#### **Supporting Information**

12 pgs, 4 figures, 4 tables

# Predicting polycyclic aromatic hydrocarbon concentrations in resident aquatic organisms using passive samplers and partial least squares calibration

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### MATERIAL AND METHOD DETAILS

**Crayfish sample preparation and chemical analysis**. The analytical method used to analyze PAHs in crayfish tissues was adapted from previous work (1). Briefly, 200 - 400 mg of tissue were transferred to a 15 mL BD Falcon tube, thawed, and spiked with  $20 \ \mu$ L of a PAH surrogate standard mixture containing acenapthene-D10, pyrene-D10, and indeno[1,2,3-*cd*]pyrene-D12. Samples were subsequently spiked with  $500 \ \mu$ L of H<sub>2</sub>O, capped, and hand shaken for 1 min. A 10 mL aliquot of ethyl acetate, acetone, and iso-octane (2:2:1, v/v/v) was added to each tube and the resulting solution was shaken for 5 min. QuEChERS salts (650 mg) were added to the tube, mixed for five minutes by hand shaking, and tubes were then centrifuged at 3800 g for 5 min. Extracts (9mL) were transferred to 15 mL volumetric conical glass tubes, solvent exchanged to ~ 400  $\mu$ L of *n*-hexane, and cleaned using *n*-hexane conditioned solid-phase extraction cartridges containing primary-secondary amines (Agilent Technologies, Santa Clara, CA). Analytes were

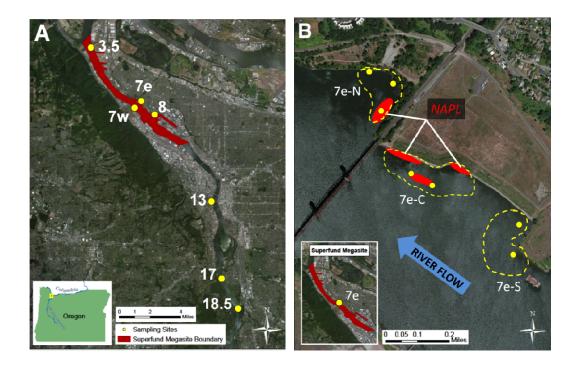
vacuum eluted with 7 mL of *n*-hexane at a flow rate of 2-3 drops/s, concentrated to a final volume of 100  $\mu$ L, and spiked with 10  $\mu$ L of recovery internal standard mixture composed of naphthalene-D8, acenaphthylene-D8, phenanthrene-D10, fluoranthene-D10, chrysene-D12, benzo[*a*]pyrene-D12, and benzo[*ghi*]perylene-D12.

Crayfish samples were chemically analyzed using an Agilent 5975B GC-MS (Santa Clara, CA) equipped with electron impact ionization (70 eV) source and a DB-5MS capillary column (30 m length, 0.25 µm film thickness, 0.25 mm I.D., Agilent J&W). The GC injection port received a 1 µL injection and was operated at 300 °C in pulsed splitless mode. PAHs were chromatographically resolved using the following temperature program: initial oven temperature was 70 °C, 1 min hold, ramp to 300 °C at 10 °C/min, 4 min hold, ramp to 310 °C at 10 °C/min, 7 min hold for a total run time of 36 min. Seven point internal standard calibration curves ranged from 1 to 1000 ng/mL and were plotted as relative response ratios.

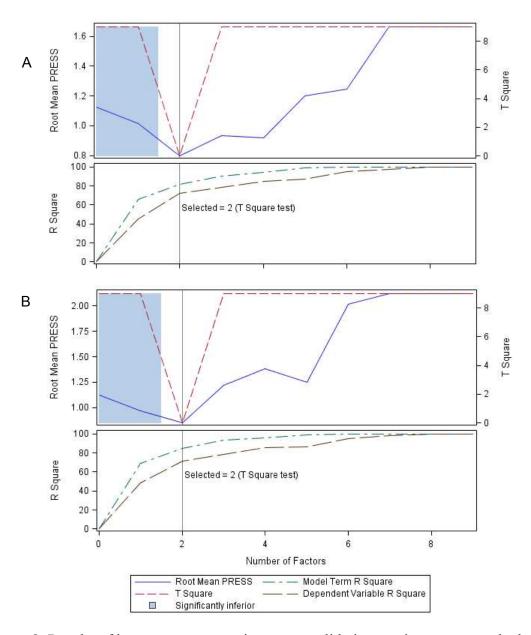
**Quality control**. PAHs were not detected in blank field or trip SPMDs, and RSDs between duplicate site averages were  $\leq 15\%$ . Analytical batches consisted of > 30% quality control samples including method blanks, instrument blanks, and continuing calibration verification standards. Crayfish method blanks contained trace level (< 5.8 ng/mL) background responses for NAP, FLO, and PHE, while all other target analytes were below reporting limits. PAH responses were at least three times greater than sample background, below detection in instrument blanks, and continuing calibration verification samples were within +/- 20% of expected values. Surrogate standard recoveries in crayfish samples were  $62 \pm 15$ ,  $75 \pm 9$ , and  $76 \pm 16\%$  of expected values for ACE-D10, PYR-D10, and IPY-D12 respectively, while average surrogate recoveries in SPMD extracts ranged from 35% to 95% for NAP and BPL (2). All results were surrogate recovery corrected.

## **RESULTS AND DISCUSSION DETAILS**

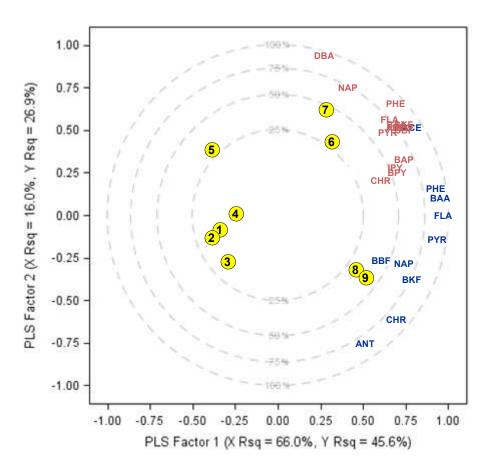
PAH concentrations in the freely dissolved phase. The spatial variation of dissolved PAHs across the study area was investigated from 2002 to 2006 and coincided with crayfish collection reported in the present study (2). The fall 2003 subset of SPMD data was paired with crayfish collection and is presented here for direct comparison to crayfish. Similar to crayfish, median freely dissolved concentrations of  $\Sigma$ PAHs (350 ng/L) and  $\Sigma$ C-PAHs (132 ng/L) were significantly higher at sites within the Superfund compared to up-river sites ( $\Sigma$ PAH and  $\Sigma$ C-PAH = 100 and 18 ng/L, respectively). SPMDs from the McCormick and Baxter Superfund site at river mile 7eC were the only samples with significantly greater levels of individual PAHs compared to up-river samples, while median  $\Sigma$ C-PAH concentrations were higher down-stream at river mile 3.5 (260 ng/L compared to 19 ng/L at up-river sites). Though a number of significant site-specific differences were observed by Sower and Anderson (2), differences between sites in this study were not as stark, likely due in part to decreased statistical power resulting from fewer replicate measurements. Across all sites, crayfish  $\Sigma$ PAH concentrations were on average 280 (range from 21 to 930) times greater than freely dissolved  $\Sigma$ PAH concentrations.



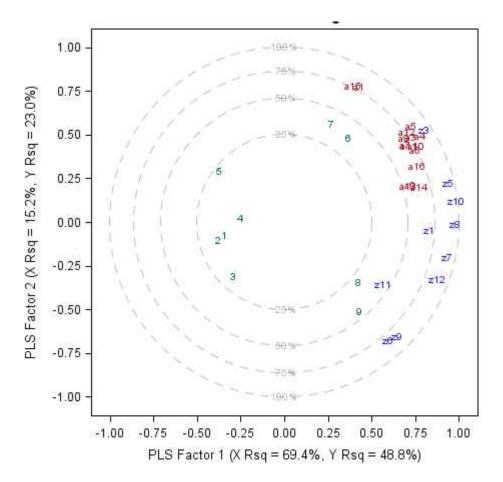
SI Figure 1. Approximate location (yellow dots) of SPMD and crayfish sampling sites in A) the lower 18.5 miles of the Willamette River and at B) river mile 7e during the fall 2003 sampling period. SPMDs and crayfish within yellow dashed circles in B) were paired. The portion of river highlighted red in A) represents the Superfund Mega-site boundary, while numbers indicate river mile. NAPL = non-aqueous phase liquid.



SI Figure 2. Results of leave-one-out-at-a-time cross-validation on site averages calculated by substituting A) 'zero' or B) method detection limits for measurements that were  $\leq$  MDLs. Prior to PLS modeling, predictor variables were 4<sup>th</sup> root transformed then averaged by site, while response variables were averaged by site then 4<sup>th</sup> root transformed. Cross-validation revealed that a two-*factor* PLS model was optimal regardless of how values  $\leq$  MDL were treated.



SI Figure 3. Partial least squares (PLS) correlation loading plot for the first two PLS *factors* modeled on 'zero' substituted data. Shown are sample scores labeled by site number, response variable loadings labeled with red letters, and predictor variable loadings labeled with blue letters. PLS factors 1 and 2 explain 72.5% of the variation in the response variables and 82% of the variation in the predictor variables. The amount of individual variable variation explained corresponds to loading vector magnitudes which can be estimated using the dashed-line circles. Sampling sites are indicated by river mile: 1 - RM 18.5, 2 - RM 17, 3 - RM 13, 4 - RM 8, 5 - RM 7eS, 6 - RM 7eC, 7 - RM 7eN, 8 - RM 7w, 9 - RM 3.5.



SI Figure 4. Partial least squares (PLS) correlation loading plot for the first two PLS factors modeled on 'detection limit' substituted data. Shown are sample scores labeled by site number, response variable loadings 'a' labeled with red letters (crayfish), and predictor variable loadings 'z' labeled with blue letters (freely dissolved). PLS *factors* 1 and 2 explain 71.8% of the variation in the response variables and 84.4% of the variation in the predictor variables. Sampling site are indicated by river mile: 1 – RM 18.5, 2 – RM 17, 3 – RM 13, 4 – RM 8, 5 – RM 7eS, 6 – RM 7eC, 7 – RM 7eN, 8 – RM 7w, 9 – RM 3.5. Numbers associated with loadings identify each PAH; NAP(1), ACE(3), FLO(4), PHE(5), ANT(6), PYR(7), FLA(8), CHR(9), BAA(10), BBF(11), BKF(12), IPY(13), BPY(14), DBA(15), BAP(16).

	S	ex	Pooled morphology (mean $\pm$ SEM)				
Sampling site (river mile)	Male Female		Body length (cm)	Carapace length (cm)	Wet weight (g)		
Up-river							
18.5	3	2	8.1 ± 0.9	3.9 ± 0.5	21.0 ± 10		
17	5	3	9.2 ± 0.4	4.7 ± 0.2	28.4 ± 4.7		
13	3	0	9.4 ± 0.4	$4.5 \pm 0.4$	28.1 ±4.5		
Superfund							
8	0	1	7.5	3.7	13.3		
7e-S	3	4	9.5 ± 0.5	4.6 ± 0.2	20.6 ± 2.3		
7e-C	1	4	8.5 ± 0.5	4.1 ± 0.2	13.9 ± 2.0		
7e-N	11	14	9.3 ± 0.3	4.6 ± 0.2	22.9 ± 2.5		
7w	2	1	9.8 ± 1.2	4.8 ± 0.6	27.4 ± 8.9		
3	0	3	10.5 ± 0.3	5.2 ± 0.1	30.5 ± 2.2		
All sites	28	32	9.2 ± 0.2	4.5 ± 0.1	23.1 ± 1.6		

SI Table 1. Signal crayfish (*P. leniusculus*) sex and morphological characteristics grouped by site for the fall 2003 collection period.

		River	mile 18.	5		River mile 17			River mile 13			
compound	moon	9E	modian	n > MDL	moon	e E	median	n > MDL	moon	e E	median	n > MDL
compound	mean	9E	median	( <i>n</i> = 5)	mean	9E	median	(n = 8)	mean	SE	median	(n = 3)
NAP	23.6	9.0	14.0	5	3.6	1.1	4.9	5	7.8	4.9	6.5	2
ACY	<0.05			0	<0.05			0	<0.05			0
ACE	<0.25	4.0	.0.47	0	< 0.25			0	5.6	2.9	7.1	2
FLO	1.8	1.2	<0.17	2	<0.17	0.4	-0.10	0	<0.17	07	-0.10	0
PHE ANT	3.6 0.4	1.5 0.3	3.4 <0.05	4 2	0.6 <0.05	0.4	<0.19	2 0	0.7 <0.05	0.7	<0.19	1 0
PYR	2.2	1.2	1.9	3	1.0	0.3	1.3	5	1.3	0.7	1.8	2
FLA	1.4	1.4	<0.25	1	0.4	0.3	<0.25	2	0.7	0.7	<0.25	1
CHR	11.8	4.1	7.2	5	1.9	1.1	<0.25	3	2.9	0.5	3.0	3
BAA	<0.15			0	<0.15			0	<0.15			0
BBF	<0.2			0	<0.2			0	<0.2			0
BKF	<0.24			0	<0.24			0	<0.24			0
BAP	<0.21			0	0.1	0.1	<0.21	1	0.3	0.3	< 0.21	1
IPY BPY	<0.19			0 0	<0.19 <0.13			0 0	0.5 0.6	0.5 0.6	<0.19 <0.13	1 1
DBA	<0.13 <1.25			0	<1.25			0	<1.25	0.0	<b>~0.13</b>	0
DDA	\$1.20			0	\$1.20			0	\$1.20			0
-		Rive	er mile 8			River	mile 7eS			River	mile 7eC	
compound	mean	SE	median	n > MDL (n = 1)	mean	SE		n > MDL (n = 7)	mean	SE	median	n > MDL (n = 5)
	20.0		20.0	1	10.0	6.0	12.0	7	65.0	0.0	64.0	F
NAP ACY	29.0 <0.05		29.0	1 0	19.0 <0.05	6.2	12.9	7 0	65.9 <0.05	8.2	64.9	5 0
ACE	<0.05			0	<0.03 17.7	11.2	5.9	6	<0.03 57.8	13.7	58.9	5
FLO	<0.17			0 0	6.8	2.8	5.3	6	47.3	10.9		5
PHE	5.0		5.0	1	21.8	10.2		7	60.7	10.3		5
ANT	3.4		3.4	1	8.3	3.6	4.4	7	18.9	3.5	21.1	5
PYR	<0.25			0	21.0	8.2	9.6	7	24.3	2.7	27.4	5
FLA	<0.25			0	25.1	10.5	10.9	7	32.2	4.2	35.8	5
CHR	< 0.25			0	9.6	3.9	7.4	6	18.9	4.4	19.7	5
BAA BBF	<0.15 <0.2			0 0	8.9 7.3	2.5 2.0	7.1 5.4	7 7	12.3 5.9	1.4 1.2	13.8 4.6	5 5
BKF	<0.2 <0.24			0	2.4	2.0	1.8	6	2.8	0.4	4.0 2.5	5
BAP	<0.24			0	2.4	0.9	1.5	5	2.0	0.5	1.7	5
IPY	<0.19			0	2.6	1.0	1.7	5	1.1	0.7	<0.19	2
BPY	<0.13			0	1.8	0.9	<0.13	3	1.6	1.2	<0.13	2
DBA	<1.25			0	0.3	0.3	<1.25	1	0.7	0.7	<1.25	1
		River	mile 7eN	I		Rive	r mile 7w			Rive	r mile 3e	
-				n > MDL				n > MDL	-			n > MDL
compound	mean	SE	median	(n = 25)	mean	SE	median	( <i>n</i> = 3)	mean	SE	median	(n = 3)
NAP	91.7		49.5	25		16.3	12.6		6.7	1.2		3
ACY	4.9	2.9	<0.05	4	<0.05			0	5.4	5.4	<0.05	1
ACE	119	34.4	50.2	22	18.1	9.8	12.2	3	20.8	13.8	8.4	3
FLO	98.2	30.0	39.4	24	9.5	4.1	6.8	3	27.6	20.5	10.3	3
PHE ANT	179 42.0	96.8 14.2	57.9 17.6	24 25	16.1 5.0	7.8 2.7	10.3 2.5	3 3	47.0 30.8	21.5 27.8	52.1 3.7	3 3
PYR	42.0 77.2	42.7	22.8	25	13.4	2.7 3.1	12.5	3	27.1	7.3	24.8	3
FLA	115	66.3	33.1	25	12.6	2.9	12.5	3	33.2	11.9	32.2	3
CHR	34.9	21.1	10.0	23	22.7	11.8	15.7	3	21.4	8.0	16.5	3
BAA	35.9	22.0	7.8	23	8.1	2.1	7.6	3	12.8	7.1	6.6	3
BBF	29.1	20.0	4.6	17	6.2	2.3	7.3	3	7.8	3.7	5.7	3
BKF	8.6	5.7	1.2	13	1.8	1.0	2.1	2	2.3	1.5	1.8	2
BAP	7.9	5.1	1.4	14	3.9	1.2	4.7	3	3.1	1.4	2.5	3
IPY	6.4	3.8	<0.19	12	3.5	1.1	3.7	3	3.4	1.2	2.8	3
BPY	3.9	2.8	< 0.13	8	3.4	1.7	4.6	2	2.3	1.1	3.3	2
DBA	1.2	1.2	<1.25	1	<1.25			0	<1.25			0

SI Table 2. Spatial distribution of PAHs (ng/g w.w.) in resident crayfish collected from within and outside of the Portland Harbor Superfund Mega-site, OR.

$\begin{array}{c c c c c c c c c c c c c c c c c c c $	MDL = 1) 0 1 0 1 1 1 1 1 0 0 0 1 1 0 0 0 0			
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	0 1 0 1 1 1 1 0 0 0 1 0			
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	1 0 1 1 1 1 0 0 0 0 1			
ACE <0.14 0 <0.14 0 <0.14   FLO NQ <sup>b</sup> NQ NQ NQ   PHE 20.9 4.3 20.9 2 18.2 6.4 18.2 2 28.8 28.8   ANT 3.4 3.4 3.4 1 3.9 3.9 3.9 1 8.3 8.3   PYR 16.9 0.9 16.9 2 13.4 0.8 13.4 2 20.4 20.4   FLA 17.4 1.3 17.4 2 14.8 1.8 14.8 2 22.6 22.6   CHR <0.94 0 <0.94 0 <0.94 0 <0.94   BAA <0.36 0 <0.36 0 <0.36 <0.36	0 1 1 1 0 0 0 1			
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	1 1 1 0 0 0 1			
PHE   20.9   4.3   20.9   2   18.2   6.4   18.2   2   28.8   28.8     ANT   3.4   3.4   1   3.9   3.9   3.9   1   8.3   8.3     PYR   16.9   0.9   16.9   2   13.4   0.8   13.4   2   20.4   20.4     FLA   17.4   1.3   17.4   2   14.8   1.8   14.8   2   22.6   22.6     CHR   <0.94	1 1 0 0 0 1			
ANT 3.4 3.4 3.4 1 3.9 3.9 3.9 1 8.3 8.3   PYR 16.9 0.9 16.9 2 13.4 0.8 13.4 2 20.4 20.4   FLA 17.4 1.3 17.4 2 14.8 1.8 14.8 2 22.6 22.6   CHR <0.94	1 1 0 0 0 1			
PYR   16.9   0.9   16.9   2   13.4   0.8   13.4   2   20.4   20.4     FLA   17.4   1.3   17.4   2   14.8   1.8   14.8   2   22.6   22.6     CHR   <0.94	1 1 0 0 1 0			
FLA   17.4   1.3   17.4   2   14.8   1.8   14.8   2   22.6   22.6     CHR   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0   <0.94   0 <td>1 0 0 1</td>	1 0 0 1			
CHR   <0.94   0   <0.94   0   <0.94     BAA   <0.36	0 0 0 1			
BAA <0.36 0 <0.36 0 <0.36	0 0 1 0			
BBE <0.83 0 <0.83 0 <0.83	1 0			
	0			
BKF 1.3 1.3 1.3 1 1.0 1.0 1.0 1 2.8 2.8				
BAP NQ NQ NQ				
	0			
	0			
DBA <7.0 0 <7.0 0 <7.0	J			
River mile 8River mile 7eSRiver mile 7eCn > MDLn > MDLn >	River mile 7eC n > MDL			
n > MDL $n > MDL$ $n >compound mean SE median (n = 2) mean SE median (n = 1) mean SE median (n = 1)$				
NAP 2.2 2.2 2.2 1 <1.4 0 27.2 11.7 <1.4	4			
	5			
	9			
FLO NQ NQ NQ				
	9			
	6			
	8			
	8 0			
	9			
	1			
	7			
BAP NQ NQ NQ				
	0			
	0			
DBA <7.0 0 <7.0 0 <7.0	0			
River mile 7eN River mile 7w River mile 3e				
n > MDL $n > MDL$ $n >compound mean SE median (n = 4) mean SE median (n = 2) mean SE median (n$	MDL = 3)			
	•			
	2			
	1 2			
FLO NQ NQ NQ	-			
	3			
	3			
PYR 73.6 2.3 74.2 4 151 85 151 2 204 65 268	3			
	3			
	2			
	3			
	1 3			
BKF 4.7 1.5 6.0 4 5.8 1.5 5.8 2 7.2 1.4 8.6 BAP NQ NQ NQ NQ	5			
	0			
	0			
<u>DBA &lt;7.0 0 &lt;7.0 0 &lt;7.0</u>	0			

SI Table 3. Spatial distribution of freely dissolved PAHs (ng/L) collected by SPMDs from within and outside of the Portland Harbor Superfund mega-site<sup>a</sup>

<sup>*a*</sup>Data were adapted from Sower and Anderson (2008), <sup>*b*</sup>'NQ' = not quantified due to chromatographic interferences.

	Superfund	vs up-river <sup>a</sup>	Individual within Superfund sites vs control sites <sup>b</sup>				
РАН	Crayfish	$H_2O_{PSD}$	Crayfish	River miles with P<0.05	$H_2O_{PSD}$	River miles with P<0.05	
NAP	<0.001* <sup>c</sup>	0.6	<0.001*	7eN, 7eC	0.8		
ACY	0.2	0.4	0.3		0.8		
ACE	<0.001*	0.01*	<0.001*	7eN, 7eC	0.01*	7eC	
FLO	<0.001*	NQ <sup>d</sup>	<0.001*	7eN, 7eC	NQ		
PHE	<0.001*	0.007*	<0.001*	7eN, 7eC	0.02*	7eC	
ANT	<0.001*	0.9	<0.001*	7eN, 7eC	0.4		
PYR	<0.001*	0.005*	<0.001*	3.5, 7eN, 7eC, 7eS	0.06		
BPY	0.01*	BDL <sup>e</sup>	0.1		BDL		
ΣNC-PAH <sup>f</sup>	<0.001*	0.011*	<0.001*	7eN, 7eC	0.037*	7eC	
FLA	<0.001*	0.005*	<0.001*	3.5, 7eN, 7eC, 7eS	0.08		
CHR	0.002*	0.4	0.01	None <sup>g</sup>	0.02*	None	
BAA	<0.001*	0.003*	<0.001*	7eN, 7eC	0.02*	None	
BBF	<0.001*	0.5	0.01*	None	0.7		
BKF	<0.001*	0.07	0.003*	7eN, 7eC	0.2		
BAP	<0.001	NQ	0.003*	7w	NQ		
IPY	<0.001*	BDL	0.008	None	BDL		
DBA	0.3	BDL	0.6		BDL		
ΣC-PAH <sup>h</sup>	<0.001*	0.002*	<0.001*	3.5, 7eN, 7eC	0.02*	3.5	
ΣΡΑΗ <sup>ί</sup>	<0.001*	0.007*	<0.001*	7eN, 7eC	0.03*	None	

SI Table 4. Summary of significant site-specific differences in median crayfish (ng/g w.w.) and freely dissolved water concentrations (ng/L) for the fall 2003 Portland Harbor, OR study

<sup>a</sup>Mann-Whitney rank sum tests between sampling sites, <sup>b</sup>Kruskal-Wallis one way ANOVA on ranks using Dunn's method for multiple comparisons between individual sites within the Superfund mega-site and up-river 'control' sites (river miles 13, 17, and 18.5), <sup>c</sup>Asterisk (\*) indicates significance at  $\alpha = 0.05$ , <sup>d</sup>Not quantified due to chromatographic interference, <sup>e</sup>Measurement was below method reporting limits, <sup>f</sup>ΣNC-PAH = the summed concentration of non-carcinogenic PAHs excluding FLO, <sup>g</sup>Indicates that no statistically significant differences were found during multiple-comparisons to controls, <sup>h</sup>ΣC-PAH = the summed concentration of carcinogenic PAHs excluding FAHs excluding FLO and BAP.

## REFERENCES

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(2) Sower, G. J.; Anderson, K. A. Spatial and temporal variation of freely dissolved polycyclic aromatic hydrocarbons in an urban river undergoing Superfund remediation. *Environ. Sci. Technol.* **2008**, *42* (24), 9065-9071.