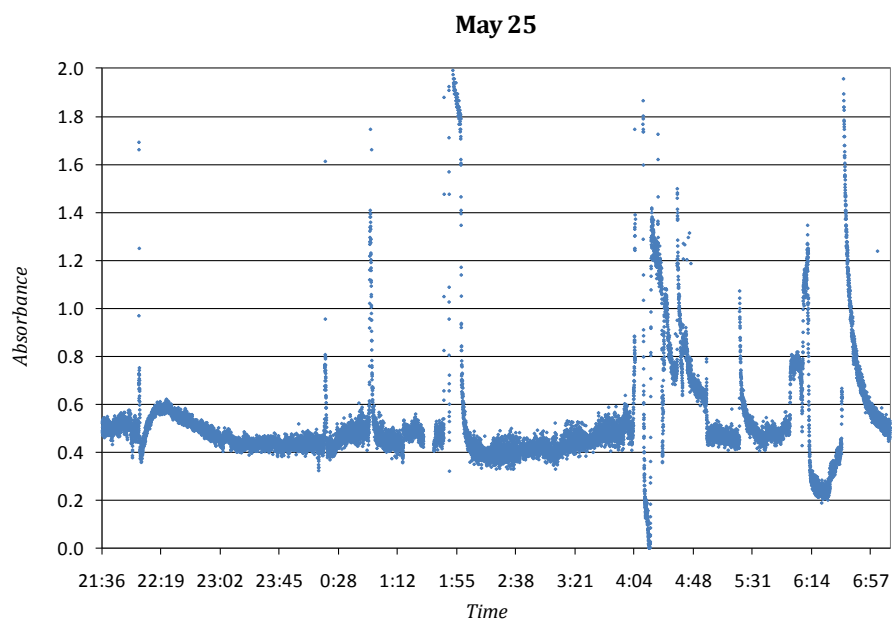


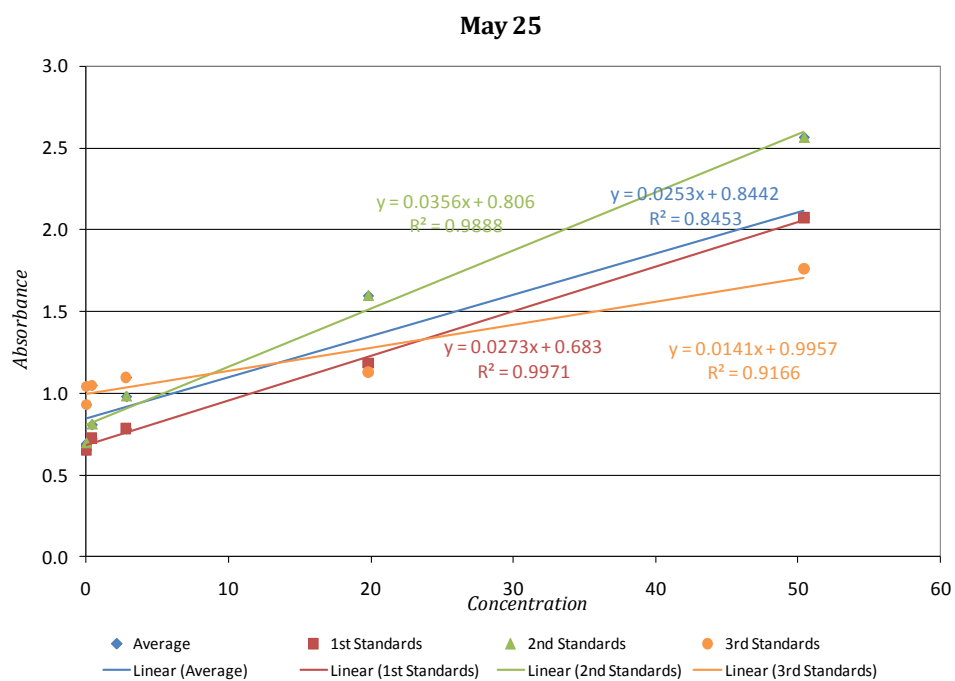
Figure 14: Absorbance (SXS2\_srvy1)

## Results

**Table 5: SXS2\_srvy1 standards**

Summary						
Standard	Absorbance	Stdev	Drift (Abs 3-Abs1)		Detection Limit:	Slope (avg)
MilliQ	0.8015	0.2796	0.2761		5.066	0.0239
Std 1	0.9286	0.2620	0.4102		(2st Std1)	Slope 1:
Std 2	0.9890	0.0468	0.3065			0.0273
Std 3	1.7363	0.1518	0.0520			Slope 2:
Std 4	2.0416	0.2648	-0.4264			0.0321
Avg		0.1813				Slope 3:
						0.0139

**Internal Lag time:**  $\approx 7$  minutes



**Figure 15: SXS2\_srvy1 standards**

## Results

**Table 6: SXS2\_srvy1 standards**

<i>Standard</i>	<i>Concentration</i>	<i>Date</i>	<i>Time</i>	<i>Absorbance</i>	<i>Stdev</i>
Std 1	0.4	5/25/2009	15:39	0.7248	0.04925
Std 3	19.8	5/25/2009	15:49	1.1840	0.03147
Std 2	2.81	5/25/2009	16:00	0.7831	0.04634
Std 4	50.41	5/25/2009	16:12	2.0746	0.06301
MilliQ	0	5/25/2006	16:26	0.6546	0.03609
MilliQ	0	5/25/2009	20:38	0.6947	0.03698
Std 3	19.8	5/25/2009	20:50	1.5953	0.05201
Std 1	0.4	5/25/2009	21:03	0.8091	0.03851
Std 4	50.41	5/25/2009	21:15	2.5615	0.07372
Std 2	2.81	5/25/2009	21:25	0.9807	0.05481
MilliQ	0	5/26/2009	5:10	1.0434	0.03640
Std 1	0.4	5/26/2009	5:37	1.0492	0.04120
Std 2	2.81	5/26/2009	5:47	1.0978	0.04550
Std 3	19.8	5/26/2009	5:53	1.1288	0.04339
Std 4	50.41	5/26/2009	6:02	1.7623	0.03743
MilliQ	0	5/26/2009	7:36	0.9307	0.69332

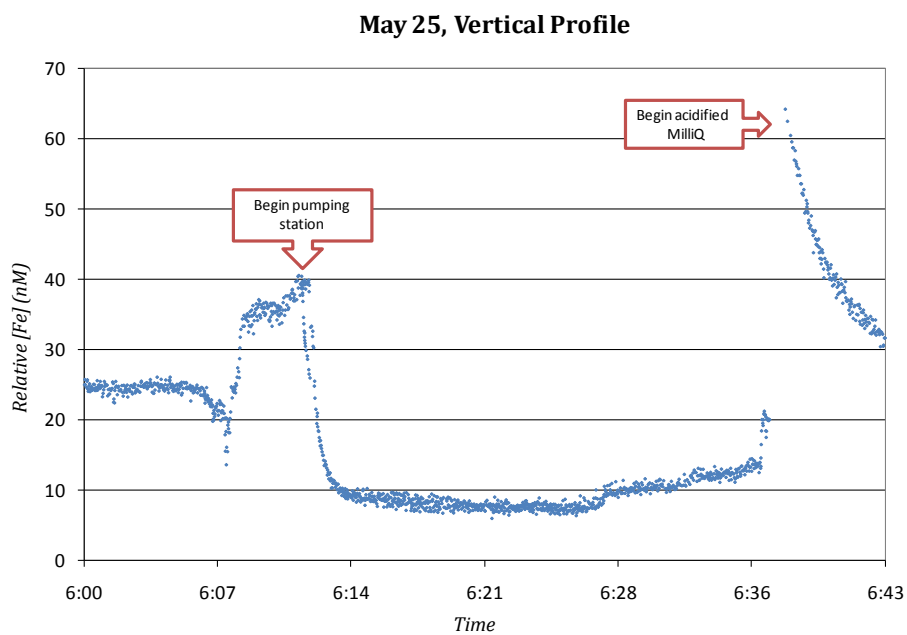
**Table 7: SXS2\_srvy1 discrete samples**

<b>Discrete Samples</b>							
<i>Date</i>	<i>Time</i>	<i>Sample</i>	<i>Abs</i>	<i>Blank Corr</i>	<i>Stdev</i>	<i>Conc.</i>	<i>Date run</i>
5/25/2009	19:13	SXS Consistency Standard					
5/25/2009	??	#111	0.9880	n/a	0.05280	39.52	5/25/2009
5/25/2009	22:31	#128	1.0190	n/a	0.03070	40.76	5/25/2009
5/26/2009	1:40	#119					
5/26/2009	6:25	#122					

## Results

**Table 8: SXS2\_srvy1 notes**

5/25/09	15:05	begin reagents
5/25/09	15:26	begin standards
5/25/09	17:01	increased temperature of acidification heater
5/25/09	17:20	temperature stabilized at 50°C
5/25/09	17:50	adjusted connections, switched to acid with brij
5/25/09	19:13	changed from 1.3 ID bubble pump tubing to 0.86ID
5/25/09	19:23	turned temperature back down to 40°C
5/25/09	20:28	increased temperature to 50°C
5/25/09	20:36	begin standards
5/25/09	21:56	begin supersucker flow
5/25/09	22:31	begin discrete samples #128 and #111
5/26/09	0:12	unfiltered line
5/26/09	0:43	filtered line
5/26/09	1:32	program froze
5/26/09	1:38	program restart
5/26/09	1:50	valve problem; fixed
5/26/09	4:13	begin standards
5/26/09	6:04	TFF vertical profile followed by surface test
5/26/09	6:28	acidified milliQ



**Figure 16: May 25th vertical profile**

## Results

### 4.3.1.1.3 May 26—Patch day 2 (SXS2\_srvy2)

May 26, 2009 the second day of patch surveying. The signal exhibited little to no variation and was very noisy. The TFF and unfiltered lines were both run. The standards were fairly consistent throughout the entire day. A discrete sample was collected and ran immediately after collection but the sample was off.

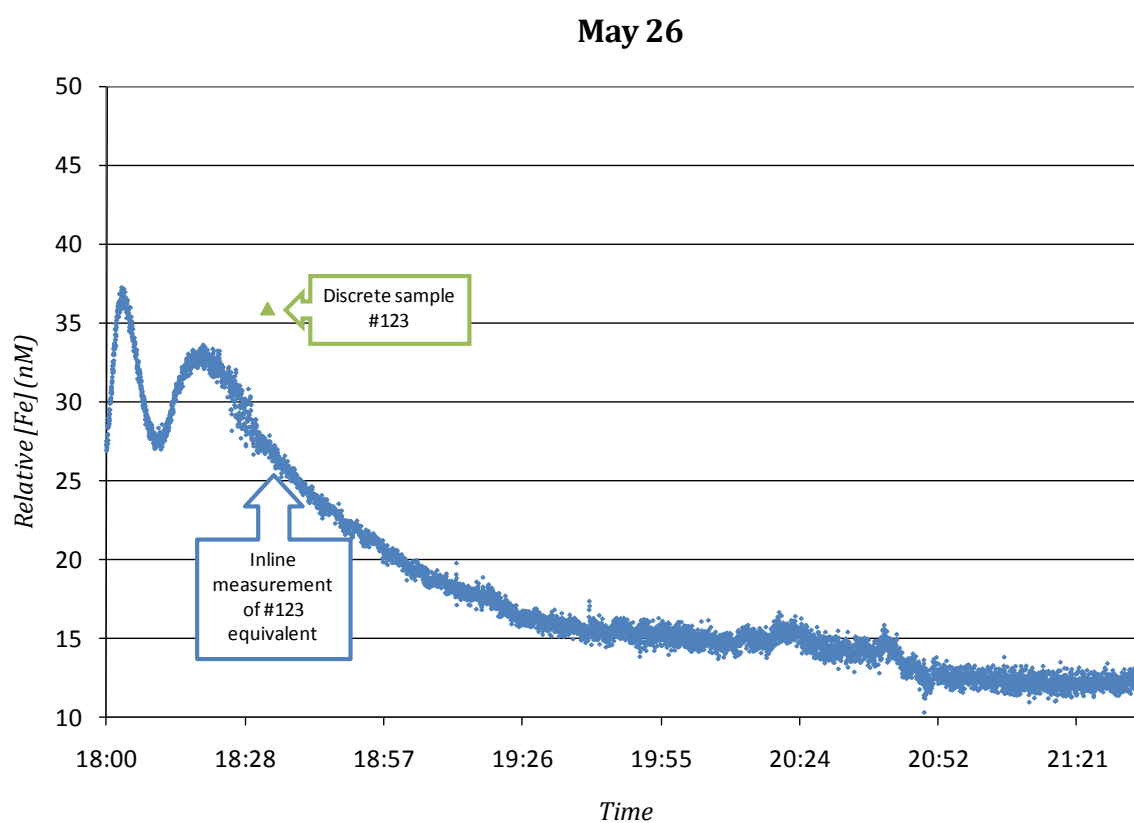


Figure 17: SXS2\_srvy2 discrete discrepancy

## Results

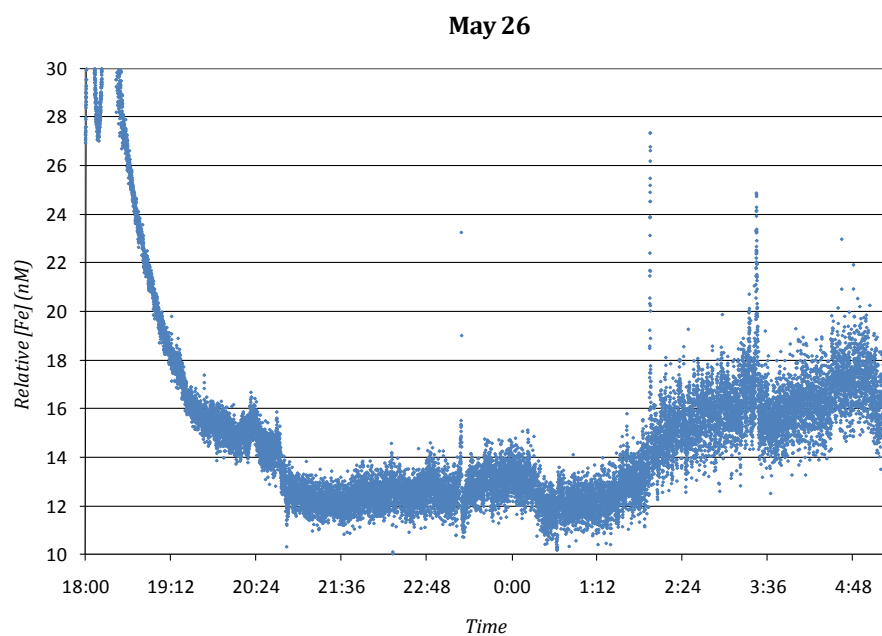


Figure 18: Relative [Fe] (SXS2\_srvy2)

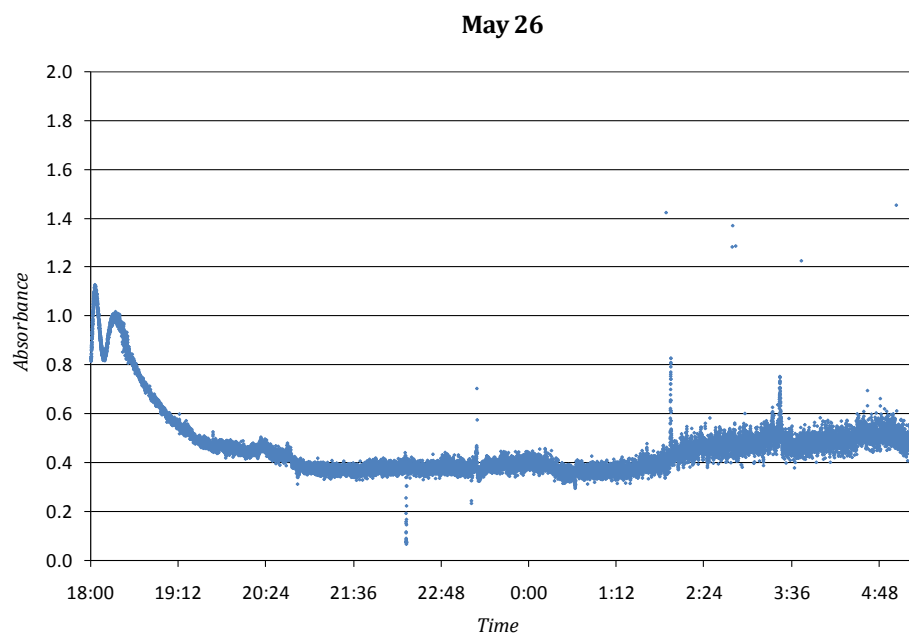


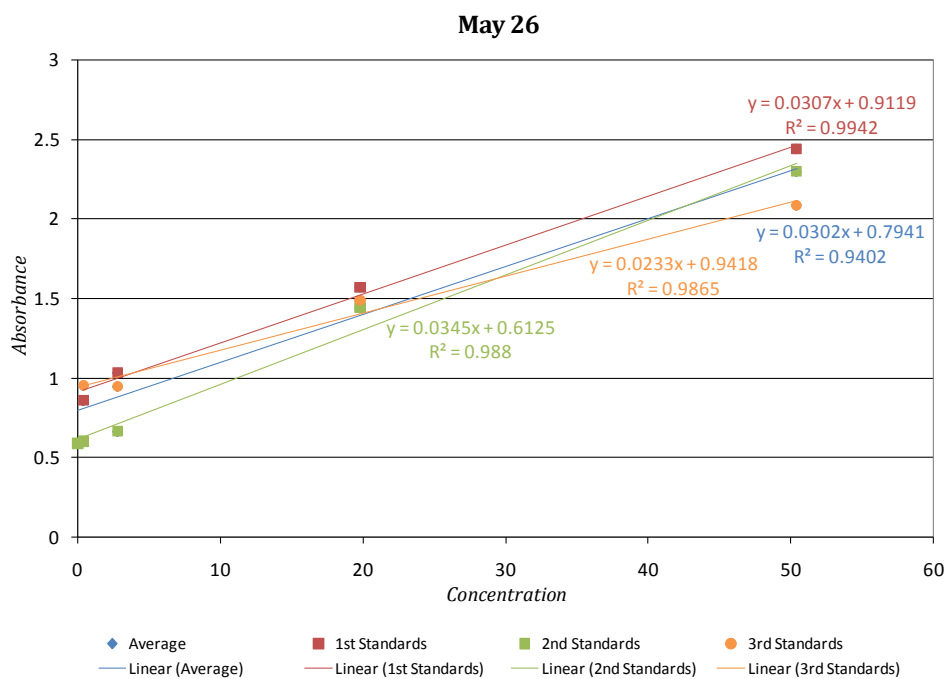
Figure 19: Absorbance (SXS2\_srvy2)

## Results

**Table 9: SXS2\_srvy2 summary**

	Absorbance	Stdev	Drift (Abs 3-Abs1)	Detection Limit:	Slope (avg)
MilliQ	0.5900	0.0449	n/a	4.46	0.0302
Std 1	0.8053	0.0449	-0.0363	(Std1 avg)	Slope 1:
Std 2	0.8825	0.1343	-0.0870		0.0207
Std 3	1.5004	0.0559	-0.0825		Slope 2:
Std 4	2.2742	0.1267	-0.3537		0.0342
Average		0.1127			Slope 3:
					0.0233

**Internal Lag time:**  $\approx 7$  minutes



**Figure 20: SXS2\_srvy2 standards**

## Results

**Table 10: SXS2\_srvy2 standards**

<i>Standard</i>	<i>Concentration</i>	<i>Date</i>	<i>Time</i>	<i>Absorbance</i>	<i>Stdev</i>
Std 3	19.8	5/26/09	14:12	1.5714	0.07189
Std 1	0.4	5/26/09	14:23	0.8585	0.06954
Std 4	50.41	5/26/09	14:32	2.4394	0.14349
Std 2	2.81	5/26/09	14:47	1.0345	0.33084
Std 1	0.4	5/26/09	16:33	0.6020	0.03182
MilliQ	0	5/26/09	16:57	0.5900	0.37923
Std 2	2.81	5/26/09	17:04	0.6654	0.03042
Std 3	19.8	5/26/09	17:18	1.4408	0.04760
Std 4	50.41	5/26/09	17:27	2.2975	0.13580
Std 1	0.4	5/27/09	7:40	0.9553	0.03324
Std 2	2.81	5/27/09	7:50	0.9475	0.04161
Std 3	19.8	5/27/09	8:04	1.4889	0.04835
Std 4	50.41	5/27/09	8:14	2.0857	0.10090

**Table 11: SXS2\_srvy2 discrete samples**

<i>Date</i>	<i>Time</i>	<i>Sample</i>	<i>Abs</i>	<i>Blank Corr</i>	<i>Stdev</i>	<i>Concentration</i>	<i>Date run</i>
5/26/09	18:27	#123	1.0843		0.02217	35.91	5/26/09
5/26/09	20:23	#113	1.3857		0.01494	76.32	6/22/09
5/26/09	21:25	#124	1.3617		0.01543	74.77	6/22/09
5/26/09		#131	1.3169		0.03264	71.88	6/22/09
5/26/09	22:39	#117	1.6391		0.02712	92.66	6/22/09
5/27/09	0:56	#115	1.2897		0.05613	70.12	6/22/09
5/27/09	0:01	#118	1.2689		0.01706	68.78	6/22/09
5/27/09	1:49	#120	1.2505		0.01518	67.59	6/22/09

**Table 12: SXS2\_srvy2 notes**

5/26/09	13:29	begin reagents
5/26/09	13:51	begin standards
5/26/09	14:39	running bulk acidified seawater while waiting for supersucker
5/26/09	16:21	standards
5/26/09	17:23	begin supersucker flow
5/26/09	18:27	running discrete sample #123
5/27/09	1:48	unfiltered line
5/27/09	3:20	TFF line
5/27/09	5:11	begin standards

## Results

### 4.3.1.1.4 May 27—Patch day 3 (SXS2\_srvy3)

May 27 the third day of patch surveying during SXS2. The day ran without many major issues beyond a program freeze. The system was alternated between the TFF and the unfiltered line. Overall, the system seems to have been more variable than usual despite the relatively low detection limit.

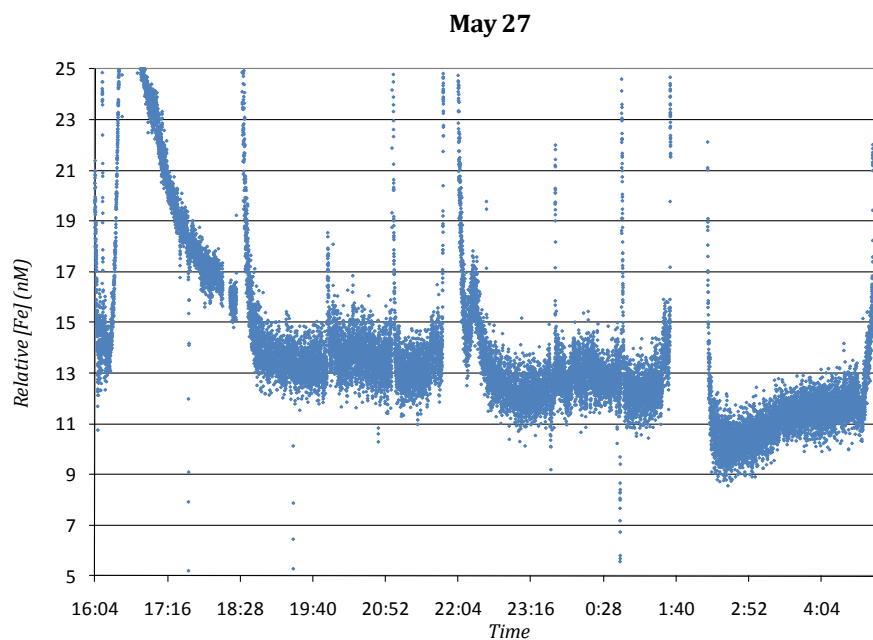


Figure 21: Relative [Fe] (SXS2\_srvy3)

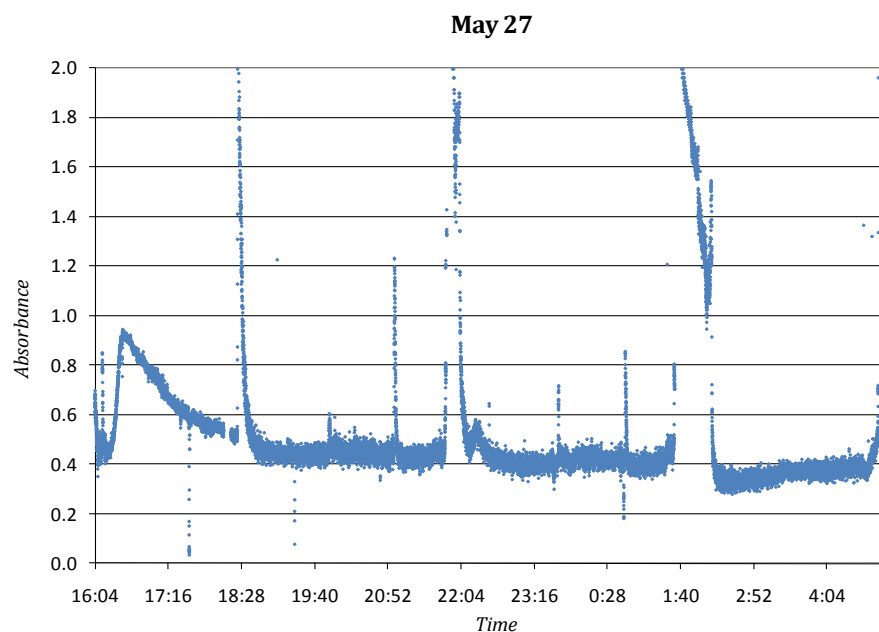


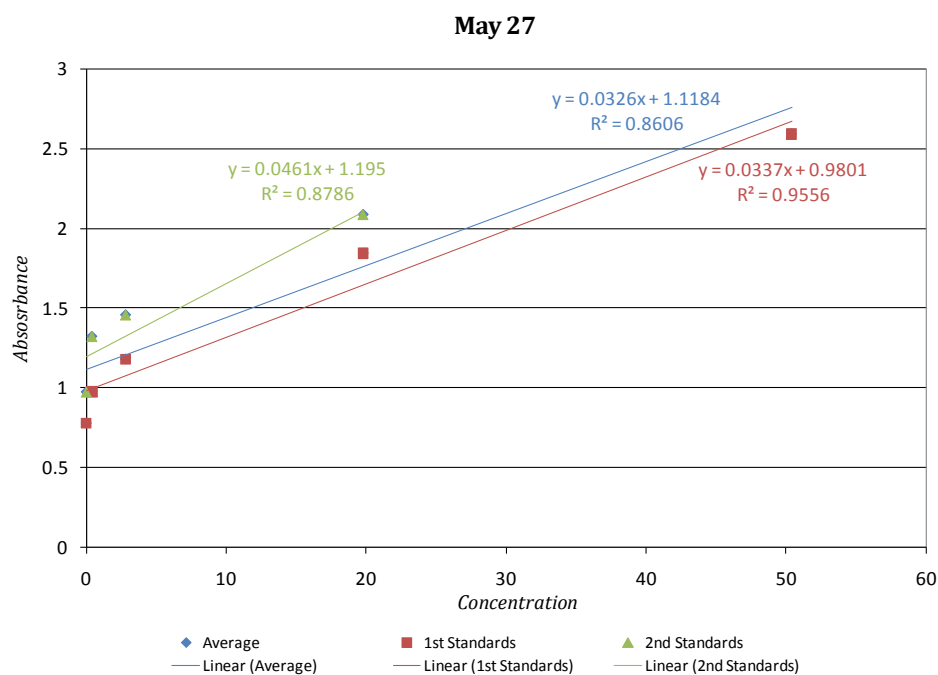
Figure 22: Absorbance (SXS2\_srvy3)

## Results

**Table 13: SXS2\_srvy3 summary**

	Absorbance	Stdev	Drift (Abs 2-Abs1)	Detection Limit:	Slope (avg)
MilliQ	0.8766	0.0686	0.1953	5.12	0.0326
Std 1	1.1487	0.0556	0.3472	(Std1 Avg)	Slope 1:
Std 2	1.3187	0.2153	0.2776		0.0337
Std 3	1.9657	0.1335	0.2427		Slope 2:
Std 4	2.5933	0.1356	2.5933		0.0461
Average		1.4681			

**Internal Lag time:  $\approx$  7 minutes**



**Figure 23: SXS2\_srvy3 standards**

**Table 14: SXS2\_srvy3 standards**

Standard	Concentration	Date	Time	Absorbance	Stdev
MilliQ	0	5/27/09	15:03	0.7789	0.03940
Std 2	2.81	5/27/09	15:20	1.1799	0.06014
Std 3	19.8	5/27/09	15:45	1.8443	0.19711
Std 4	50.41	5/27/09	15:55	2.5933	0.13557
Std 1	0.4	5/27/09	15:34	0.9751	0.05648
MilliQ	0	5/28/09	6:32	0.9742	0.09781
Std 1	0.4	5/28/09	6:58	1.3223	0.05471
Std 2	2.81	5/28/09	7:07	1.4575	0.37047
Std 3	19.8	5/28/09	7:15	2.0870	0.06989

## Results

**Table 15: SXS2\_srvy3 discrete samples**

<i>Date</i>	<i>Time</i>	<i>Sample</i>	<i>Abs</i>	<i>Blank Corr</i>	<i>Stdev</i>	<i>Concentration</i>	<i>Date run</i>
5/27/09	15:55	SXS Cons. Std	1.0488		0.05746	32.17	5/27/09
5/27/09	18:40	#34					
5/27/09	20:17	#31	1.3632		0.0123	74.86	6/22/09
5/27/09	21:28	#28	1.2414		0.0382	67.01	6/22/09
5/27/09	21:43	SXS Cons. Std					**no plateau
5/27/09	23:55	#27					
5/28/09	2:55	#38	1.1223		0.0119	59.32	6/22/09
5/28/09	4:43	#48					

**Table 16: SXS2\_srvy3 notes**

5/27/09	14:05	reagents flowing
5/27/09	14:42	standards
5/27/09	15:55	SXS Consistency Std
5/27/09	16:07	TFF
5/27/09	18:15	labview error
5/27/09	18:19	program restart
5/27/09	19:48	unfiltered line
5/27/09	20:52	TFF
5/27/09	21:43	SXS Consistency Std
5/27/09	21:58	TFF
5/27/09	23:35	unfiltered line
5/28/09	0:40	TFF
5/28/09	1:27	lab standards
5/28/09	2:05	TFF
5/28/09	4:48	standards

### 4.3.1.1.5 May 28—Patch day 4 (SXS2\_srvy4)

May 28 was the fourth day of surveying the tracer patch during SXS2. The run showed very little variation in concentration which may be partially attributable to the deterioration of the bubbles throughout the day (the bubble deterioration may have been caused by aging pump tubing for bubble injection). NASS-5 was also run twice as a benchmark for the day.

## Results

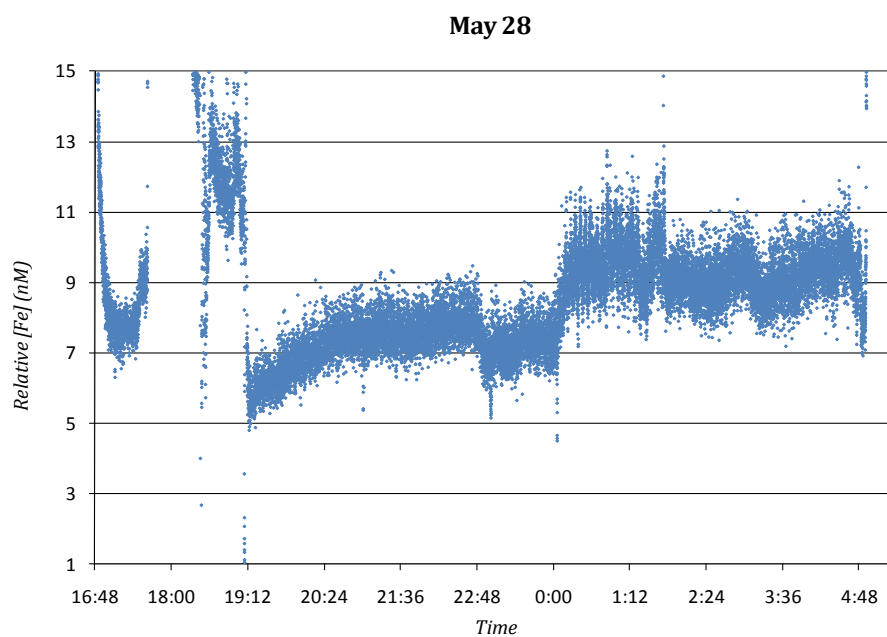


Figure 24: Relative [Fe] (SXS2\_srvy4)

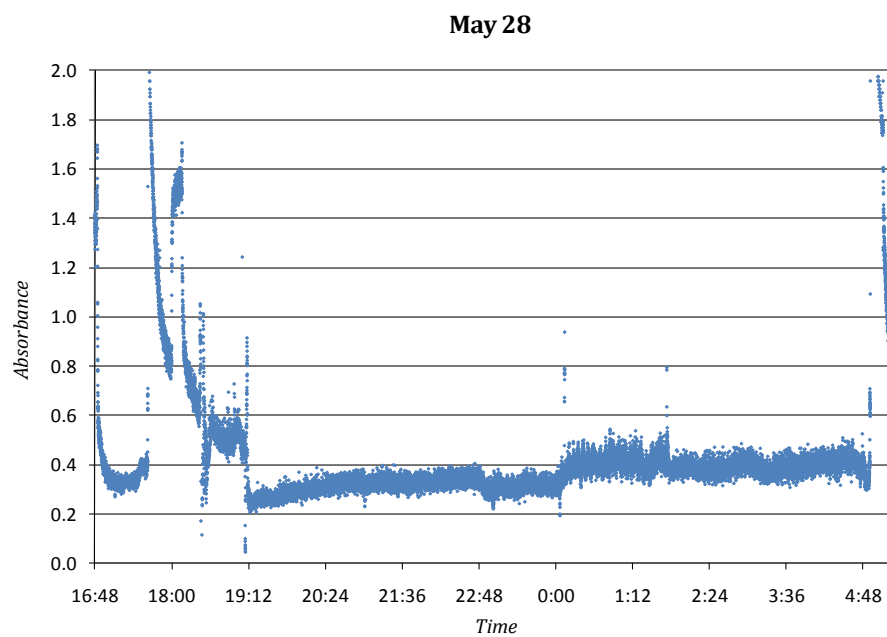


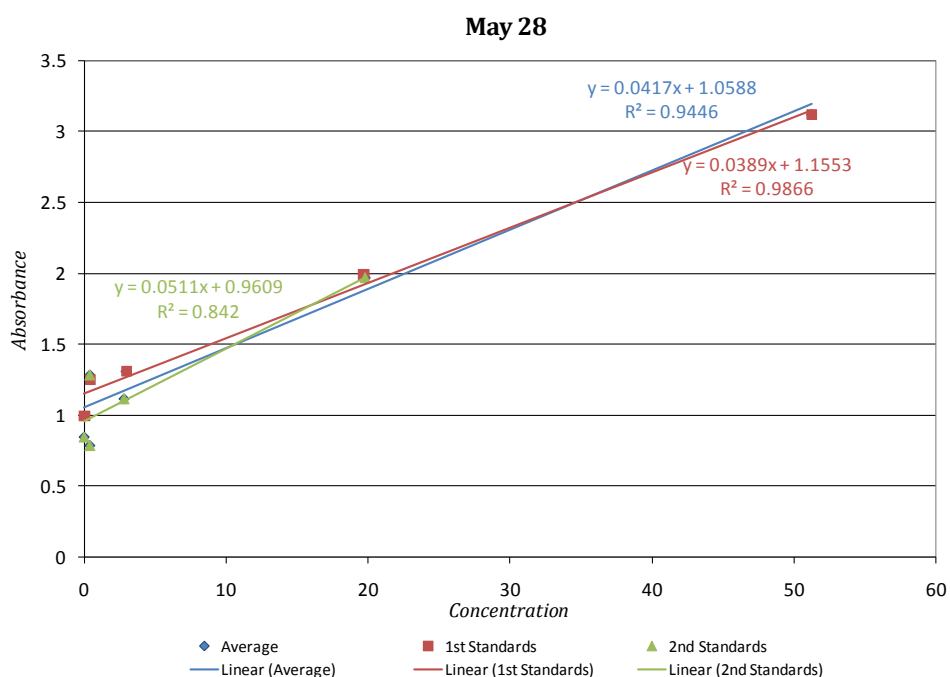
Figure 25: Absorbance (SXS2\_srvy4)

## Results

**Table 17: SXS2\_srvy4 summary**

	Absorbance	Stdev	Drift (Abs 2-Abs1)		Detection Limit:		Slope (avg)
MilliQ	0.9210	0.05428	-0.149		3.92		0.0417
Std 1	1.2679	0.05031	0.031		(Std1 Avg)		Slope 1:
Std 2	1.0476	0.04141	-0.523				0.0389
Std 3	1.9796	0.05297	-0.021				Slope 2:
Std 4	2.1170	0.07288	-2.003				0.0511
Average		0.05437					

**Internal Lag time:**  $\approx 7$  minutes



**Table 18: SXS2\_srvy4 standards**

Standard	Concentration	Date	Time	Absorbance	Stdev
MilliQ	0	5/28/09	15:47	0.9954	0.08548
Std 1	0.41	5/28/09	16:08	1.2524	0.05451
Std 2	2.95	5/28/09	16:16	1.3089	0.06036
Std 3	19.67	5/28/09	16:39	1.9899	0.06076
Std 4	51.24	5/28/09	16:47	3.1185	0.09286
Std 1	0.4	5/29/09	6:40	1.2834	0.04611
Std 3	19.8	5/29/09	6:58	1.9692	0.04518
Std 4	2.81	5/29/09	7:19	1.1154	0.05290
Std 2	0.4	5/29/09	7:36	0.7863	0.02247
MilliQ	0	5/29/09	8:31	0.8466	0.02308

## Results

**Table 19: SXS2\_srvy4 discrete samples**

<i>Date</i>	<i>Time</i>	<i>Sample</i>	<i>Abs</i>	<i>Blank Corr</i>	<i>Stdev</i>	<i>Concentration</i>	<i>Date run</i>
5/28/09	17:08	#40					
5/28/09	18:50	NASS-5	1.2315		0.07870	4.21	5/28/2009
5/29/09	4:46	#44					
5/29/09	5:50	NASS-5	1.9605		0.07301	21.99	5/29/2009

**Table 20: SXS2\_srvy4 notes**

5/28/09	14:25	begin standards
5/28/09	16:44	begin TFF flow
5/28/09	22:49	bubbles deteriorate; need to replace tubing
5/29/09	0:01	unfiltered line
5/29/09	1:36	TFF
5/29/09	5:10	bulk seawater
5/29/09	5:50	standards

### 4.3.1.1.6 May 29—Patch day 5 (SXS2\_srvy5)

May 29 was the final patch tracing day for SXS2. The system was run with little interruption except for a supersucker problem about 22:43. The system was switched to the unfiltered line between 00:36 and 3:50 but otherwise ran uneventfully on the TFF.

Variations in iron were measured throughout the transect, particularly at the end of the day. The dual slow increases of iron (one on either side of the supersucker problem at 22:43) are interesting to note. They could indicate some sort of iron accumulation issue either in our system or in the supersucker.

## Results

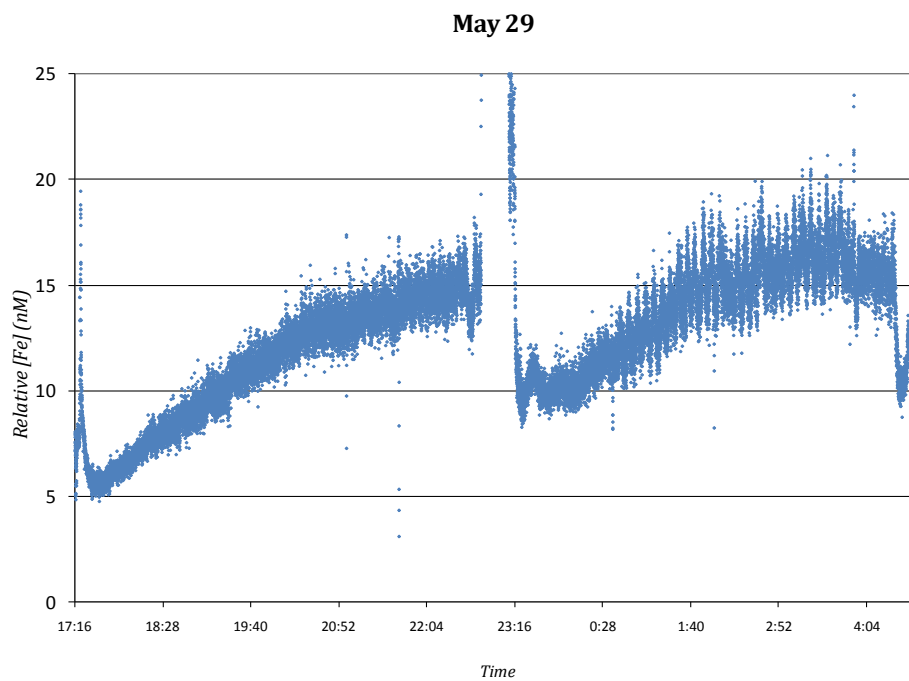


Figure 27: Relative [Fe] (SXS2\_srvy5)

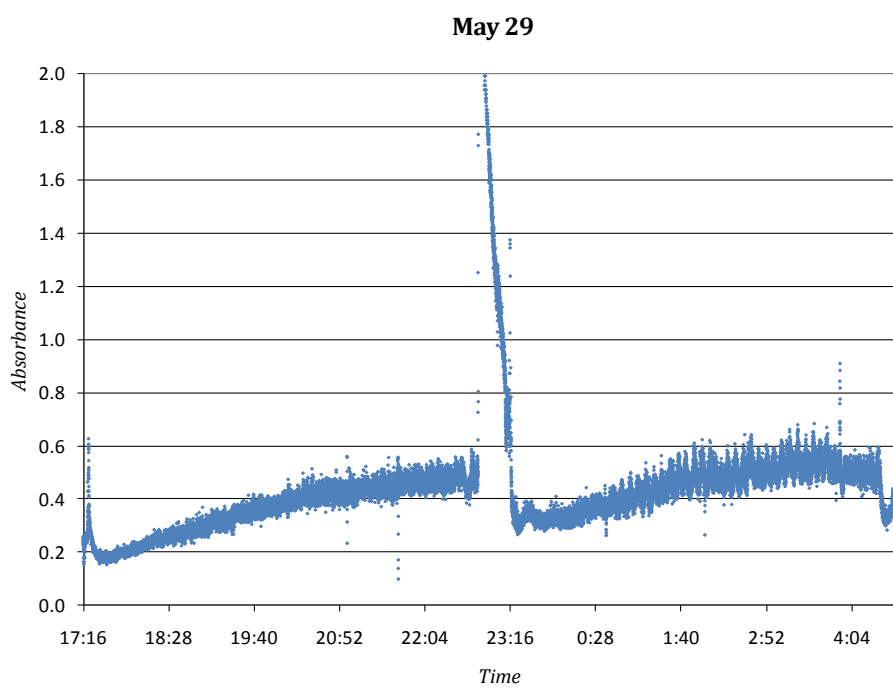


Figure 28: Absorbance (SXS2\_srvy5)

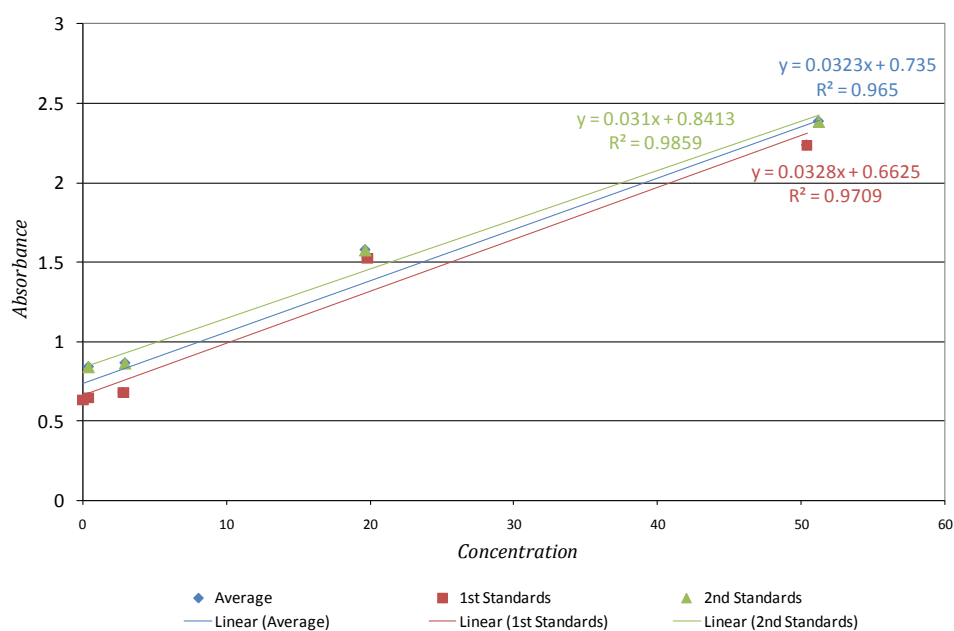
## Results

**Table 21: SXS2\_srvy5 summary**

	Abs	Stdev	Drift (Abs 2-Abs1)		Detection Limit:		Slope (avg)
MilliQ	0.6322	0.0225	n/a		2.21		0.0323
Std 1	0.7435	0.2151	0.1945		(Std1 Avg)		Slope 1:
Std 2	0.7720	0.0254	0.1851				0.0328
Std 3	1.5516	0.0461	0.0500				Slope 2:
Std 4	2.3104	0.0576	0.1496				0.031
Average		0.0365					

**Internal Lag time:** ≈ 5 minutes

**May 29**



**Figure 29: SXS2\_srvy5 standards**

**Table 22: SXS2\_srvy5 standards**

Standard	Concentration	Date	Time	Absorbance	Stdev
Std 1	0.4	5/29/09	16:18	0.6463	0.02022
Std 2	2.81	5/29/09	16:29	0.6795	0.01827
Std 3	19.8	5/29/09	16:42	1.5266	0.04547
Std 4	50.41	5/29/09	16:52	2.2356	0.06035
MilliQ	0	5/29/09	17:10	0.6322	0.02251
Std 4	51.24	5/30/09	7:10	2.3852	0.05492
Std 3	19.67	5/30/09	7:21	1.5766	0.04682
Std 2	2.95	5/30/09	7:44	0.8646	0.03249
Std 1	0.41	5/30/09	8:10	0.8407	0.02743

## Results

**Table 23: SXS2\_srvy5 discrete samples**

Date	Time	Sample	Abs	Blank Corr	Stdev	Concentration	Date run
5/30/09	4:38	Fe Check, surface					

**Table 24: SXS2\_srvy5 notes**

5/29/09	15:53	begin standards
5/29/09	17:16	TFF
5/29/09	22:43	bulk seawater
5/29/09	23:11	TFF
5/30/09	0:36	unfiltered line
5/30/09	3:50	TFF
5/30/09	4:43	bulk seawater
5/30/09	6:59	begin standards

### 4.3.1.1.7 May 30--45°N Transect (SXS2\_xsct2)

The May 30<sup>th</sup> data is a transect of 45°N from the 31m isobath to the 250m isobaths. This transect shows encouraging changes in the iron concentration that match the movements of the supersucker in the water column. NASS-5 was run twice as a benchmark during this run.

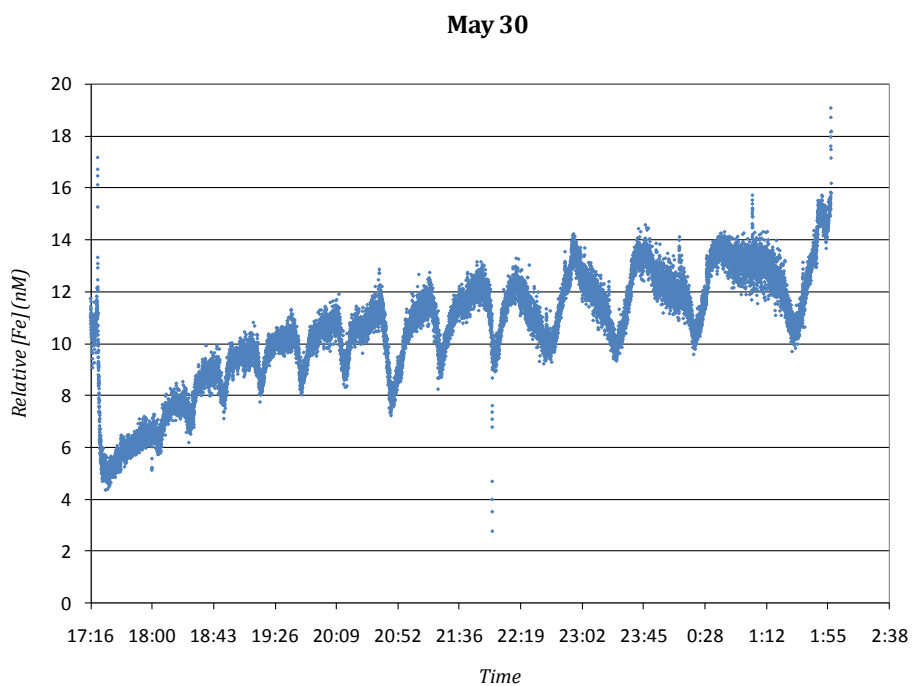


Figure 30: Relative [Fe] (SXS2\_xsct2)

## Results

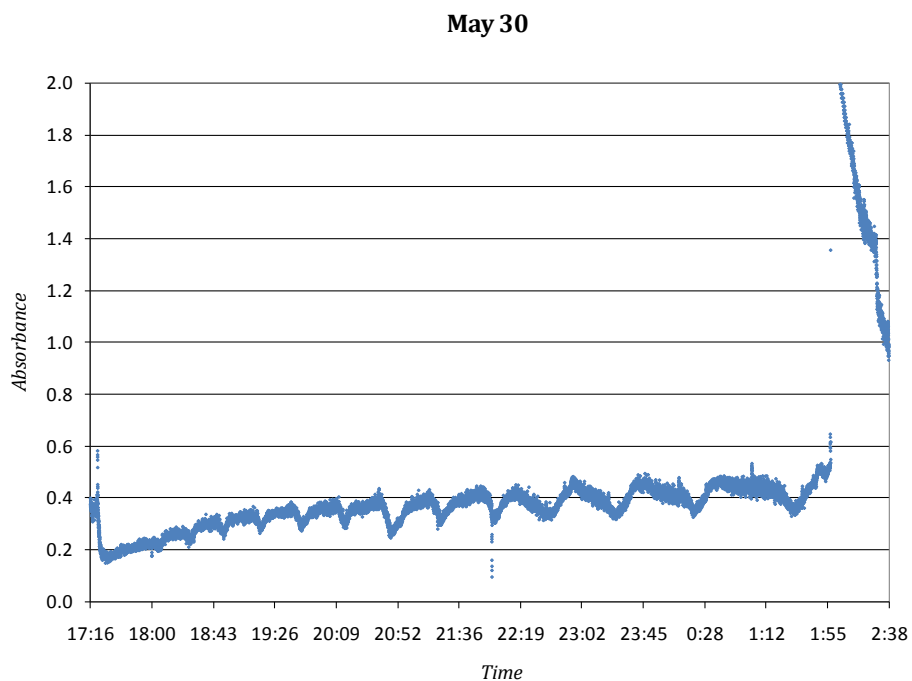


Figure 31: Absorbance (SXS2\_xsct2)

**Table 25: SXS2\_xsct2 summary**

	<i>Absorbance</i>	<i>Stdev</i>	<i>Drift (Abs 2-Abs1)</i>	<i>Detection Limit:</i>	<i>Slope (avg)</i>
MilliQ	0.6003	0.0167	-0.0681	2.22	0.0338
Std 1	1.0015	0.0250	0.1136	<i>(Std1 Avg)</i>	<i>Slope 1:</i>
Std 2	0.9992	0.0376	0.1817		0.0305
Std 3	1.7457	0.0532	0.1737		<i>Slope 2:</i>
Std 4	2.5225	0.0989	0.4041		0.0372
<i>Average</i>		0.0463			

**Internal Lag time:** ≈ 6 minutes

## Results

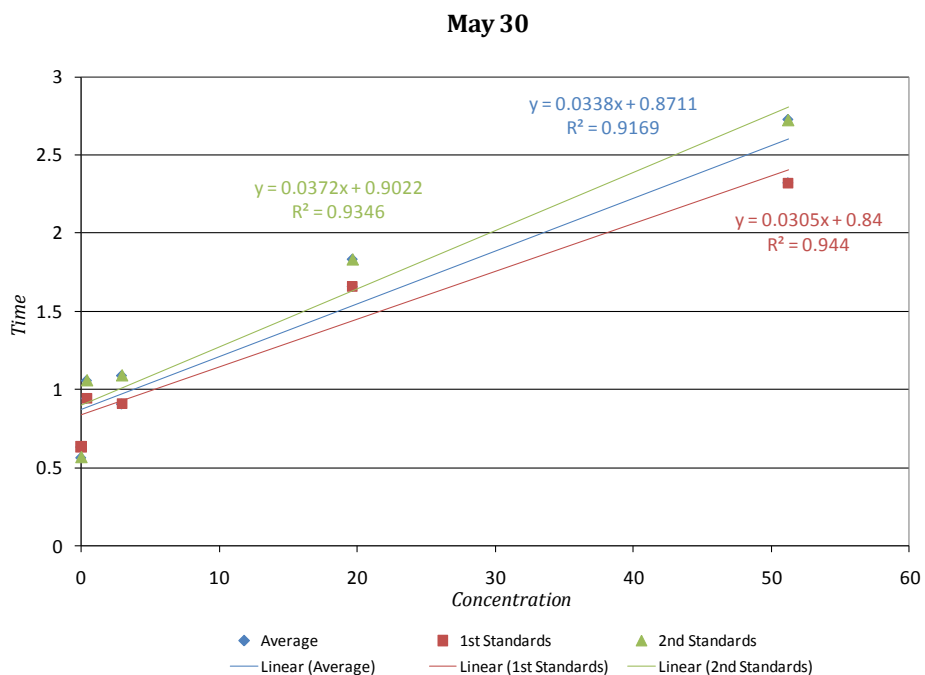


Figure 32: SXS2\_xsct2 standards

**Table 26: SXS2\_xsct2 standards**

Standard	Concentration	Date	Time	Absorbance	Stdev
MilliQ	0	5/30/09	16:04	0.6343	0.01782
4	51.24	5/30/09	16:12	2.3204	0.05724
3	19.67	5/30/09	16:26	1.6588	0.05546
2	2.95	5/30/09	16:56	0.9083	0.04091
1	0.41	5/30/09	17:09	0.9447	0.03422
4	51.24	5/31/09	2:46	2.7245	0.14054
3	19.67	5/31/09	3:02	1.8325	0.05092
2	2.95	5/31/09	3:26	1.0900	0.03438
1	0.41	5/31/09	3:44	1.0583	0.01581
MilliQ	0	5/31/09	4:46	0.5662	0.01607

**Table 27: SXS2\_xsct2 discrete samples**

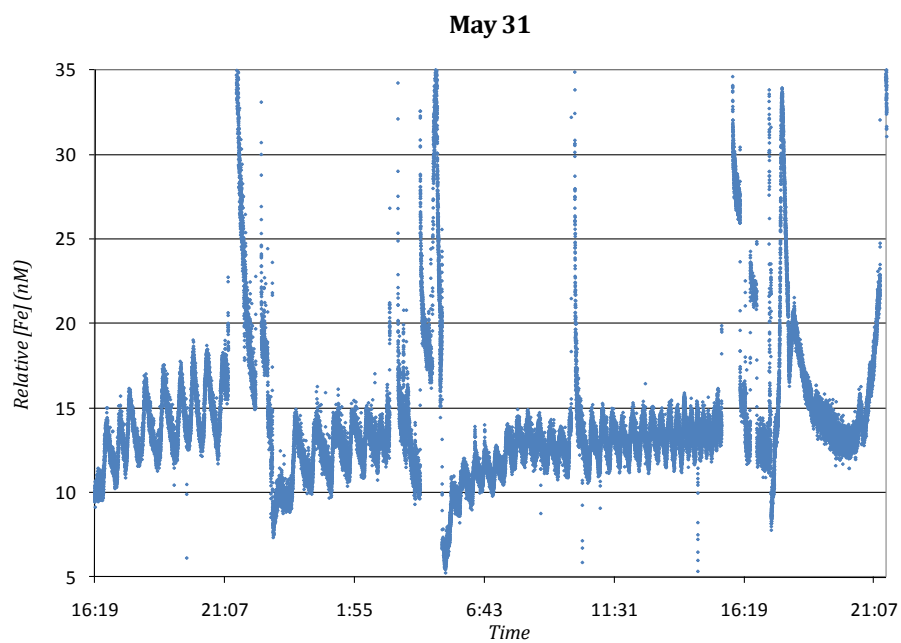
Date	Time	Sample	Abs	Blank Corr	Stdev	Conc.	Date run
5/30/09	17:05	NASS-5	0.8250	n/a	0.03673	1.09	5/20/09
5/31/09	3:31	NASS-5	1.0686	n/a	0.02478	0.73	5/31/09

## Results

**Table 28: SXS2\_xsct2 notes**

5/30/09	15:38	milliQ valve comparison
5/30/09	16:03	begin standards
5/30/09	17:05	NASS-5
5/30/09	17:16	TFF
5/31/09	1:51	begin standards
5/31/09	3:19	NASS-5

### 4.3.1.1.8 May 31—43.9°N Transect (SXS2\_xsct3)



**Figure 33: Relative [Fe] (SXS2\_xsct3)**

## Results

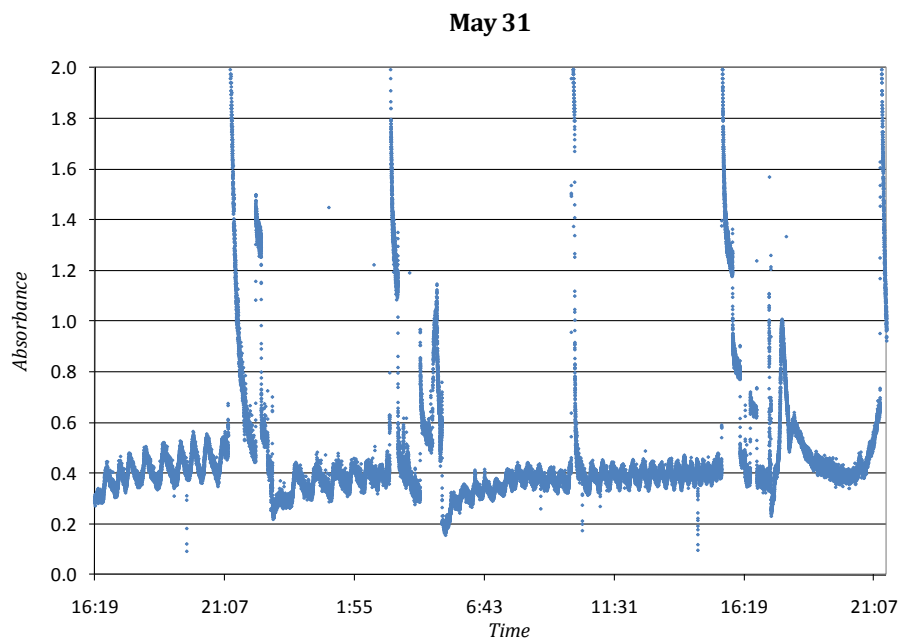


Figure 34: Absorbance (SXS2\_xsct3)

**Table 29: SXS2\_xsct3 summary**

	<i>Absorbance</i>	<i>Stdev</i>	<i>Drift (Abs 2-Abs1)</i>	<i>Detection Limit:</i>	<i>Slope (avg)</i>
MilliQ	n/a	n/a		6.17	0.0297
Standard 1	0.9796	0.0611	0.8408		<i>Slope 1:</i>
Standard 3	1.1540	0.0421	0.7458		0.0212
Standard 4	2.5178	0.0869	1.2485		<i>Slope 2:</i>
<i>Average</i>		0.0634			0.0417
					<i>Slope 3:</i>
					0.0301
					<i>Slope 4:</i>
					0.0003

**Internal Lag time:** ≈ 5 minutes

## Results

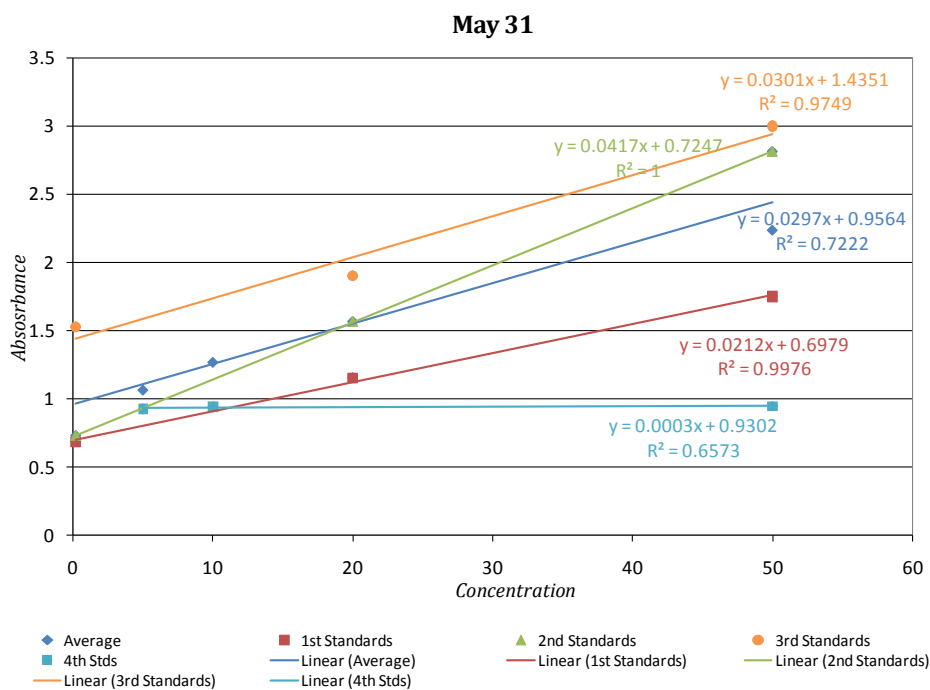


Figure 35: SXS2\_xsct3 standards

Table 30: SXS2\_xsct3 standards

**Standards made onboard					
Standard	Conc.	Date	Time	Absorbance	Stdev
Std 3	20	5/31/2009	15:33	1.1528	0.04191
Std 1	0.2	5/31/2009	15:44	0.6840	0.02932
Std 4	50	5/31/2009	16:04	1.7480	0.12267
Std 4	50	6/1/2009	3:25	2.8090	0.07139
Std 1	0.2	6/1/2009	4:33	0.7298	0.13557
Std 3	20	6/1/2009	5:01	1.5646	0.04382
Std 4	50	6/1/2009	15:44	2.9965	0.06667
Std 3	20	6/1/2009	16:02	1.8986	0.04062
Std 1	0.2	6/1/2009	16:36	1.5248	0.01847
consistency seawater + 5nM	5	6/1/2009	22:14	1.0617	0.01931
consistency seawater + 10nM	10	6/1/2009	22:33	1.265	0.033044
consistency seawater + 50nM		6/1/2009	22:42	2.2322	0.16475

## Results

**Table 31: SXS2\_xsct3 discrete samples**

<i>Date</i>	<i>Time</i>	<i>Sample</i>	<i>Abs</i>	<i>Blank Corr</i>	<i>Stdev</i>	<i>Conc.</i>	<i>Date run</i>
5/31/2009	15:33	NASS-5	0.60845	n/a	0.020408	18.66	5/31/2009
6/1/2009	3:24	SXS Consis. Std.	0.04054	n/a	0.04054	1.24	6/1/2009
5/31/2009	21:11	TFF sample	2.1839	n/a	0.0723	66.99	6/1/2009
6/1/2009	16:05	NASS-5	1.0612	n/a	0.024388	32.55	6/1/2009
6/1/2009	16:26	SXS Consis. Std.	0.89321	n/a	0.045277	27.40	6/1/2009
6/1/2009	16:42	SXS Consis. Std.	0.84425	n/a	0.025607	25.90	6/1/2009
6/1/2009	22:42	NASS-5	0.6881	n/a	0.021934	21.11	6/1/2009

**Table 32: SXS2\_xsct3 notes**

5/31/2009	14:35	reagents flowing
5/31/2009	15:02	heaters weren't on
5/31/2009	16:02	TFF
5/31/2009	21:05	acidified MilliQ
5/31/2009	22:48	TFF
6/1/2009	3:05	standard, const. std.; reagent top off
6/1/2009	5:04	TFF
6/1/2009	10:00	brief switch to position 1, no signal
6/1/2009	15:48	standards
6/1/2009	17:15	TFF
6/1/2009	18:45	DPD top off
6/1/2009	21:15	standards

### 4.3.1.2 Discussion

SXS 2 highlighted the need to resolve issues between the discrete samples and the inline data before SXS 3. It also raised concerns that the system may not be responding rapidly enough to changes in concentration. The bubble irregularities were believed to be the root cause of the slow response as well as creating unacceptable amounts of noise in the data.

**Table 33: SXS 2 Summary**

<i>Date</i>	<i>What?</i>	<i>Slope</i>	<i>R<sup>2</sup></i>	<i>Detection limit</i>
23-May	45N transect	0.0466	0.999	0.92
25-May	Patch	0.0237	0.997	5.07
26-May	Patch	0.0302	0.940	4.46
27-May	Patch	0.0326	0.861	5.12
28-May	Patch	0.0427	0.945	3.92
29-May	Patch	0.0323	0.965	2.21
30-May	45N transect	0.0338	0.917	2.22
31-May	43.9N transect	.0297	.722	6.17

## Results

### 4.3.1.2.1 High Resolution Data SXS2

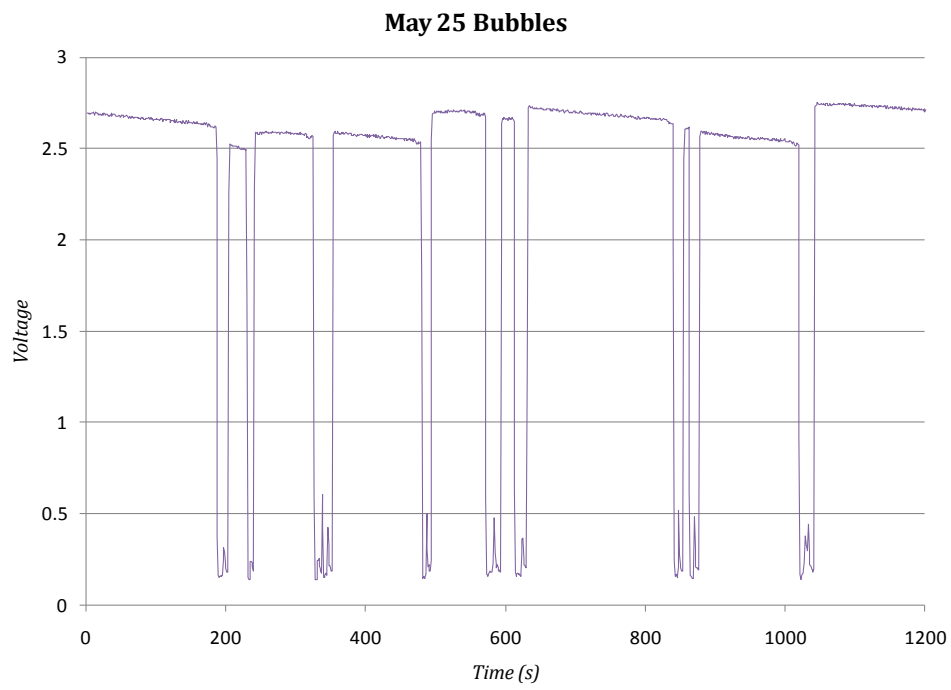


Figure 36: Early SXS2 bubble pattern (20 minutes)

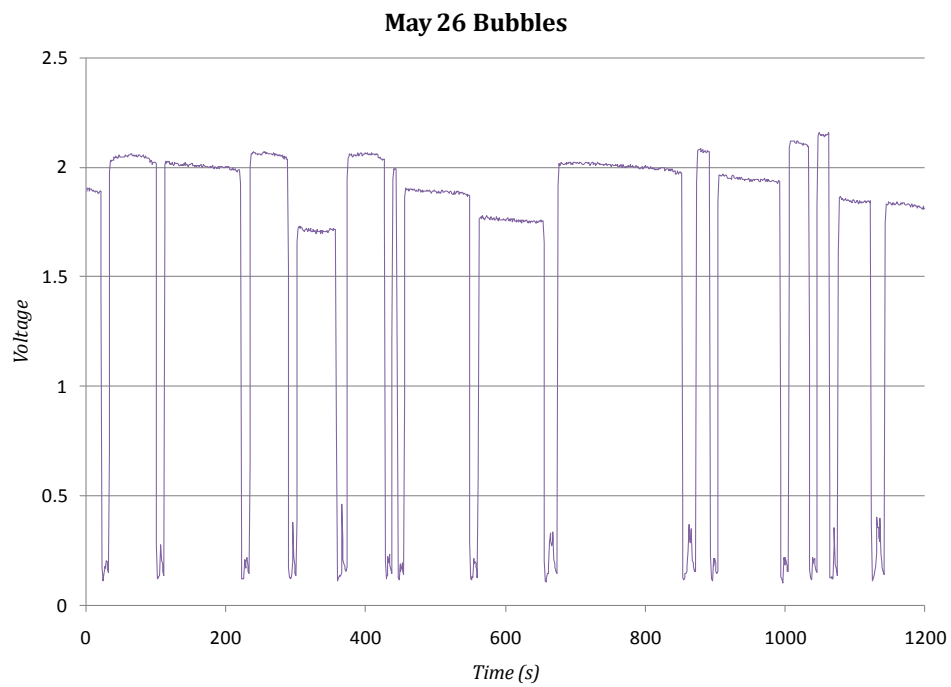


Figure 37: Mid-SXS2 bubble pattern (20 minutes)

## Results

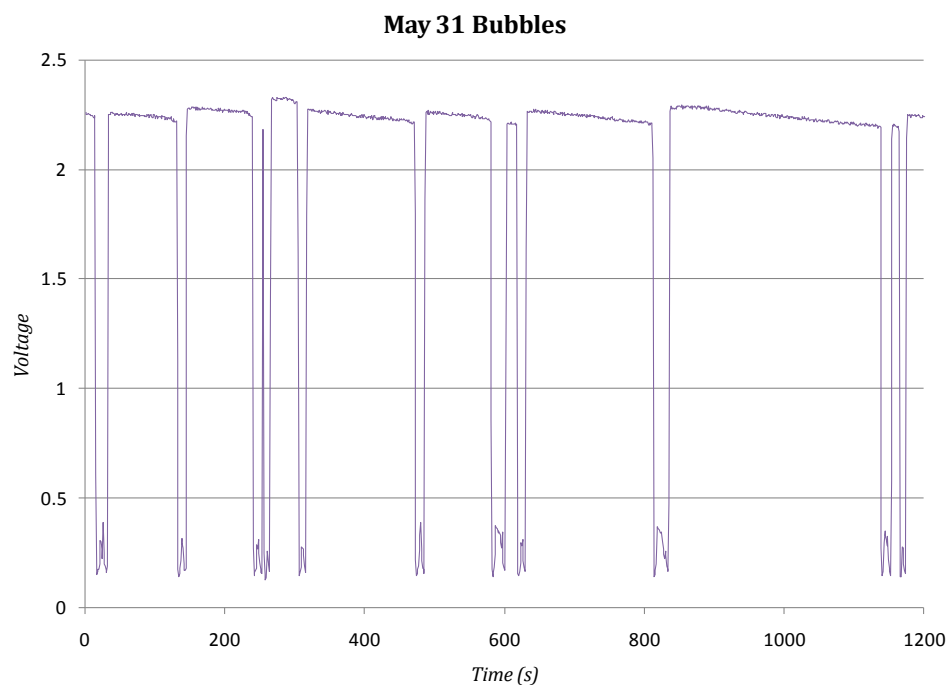


Figure 38: End SXS2 bubble pattern (20 minutes)

Above are three plots comparing the bubbles during SXS2. The plot of the May 26<sup>th</sup> (SXS2\_srvy2) is perhaps most representative of the bubbles during this cruise. By the end of the cruise, the May 30 (SXS2\_xsct2) and May 31 (SXS2\_xsct3) bubbles were looking better, although not completely consistent. The improvements were largely due to improving the connections between glass tubing sections and eliminating some of the unnecessary acidification tubing.

## Results

### 4.3.1.2.2 1 Hour Samples SXS2

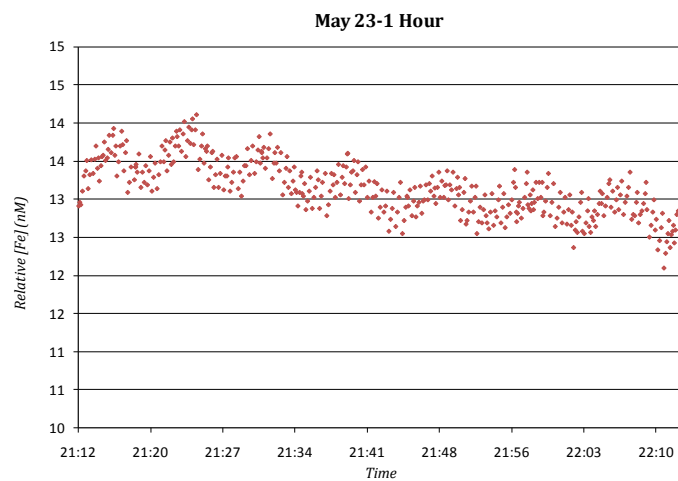


Figure 39: May 23 1 Hour Data

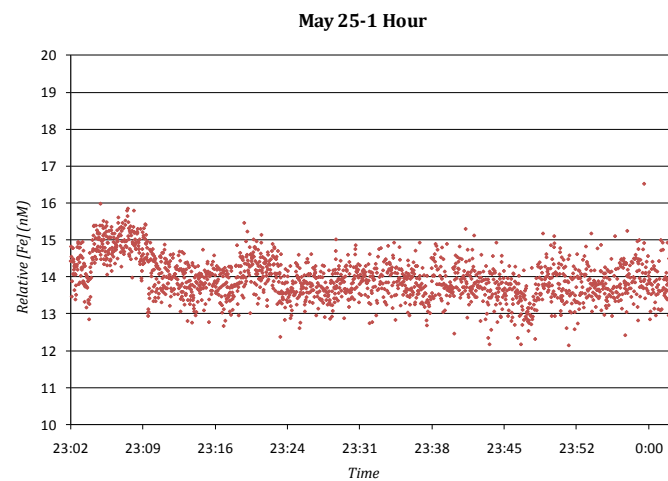


Figure 40: May 25 1 Hour Data

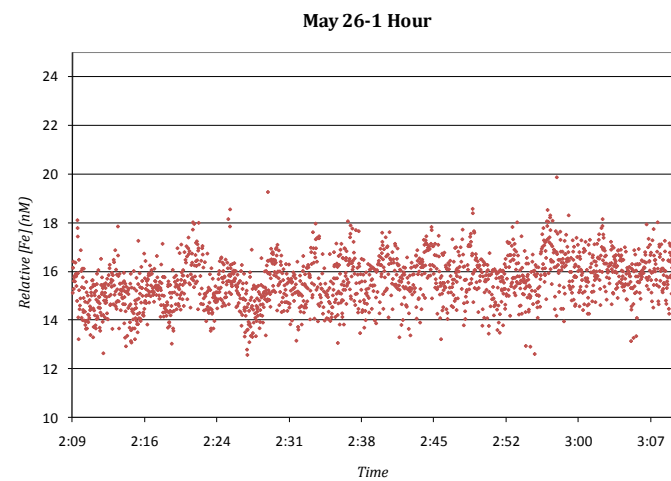


Figure 42: May 26 1 Hour Data

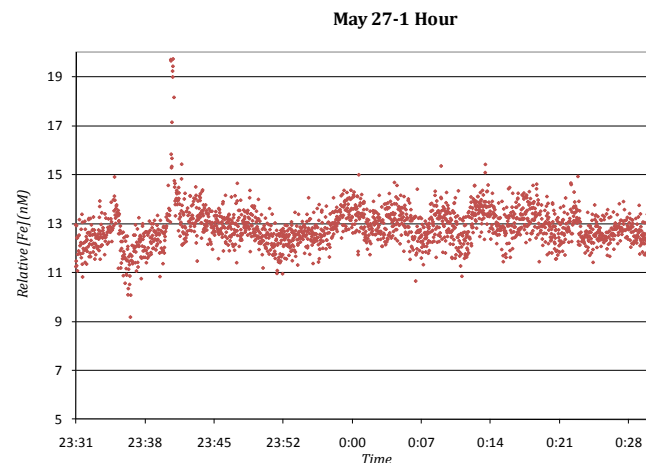


Figure 41: May 27 1 Hour Data

## Results

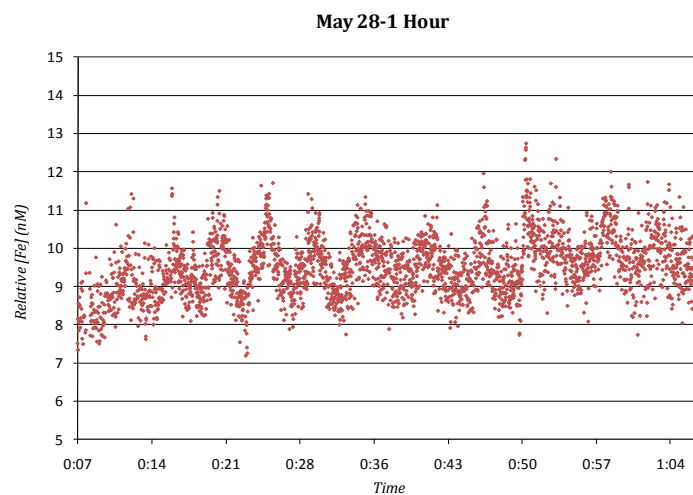


Figure 43: May 28 1 Hour Data

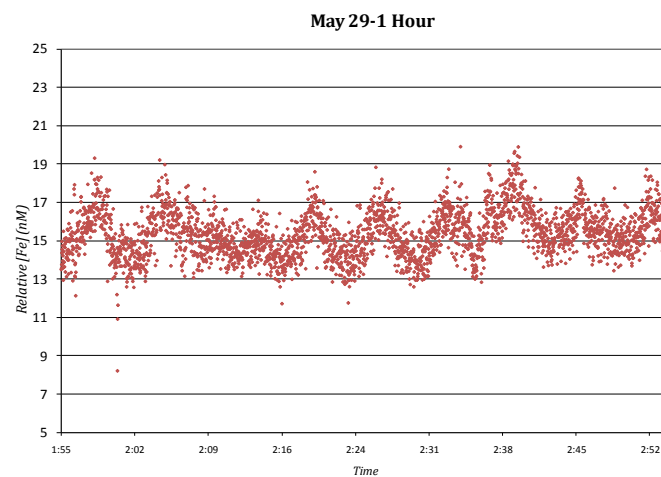


Figure 44: May 29 1 Hour Data

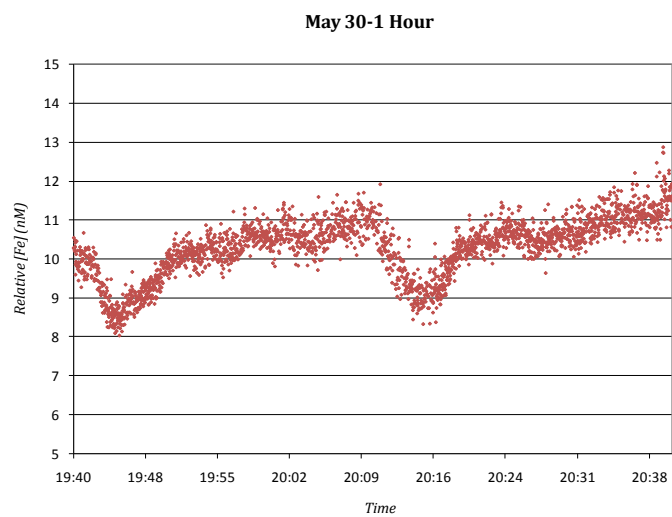


Figure 46: May 30 1 Hour Data

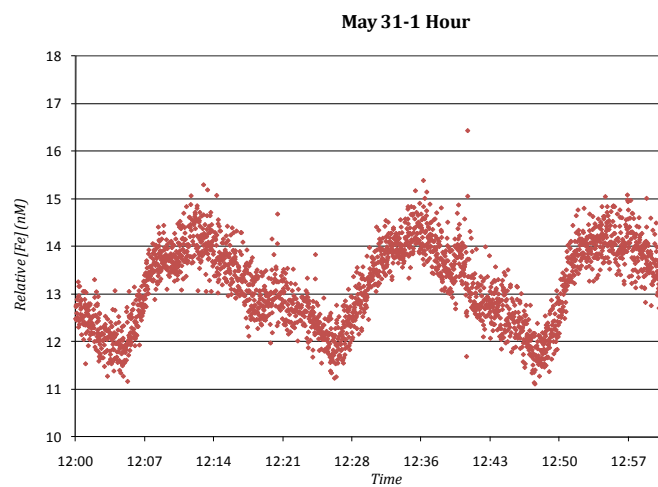


Figure 45: May 31 1 Hour Data

## Results

### 4.3.1.2.3 Standard Voltages

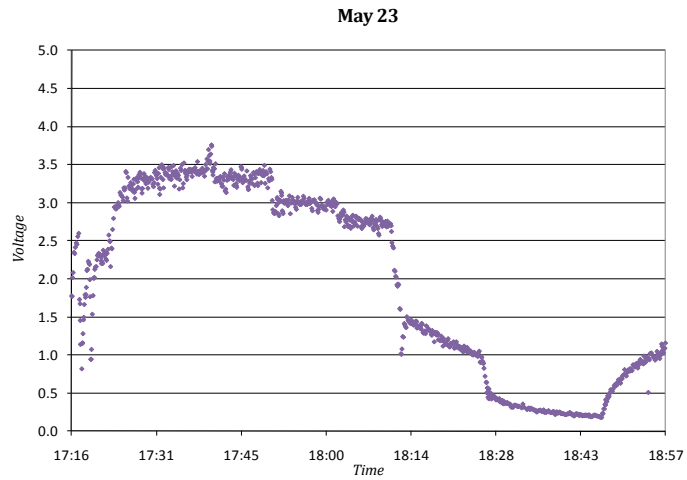


Figure 47: May 23 standards (2nd set)

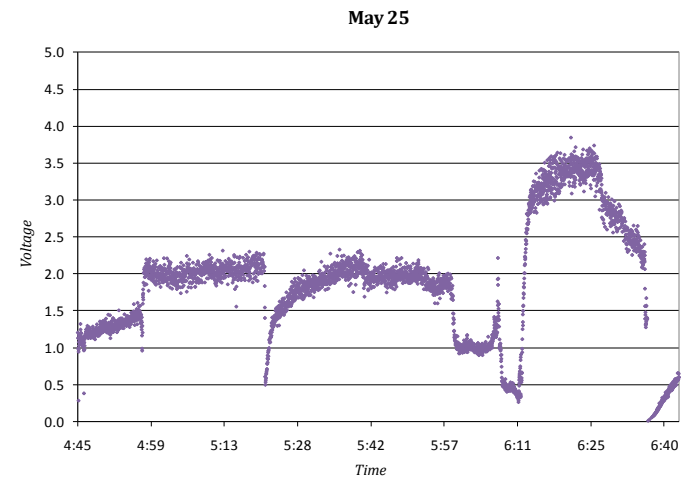


Figure 48: May 25 standards

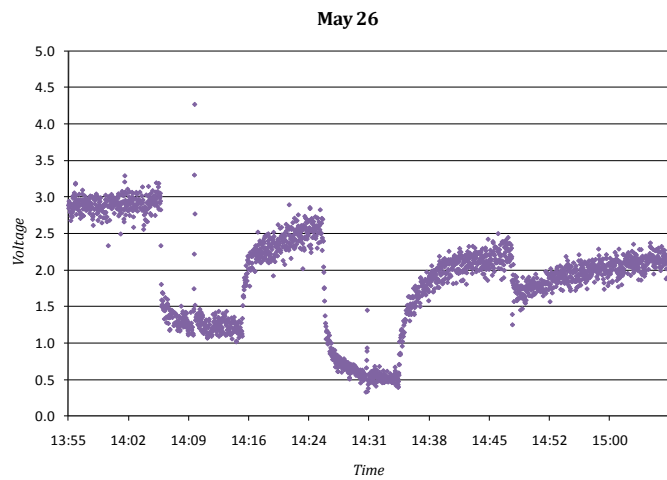


Figure 49: May 26 standards (1st set)

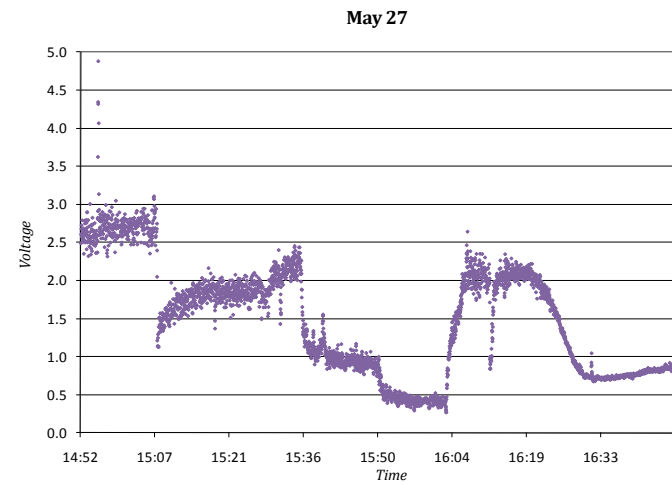


Figure 50: May 27 standards (1st set)

## Results

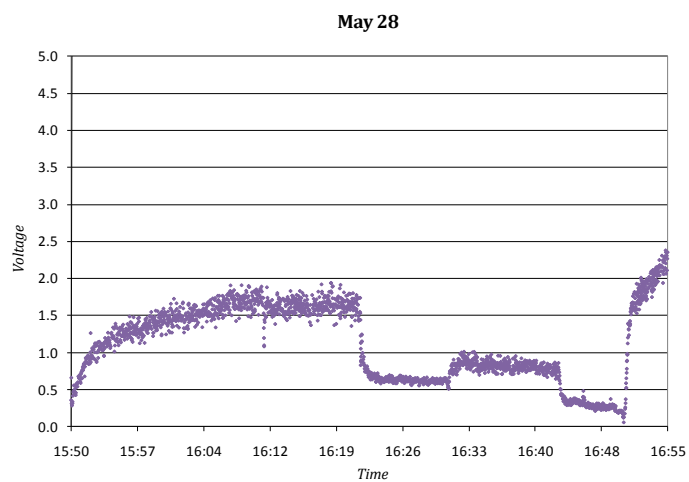


Figure 51: May 28 standards (2nd set)

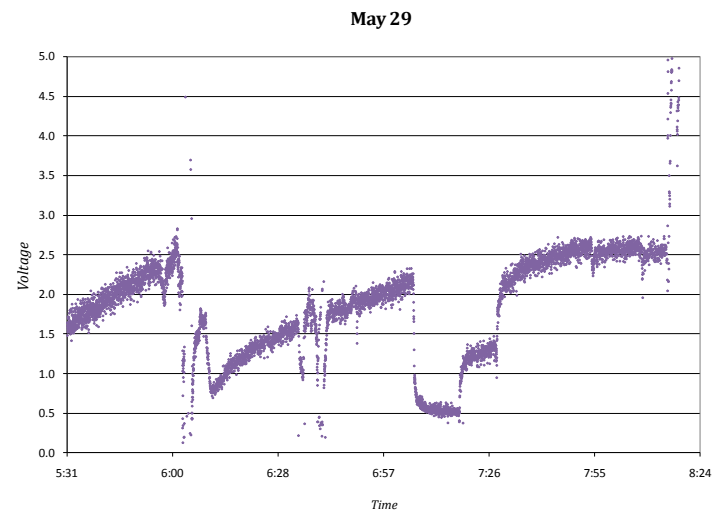


Figure 52: May 29 standards (1st set)

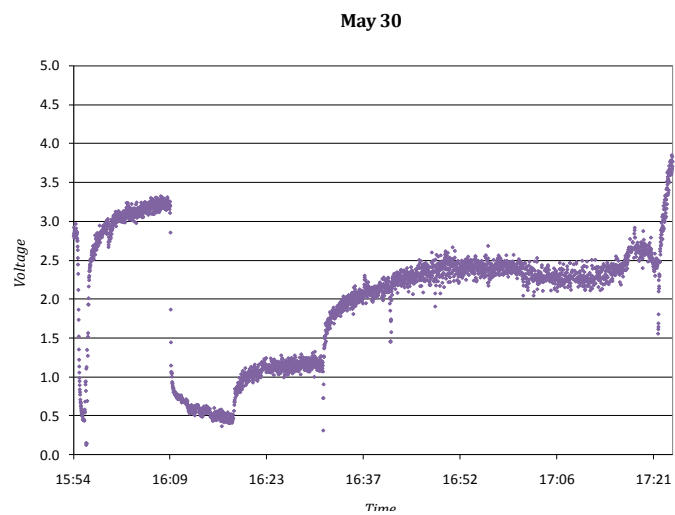


Figure 53: May 30 standards (1st set)

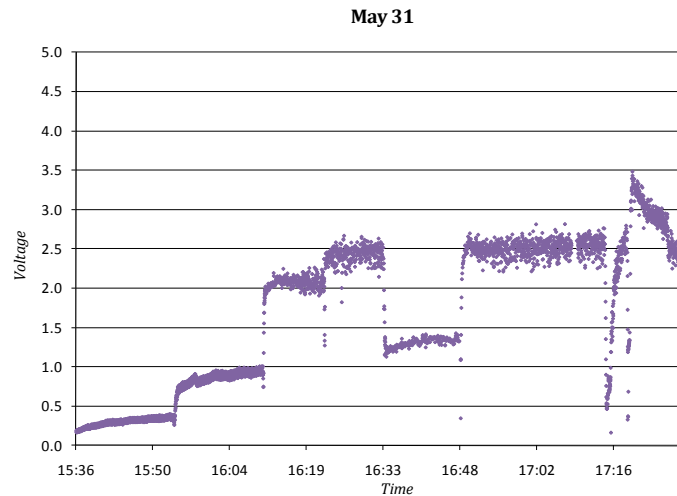


Figure 54: May 31 standards (3rd set)

## Results

### 4.3.2 SXS3 (August) Cruise

#### 4.3.2.1 Daily Results

\*Times given in the notes section reflect the time the change was expressed in the data.

##### 4.3.2.1.1 August 2—Patch tracing day 1 (SXS3\_srvy1)

August 2, 2009 the first day of patch surveying during SXS3. A discrete sample (approximately 30 mL) was collected from the TFF from 21:46:42-21:49:25. This sample was immediately run unacidified (starting at 21:49:45 and measured at 22:25). The sample was then poured off into a new bottle, acidified, and then ran (starting 22:24:18 and measured 22:49). These two samples had drastically different absorbances. This measurement was ambiguous due to the small sample size as the system can take quite awhile to reequilibrate to discrete samples after running from the TFF and the voltage from the acidified sample was still trending upwards when the sample was gone (maximum measured voltage of the acidified sample was ~0.55V compared to a maximum voltage of ~1.1V for the unacidified sample).

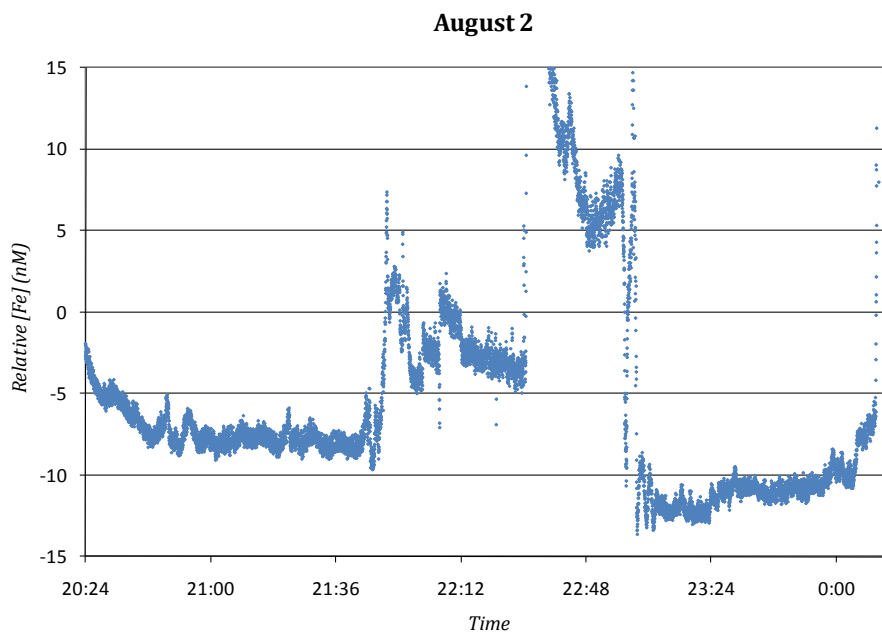


Figure 55: Relative [Fe] (SXS3\_srvy1)

## Results

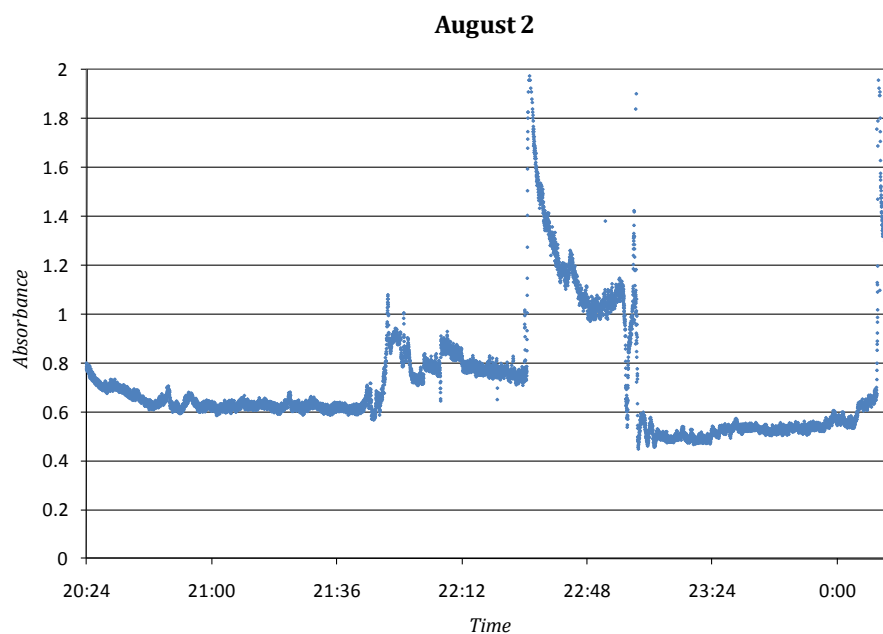


Figure 56: Absorbance (SXS3\_srvy1)

**Table 34: SXS3\_srvy1 summary**

	<i>Absorbance</i>	<i>Stdev</i>	<i>Drift</i>	<i>Detection Limit:</i>	<i>Slope (avg)</i>
Std 1	0.8218	0.0140	n/a		0.0301
Std 2	0.9479	0.0167	-0.0575		<i>Slope 1:</i>
Std 3	1.5102	0.0215	0.3921		0.0233
Std 4	2.2824	0.0342	0.5592		<i>Slope 2:</i>
<i>Average</i>		0.0227			0.0366

**Internal Lag time:**  $\approx$  4 minutes

## Results

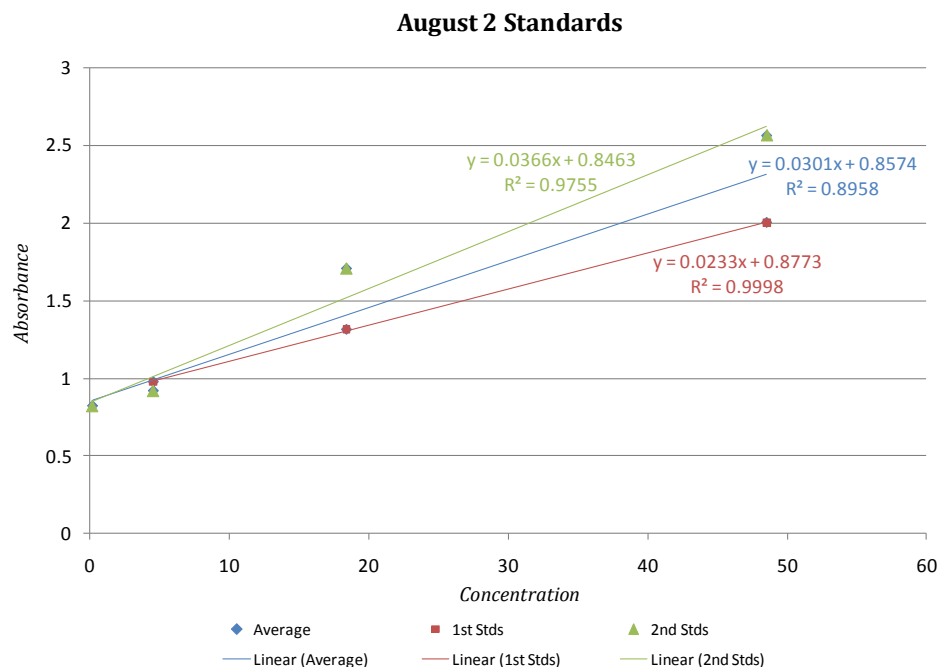


Figure 57: SXS3\_srvy1 standards

**Table 35: SXS3\_srvy1 standards**

Standard	Concentration	Date	Time	Absorbance	Stdev
Std 2	4.54	8/2/09	15:59	0.9767	0.01565
Std 3	18.39	8/2/09	16:18	1.3141	0.01836
Std 4	48.52	8/2/09	16:30	2.0028	0.02411
Std 4	48.52	8/3/09	0:48	2.5620	0.04425
Std 3	18.39	8/3/09	1:12	1.7062	0.02467
Std 2	4.54	8/3/09	3:01	0.9191	0.01782
Std 1	0.18	8/4/09	3:55	0.8218	0.01398

**Table 36: SXS3\_srvy1 discrete samples**

Date	Time	Sample	Abs.	Blank Corr. Abs	Stdev	Conc.	Date run
8/2/09	22:25	Discrete sample (unacidified)	1.7375	n/a	0.04000	29.24	
8/2/09	22:49	Discrete sample (acidified)	2.3366	n/a	0.05338	49.14	

## Results

**Table 37: SXS3\_srvy1 notes**

8/2/2009	15:15	Begin standards. Standards seem fairly quick but exhibited the same shallow slope seen throughout the cruise
8/2/2009	17:23	Acidified seawater. Lots of noise in this sample. Noise seemed to decrease throughout the cruise reaching a plateau during the August 8 data
8/2/2009	20:09	Changed buffer & took a sample from the TFF
8/2/2009	21:57	TFF sample—unacidified
8/2/2009	22:51	TFF sample—acidified in new bottle
8/2/2009	23:02	Return to SS flow
8/3/2009	00:45	Begin standards
8/3/2009	04:37	End standards

### 4.3.2.1.2 August 3—Patch tracing day 2 (SXS3\_srvy2)

August 3 was the second day of the patch tracing during SXS3. The calculated absorbancies were quite high during this run, usually the absorbance ranges from 0-2 instead of the 2-12 seen here. This is probably due to samples taken during the day which cause disruptance to the flow of data.

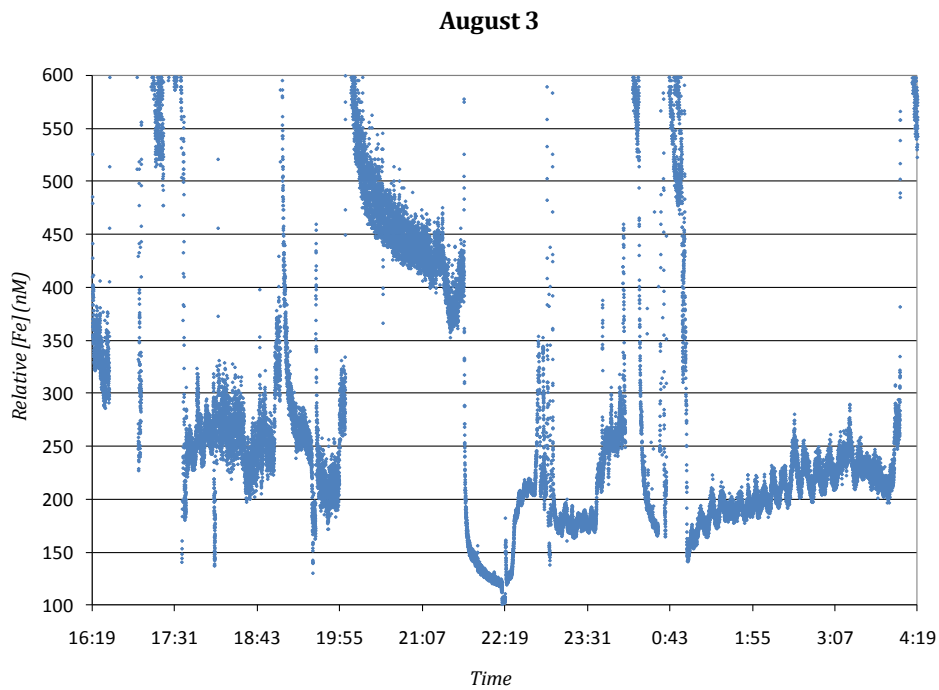


Figure 58: Relative [Fe] (SXS3\_srvy2)

## Results

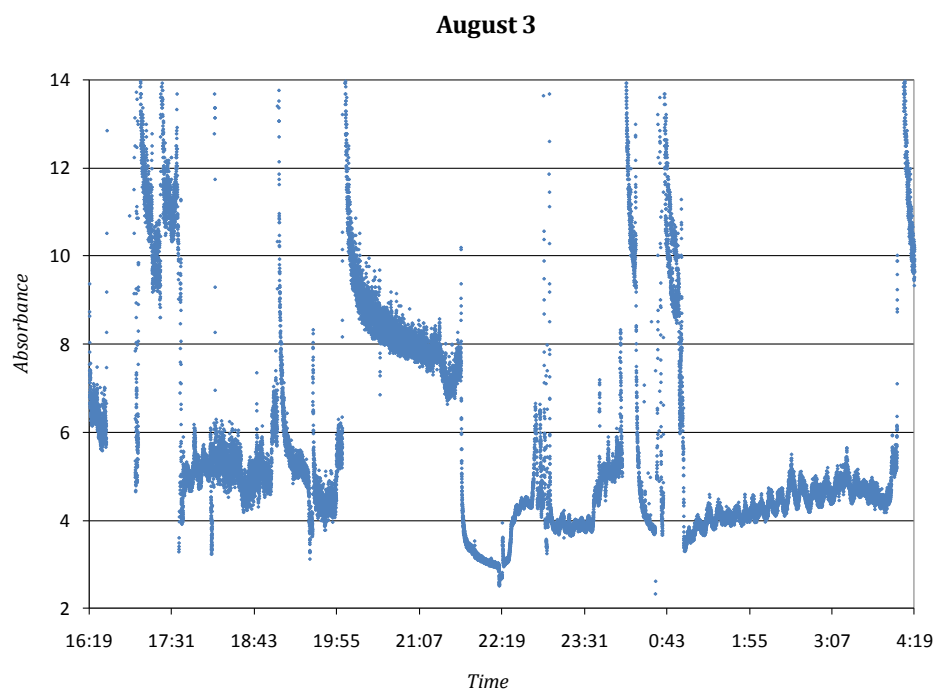


Figure 59: Absorbance (SXS3\_srvy2)

**Table 38: SXS3\_srvy2 summary**

	<i>Absorbance</i>	<i>Stdev</i>	<i>Drift (Abs 2-Abs1)</i>	<i>Detection Limit:</i>	<i>Slope (avg)</i>
Std 1	1.0460	0.0178	-0.0340	3.38	0.0158
Std 2	1.1340	0.0194	-0.0095	(Std1 Avg)	Slope 1:
Std 3	1.4348	0.0226	0.0629		0.0081
Std 4	1.8167	0.0264	0.6929		Slope 2:
Average		0.0215			0.0235

**Internal Lag time:** ≈ 5 minutes

## Results

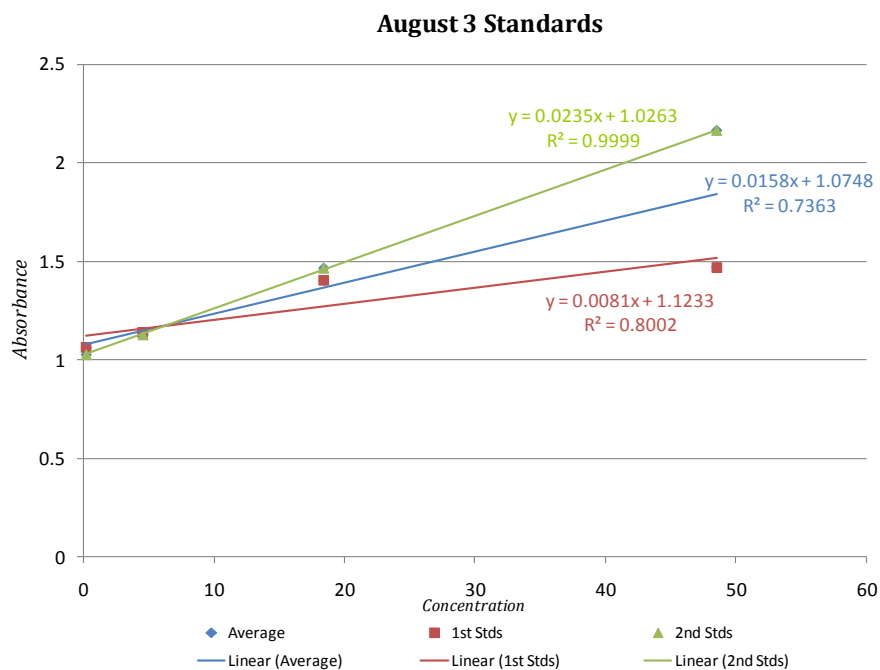


Figure 60: SXS3\_srvy2 standards

**Table 39: SXS3\_srvy2 standards**

Standard	Concentration	Date	Time	Absorbance	Stdev
Std 4	48.52	8/3/09	14:58	1.4702	0.02702
Std 3	18.39	8/3/09	15:03	1.4033	0.02161
Std 2	4.54	8/3/09	15:20	1.1387	0.02278
Std 1	0.18	8/3/09	15:30	1.0630	0.02091
Std 4	48.52	8/4/09	4:32	2.1631	0.02573
Std 3	18.39	8/4/09	4:54	1.4662	0.02358
Std 2	4.54	8/4/09	5:30	1.1292	0.01604
Std 1	0.18	8/4/09	5:55	1.0290	0.01472

**Table 40: SXS3\_srvy2 discrete samples**

Date	Time	Sample	Abs	Blank Corr	Stdev	Conc.	Date run
8/4/09	3:59	#31	0.6855	0.6111	0.0066	77.97	9/14/09
8/3/09	17:15		2.2886	n/a	0.0402		8/3/09
8/3/09	17:30		2.3878	n/a	0.0360		8/3/09
8/3/09	23:52		1.6395	n/a	0.0285		8/3/09
8/4/09	0:50		2.2446	n/a	0.0743		8/3/09

## Results

**Table 41: SXS3\_srvy2 notes**

8/3/09	14:33	begin standards
8/3/09	15:35	end standards
8/3/09	16:32	heater issues. Very hot; collected acidification line sample while cooling
8/3/09	17:41	acidification line test sample followed by double acidified acidification line sample
8/3/09	18:38	TFF
8/3/09	18:58	Surface TFF
8/3/09	19:59	bulk acidified seawater
8/3/09	21:43	milliQ
8/3/09	22:54	TFF
8/3/09	23:38	milliQ
8/4/09	0:03	begin acidification line tests
8/4/09	0:59	end acidification line tests; begin TFF
8/4/09	4:03	standards

### 4.3.2.1.3 August 4—Patch tracing day 3 (SXS3\_srvy3)

August 4 was the third day of the patch survey for SXS3. The system was run alternating between the surface fish flow (known to be very low in iron) and the supersucker flow. A discrete sample was collected from the surface at about 21:59 and run unacidified. The sample was then poured into a new bottle, acidified, and ran on the system. The results, as usual, did not match.

## Results

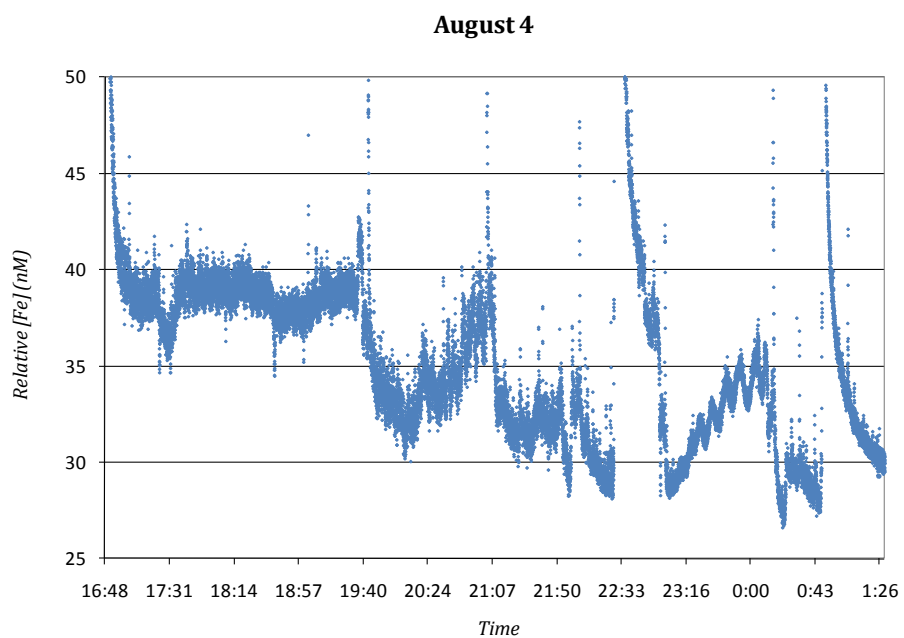


Figure 61: Relative [Fe] (SXS3\_srvy3)

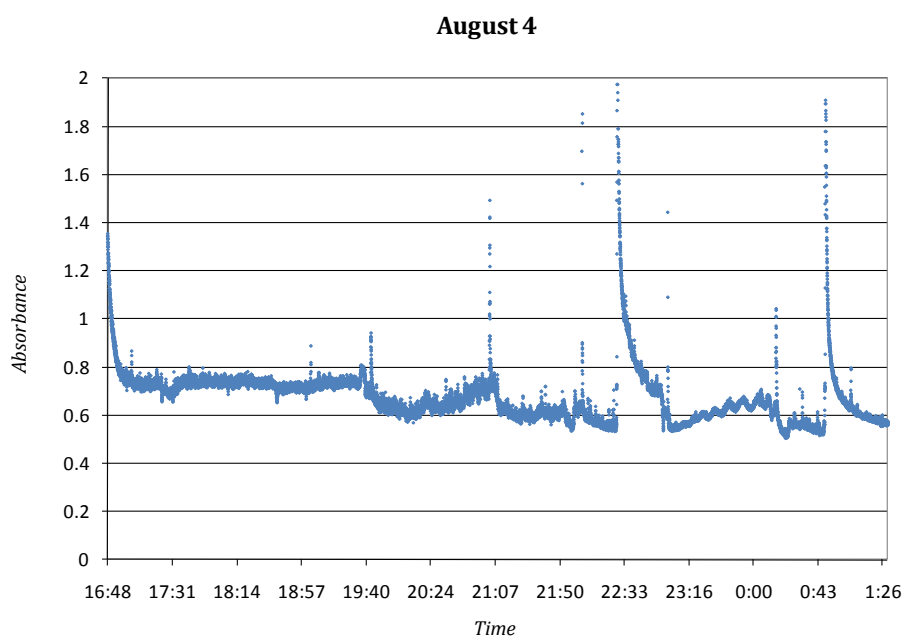


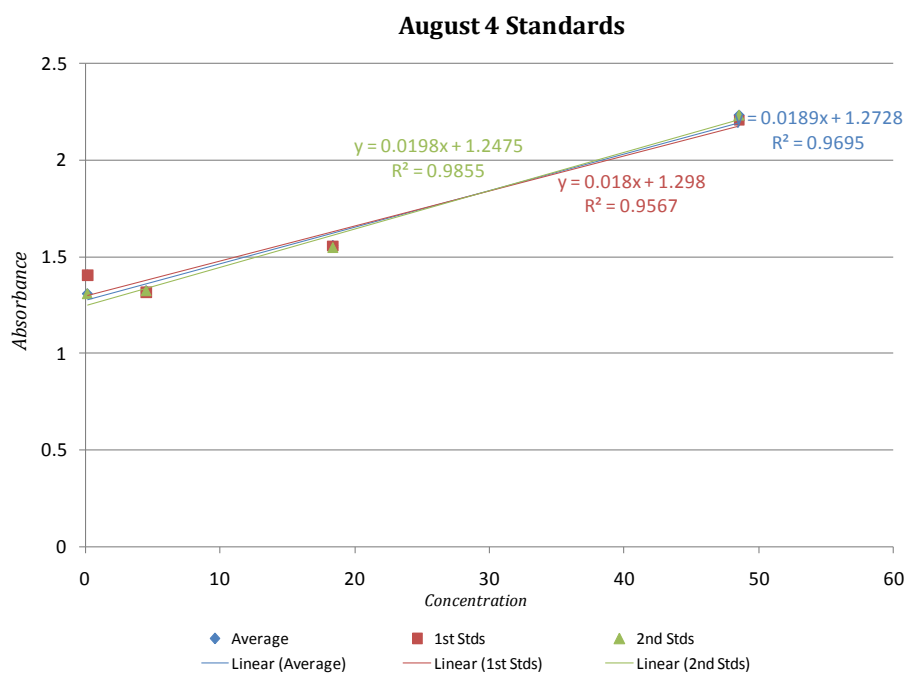
Figure 62: Absorbance (SXS3\_srvy3)

## Results

**Table 42: SXS3\_srvy3 summary**

	Absorbance	Stdev	Drift (Abs 2-Abs1)	Detection Limit:	Slope (avg)
Std 1	1.3553	0.0232	-0.0164	3.68	0.0189
Std 2	1.3191	0.0169	-0.0098	(Std1 Avg)	Slope 1:
Std 3	1.5517	0.0211	-0.0038		0.0198
Std 4	2.2207	0.0396	0.0190		Slope 2:
Average		0.0252			0.018

**Internal Lag time:** ≈ 5 minutes



**Figure 63: SXS3\_srvy3 standards**

**Table 43: SXS3\_srvy3 standards**

Standard	Concentration	Date	Time	Absorbance	Stdev
Std 4	48.52	8/4/09	15:47	2.2066	0.03008
Std 3	18.39	8/4/09	16:09	1.5572	0.02303
Std 2	4.54	8/4/09	16:25	1.3163	0.02179
Std 1	0.18	8/4/09	21:43	1.4044	0.03141
Std 1	0.18	8/5/09	1:28	1.3062	0.01501
Std 2	4.54	8/5/09	1:50	1.3218	0.01202
Std 3	18.39	8/5/09	2:19	1.5462	0.01920
Std 4	48.52	8/5/09	3:05	2.2347	0.04910

## Results

**Table 44: SXS3\_srvy3 discrete samples**

Date	Time	Sample	Abs	Blank Corr	Stdev	Conc.	Date run
8/4/09	22:22	surface sample (unacidified)	1.2666	n/a	0.02051	67.02	8/4/09
8/4/09	22:50	surface sample (acidified)	1.6357	n/a	0.01681	86.54	8/4/09

**Table 45: SXS3\_srvy3 notes**

8/4/2009	15:06	Began replacing tubing
8/4/2009	15:25	Finished replacing tubing
8/4/2009	15:41	Standards
8/4/2009	16:47	TFF
8/4/2009	19:45	Surface TFF
8/4/2009	22:30	Surface discrete sample collected
8/5/2009	0:49	Supersucker TFF
8/5/2009	1:28	Begin standards

### 4.3.2.1.4 August 5—Patch tracing day 4 (SXS3\_srvy4)

August 5, 2009 was day four of the patch survey. The GCFA was run constantly with no discrete samples run during the day. A discrete sample was collected from the supersucker near the surface via the sink port and was very high in iron compared to in the inline samples even when taking into consideration the issues with NASS-5 when the sample was run in the lab on September 14, 2009.

## Results

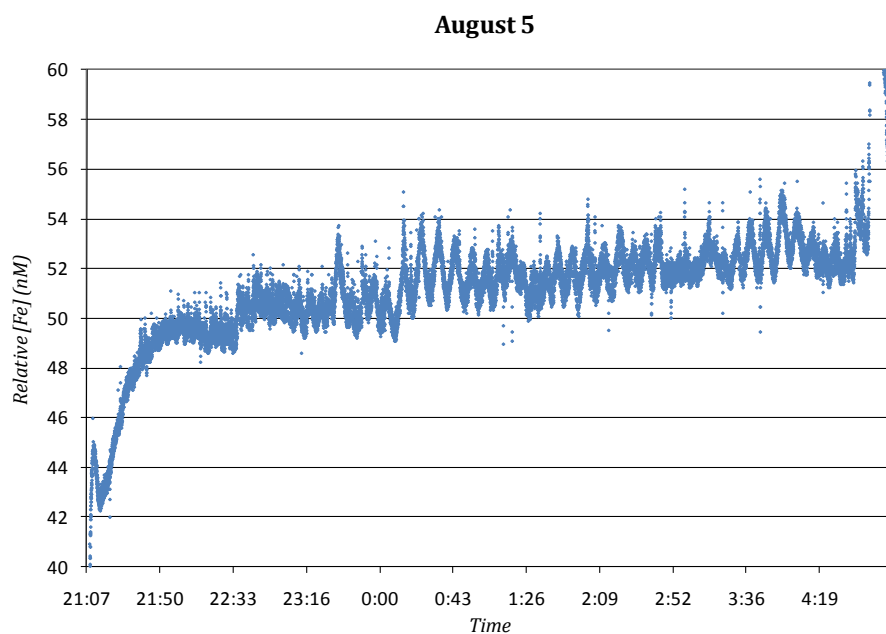


Figure 64: Relative [Fe] (SXS3\_srvy4)

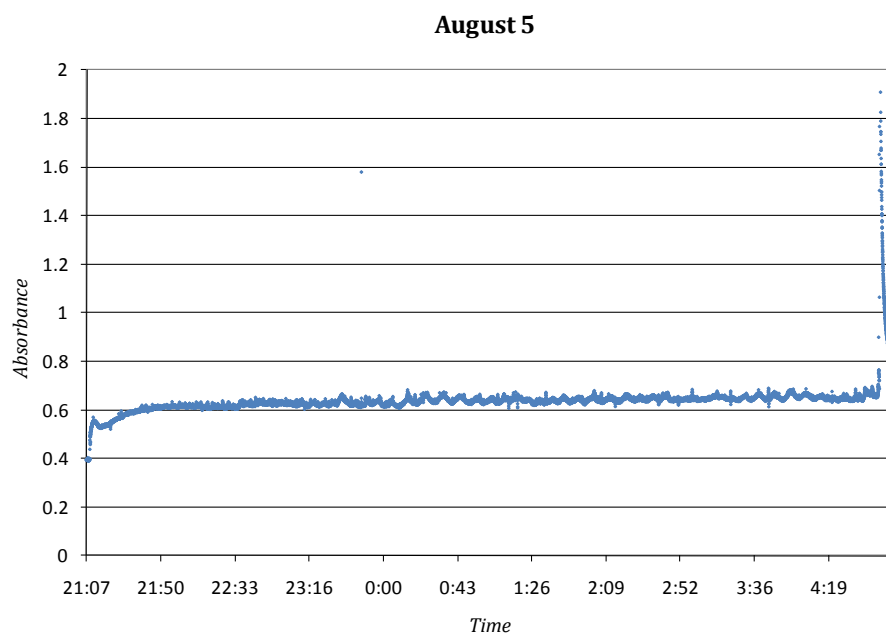


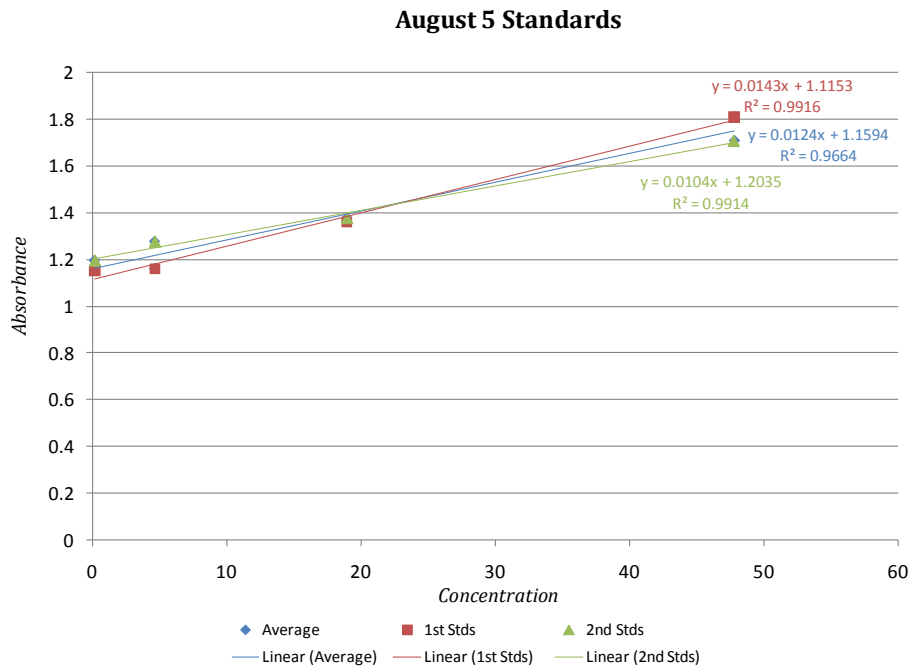
Figure 65: Absorbance (SXS3\_srvy4)

## Results

**Table 46: SXS3\_srvy4 summary**

	Absorbance	Stdev	Drift (Abs 2-Abs1)	Detection Limit:	Slope (avg)
Std 1	1.1741	0.0171	0.0171	4.13	0.0124
Std 2	1.2196	0.0089	0.0035	(Std1 Avg)	Slope 1:
Std 3	1.3694	0.0180	0.0097		0.0142
Std 4	1.7601	0.0137	-0.0038		Slope 2:
Average		0.0144			0.0104

**Internal Lag time:** ≈ 5 minutes



**Figure 66: SXS3\_srvy4 standards**

**Table 47: SXS3\_srvy4 standards**

Standard	Concentration	Date	Month	Absorbance	Stdev
Std 1	0.18	8/5/09	17:32	1.1518	0.00852
Std 2	4.65	8/5/09	17:43	1.1617	0.00716
Std 3	18.97	8/5/09	18:03	1.3604	0.01316
Std 4	47.8	8/5/09	18:20	1.8109	0.01563
Std 2	4.65	8/6/09	5:37	1.2775	0.01064
Std 1	0.18	8/6/09	5:57	1.1964	0.02558
Std 3	18.97	8/6/09	6:13	1.3784	0.02291
Std 4	47.8	8/6/09	6:27	1.7092	0.01180

## Results

**Table 48: SXS3\_srvy 4 discrete samples**

<i>Date</i>	<i>Time</i>	<i>Sample</i>	<i>Abs</i>	<i>Blank Corr</i>	<i>Stdev</i>	<i>Conc.</i>	<i>Date run</i>
8/6/09	4:38	#45	0.7294	0.6551	0.00843	83.19	9/14/09

**Table 49: SXS3\_srvy4 notes**

8/5/2009	17:04	Begin standards
8/5/2009	18:27	End standards; begin milliQ
8/5/2009	21:09	End milliQ; begin TFF
8/6/2009	4:46	Begin standards
8/6/2009	6:30	End standards

### 4.3.2.1.5 August 6—Patch tracing day 5 (SXS3\_srvy5)

August 6, 2009 was the fifth day of following the patch during SXS3. The GCFA was run without interruption and a discrete TFF sample was taken at 15:14. This sample was acidified and run along with the standards at the end of the day.

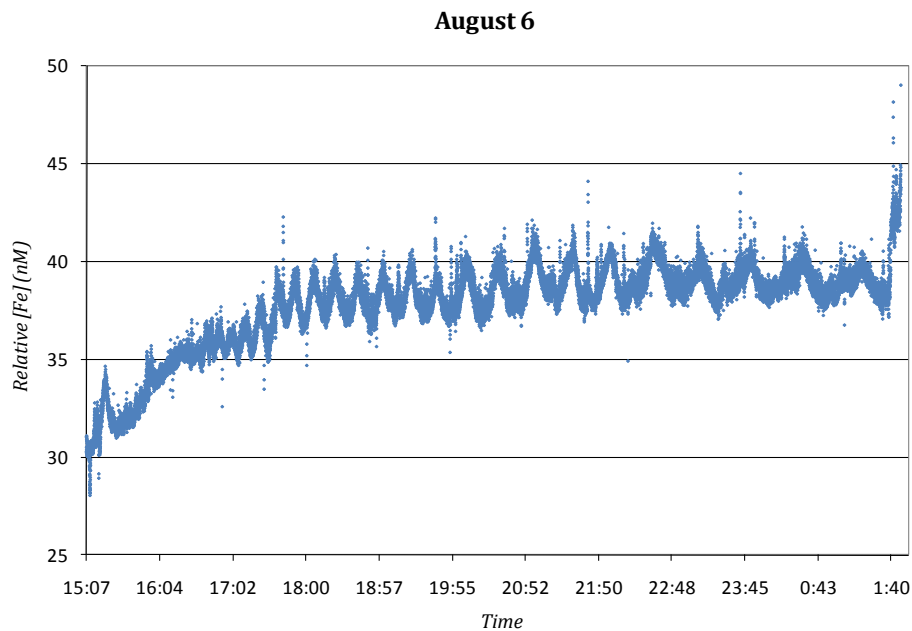


Figure 67: Relative [Fe] (SXS3\_srvy5)

## Results

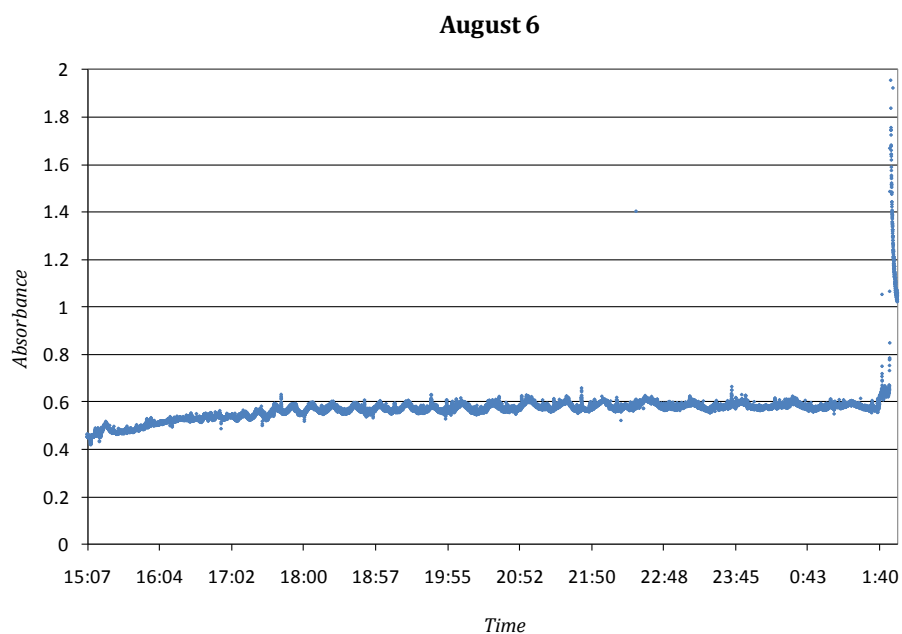


Figure 68: Absorbance (SXS3\_srvy5)

**Table 50: SXS3\_srvy5 summary**

	<i>Absorbance</i>	<i>Stdev</i>	<i>Drift (Abs 2-Abs1)</i>		<i>Detection Limit:</i>	<i>Slope (avg)</i>
MilliQ	0.8104	0.0035	n/a		1.91	0.0149
Std 1	1.0499	0.0095	0.1670		(Std1 Avg)	Slope 1:
Std 2	1.1083	0.1771	0.2005			0.0101
Std 3	0.0153	0.0153	0.4356			Slope 2:
Std 4	0.0156	0.0156	0.6238			0.0186
Average		0.0487				

**Internal Lag time:** ≈ 5 minutes

## Results

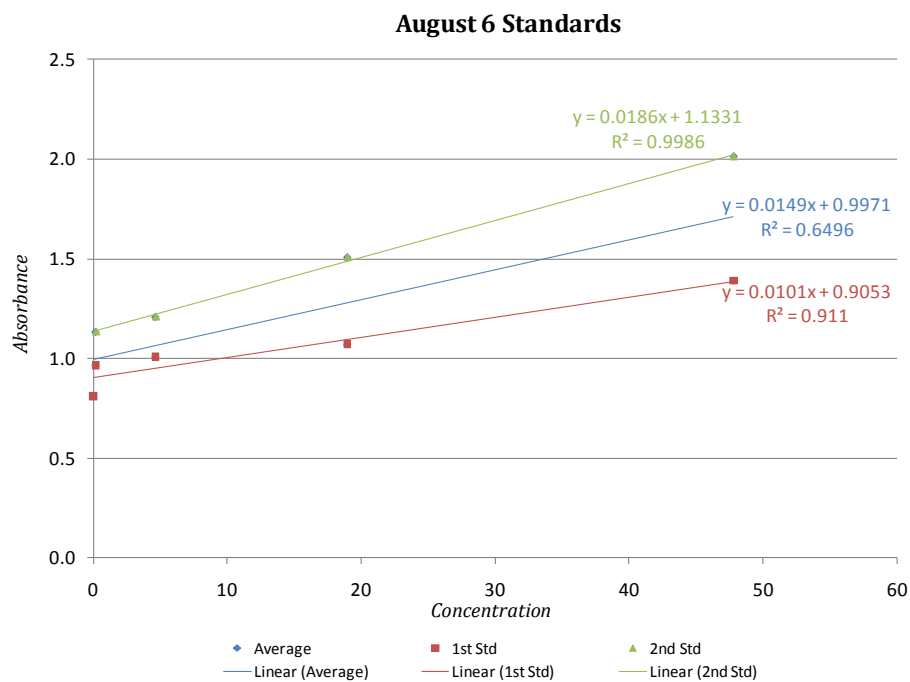


Figure 69: SXS3\_srvy 5 standards

**Table 51: SXS3\_srvy5 standards**

Standard	Concentration	Date	Time	Absorbance	Stdev
MilliQ	0	8/6/09	13:50	0.8104	0.00355
1	0.18	8/6/09	14:07	0.9664	0.00879
2	4.65	8/6/09	14:32	1.0080	0.34019
3	18.97	8/6/09	14:43	1.0720	0.01162
4	47.8	8/6/09	14:54	1.3907	0.01126
4	47.8	8/7/09	2:08	2.0145	0.02003
3	18.97	8/7/09	2:43	1.5076	0.01894
2	4.65	8/7/09	3:15	1.2085	0.01401
1	0.18	8/7/09	3:29	1.1334	0.01021

**Table 52: SXS3\_srvy5 discrete samples**

Date	Time	Sample	Abs	Blank Corr	Stdev	Conc.	Date run
8/6/09	15:14	Acidified TFF sample	1.5908	n/a	0.01682	106.77	8/7/09
8/7/09	1:47	#51	0.8608	0.7864	0.00984	98.83	9/14/09

## Results

**Table 53: SXS3\_srvy5 notes**

8/6/09	13:51	acidified milliQ
8/6/09	14:12	begin standards
8/6/09	15:12	TFF
8/7/09	14:28	TFF sample taken
8/7/09	1:40	begin standards
8/7/09	3:32	begin running TFF sample (collected 8/6 15:28)

### 4.3.2.1.6 August 7—45°N Transect (SXS3\_xsct4)

The transect at 45°N was run uninterrupted. A sample (#61) was collected and run both unacidified and acidified at the end of the day. A plateau was never reached for the sample when it was acidified (only ~30mL were taken). Several other samples were collected, acidified, and analyzed in the lab.

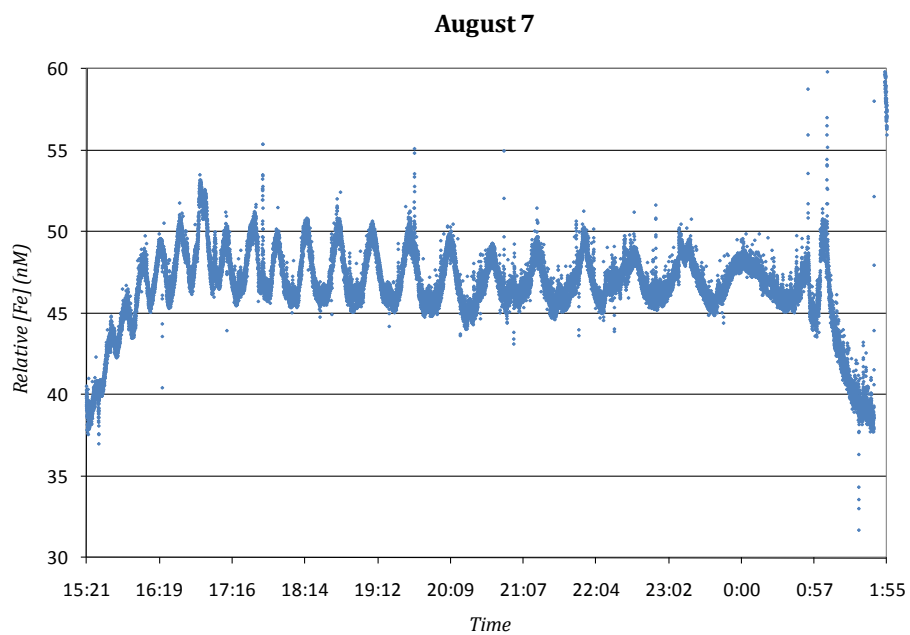


Figure 70: Relative [Fe] (SXS3\_xsct4)

## Results

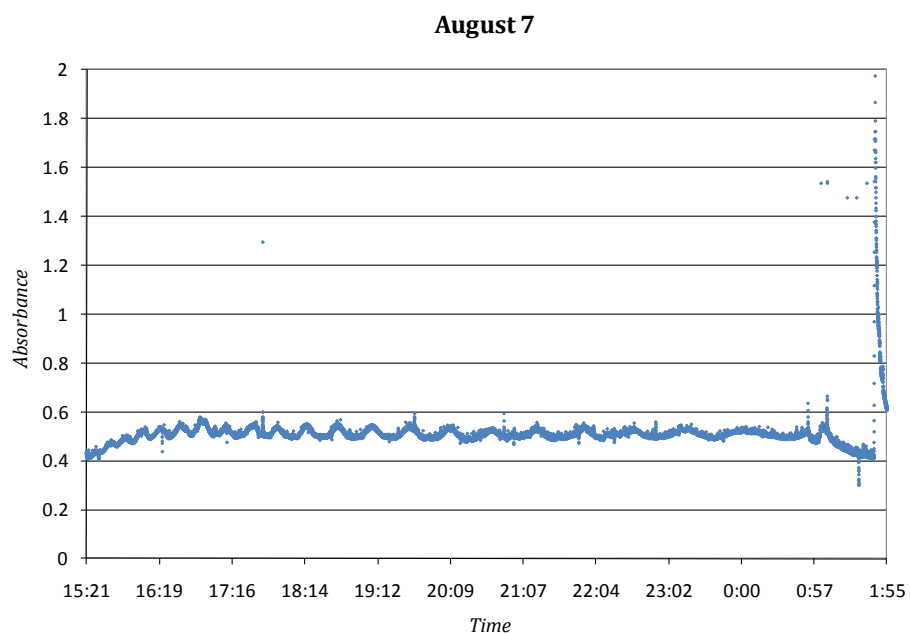


Figure 71: Absorbance

**Table 54: SXS3\_xsct4 summary**

	<i>Absorbance</i>	<i>Stdev</i>	<i>Drift (Abs 2-Abs1)</i>	<i>Detection Limit:</i>	<i>Slope (avg)</i>
MilliQ	n/a	n/a	n/a	3.32	0.0108
Std 1	0.8838	0.0119	0.0020	(Std1 Avg)	Slope 1:
Std 2	0.9665	0.0326	0.0448		0.0103
Std 3	1.0606	0.0176	0.0096		Slope 2:
Std 4	1.4153	0.0234	0.0101		0.0114
Average		0.0214			

**Internal Lag time:**  $\approx$  7 minutes

## Results

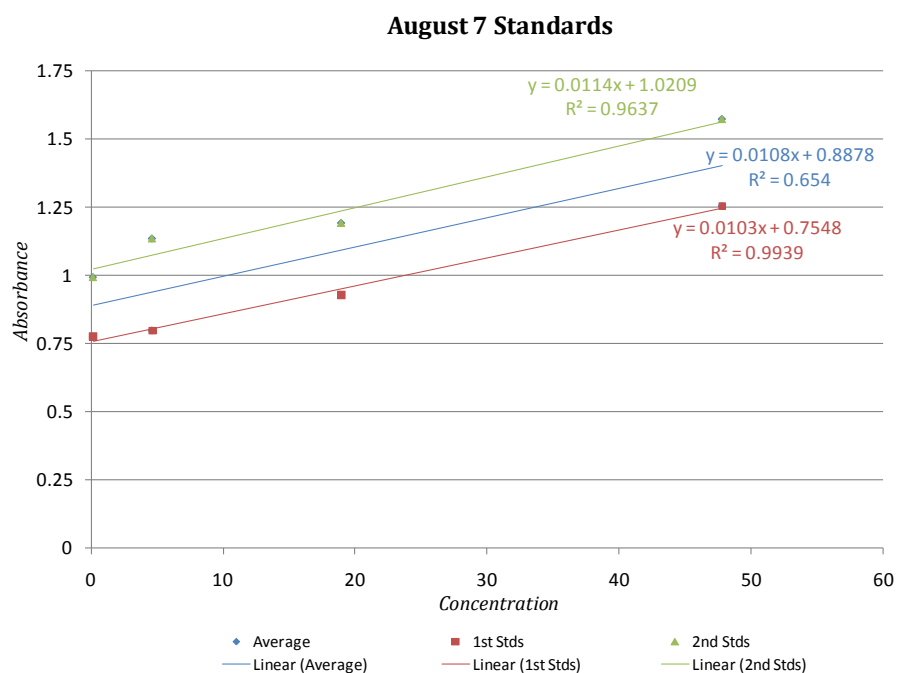


Figure 72: SXS3\_xsct4 standards

**Table 55: SXS3\_xsct4 standards**

Standard	Concentration	Date	Time	Absorbance	Stdev
Std 2	4.65	8/7/09	14:09	0.7973	0.01021
Std 1	0.18	8/7/09	14:20	0.7745	0.01094
Std 3	18.97	8/7/09	14:43	0.9282	0.01282
Std 4	47.8	8/7/09	15:03	1.2551	0.01789
Std 4	47.8	8/8/09	2:09	1.5756	0.02888
Std 2	4.65	8/8/09	2:40	1.1357	0.05504
Std 3	18.97	8/8/09	2:45	1.1930	0.02239
Std 1	0.18	8/8/09	3:09	0.9932	0.01295

## Results

**Table 56: SXS3\_xsct4 discrete samples**

<i>Date</i>	<i>Time</i>	<i>Sample</i>	<i>Abs</i>	<i>Blank Corr</i>	<i>Stdev</i>	<i>Concentration</i>	<i>Date run</i>
8/7/09	23:22	#58	0.5629	0.4885	0.01328	63.37	9/14/09
8/8/09	1:25	#59	0.9364	0.8621	0.72036	107.84	9/14/09
8/8/09	1:32	#60	1.0983	1.0875	0.00652	161.12	9/15/09
8/8/09	1:38	Fe #61, unacidified	0.9819	n/a	0.02225	90.92	8/8/09
8/8/09		Fe #61, acidified no plateau					8/8/09

**Table 57: SXS3\_xsct4 notes**

8/7/09	13:57	begin standards
8/7/09	15:11	begin TFF
8/8/09	1:08	begin Fe #61 (unacidified)
8/8/09	1:44	begin Fe #61 (added acid to bottle)
8/8/09	2:16	standards

### 4.3.2.1.7 August 8—43.9°N Transect (SXS3\_xsct5)

Transect 5 along 43.9°N was conducted on August 8-9, 2009. The data was collected relatively uninterrupted with a break at 8/9 3:06 when the supersucker flow was stopped. There was one other small problem with the GCFA at about 8/8 10:15 which was resolved and then standards were run. A discrete sample was also collected from the supersucker flow at 8/8 16:43 and the discrete sample was measured as having much higher iron than the inline sample. The source of this sample was not recorded, however it was most likely from the TFF but may possibly have been from the sink port.

Results

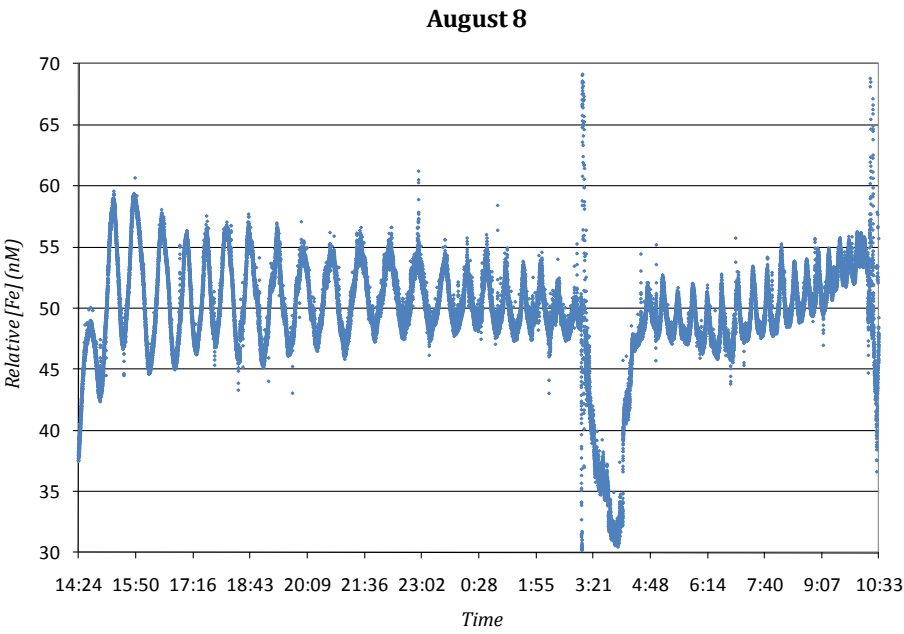


Figure 73: Relative [Fe] (SXS3\_xsct5)

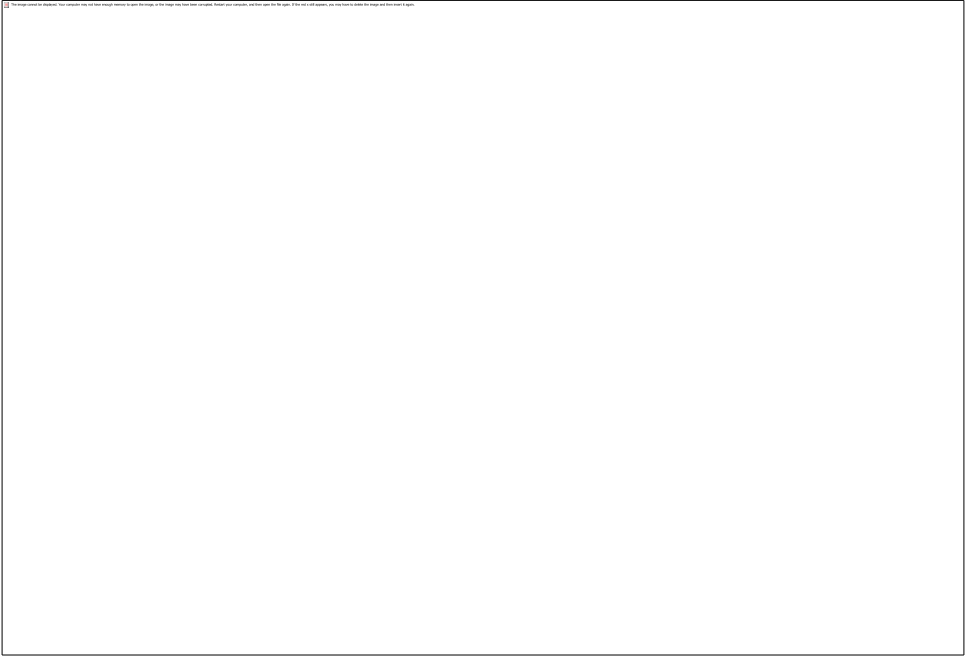


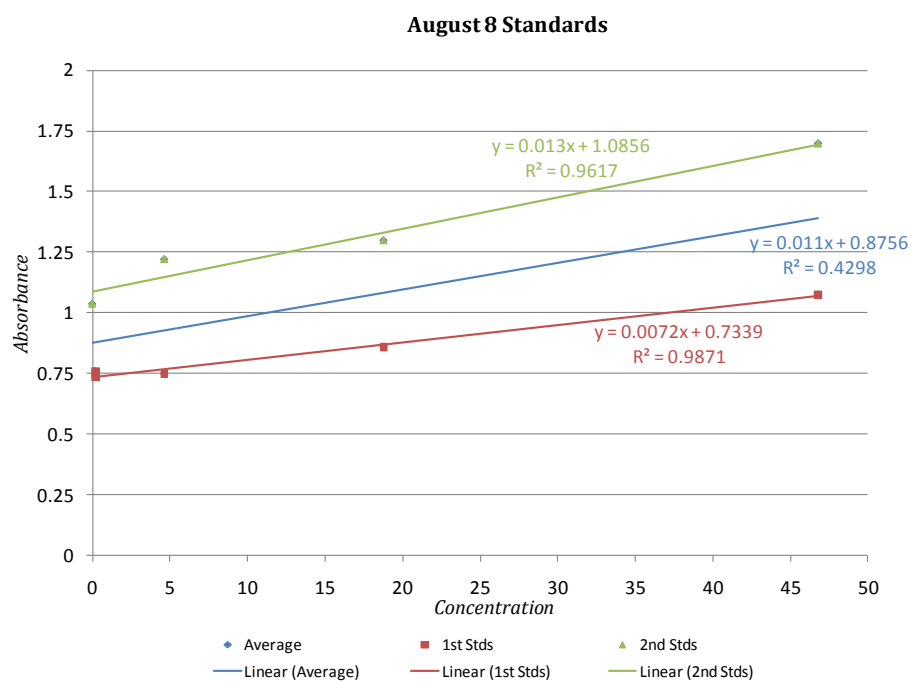
Figure 74: Absorbance (SXS3\_xsct5)

## Results

**Table 58: SXS3\_xsct5 summary**

	Absorb.	Stdev	Drift (Abs 2-Abs1)	Detection Limit:	Slope (avg)
MilliQ	1.0366	0.0130		2.45	0.011
Std 1	0.7465	0.0090	-0.0252	(Std1 Avg)	Slope 1:
Std 2	0.9845	0.0120	0.4728		0.0072
Std 3	1.0791	0.0165	0.4397		Slope 2:
Std 4	1.3873	0.0167	0.0052		0.013
Average		0.0135			

**Internal Lag time: ≈ 5 minutes**



**Figure 75: SXS3\_xsct5 standards**

## Results

**Table 59: SXS3\_xsct5 standards**

<i>Standard</i>	<i>Concentration</i>	<i>Date</i>	<i>Month</i>	<i>Absorbance</i>	<i>Stdev</i>
Std 1	0.18	8/8/09	13:23	0.7591	0.00710
Std 1	0.19	8/8/09	13:36	0.7340	0.01084
Std 2	4.64	8/8/09	13:46	0.7481	0.00801
Std 3	18.77	8/8/09	14:04	0.8593	0.01456
Std 4	46.78	8/8/09	14:09	1.0748	0.01414
Std 2	4.64	8/9/09	11:59	1.2209	0.01592
Std 3	18.77	8/9/09	12:14	1.2990	0.01854
Std 4	46.78	8/9/09	12:41	1.6997	0.01931
MilliQ	0	8/9/09	12:51	1.0366	0.01299

**Table 60: SXS3\_xsct5 discrete samples**

<i>Date</i>	<i>Time</i>	<i>Sample</i>	<i>Abs</i>	<i>Blank Corr</i>	<i>Stdev</i>	<i>Conc.</i>	<i>Date run</i>
8/8/09	16:53	#83	1.0295	0.9551	0.00740	118.92	9/14/09

**Table 61: SXS3\_xsct5 notes**

8/8/09	13:00	standards
8/8/09	14:10	Begin TFF flow
8/9/09	2:15	topped off DPD and hydrogen peroxide
8/9/09	3:06	Switched to surface fish TFF (SS problem)
8/9/09	4:02	Switched back to TFF
8/9/09	10:16	Sample line came off, fixed about 10:25
8/9/09	10:39	acidified CSW seawater
8/9/09	11:56	fixed flow problem (bubble stuck in detector)
8/9/09	11:34	begin standards

### 4.3.2.1.8 August 9—Waldport Line (SXS3\_xsect6)

The Waldport line was run on August 9-10, 2009. There were a couple small problems with the supersucker (switched to the surface fish TFF at 04:15 and back to supersucker at 04:49) and with the GCFA (bubble in the flow cell at 6:13). When switching to standards at the end of the run an airblock was created in the sample line—milliQ was forced into the tubing with a syringe to clear. The signal resumed but it was rather noisy which was problematic for the final set of standards.

## Results

A series of discrete samples were also collected at the TFF. Some discrete samples were measured as being lower in concentration than the inline samples while others were measured as higher than their inline counterparts.

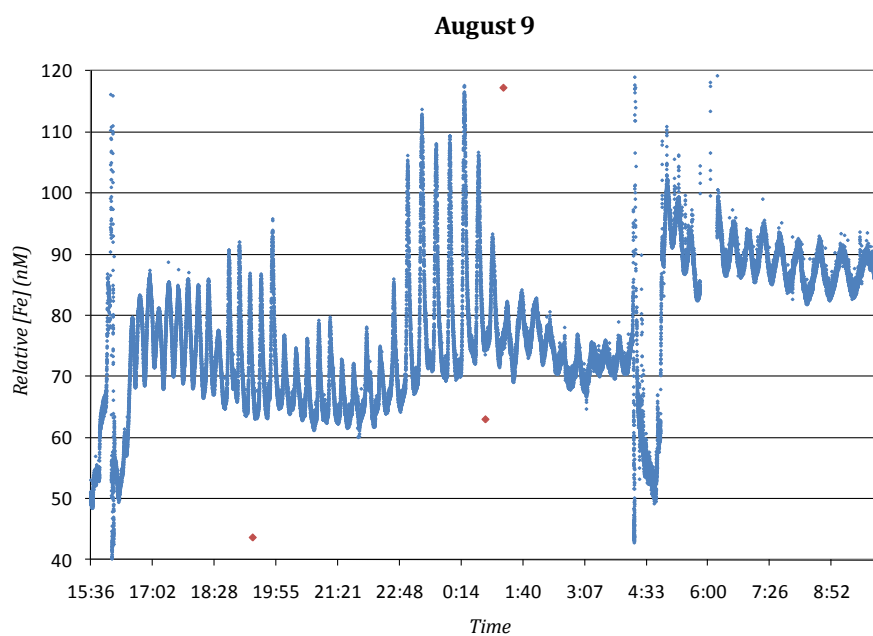


Figure 76: Relative [Fe] (SXS3\_xsct6)

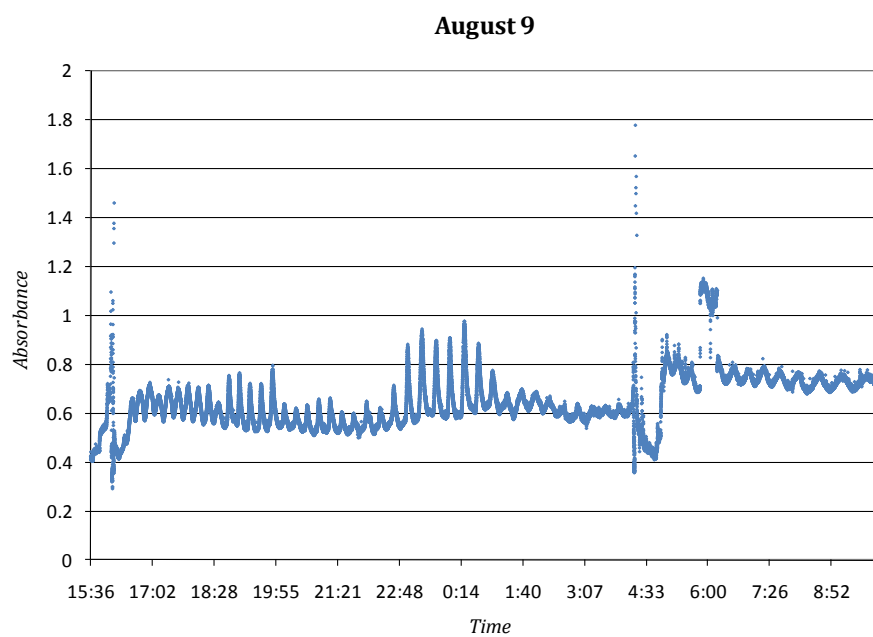


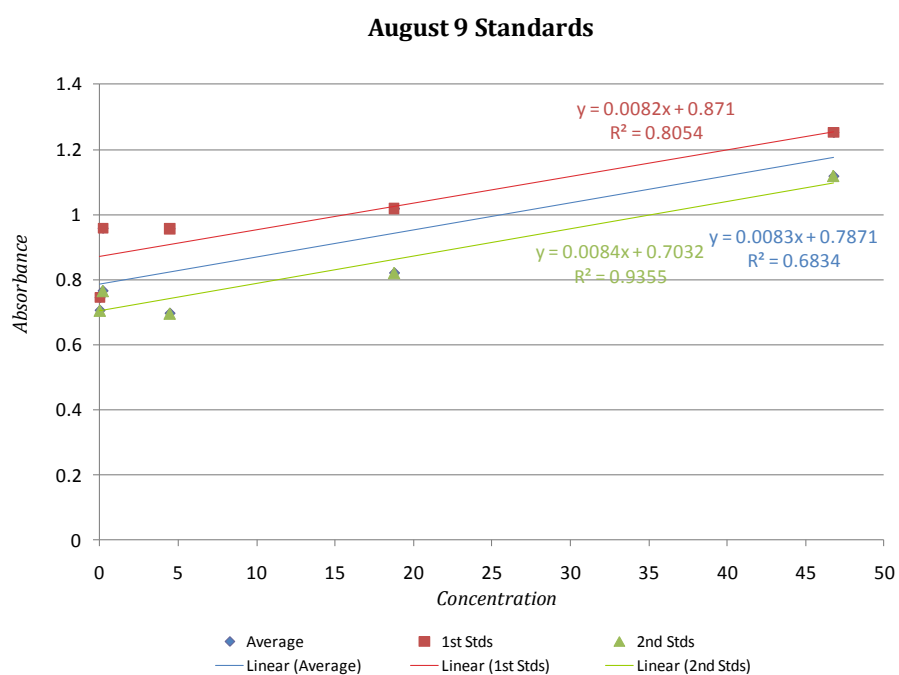
Figure 77: Absorbance (SXS3\_xsct6)

## Results

**Table 62: SXS3\_xsct6 summary**

	Absorbance	Stdev	Drift (Abs 2-Abs1)	Detection Limit:	Slope (avg)
MilliQ	0.7256	0.0143	-0.0391	7.88	0.0083
Std 1	0.8621	0.0218	-0.1920	(Std1 Avg)	Slope 1:
Std 2	0.8270	0.0191	-0.2599		0.0082
Std 3	0.9197	0.0215	-0.1974		Slope 2:
Std 4	1.1854	0.0259	-0.1344		0.0084
Average		0.0205			

**Internal Lag time: ≈ 5 minutes**



**Figure 78: SXS3\_xsct6 standards**

## Results

**Table 63: SXS3\_xsct6 standards**

<i>Standard</i>	<i>Concentration</i>	<i>Date</i>	<i>Time</i>	<i>Absorbance</i>	<i>Stdev</i>
MilliQ	0	8/9/2009	14:39	0.7451	0.00579
Std 1	0.19	8/9/2009	15:17	0.9581	0.01284
Std 2	4.46	8/9/2009	15:32	0.9569	0.01179
Std 3	18.77	8/9/2009	15:41	1.0184	0.01276
Std 4	46.78	8/9/2009	15:55	1.2526	0.01485
MilliQ	0	8/10/2009	11:28	0.7060	0.02276
Std 1	0.19	8/10/2009	12:06	0.7661	0.03078
Std 2	4.46	8/10/2009	12:17	0.6970	0.02644
Std 3	18.77	8/10/2009	12:40	0.8210	0.03019
Std 4	46.78	8/10/2009	12:57	1.1182	0.03699

**Table 64: SXS3\_xsct6 discrete samples**

<i>Date</i>	<i>Time</i>	<i>Sample</i>	<i>Abs</i>	<i>Blank Corr</i>	<i>Stdev</i>	<i>Concentration</i>	<i>Date run</i>
8/9/09	19:23	#91	0.3970	0.322644	0.010673	43.62	9/14/09
8/10/09	0:48	#94	0.5595	0.485114	0.010914	62.96	9/14/09
8/10/09	1:10	#95	1.1455	1.071144	0.012405	132.73	9/14/09
8/10/09	1:13	#96	1.0155	0.941144	0.007313	117.25	9/15/09

**Table 65: SXS3\_xsct6 notes**

8/9/09	14:50	begin standards
8/9/09	16:16	TFF
8/10/09	4:20	surface TFF
8/10/09	5:48	refilled buffer bottle
8/10/09	6:06	bubble in flow cell
8/10/09	10:52	standards

### 4.3.2.1.9 August 10—Newport Line (SXS3\_xsct7)

The transect run on August 10-August 11 covered the Newport line going from west to east. This run went quite smoothly aside from some issues with the second set of standards.

## Results

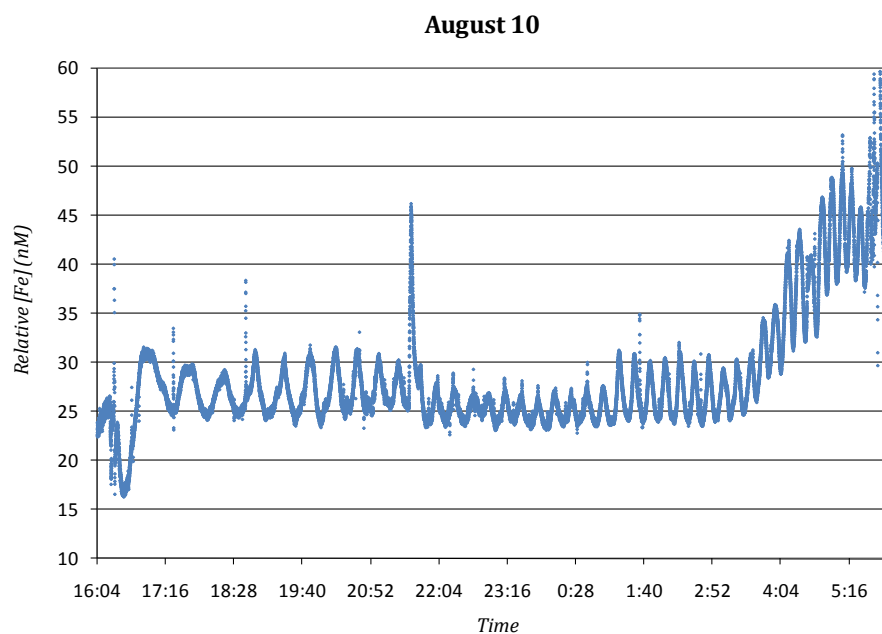


Figure 79: Relative [Fe] (SXS3\_xsct7)

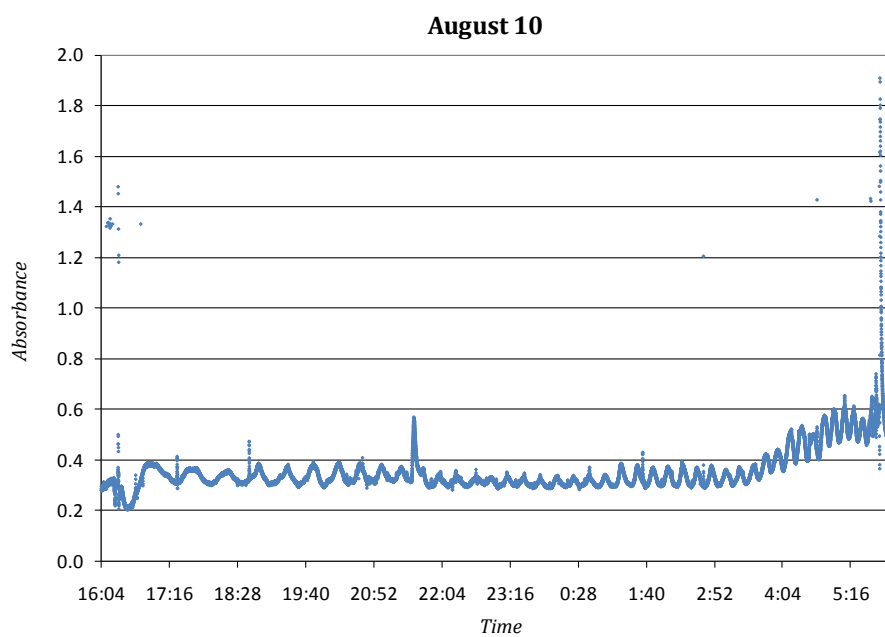


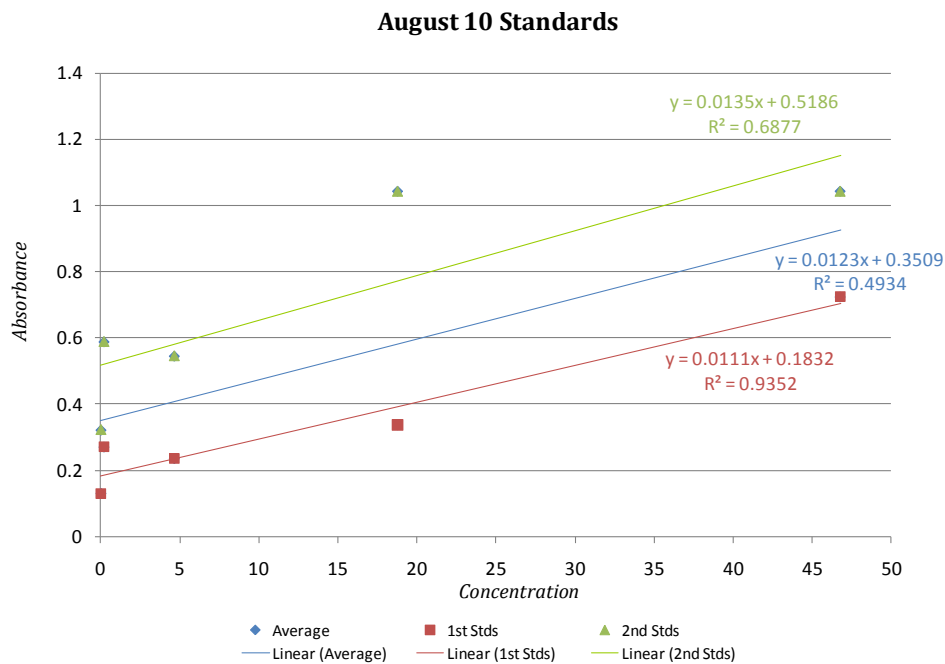
Figure 80: Absorbance (SXS3\_xsct7)

## Results

**Table 66: SXS3\_xsct7 summary**

	Absorbance	Stdev	Drift (Abs 2-Abs1)	Detection Limit:	Slope (avg)
MilliQ	0.2259	0.0094	0.1911	2.29	0.0123
Std 1	0.4293	0.1568	0.3180	(MilliQ Avg)	Slope 1:
Std 2	0.3903	0.0140	0.3096		0.0135
Std 3	0.6906	0.0060	0.7065		Slope 2:
Std 4		0.0069	1.0438		0.0111
Average		0.0386			

**Internal Lag time: ≈ 5 minutes**



**Figure 81: SXS3\_xsct7 standards**

**Table 67: SXS3\_xsct7 standards**

Standard	Concentration	Date	Time	Absorbance	Stdev
MilliQ	0	8/10/09	15:03	0.1304	0.00825
Std 1	0.19	8/10/09	15:14	0.2703	0.01273
Std 2	4.64	8/10/09	15:25	0.2355	0.00942
Std 3	18.77	8/10/09	15:45	0.3373	0.01196
Std 4	46.78	8/10/09	16:17	0.7242	0.01371
Std 4	46.78	8/11/09	6:03	1.0438	0
Std 3	18.77	8/11/09	6:25	1.0438	0
Std 2	4.64	8/11/09	6:40	0.5452	0.01849
Std 1	0.19	8/11/09	6:44	0.5883	0.30089
MilliQ	0	8/11/09	6:57	0.3214	0.01055

## Results

**Table 68: SXS3\_xsct7 discrete samples**

Date	Time	Sample	Abs	Blank Corr	Stdev	Conc.	Date run
8/11/09	5:47	#115	1.7566	1.7458	0.00751	260.86	9/15/09
8/11/09	5:47	#118	1.8233	1.8125	0.01519	270.97	9/15/09

**Table 69: SXS3\_notes**

8/10/09	14:56	begin standards
8/10/09	16:34	TFF
8/11/09	5:50	standards

### 4.3.2.2 Discussion

#### 4.3.2.2.1 High Resolution Data SXS3

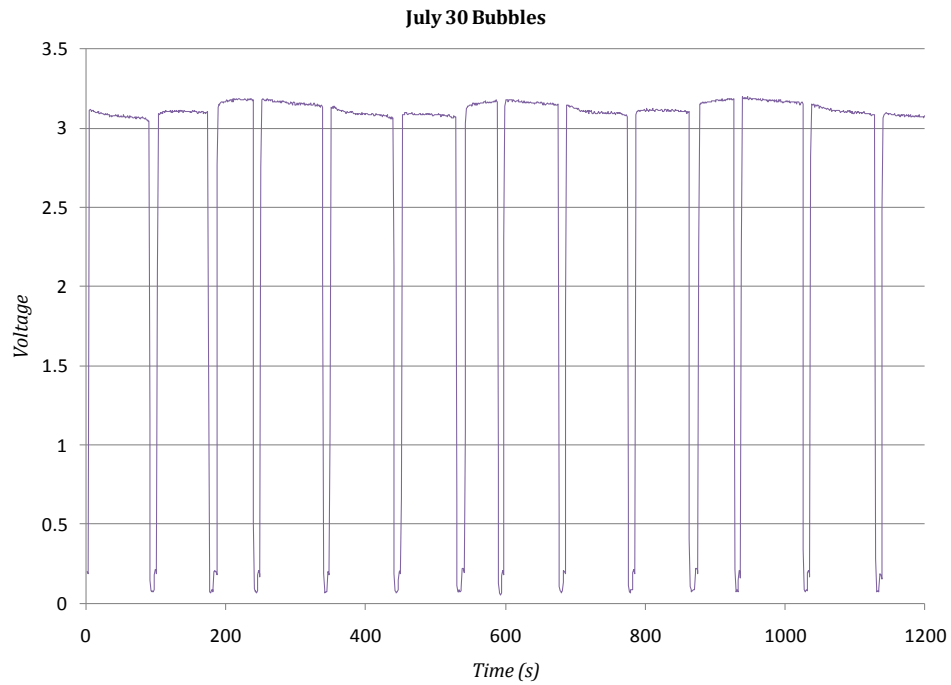


Figure 82: Early SXS3 bubble pattern (20 minutes)

## Results

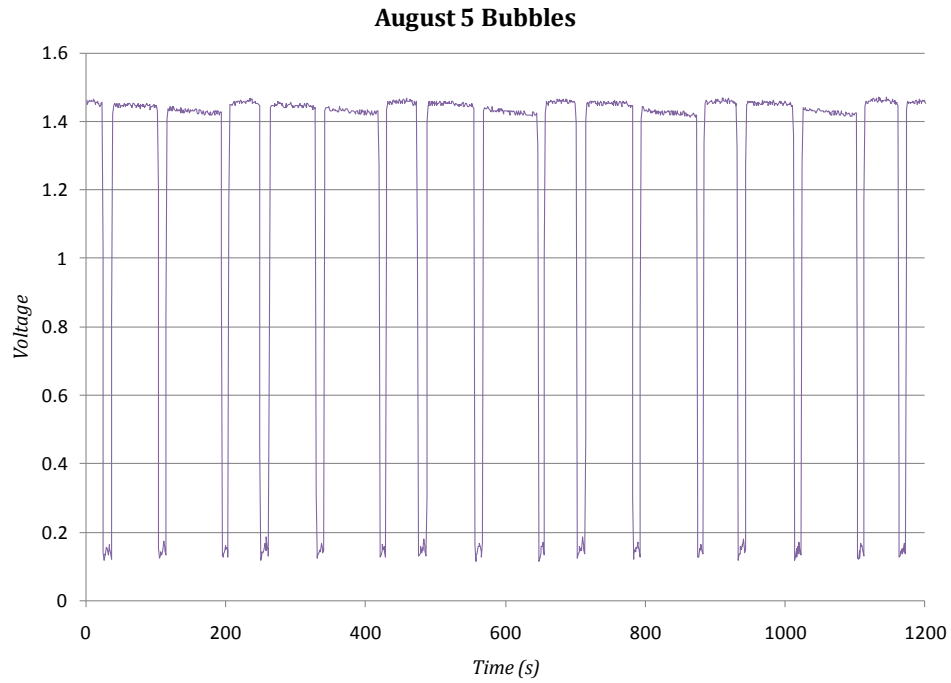


Figure 83: Mid-SXS3 bubble pattern (20 minutes)

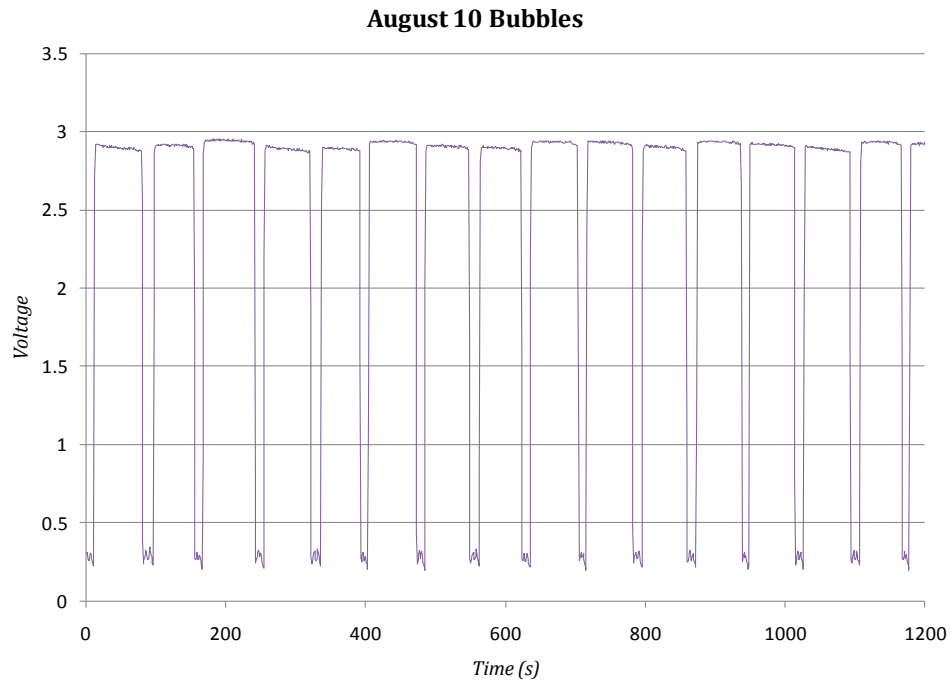


Figure 84: End SXS3 bubble pattern (20 minutes)

The changes in bubble patterns during the August cruise are different than during the May cruise. The rolling variation in the July 30 data is characteristic of early in the cruise. It began to dissipate (as in the August 5 data) and eventually subsided into the best bubble patterns seen on any of the SUCCES cruises as seen in the August 10 data.

## Results

### 4.3.2.2.2 1 Hour Data SXS3

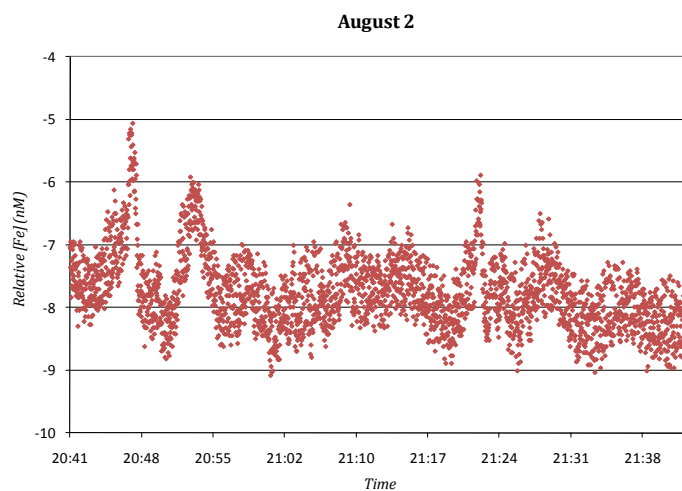


Figure 85: August 2, 1 Hour Data

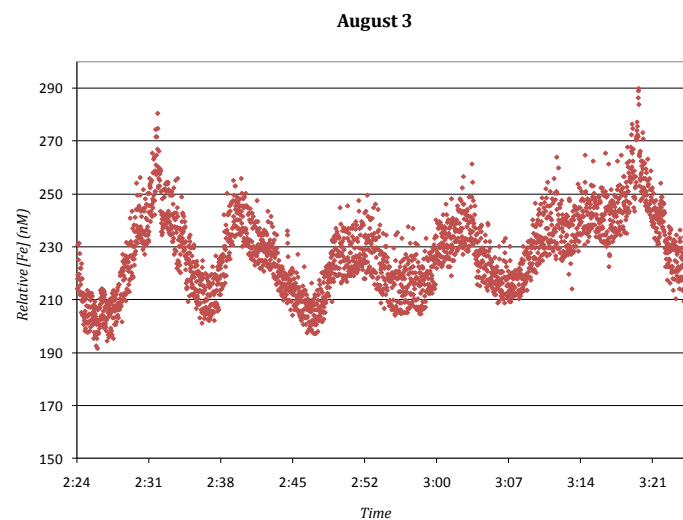


Figure 86: August 3, 1 Hour Data

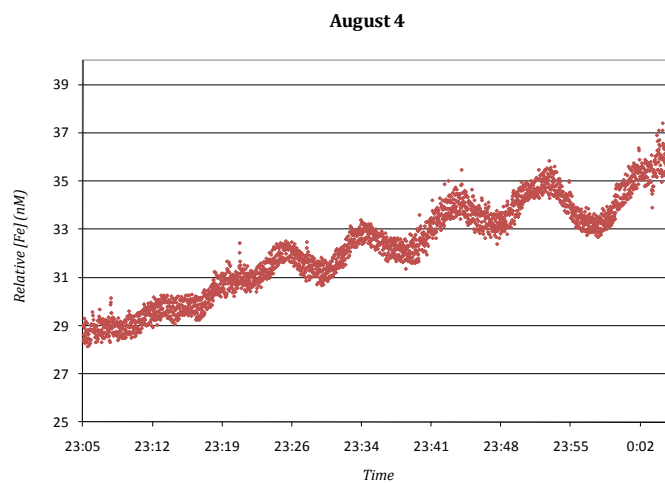


Figure 88: August 4, 1 Hour Data

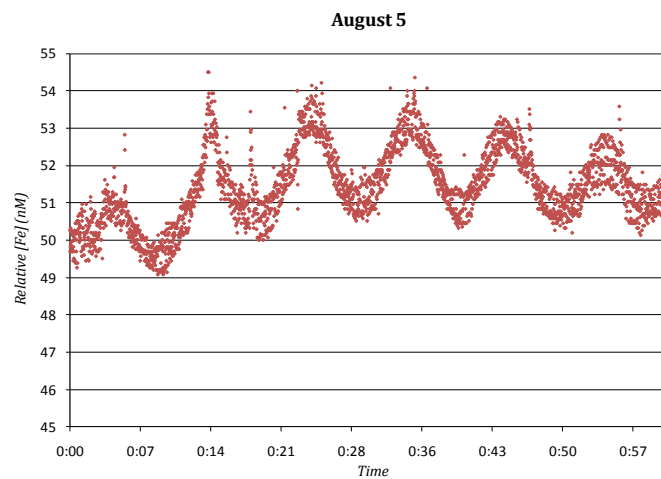


Figure 87: August 5, 1 Hour Data

## Results

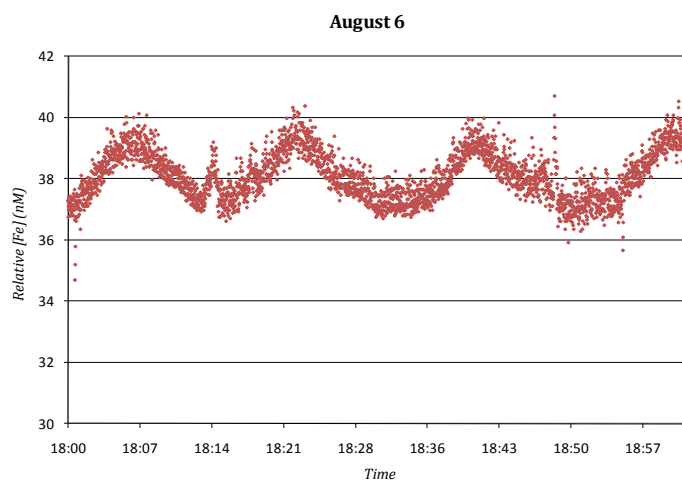


Figure 89: August 6, 1 Hour Data

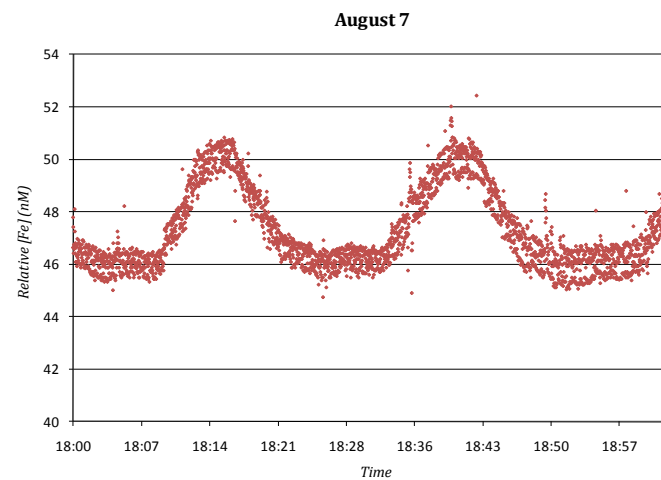


Figure 90: August 7, 1 Hour Data

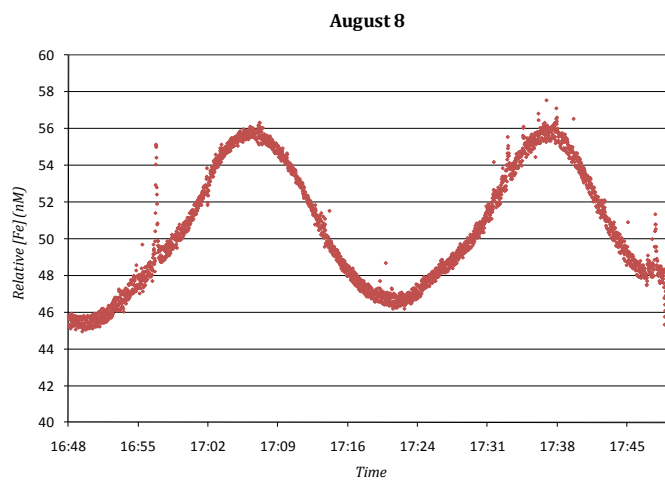


Figure 91: August 8, 1 Hour Data

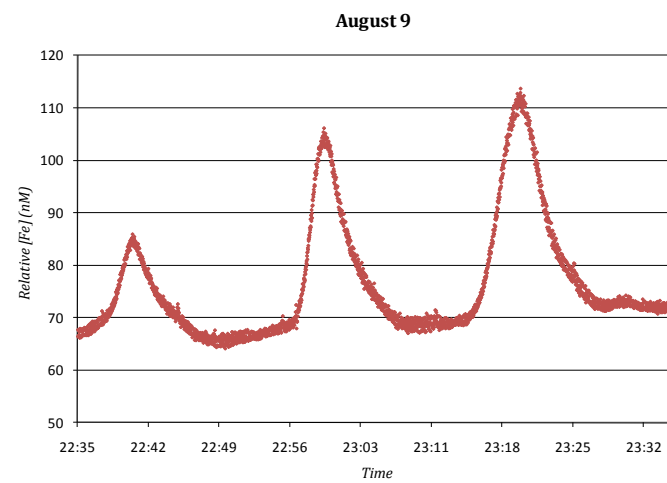


Figure 92: August 9, 1 Hour Data

## Results

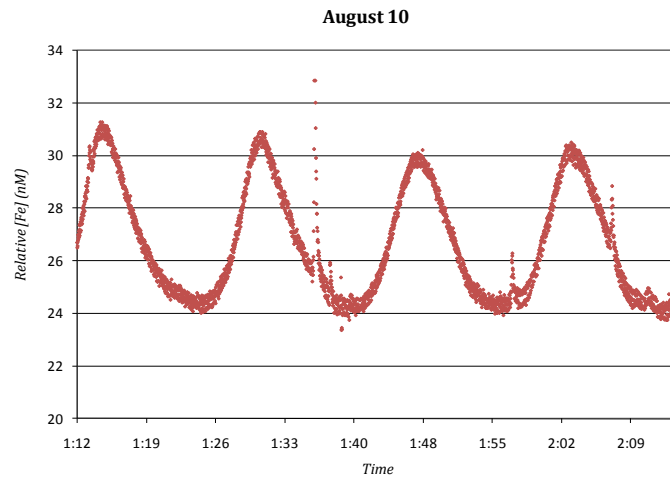


Figure 93: August 10, 1 Hour Data

## Results

### 4.3.2.2.3 Standard Voltages

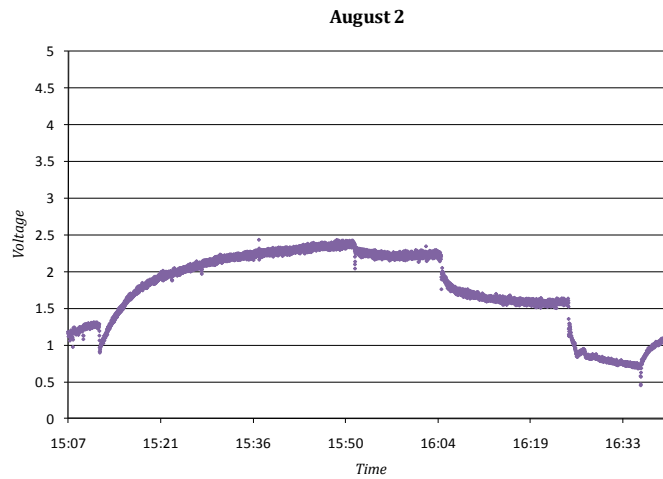


Figure 94: August 2 Standards (1st set)

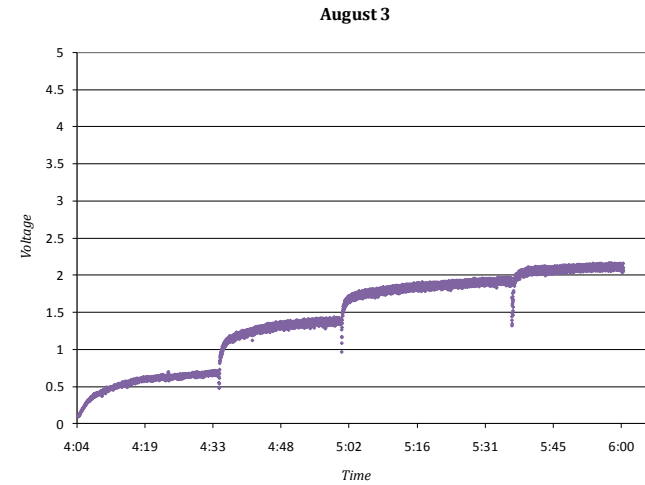


Figure 95: August 3 Standards (2nd set)

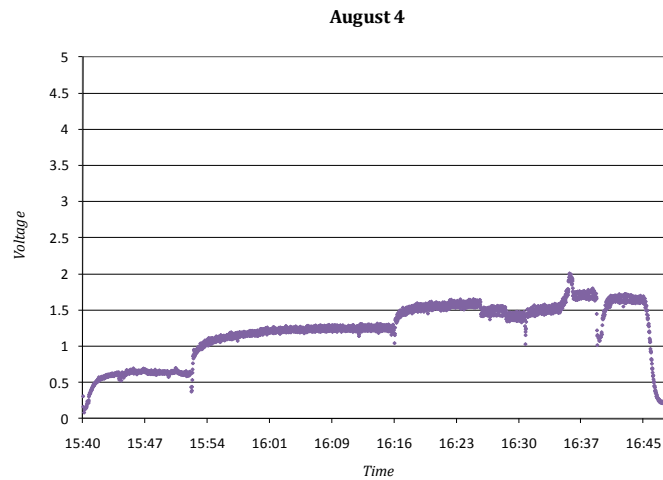


Figure 96: August 4 Standards (1st set)

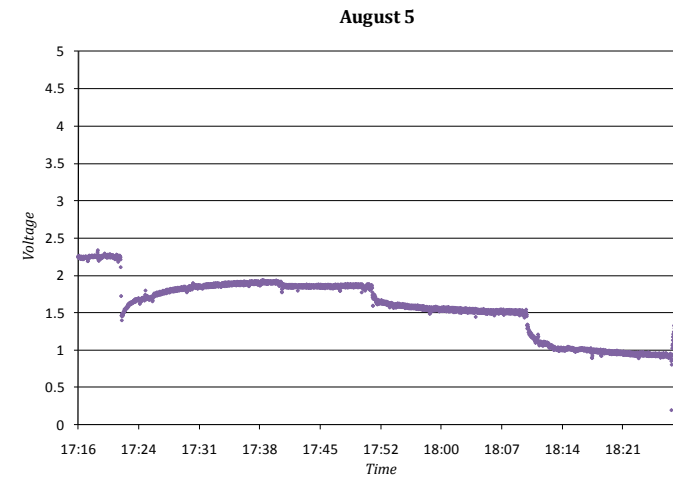


Figure 97: August 5 Standards (1st set)

## Results

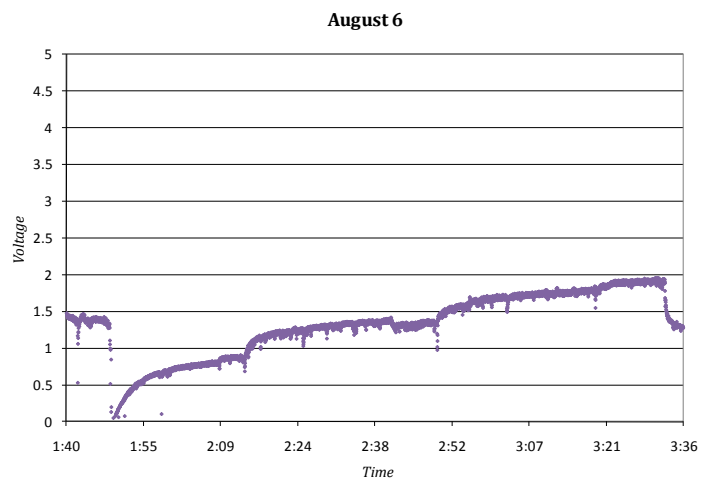


Figure 98: August 6 Standards (2nd set)

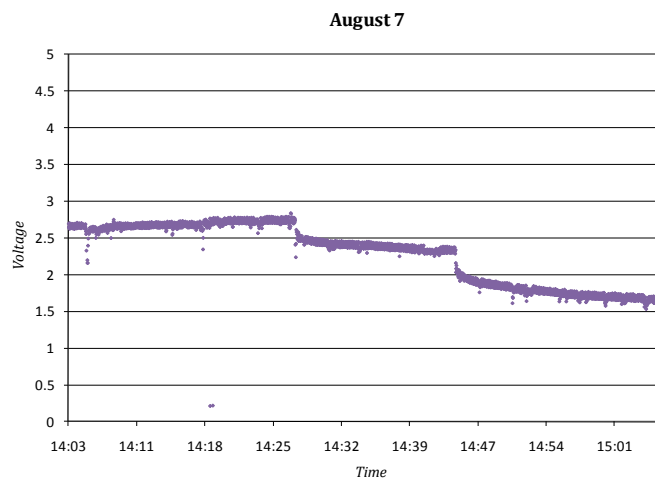


Figure 99: August 7 Standards (1st set)

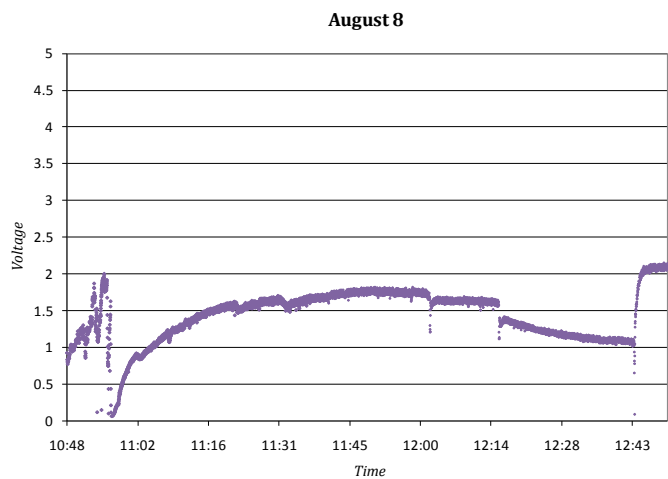


Figure 101: August 8 Standards (1st set)

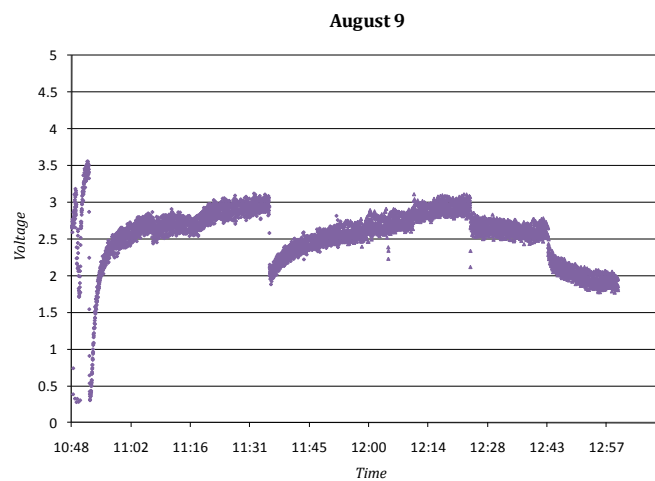


Figure 100: August 9 Standards (1st set)

## Results

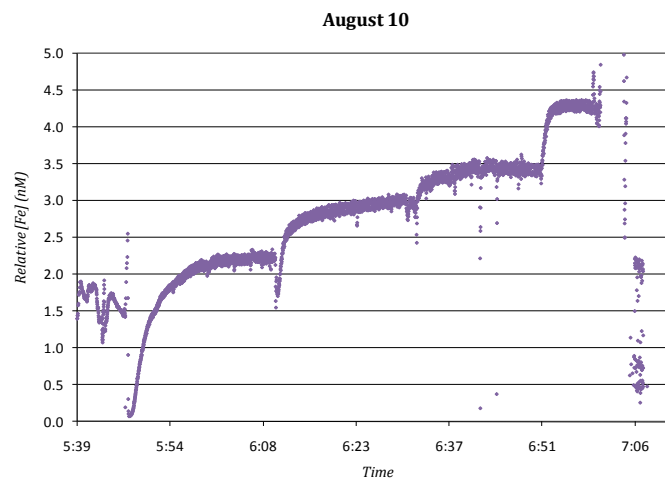


Figure 102: August 10 Standards (2nd set)

### 4.3.3 Cruise Data

**Table 70: Cruise data files**

<b><i>Transect</i></b>		<b><i>File</i></b>	<b><i>Descriptions</i></b>
<b>SXS2</b>			
May 23	45°N Transect SXS2_xsct1	May23.xlsx	May 23 summary of standards and discrete samples, charts of [Fe] and absorbance
		May23data.csv	Processed data file from LabView
		May23times.csv	Peak picking times used in Matlab
May 25	Patch, Day 1 SXS2_srvy1	May25.xlsx	May 25 summary file
		May25data.csv	Processed data file from LabView
		May25times.csv	Peak picking times used in Matlab
May 26	Patch, Day 2 SXS2_srvy2	May26.xlsx	May 26 summary file
		May26data.csv	Processed data file from LabView
		May26times.csv	Peak picking times used in Matlab
May 27	Patch, Day 3 SXS2_srvy3	May27.xlsx	May 27 summary file
		May27data.csv	Processed data from LabView
		May27times.csv	Peak picking times used in Matlab
May 28	Patch, Day 4 SXS2_srvy4	May28.xlsx	May 28 summary file
		May28data.csv	Processed data from LabView
		May28times.csv	Peak picking times used in Matlab
May 29	Patch, Day 5 SXS2_srvy5	May29.xlsx	May 29 summary file
		May29data.csv	Processed data from LabView
		May29times.csv	Peak picking times used in Matlab
May 30	45°N Transect SXS2_xsct2	May30.xlsx	May 30 summary file
		May30data.csv	Processed data from LabView
		May30times.csv	Peak picking times used in Matlab
May 31	43.9°N Transect SXS2_xsct3	May31.xlsx	May 31 summary file
		May31data.csv	Processed data from LabView
		May31times.csv	Peak picking times used in Matlab
<b>SXS3</b>			
August 2	Patch, Day 1 SXS3_srvy1	Aug2.xlsx	August 2 summary of standards and discrete samples, charts of [Fe] and absorbance
		Aug2data.csv	Processed data file from LabView
		Aug2times.csv	Peak picking times used in Matlab
August 3	Patch, Day 2 SXS3_srvy2	Aug3.xlsx	August 3 summary file
		Aug3data.csv	Processed data file from LabView
		Aug3times.csv	Peak picking times used in Matlab
August 4	Patch, Day 3 SXS3_srvy3	Aug4.xlsx	August 4 summary file
		Aug4data.csv	Processed data file from LabView
		Aug4times.csv	Peak picking times used in Matlab

## Discussion

August 5	Patch, Day 4 SXS3_srvy4	Aug5.xlsx	August 5 summary file
		Aug5data.csv	Processed data file from LabView
		Aug5times.csv	Peak picking times used in Matlab
August 6	Patch, Day 5 SXS3_srvy5	Aug6.xlsx	August 6 summary file
		Aug6data.csv	Processed data file from LabView
		Aug6times.csv	Peak picking times used in Matlab
August 7	45°N Transect SXS3_xsct4	Aug7.xlsx	August 7 summary file
		Aug7data.csv	Processed data file from LabView
		Aug7times.csv	Peak picking times used in Matlab
August 8	43.9°N Transect SXS3_xsct5	Aug8.xlsx	August 8 summary file
		Aug8data.csv	Processed data file from LabView
		Aug8times.csv	Peak picking times used in Matlab
August 9	Waldport Line SXS3_xsct6	Aug9.xlsx	August 9 summary file
		Aug9data.csv	Processed data file from LabView
		Aug9times.csv	Peak picking times used in Matlab
August 10	Newport Line SXS3_xsct7	Aug10.xlsx	August 10 summary file
		Aug10data.csv	Processed data file from LabView
		Aug10times.csv	Peak picking times used in Matlab

## 5 Discussion

As it was deployed on SUCCES2 (May 2009) and SUCCES3 (August 2009) there were both failures and successes with the GCFA. The system was improved from the May cruise to the August cruise by focusing on the need to have uniformly sized bubbles that would proceed through the system without breaking or smearing. These improvements seemed to boost the system's ability to show changes in iron concentration.

The system still exhibits a high and widely varying detection limit. During the cruises, the detection limit varied from about 1.6 nM to as high as 5.1 nM. In the lab, the detection limit varied just as much (see chart).

## Discussion

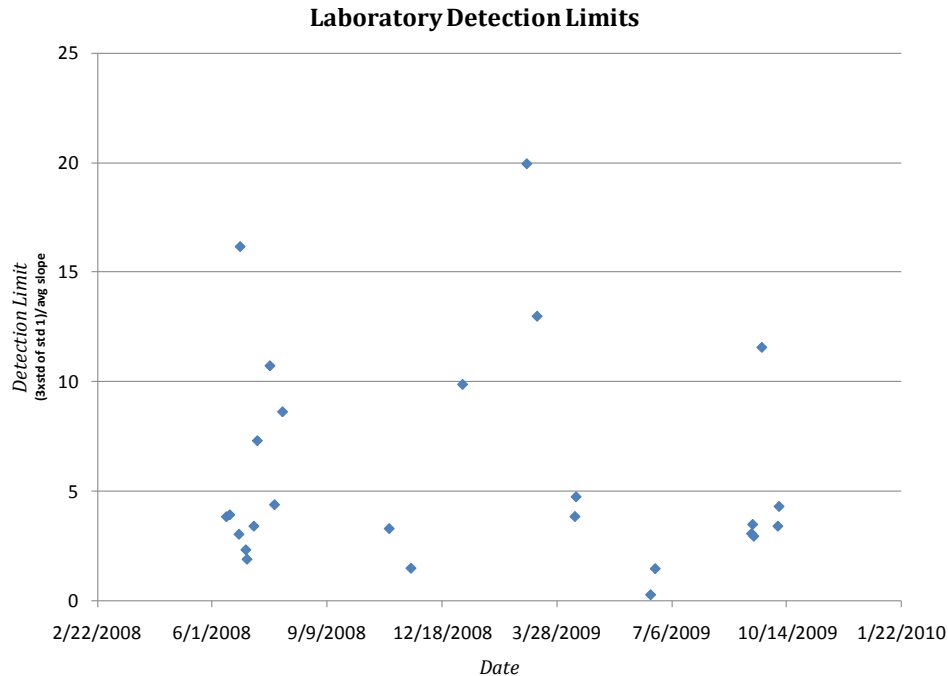


Figure 103: Laboratory detection limits

Issues with the detection limit may be attributable to the inability to achieve consistent bubbles. When the bubbles lack consistency, the reagents may not be mixed in the same proportion in each sample “packet” leading to different measurements in each one. This variation would boost the standard deviation from the mean and lead to a higher detection limit.

The system also has a response time issue that has not been completely quantified. Sometimes it appears that the system responds quite quickly to a change in concentration while others it seems to respond excruciatingly slowly. The pattern of response does not change, however. A change in concentration is characterized by a large and fast jump that accounts for as much as 70% of the total change in voltage. This initial change is followed by a much longer, slower, and lower magnitude “ramping up” or “ramping down” of voltage. The reasons for the slow response time have not been established. Iron could be “sticking” to the inside of the tubing although the concentration is seen as ramping down when going from a low standard to a high one. There is also the possibility that the glass tubing heightens the sticking issue. Glass tubing is often rejected in iron work as it is known to have a high iron background. For our work however, the glass tubing is crucial to maintaining consistent bubbles. The reason for this ramping may also not be thought of yet.

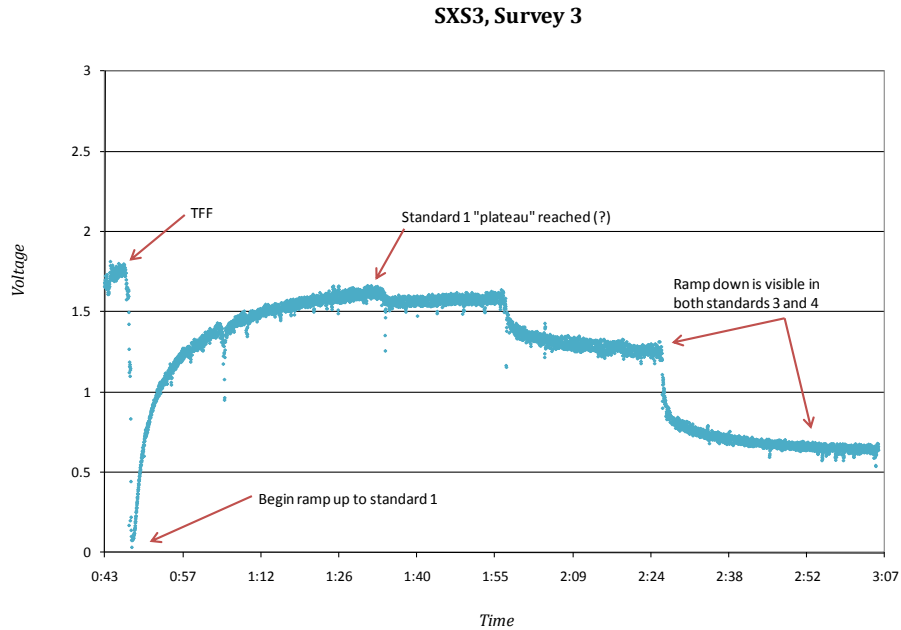


Figure 104: Standard ramping

The chart above illustrates the final set of standards on day 3 of surveying during SXS3. The "ramping down effect" is seen in the changes to the 3rd and 4th standards. Standard 4 particularly exemplifies the fast initial drop followed by a long and slow secondary change. The chart also illustrates a problem that may be connected to the response time issue. The system does not seem to transition well from water flowing from the TFF to the first standard. The long slow ramp up to standard 1's plateau limits our ability to run standards in the middle of the day because it takes up to an hour and a half. From the time the instrument first detected the beginning of standard 1 to the time it reached its plateau was almost 50 minutes.

On board the ship, the system appears to respond fast enough to catch a signal as the supersucker was moved from the bottom boundary layer to the surface. It was discovered, accidentally, that there is enough pressure from the TFF to push water over the peristaltic pump even when it is not operating. If the sample was pushed through the system at a higher rate than the standards or samples in the lab, this could possibly have had the effect of "sweeping" iron through the system before it had a chance to stick to anything.

The system also occasionally has problems accurately measuring the concentration of standard and internal reference materials. NASS-5 was more often than not measured at concentrations of about 10 nM when in reality it is about 3.7 nM. This is partially accounted for by the poor

## Discussion

detection limit but also points towards some other lingering issues with standards and general feasibility of the method.

The most damning problem may be the discrepancy between the discrete and inline samples. Several of these samples were run during the cruises at the end of a day's run or as part of the next day's standards and did not match up with their inline counterpart. The majority of the samples from SXS2 and SXS3 were run in the lab. While it was evident the system was having some problems when the discrete samples from SXS3 were run in lab (the NASS-5 concentration was off by as much as a factor of 5) the discrete samples did not remotely resemble the concentrations measured inline by the GCFA.

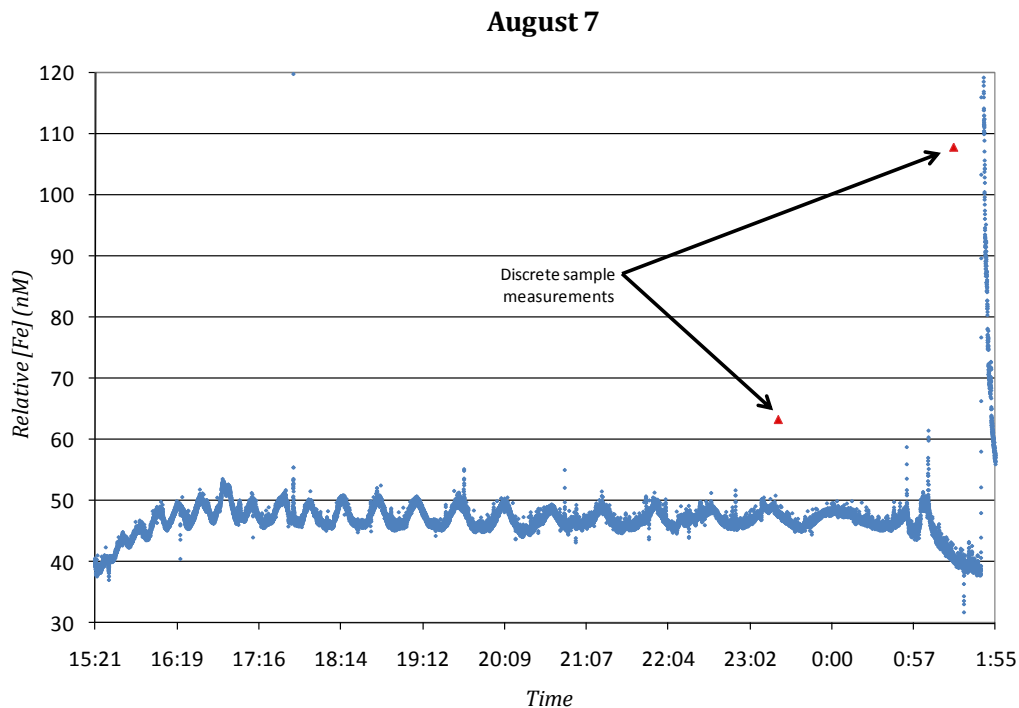


Figure 105: Inline vs discrete discrepancies

The data from the May cruise is not complete but the samples run from SXS2 do not appear to match their inline counterparts either. With inline acidification, temperature, and pH eliminated in the laboratory this requires a new way of looking at it to determine the issues. Perhaps the increased flow rates for sample cause the sample to be under-acidified with respect to both concentration of HCl and time.

## Discussion

In order for the GSCFA to be deployed successfully the offsets between the discrete samples and the inline samples must be resolved. Additionally, work should be done to quantify and improve the response time as well as decreasing the detection limit.

## References

## 6 References

- Bowie, A.R., E.P. Achterberg, P.L. Croot, H.J.W. de Baar, P. Laan, J.W. Moffett, S. Ussher, and P.J. Worsfold. 2006. A community-wide intercomparison exercise for the determination of dissolved iron in seawater. *Mar. Chem.* 98:81-99.
- Chase, Z., A. van Geen, P.M. Kosro, J. Marra, P.A. Wheeler. 2002. Iron, nutrient, and phytoplankton distributions in Oregon coastal waters. *J. Geophys. Res.* 107(C10), 3174, doi:10.1029/2001JC000977.
- Chase, Z., B. Hales, T. Cowles, R. Schwartz, A. van Geen. 2005. Distribution and variability of iron input to Oregon coastal waters during the upwelling season. *J. Geophys. Res.* 110, C10S12, doi:10.1029/2004JC002590.
- Chase, Z., K.S. Johnson, V.A. Elrod, J.N. Plant, S.E. Fitzwater, L. Pickell, C.M. Sakamoto. 2005. Manganese and iron distributions off central California influenced by upwelling and shelf width. *Mar. Chem.* 95:235-254.
- Chase, Z., P.G. Strutton, B.H. Hales. 2007. Iron links river runoff and shelf width to phytoplankton biomass along the U.S. West Coast. *Geophys. Res. Lett.* 34, L04607, doi:10.1029/2006GL028069.
- Hales, B., L. Karp-Boss, A. Perlin, P.A. Wheeler. 2006. Oxygen production and carbon sequestration in an upwelling coastal margin. *Global Biogeochem. Cycles* 20, GB2001, doi:10.1929/2005GB002517.
- Hales, B., T. Takahasi, L. Bandstra. 2005. Atmospheric CO<sub>2</sub> uptake by a coastal upwelling system. *Global Biogeochem. Cycles* 19, GB1009, doi:10.1029/2004GB002295.
- Hirayama, K. and N. Unohara. 1988. Spectrophotometric catalytic determination of an ultratrace amount of iron(III) in water based on the oxydation of *n,n*-dimethyl-*p*-phenylenediamine by hydrogen peroxide. *Anal. Chem.* 60:2573-2577.
- Lohan, M.C., A.M. AguilarOslas, R.P. Franks, K.W. Bruland. 2005. Determination of iron and copper in seawater at pH 1.7 with a new commercially available chelating resin, NTA Superflow. *Anal. Chem. Acta* 530:121-129.
- Obata, H. and C.M.G. van den Berg. 2001. Determination of picomolar levels of iron in seawater using catalytic cathodic stripping voltammetry. *Anal. Chem.* 73:2522-2528.
- Obata, H., H. Karatani, E. Nakayama. 1993. Automated determination of iron in seawater by chelating resin concentration and chemiluminescence detection. *Anal. Chem.* 65:1524-1528.
- Measures, C.I., J. Yuan, J.A. Resing. 1995. Determination of iron in seawater by flow injection analysis using in-line preconcentration and spectrophotometric detection. *Mar. Chem.* 50:3-12.

## References

Wu, J., and E.A. Boyle. 1998. Determination of iron in seawater by high-resolution isotope dilution inductively coupled plasma mass spectrometry after  $\text{Mg}(\text{OH})_2$  coprecipitation. *Anal. Chim. Acta* 367:183-191.

Zhang, J., C. Kelble, F.J. Millero. 2001. Gas-segmented continuous flow analysis of iron in water with a long liquid waveguide capillary flow cell. *Anal. Chim. Acta* 438:59-57.