

**Supplementary Figure 1** - *In situ* hybridization images of 120 hpf wild-type larval zebrafish paraffin sections showing expression of otoferlin a in the mid-brain (MB) and retinal ganglion cell layer (RGL) . upper panel - otoferlin a probe and corresponding bright-field (bf) images. lower panel - no-probe control and corresponding bright-field (bf) images.

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120 hpf							120 hp	f					

## morpholino knockdown

cross-expression

**Supplementary Figure 2** - Morpholino knockdown of otoferlin in zebrafish larvae. **A)** RT-PCR gel image of otoferlin KD zebrafish larvae at 120 hpf, (lane 1) = molecular weight marker ; (lane 2) = negative control tested for otoferlin b; (lane 3) = negative control injected tested for otoferlin a; (lane 4) = otoferlin b KD tested for otoferlin b; (lane 5) = otoferlin a KD tested for otoferlin a; (lane 6) = otoferlin b+a double KD tested for otoferlin b; (lane 7) = otoferlin b+a double KD tested for otoferlin a. **B)** Cross expression studies with 120 hpf zebrafish larvae. RT-PCR gel image shows expression of: (lane 1)= molecular weight marker; (lane 2) = otoferlin a in control; (lane 3)= otoferlin a in otoferlin b KD; (lane 4) = otoferlin b in otoferlin b KD; (lane 5) = otoferlin b in control; (lane 6)= otoferlin a in otoferlin a KD; (lane 7) = otoferlin a KD. (Inj. C – injected control, KD – Knockdown, otof a – otoferlin a, otof b – otoferlin b, otof b+a – otoferlin b+a)



Supplementary Figure 3 - Compressed z-stack through the ear region showing otoferlin expression in 120 hpf injected control (A) larval zebrafish, and, otoferlin expression in double knockdown (B) 120 hpf larval fish (white hemicircles denotes the eye). Red clusters in Figure are otoferlin positively stained hair cell neuromasts.



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Supplementary Figure 4 - A) Observable phenotypes associated with the otoferlin b+a KD in120 hpf larval zebrafish with second set of morpholinos (e11i11 - otoferlin a and e2i2- otoferlin b). Arrow indicates swim bladder.
B) qPCR showing relative expression of shha in 72 hpf otoferlin control and double morphants normalized to beta-actin expression. Data shows no statistically significant differences in the expression of shha in control and double morphants. The statistical significance is calculated through Mann-Whitney test.

## 96 hpf otoferlin b+a KD co-injected with mouse-FL otoferlin construct





G)	Groups	Number of fish	Mean distance moved	sem					
	Control	23	63.94	5.69					
	Otoferlin a KD	18	66.03	5.73					
	Otoferlin b KD	17	66.79	7.53					
	Otoferlin b+a KD	16	22.92	2.65					

Supplementary Figure 5 - (A-C) Whole-mount *in situ* hybridization on otoferlin double morphants co-injected with the full-length mouse otoferlin construct under the hair cell specific promoter at 96hpf. A, B, and C shows mRNA expression in the anterior lateral line (aLL), posterior lateral line (pLL), and otic vesicle. (E-F) Confocal images of whole mount immunohistochemistry of 72 hpf larval zebrafish showing mauthner cells. E) Injected control, F) Otoferlin b+a double morphants. Arrowhead indicates the mauthner cells.G) Summary statistics of the startle between otoferlin morphants and control groups, sem - Std err of mean.



**Supplementary Figure 6** - Dark-light behavioral assay : Distances travelled (in mm) during the dark-phase by larvae are shown. Mean distances moved in mm: Injected control (n=72) = 62.73, otoferlin b KD (n=72) = 45.67, otoferlin a KD (n=72) = 46.68, otoferlin b+a KD (n=72) = 35.61, Rescue FL (n=72) = 59.89, Rescue  $\Delta$ ABC (n=48) = 59.81. Dunn's multiple comparison test with standard 5% significance level shows significant difference between control and KD groups and no significant difference between control and rescue groups. Error bars indicate 95% confidence interval of the sample mean.



**Supplementary Figure 7** - Rescue of zebrafish otoferlin KD with mouse otoferlin constructs. Rescue of swim bladder defect in 120 hpf otoferlin b+a KDs with mouse otoferlin, **A)** del-ABCDE, **B)** del-ABCD constructs. Arrow indicates inflated swim bladder.