AN ABSTRACT OF THE THESIS OF

Michael Gene O'Rand		for the	Master of Science		
**************************************	(Name)		(Degree)		
in	Zoology	presented on	August 8, 1969 (Date)		
	(Major)		(Date)		
Title:	tle: AN ULTRASTRUCTURAL STUDY OF TENTACLE				
	DEVELOPMENT IN	CORDYLOPHO	DRA LACUSTRIS		
Abstract approved: Redacted for Privacy					
Patriçia J. Harris					

Morphological changes which occur during tentacle formation have been described in the colonial athecate hydroid Cordylophora lacustris. These changes involve an outpocketing of mesoglea into a finger-like extension of the hydranth. Gastrodermal cells line the inside of this extension of mesoglea while ectodermal cells cover the outside. Electron micrographs reveal the presence of several distinct cell types. Their function and possible role in tentacle differentiation is discussed.

An Ultrastructural Study of Tentacle Development in Cordylophora lacustris

by

Michael Gene O'Rand

A THESIS

submitted to

Oregon State University

in partial fulfillment of the requirements for the degree of

Master of Science

June 1970

APPROVED:

Redacted for Privacy

Associate Professor of Zoology

In Charge of Major

Redacted for Privacy

Chairman of Department of Zoology

Redacted for Privacy

Dean of Graduate School

Date thesis is presented	August 8, 1969	
Typed by Nancy Kerley for	Michael Gene O'Rand	

ACKNOWLEDGMENT

I wish to express my appreciation to Dr. Patricia J. Harris for her constant encouragement and insightful guidance during the course of this study. I would also like to thank Dr. R. L. Miller for his helpful discussions concerning this manuscript. This study was supported in part by a grant from the United States Public Health Service (GM - 12963) to Dr. P. J. Harris.

TABLE OF CONTENTS

		Page
I.	INTRODUCTION	1
II.	MATERIALS AND METHODS	3
III.	OBSERVATIONS	6
	The Tentacle Bud The Immature Tentacle The Adult Tentacle	6 12 15
IV.	DISCUSSION	17
	BIBLIOGRAPHY	20
	APPENDIX	23

AN ULTRASTRUCTURAL STUDY OF TENTACLE DEVELOPMENT IN CORDYLOPHORA LACUSTRIS

I. INTRODUCTION

Developmental processes concerning the hydrozoan coelenterates, have been the object of numerous investigations, yet only within the last several years have ultrastructural studies been conducted to lay the morphological groundwork necessary for an understanding of the processes and changes that occur during development. Questions such as those being asked by Clarkson (1969), as to whether or not regeneration in hydra is based upon the existence of long term messenger ribonucleic acid, must be based on an understanding of the morphological events occurring during this time. Similarly, developmental processes in colonial hydroids must also be based on a knowledge of the animal's normal morphology. Thus, if the events responsible for the development of growth patterns and numerous specialized structures (i.e. tentacles, gonophores, dactylozooids, etc.) found in colonial forms are to be explained, then a firm morphological baseline must exist from which to work.

In studying a particular differentiation process the ultimate aim is to understand the events occurring from the time the genetic message is sent out until the adult structure is complete. Before events on the molecular level can be characterized however, those tissues

involved in the process must be characterized and their role understood. The formation of a tentacle by the hydranth of a colonial hydroid is perhaps one such differentiation process which might lend itself to analysis first on the morphological level and secondarily on a biochemical level. The present study is designed to describe the changes in morphology which occur during normal tentacle development in Cordylophora lacustris Allman.

Cordylophora lacustris, first described by Allman (1853), is a brackish-water colonial athecate hydroid which has been the subject of many developmental studies, including those of Beadle and Booth (1938), Diehl and Bouillon (1966), Fulton (1963), Kirschner (1934), Kuhn (1914), Moore (1952, 1952), Morgenstern (1901), Pauly (1902), Penzlin (1957), Schulze (1871), Tessenow (1959), and Zwilling (1963). Ultrastructural studies have been undertaken by Jha and Mackie (1967) and Overton (1963). These were concerned with nerve elements and intercellular connections respectively.

II. MATERIALS AND METHODS

The colonies used in the present study, collected at Woods Hole, Massachusetts, were provided by Dr. R. L. Miller. These colonies are grown in a controlled environment following the optimum growth conditions outlined by Fulton (1960). The colonies have been growing in the laboratory for approximately three years and have been subcultured repeatedly. They are grown on 1" x 3" glass microscope slides in 250 cc pyrex beakers in a constant temperature bath at 22°C. The pH of Fulton's culture solution was adjusted to 7.3 but light and oxygen tension were not controlled. Light was the normal daily cycle for Oregon. Each colony except those being examined after 48 hours starvation was fed Artemia nauplii every morning. batches of Artemia were hatched in sea water every other day and washed in distilled water before being given to the colonies. The entire colony was fed in such a way to insure that each individual got at least one Artemia; most received five or six. This was determined by visual observation. After feeding, the culture solution was changed and then changed again six hours later. The procedure was repeated at each feeding.

In his studies on mitosis in hydra Campbell (1967b) showed that there was a definite increase in the number of dividing cells of all cell types after feeding. Since feeding is also likely to affect mitosis and subsequent development processes in Cordylophora, the examination of all animals in this study was relative to the feeding time (time zero). Individual animals in different stages of tentacle development were examined at four or five hours and 48 hours after feeding. Tentacle development has been arbitrarily divided into three stages: stage I - the tentacle bud, stage II - the immature tentacle, stage III - the adult tentacle. For each time period four to eight animals at each stage were cut from seven different colonies growing under controlled conditions.

The animals were allowed to relax in a small dish and then fixed in cold 3% glutaraldehyde in 0.2 M phosphate buffer at pH 7.3 for two hours. They were then washed in buffer several times and stored in cold buffer overnight. Following post fixation in 1% osmium tetroxide for one hour, they were then dehydrated in a graded series of ethanol: 50% for one hour, 70% for one-half hour, two changes of 95% for one hour, and three changes of 100% for a total of one hour. Specimens were given 30 minutes in propylene oxide with one change and then a 1:1 mixture of propylene oxide and Araldite. The Araldite embedding procedure was according to Luft (1961). The animals were left overnight in the 1:1 mixture and then flat embedded in aluminum pans at 60°C for 24 hours. In each step during the preparation procedure an adult hydranth fixed with tentacles fully extended was carried along to insure that the tissues were not damaged. Rough

handling or careless pipeting resulted in tentacles becoming detached from the hydranth.

The specimens were cut on a Porter-Blum microtome and thin sections mounted on Formvar covered copper grids (75 mesh). Grids were stained with uranyl acetate and Millonig's lead hydroxide (1961) and examined in an RCA EMU 3H microscope. Thick sections were cut approximately one-half microns and stained with a modified method of Schantz and Schecter (1965). Solutions of ferric ammonium sulfate, hematoxylin, and safranin 0 were placed in separate Coplin jars and kept in the oven at 65° C. Slides with thick sections were immersed for 30 minutes in each solution. They were rinsed for ten seconds in distilled water and dried in the oven between each solution. The results clearly differentiate nucleus, nucleolus, cell membranes, mesoglea, lipid droplets from other inclusions and vesicles, and nematocysts. Electron micrographs were printed on Kodak F-5 paper and light micrographs, taken through a Zeiss phase contrast scope with Panatomic-X film, were printed on Kodak Polycontrast paper.

III. OBSERVATIONS

Examination of animals at different times after feeding revealed no significant differences in tentacle development stages. The only obvious difference was in the amount of injested material present in the digestive cells. Therefore the following observations apply to all time periods.

The Tentacle Bud

The tentacle bud stage can be most readily described as an immature hydranth with a low ridge of cells below the hypostome. The distal tentacles, which are the first to differentiate, appear as small projections on this ridge (Fig. 1, 23). In this condition the coelenteron of the stem or stolon is confluent with that of the immature hydranth and extends for approximately two-thirds of its length (Fig. 2, 3). In medial longitudinal sections the ridge can be seen as a bulge slightly distal to the coelenteron cavity (Fig. 3, 4). The tentacle will differentiate from this bulge. Medial sections of this hydranth show the cell types present to be morphologically similar to those seen in the adult hydranth.

Hydranth Ectoderm

1. Epitheliomuscular cells. Epitheliomuscular cells (Fig. 5)

often a protein body as described by Mackie et al. (1964). Usually numerous vacuoles are present, with the nucleus occupying the central portion of the cell. The nucleus is usually surrounded by a series of small peripheral vacuoles. The outer nuclear membrane appears studded with ribosomes while mitochondria, golgi, and dilated rough endoplasmic reticulum are found throughout the cytoplasm.

Numerous inclusions are often present in the proximal portions of the cell, as well as lipid droplets and heterogeneous granules similar to those seen in the gastroderm. The basal portions of the cells are characterized by myofibrils adjacent to the mesoglea. At some points they appear to insert into the mesoglea. The myofibrils are usually seen running parallel to the long axis of the animal.

Intercellular attachments between epitheliomuscular cells (Fig. 5, 6) similar to those described by Overton (1963) in the epidermis of the stolon, are present. They appear as thickenings of the adjacent cell membranes. In accordance with Overton's findings these desmosomes have not been seen in areas where the cell surface would be expected to be metabolically active (i.e. the tentacle bud).

The external surface of the epitheliomuscular cell is characterized by small finger-like projections of cytoplasm which are directly beneath the outer acellular layer. The acellular layer surrounds the

entire hydranth and is continuous with the perisarc of the stem.

2. Interstitial cells. Interstitial cells (I-cells) are cells of a non-epitheliomuscular nature found in the ectoderm. Cells of similar description are occasionally found in the gastroderm (Fig. 6, 7, 8, 9). Interstitial cells can be characterized by their abundant mitochondria, sparse dilated endoplasmic reticulum, and free ribosomes. The free ribosomes usually give the cytoplasm of the I-cell a darker appearance than that of the adjacent epitheliomuscular cells. Figures 6 and 7 show the manner in which I-cells are surrounded by the processes of epitheliomuscular cells. Interstitial cell nuclei are small, centrally located, and according to Mackie et al. (1964) lack a protein body. Generally the cytoplasm of I-cells is not vacuolated and few granules or inclusions are ever seen. Contrary to the observations of Jha and Mackie (1967), Cordylophora hydranths do have interstitial cells.

Some I-cells may be nerve cells as described by Jha and Mackie (1967), but this cannot be determined without careful serial sectioning to locate a sensory projection or neurite and comparison with silver stained material. Although not numerous, these small compact cells (approximately five microns in diameter) occur adjacent to the mesoglea and may occur in clusters or nests (Fig. 10). They are most often seen near tentacle-hydranth junctions in the mature hydranth.

3. Nematocysts. Developing and mature nematocysts have

been observed scattered throughout the hydranth ectoderm and gastroderm (Fig. 4, 7, 8, 10, 12). Developing nematocysts (cnidoblasts) are most often seen in the interstitial cell clusters near tentacle-hydranth junctions (Fig. 10). The cnidoblast appears essentially the same as the interstitial cell, having mitochondria, rough endoplasmic reticulum and few granules or inclusions. The nucleus is usually distorted somewhat due to the developing nematocyst.

Hydranth Gastroderm

The hydranth gastroderm of the tentacle bud stage showed cell types similar to those found in the adult hydranth, as mentioned previously. However, the mouth does not completely differentiate until after tentacle development so that distally, gastrodermal cells appear in close apposition to one another. Some of the cells appear as digestive cells while others appear secretory (Fig. 11).

1. Digestive cells. A digestive cell (Fig. 12, 13, 14) is characterized by its large nucleus which is usually surrounded by small peripheral vacuoles. The outer nuclear membrane is studded with ribosomes. In medial sections numerous microvilli and flagella are also characteristic of digestive cells (fig. 13, 14). The cytoplasm is often seen as thin strands surrounding large vacuoles which may be filled with various injested material depending upon the nutritive state at fixation. Figure 9 shows the basal portions of digestive cells

where each large vacuole is separated from the adjacent vacuole by the cytoplasm of two different cells. The cytoplasm contains rough endoplasmic reticulum, mitochondria, and golgi. Granules of varying density may be seen in the basal regions of digestive cells in the proximal regions of the hydranth where it appears that large and small granules are connected or are coalescing (Fig. 9). In the apical regions of the cell (Fig. 13, 14) and the more distal regions of the hydranth (Fig. 11), digestive cell nuclei appear surrounded by granules of varying density. Intercellular connections are often seen between gastrodermal cells in the more stable regions of the hydranth (Fig. 12, 13).

2. Secretory cells. In the adult hydranth numerous types of secretory cells may be present as noted in other hydroids by Bouillion (1966), Rose and Burnett (1968), and Brock, Strehler, and Brandes (1968). However, in the hydranths investigated during tentacle development, the most obvious secretory cells were those characterized by a small nucleus, compact cytoplasm filled with dilated rough endoplasmic reticulum often seen in stacks (Fig. 15), and zymogen-like granules. This cell type was never observed to have microvilli or flagella. In the tentacle bud hydranth many secretory cells appear as in Figure 11. Here they are situated at the apex of the hydranth gastroderm and appear smaller than those in the adult hydranth (Fig. 15). There are also some small compact cells seen

in the gastroderm which are not easily identified. They may be termed interstitial as in Figure 6 or secretory as in Figures 7 and 8. They are surrounded by cytoplasmic processes of digestive cells much as interstitial cells in the ectoderm are surrounded by epitheliomuscular cell processes. Their exact role is not known. Figure 12 shows another small cell which is occasionally seen in the gastrodermis. Its role is also unknown.

Mesoglea

The mesoglea present in the tentacle bud hydranth courses its way between the gastroderm and ectoderm as in the adult. It can be characterized by its fibrillar nature, low electron opacity, and thin projections into both gastroderm and ectoderm (Fig. 5, 6, 7, 8, 9). Small particles can be seen scattered throughout the fibrils which appear to run parallel to the long axis of the hydranth.

Tentacle Bud

In the electron microscope the tentacle bud (Fig. 4, 16) is seen as an outpocketing of the mesoglea filled with gastroderm cells on the inner side and ectoderm cells on the outer side. The ectoderm of this bud region is morphologically similar to hydranth ectoderm. The epitheliomuscular cells in the bud region appear vacuolated but are often filled with numerous inclusions.

The gastroderm cells seem to be morphologically the most similar to hydranth digestive cells although few microvilli or flagella are ever seen. Large numbers of ribosomes are seen in the cytoplasm, but most characteristic is the presence of granules of variable density. The variation in density within these granules (Fig. 16) appears like that of the granules seen in hydranth digestive cells (Fig. 12, 13, 14) and may represent in part, the result of compression during sectioning. They have a halo-like periphery which appears in later stages of tentacle development to be bounded by a membrane. The interior of these granules is less dense than the surrounding dense band directly inside the halo-like periphery. Their heterogeneity can be compared to the more homogeneous looking zymogenlike granules of the secretory cell seen in Figure 15. Some of the cells described under hydranth secretory cells might also be in the bud region but they are not immediately obvious. Secretory cells have not been seen in the later tentacle gastroderm.

The Immature Tentacle

In the hydranth of the immature tentacle the ridge of cells below the hypostome has enlarged considerably and four small tentacles protrude just below the hypostome (Fig. 17, 23). The hydranth cell types are as described in the tentacle bud hydranth. The immature tentacles are characterized by a further outpocketing of mesoglea away from the hydranth. The rounded apex of the previous stage is now quite pointed (Fig. 18). In a median section the hydranth gastroderm is confinent with the tentacle gastroderm (Fig. 19a), but in sagittal sections above and below the median the mesoglea separates the hydranth and tentacle gastroderm (Fig. 19b).

Tentacular Ectoderm

1. Epitheliomuscular cells. Epitheliomuscular cells present in the tentacular ectoderm are morphologically similar to those of the hydranth (Fig. 18). The nucleus is generally centrally located among numerous vacuoles of various size. It may possess a protein body as in Figure 18. Dilated rough endoplasmic reticulum is present and mitochondria are scattered about the cytoplasm.

The basal myofibrils present in the hydranth cells do not appear prominent in the immature tentacle epitheliomuscular cells. The outer surface of the ectodermal cells is covered with an acellular material which is continuous with that of the hydranth.

2. Interstitial cells. Numerous non-epitheliomuscular cells are scattered along the mesoglea of the immature tentacle. Figure 18 shows several at the tip of the immature tentacle. Some may differentiate into nematocysts as reported by Kirchner (1934) or into some other cell type while some may be nerve cells. The sensory hair seen in Figure 18 appears to be of the type described by Jha and

Mackie (1967) as being characteristic of nerve cells. Furthermore, tubules can be seen running through the cell body. Basal bodies can be seen in adjacent cells suggesting other possible sensory hairs in the immature tentacle tip.

3. Nematocysts. Developing and mature nematocysts appear in the immature tentacle in much the same manner as interstitial cells. They do not seem to be clustered or oriented toward the surface at this stage.

Tentacular Gastroderm

The tentacular gastroderm cells of the immature tentacle appear similar to those in Figure 4 or those hydranth gastroderm cells in Figure 11. The nucleus is relatively large and generally close to the median plane of the tentacle. It is surrounded by small peripheral vacuoles and the outer nuclear membrane may be studded with ribosomes. The cytoplasm has the characteristic vesicles of variable density which have been previously described. Some of the granules may be seen surrounded by a membrane and free inside the vacuoles (Fig. 19a, 19b). The cytoplasm also has numerous free ribosomes as well as rough endoplasmic reticulum and mitochondria (Fig. 19a, 19b). At this stage there is considerably more gastroderm present due to the larger mesogleal outpocketing. It appears quite compacted within the outpocketing as the vacuoles are generally small.

Mesoglea

The mesoglea is continuous with hydranth mesoglea and similar in its fibrillar structures and low electron opacity. As was previously mentioned the mesoglea does not cut off the tentacle gastroderm from the hydranth completely since a median section shows the hydranth gastroderm to be confluent with that of the tentacle. Figure 18 shows the distal end of the mesoglea in the tentacle tip.

The Adult Tentacle

The adult tentacle is shown in Figures 20 and 23. The four most distal tentacles directly below the hypostome were those investigated in this study.

Tentacular Ectoderm

The ectoderm of the adult tentacle contains the cell types as described for the immature tentacle. Figure 21 shows a medial longitudinal section of a tentacle in an advanced state of maturation. The adult tentacle is identical to this but of considerably greater length. The distal end is blunt and knob-like while the ectoderm's exterior surface is scalloped. Each scallop is filled with epitheliomuscular cells and nematocysts. They can be seen as black spots along the tentacle in Figure 20. Figure 21 also shows nematocysts

and interstitial cells scattered along the length of the tentacle. Characteristic of the adult tentacle's epitheliomuscular cells are the well developed myofibrils adjacent to the mesoglea (Fig. 22).

Tentacular Gastroderm

The tentacular gastroderm is as described previously for the immature tentacle. The gastroderm seen in Figure 21 differs from that in the immature tentacle only in its ladder-like appearance. The gastroderm vacuoles become larger and the cells appear less closely packed as they stretch out along the tentacle. The vacuoles are surrounded by a thin band of gastrodermal cytoplasm (Fig. 22). Each rung in the ladder usually consists of one or two large nuclei and their associated cytoplasm.

Mesoglea

The mesoglea seen in the adult tentacle (Fig. 21) is as described previously. The myofibrils of the epitheliomuscular cells can be seen adjacent to the mesoglea (Fig. 22). The mesoglea of the tentacle is continuous with the hydranth mesoglea except in medial sections where it is proximally discontinuous. Medial sections of the distal part of the tentacle indicate that the mesoglea is continuous in this region.

IV. DISCUSSION

The stages in the formation of a tentacle have been described in terms of morphological changes that occur during the differentiation process. The process can be thought of as an outpocketing of hydranth mesoglea into a finger-like extension from the hydranth.

The hydranth gastrodermal cells seen in the earliest mesogleal outpocketing certainly have the general morphological characteristics of the hydranth digestive cells. The fact that these cells are in the outpocketing surrounded by other cells and do not have a free surface facing the coelenteron could account for the reduced appearance of microvilli and flagella. This would also account for their lack of food inclusions. Rather their inclusions tend to be lipid or membrane-bounded granules of a heterogeneous nature. It is possible that these granules are in fact manufactured by these cells at an earlier stage prior to their arrival at the tentacle site. The dilated rough endoplasmic reticulum of these cells would certainly indicate that active synthesis is occurring.

As the outpocketing lengthens it would seem that more gastro-dermal cells would be added to the tentacle base. Although there is no evidence for this in Cordylophora, evidence from other hydroids makes this plausible. The migration of epitheliomuscular cells along the mesoglea has been shown in Hydra by Shostak and Globus (1966).

The migration of gastrodermal cells from the hydranth into the tentacle during development and later in its adult state has been shown in Hydra by Shostak (1968). Working with Tubularia croeca, Campbell (1967a) reported that "at the base of the distal tentacle whorl the gastrodermal core of the tentacles runs along the hydranth mesolamella." This investigation has established that the tentacular gastroderm is continuous with the hydranth gastroderm during all stages of tentacle development in Cordylophora. Furthermore, mitotic studies by Campbell in Hydra (1967b) and Tubularia and Hydractinia (1967a) indicate that there is little or no mitotic activity in the tentacle itself but rather at its base.

Perhaps the most interesting question which can now be asked is: What is responsible for the initial outpocketing of mesoglea? If one assumes the constancy of the gastrodermal cell population as Shostak (1968) reports for Hydra, then the initial outpocketing may be due to a specific biochemical event occurring in certain cell types. Alternatively, the physical pressure of cells packing into the hydranth's apex by continual distal migration could be responsible for such an outpocketing. Localized mitotic activity has been tentatively ruled out because no gastrodermal mitotic activity in the bud was ever observed. In any event, it seems most likely that the gastrodermal and ectodermal cells in the bud region influence the mesoglea in some manner. Indeed, the synthesis of mesogleal protein has been

suggested by Shostak, Patel, and Burnett (1965) as possibly responsible for the determination of form in Hydra.

The heterogeneous granules appearing in the tentacle gastroderm cells in the bud, which are similar to those seen in hydranth digestive cells, seem to warrant further investigation. Since they are seen free in cellular vacuoles in later stages of tentacle development, their function may be of importance in the process of differentiation.

The cluster of small compact cells in the tentacle tip in

Figure 18 would also seem to warrant further investigation. This

cluster may influence the development of the differentiating cells

of the immature tentacle in some manner.

BIBLIOGRAPHY

- Allman, G. J. 1853. On the anatomy and physiology of <u>Cordylophora</u>, a contribution to our knowledge of Tubularian zoophytes.

 Philosophical Transactions of the Royal Society (London) 143: 367-384.
- Beadle, L. C. and F. A. Booth. 1938. The reorganization of tissue masses of <u>C. lacustris</u> and the effect of oral cone grafts, with supplementary observations on <u>Obelia gelatinosa</u>. Journal of Experimental Biology 15:303-326.
- Bouillon, J. 1966. Les cellules glandulaires des hydroides et hydromeduses. Leur structure et la nature de leurs secretions. Cahiers de Biologie Marine 7:157-205.
- Brock, M. A., B. L. Strehler and D. Brandes. 1968. Ultrastructural studies on the life cycle of a short-lived metazoan, Campanularia flexuosa. I. Structure of the young adult. Journal of Ultrastructure Research 21:281-312.
- Campbell, R. 1967a. Cell proliferation and morphological patterns in the hydroids <u>Tubularia</u> and <u>Hydractinia</u>. Journal of Embryology and Experimental Morphology 17:607-616.
- 1967b. Tissue dynamics of steady state growth in H. littoralis. I. Patterns of cell division. Developmental Biology 15:487-502.
- Clarkson, S. G. 1969. Nucleic acid and protein synthesis and pattern regulation in hydra. I. Regional patterns of synthesis and changes in synthesis during hypostome formation. Journal of Embryology and Experimental Morphology 21:33-54.
- Diehl, F. A. and J. Bouillon. 1966. Observations sur les potentialities morphogenetiques des tissues irradies de Cordylophora caspia (Pallas). Bulletin de l'Academie Royale des Sciences, des Lettres et des Beaux-arts de Belgique (Brussels), Classe des Sciences 52:138-146.
- Fulton, C. 1960. Culture of a colonial hydroid under controlled conditions. Science 132:473-474.

- Fulton, C. 1963. The development of a hydroid colony. Developmental Biology 6:333-369.
- Jha, R. K. and G. O. Mackie. 1967. The recognition, distribution and ultrastructure of hydrozoan nerve elements. Journal of Morphology 123:43-62.
- Kirchner, H. A. 1934. Die Bedeutung der I-Zellen für den Aufbau von C. caspia. Zeitschrift für Zellforschung und mikroskopische Anatomie 22:1-19.
- Kuhn, A. 1914. Entwichlungsgeschichte und Verwandtschaftsbeziehungen der Hydrozoen. Ergebnisse und Fortschritte der Zoologie 4:1-284.
- Luft, J. H. 1961. Improvements in epoxy resin embedding methods. Journal of Biophysical and Biochemical Cytology 9:409-414.
- Mackie, G. O. et al. 1964. Massive accumulations of protein in nuclei of Cordylophora. Canadian Journal of Zoology 42: 1011-1016.
- Millonig, G. 1961. A modified procedure for lead staining of thin sections. Journal of Biophysical and Biochemical Cytology 11: 736-739.
- Moore, J. 1952. Induction of regeneration in the hydroid

 <u>Cordylophora lacustris</u>. Journal of Experimental Biology
 29:72-93.
- 1952. Interstitial cells in the regeneration of Cordylophora lacustris. Quarterly Journal of Microscopical Science 93:269-288.
- Morgenstern, P. 1901. Untersuchungen über der Entwicklung von C. lacustris Allman. Zeitschrift für wissenschaftliche Zoologie 70:567-591.
- Overton, J. 1963. Intercellular connections in the outgrowing stolon of Cordylophora. Journal of Cell Biology 17:661-671.
- Pauly, R. 1902. Untersuchungen über den Bau und die Lebensweise der Cordylophora lacustris Allman. Jenaische Zeitschrift für Naturwissenschaften 63:737-780.

- Penzlin, H. 1957. Experimentelle untersuchungen über die regeneration bei <u>Cordylophora caspia</u> Pallas. Wilhelm Roux' Archiv für Entwicklungsmechanik der Organismen 149:624-643.
- Rose, P. G. and A. L. Burnett. 1968. An electron microscopic and histochemical study of the secretory cells in <u>Hydra viridis</u>. Wilhelm Roux' Archiv fur Entwicklungsmechanik der Organismen 161:281-297.
- Schantz, A. and A. Schecter. 1965. Iron-hematoxylin and safarin 0 as a polychrome stain for epon sections. Stain Technology 40:297-282.
- Schulze, F. E. 1871. Über den Bau und die Entwicklung von

 <u>Cordylophora lacustris</u> (Allman). Leipzig, Wilhelm Englemar.

 52 p.
- Shostak, S. N. 1968. Growth in <u>Hydra viridis</u>. Journal of Experimental Zoology 169:431-446.
- Shostak, S. N. and M. Globus. 1966. Migration of epithelio-muscular cells in Hydra. Nature 210:218-219.
- Shostak, S. N., N. G. Patel and A. L. Burnett. 1965. The role of the mesoglea in mass cell movements in <u>Hydra</u>. Developmental Biology 12:434-450.
- Tessenow, W. 1959. Untersuchungen an den interstitiellen Zellen von Pelmatohydra oligactis Pallas und Cordylophora lacustris Pallas unter besonderer Berucksichtigung spezifischer Farbenmethoden. Protoplasma 51:563-594.
- Zwilling, E. 1963. Formation of endoderm from ectoderm in Cordylophora. Biological Bulletin 124:368-378.



- Figure 1. Stage one hydranth showing the low ridge of cells (R) below the hypostome and the tentacle buds (TB). 875 X
- Figure 2. Light micrograph of stem-hydranth junction showing the coelenteron (C) cavity lined with microvilli of the digestive cells. 7900 X

LIST OF ABBREVIATIONS

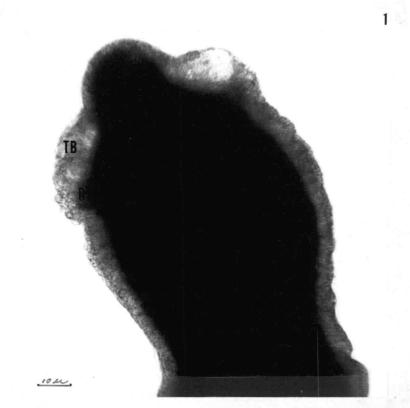
C Coelenteron D Digestive cell E Ectoderm Ep Epitheliomuscular cell Fl Flagellum G Gastroderm GI Gastrodermal inclusion G Golgi HG Heterogeneous granules Ia Intercellular attachments I Interstitial cell R Low ridge of cells below hypostome M Mesoglea Mv Microvilli Mi Mitochondria My Myofibrils Ne Nematocysts No Nucleolus N Nucleus A Outer acellular layer PB Protein body RER Rough endoplasmic reticulum S Secretory cell	ВВ	Basal body
E Ectoderm Ep Epitheliomuscular cell Fl Flagellum G Gastroderm GI Gastrodermal inclusion G Golgi HG Heterogeneous granules Ia Intercellular attachments I Interstitial cell R Low ridge of cells below hypostome M Mesoglea Mv Microvilli Mi Mitochondria My Myofibrils Ne Nematocysts No Nucleolus N Nucleus A Outer acellular layer PB Protein body RER Rough endoplasmic reticulum S Secretory cell	С	Coelenteron
Ep Epitheliomuscular cell Fl Flagellum G Gastroderm GI Gastrodermal inclusion G Golgi HG Heterogeneous granules Ia Intercellular attachments I Interstitial cell R Low ridge of cells below hypostome M Mesoglea Mv Microvilli Mi Mitochondria My Myofibrils Ne Nematocysts No Nucleolus N Nucleus A Outer acellular layer PB Protein body RER Rough endoplasmic reticulum S Secretory cell	D	Digestive cell
Fl Flagellum G Gastroderm GI Gastrodermal inclusion G Golgi HG Heterogeneous granules Ia Intercellular attachments I Interstitial cell R Low ridge of cells below hypostome M Mesoglea Mv Microvilli Mi Mitochondria My Myofibrils Ne Nematocysts No Nucleolus N Nucleus A Outer acellular layer PB Protein body RER Rough endoplasmic reticulum S Secretory cell	E	Ectoderm
G Gastroderm GI Gastrodermal inclusion G Golgi HG Heterogeneous granules Ia Intercellular attachments I Interstitial cell R Low ridge of cells below hypostome M Mesoglea Mv Microvilli Mi Mitochondria My Myofibrils Ne Nematocysts No Nucleolus N Nucleus A Outer acellular layer PB Protein body RER Rough endoplasmic reticulum S Secretory cell	Ep	Epitheliomuscular cell
GI Gastrodermal inclusion G Golgi HG Heterogeneous granules Ia Intercellular attachments I Interstitial cell R Low ridge of cells below hypostome M Mesoglea Mv Microvilli Mi Mitochondria My Myofibrils Ne Nematocysts No Nucleolus N Nucleus A Outer acellular layer PB Protein body RER Rough endoplasmic reticulum S Secretory cell	Fl	Flagellum
G Golgi HG Heterogeneous granules Ia Intercellular attachments I Interstitial cell R Low ridge of cells below hypostome M Mesoglea Mv Microvilli Mi Mitochondria My Myofibrils Ne Nematocysts No Nucleolus N Nucleus A Outer acellular layer PB Protein body RER Rough endoplasmic reticulum S Secretory cell	G	Gastroderm
HG Heterogeneous granules Ia Intercellular attachments I Interstitial cell R Low ridge of cells below hypostome M Mesoglea Mv Microvilli Mi Mitochondria My Myofibrils Ne Nematocysts No Nucleolus N Nucleus A Outer acellular layer PB Protein body RER Rough endoplasmic reticulum S Secretory cell	GI	Gastrodermal inclusion
Ia Intercellular attachments I Interstitial cell R Low ridge of cells below hypostome M Mesoglea Mv Microvilli Mi Mitochondria My Myofibrils Ne Nematocysts No Nucleolus N Nucleolus A Outer acellular layer PB Protein body RER Rough endoplasmic reticulum S Secretory cell	G	Golgi
I Interstitial cell R Low ridge of cells below hypostome M Mesoglea Mv Microvilli Mi Mitochondria My Myofibrils Ne Nematocysts No Nucleolus N Nucleolus A Outer acellular layer PB Protein body RER Rough endoplasmic reticulum S Secretory cell	HG	Heterogeneous granules
R Low ridge of cells below hypostome M Mesoglea Mv Microvilli Mi Mitochondria My Myofibrils Ne Nematocysts No Nucleolus N Nucleolus A Outer acellular layer PB Protein body RER Rough endoplasmic reticulum S Secretory cell	Ia	Intercellular attachments
M Mesoglea Mv Microvilli Mi Mitochondria My Myofibrils Ne Nematocysts No Nucleolus N Nucleolus A Outer acellular layer PB Protein body RER Rough endoplasmic reticulum S Secretory cell	I	Interstitial cell
Mv Microvilli Mi Mitochondria My Myofibrils Ne Nematocysts No Nucleolus N Nucleus A Outer acellular layer PB Protein body RER Rough endoplasmic reticulum S Secretory cell	R	Low ridge of cells below hypostome
Mi Mitochondria My Myofibrils Ne Nematocysts No Nucleolus N Nucleus A Outer acellular layer PB Protein body RER Rough endoplasmic reticulum S Secretory cell	M	Mesoglea
My Myofibrils Ne Nematocysts No Nucleolus N Nucleus A Outer acellular layer PB Protein body RER Rough endoplasmic reticulum S Secretory cell	Mv	Microvilli
Ne Nematocysts No Nucleolus N Nucleus A Outer acellular layer PB Protein body RER Rough endoplasmic reticulum S Secretory cell	Mi	Mitochondria
No Nucleolus N Nucleus A Outer acellular layer PB Protein body RER Rough endoplasmic reticulum S Secretory cell	My	Myofibrils
 Nucleus A Outer acellular layer PB Protein body RER Rough endoplasmic reticulum S Secretory cell 	Ne	Nematocysts
A Outer acellular layer PB Protein body RER Rough endoplasmic reticulum S Secretory cell	No	Nucleolus
PB Protein body RER Rough endoplasmic reticulum S Secretory cell	N	Nucleus
RER Rough endoplasmic reticulum S Secretory cell	A	Outer acellular layer
S Secretory cell	PB	Protein body
•	RER	Rough endoplasmic reticulum
m . 1 1 1	S	Secretory cell
TB Tentacle bud	TB	Tentacle bud
X Undetermined cell type	X	Undetermined cell type

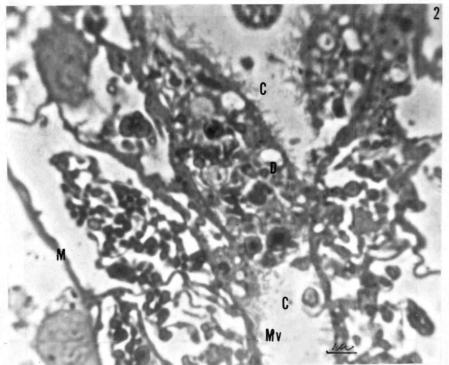
Vacuole

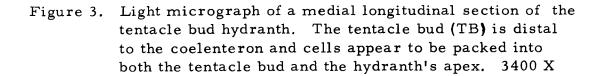
Zymogen-like granule

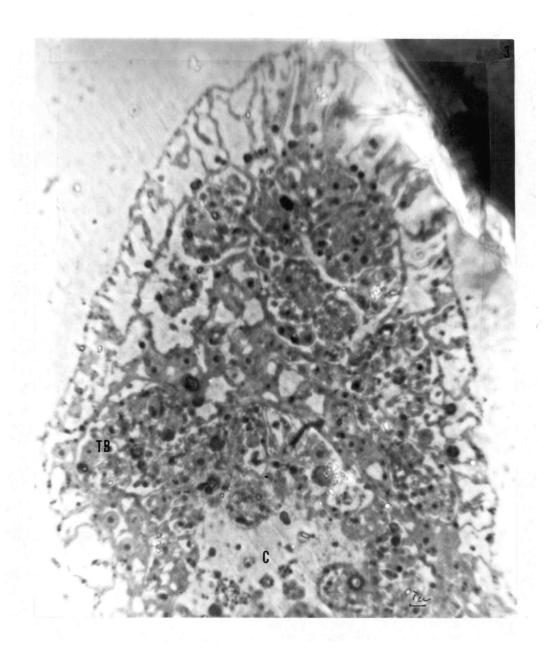
V

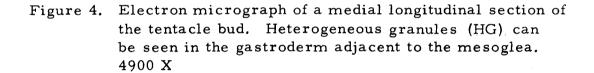
 \mathbf{Z}

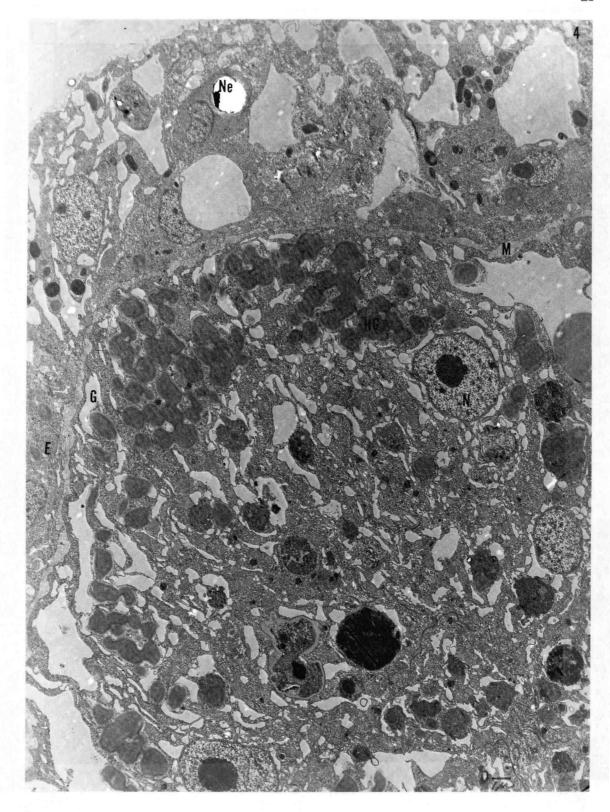












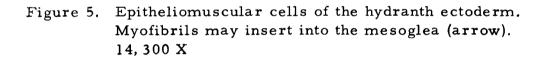
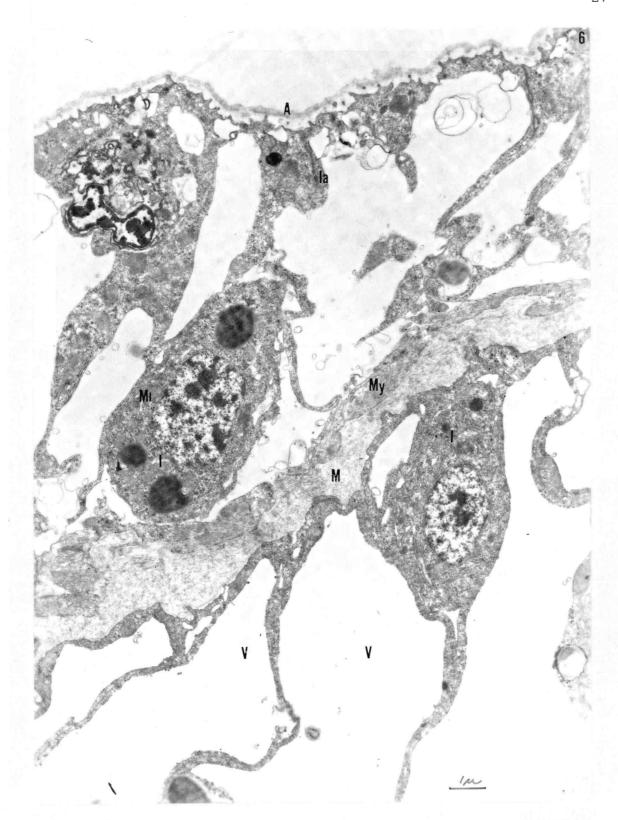
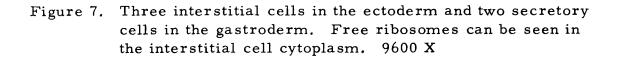
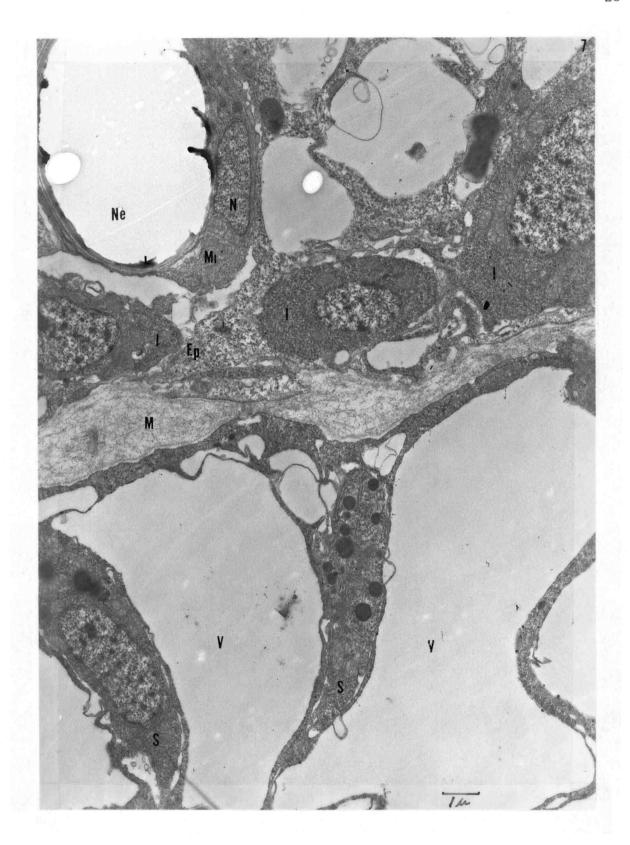


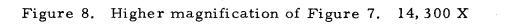


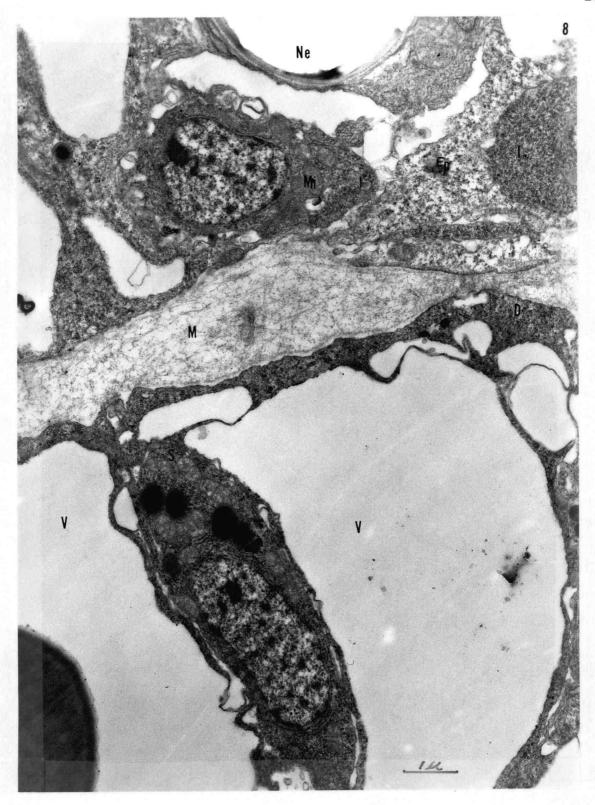
Figure 6. Interstitial cells of the hydranth ectoderm and gastroderm which are adjacent to the mesoglea and surrounded by the processes of adjacent cells. 9600 X

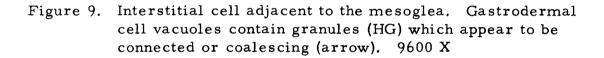












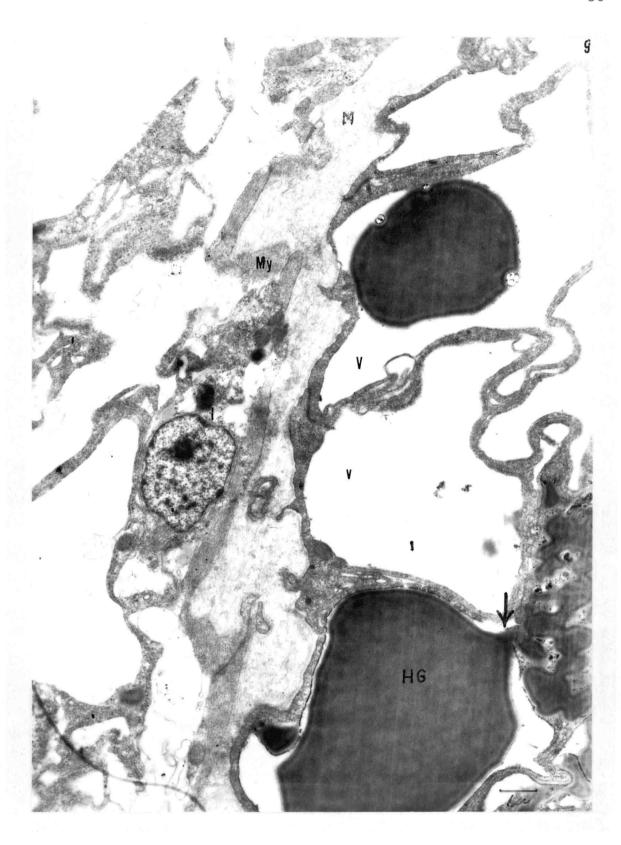
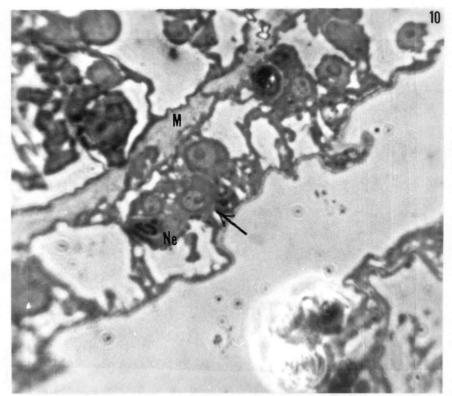
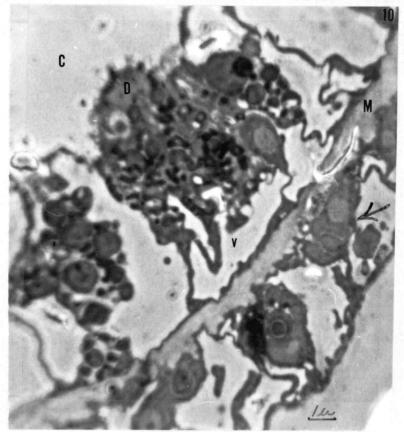
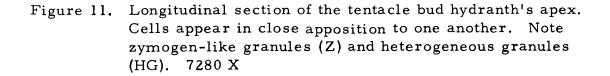
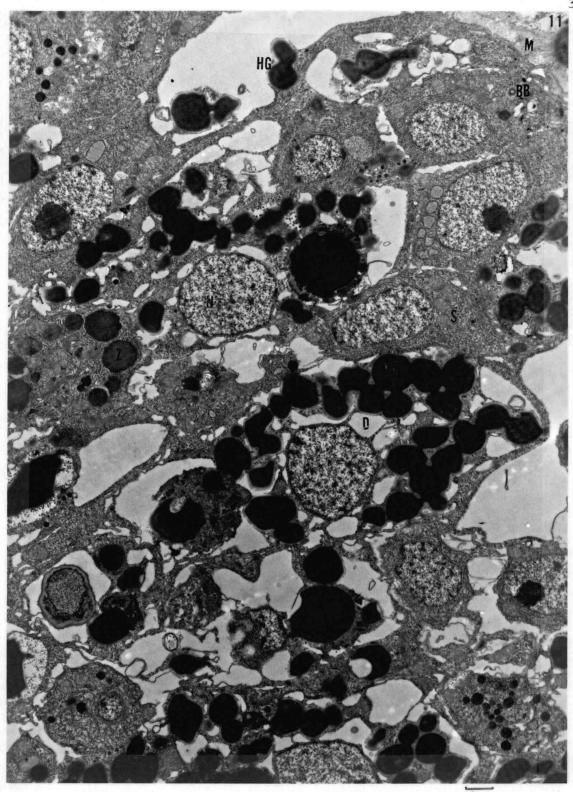


Figure 10. Light micrographs of cross sections of an adult hydranth close to a tentacle-hydranth junction showing interstitial cell clusters (arrows). Nematocysts appear as black spots in the clusters. Both micrographs 8700 X

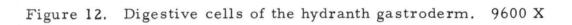


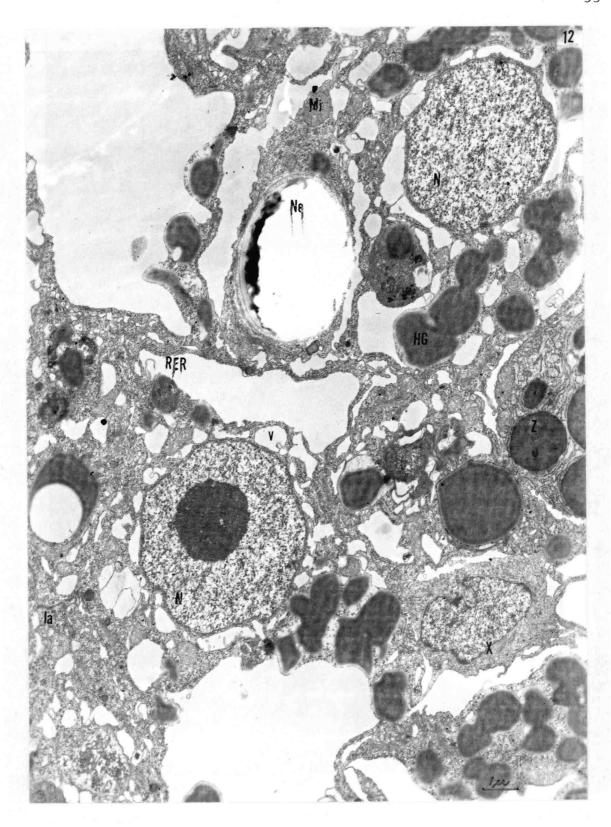


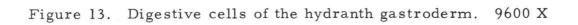


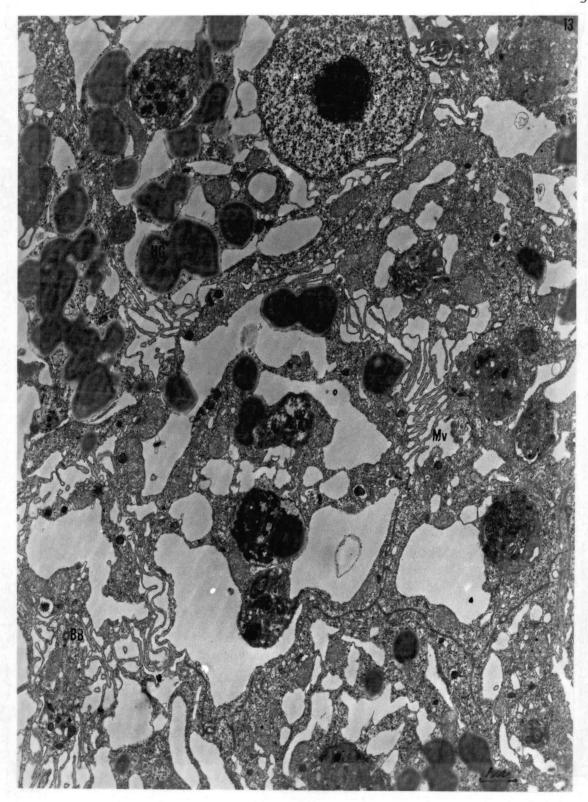


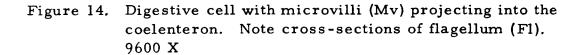
IM



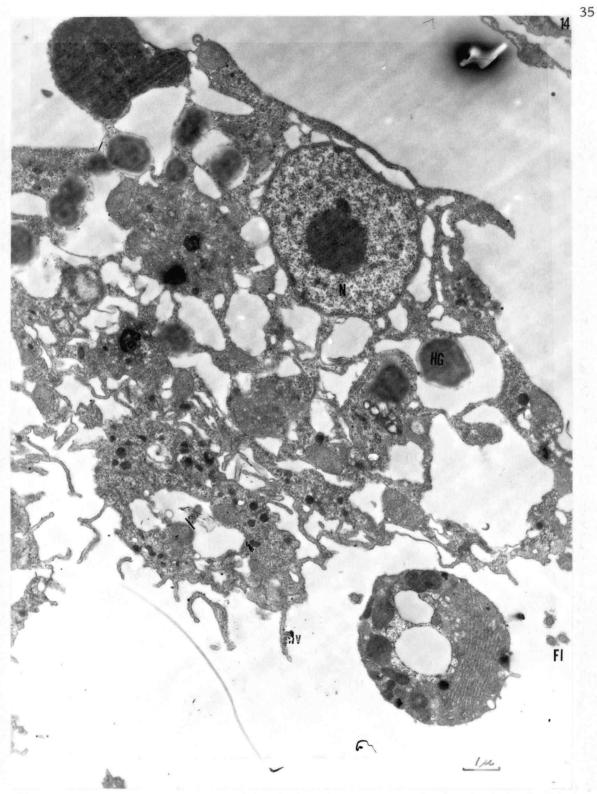


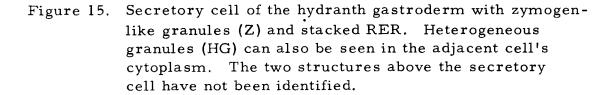












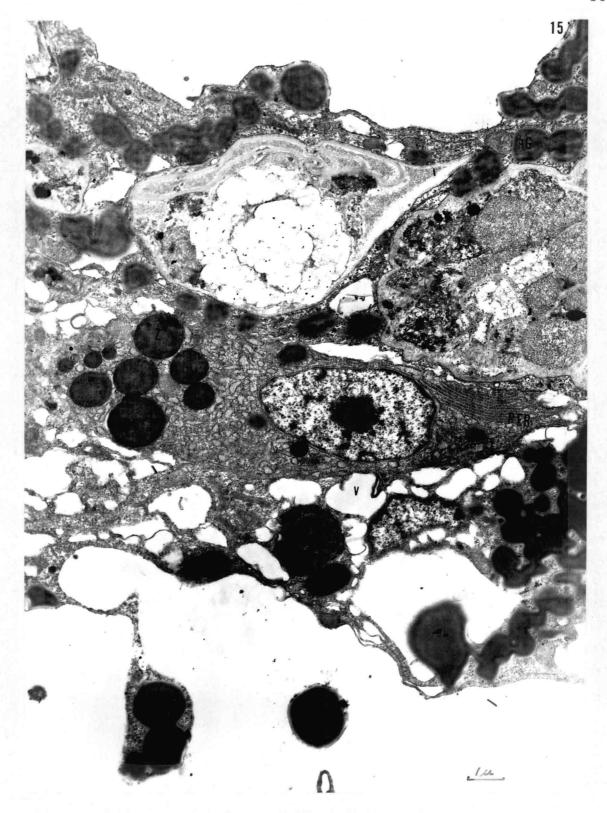
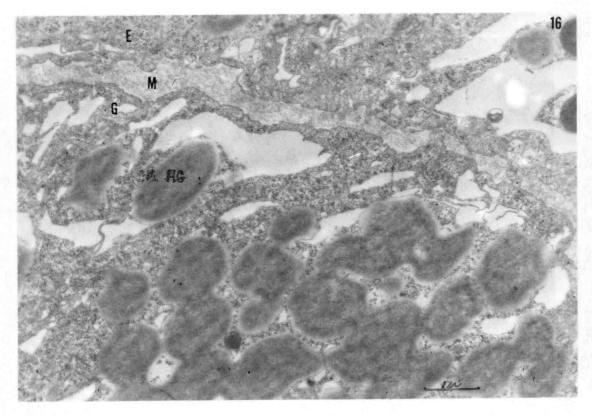
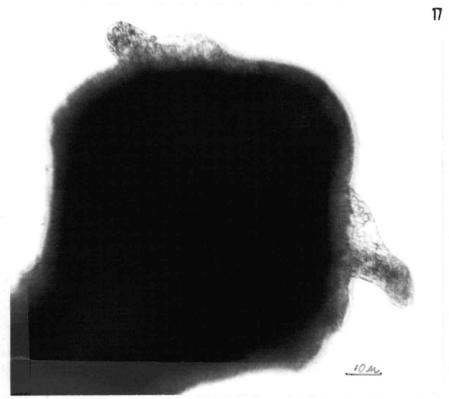
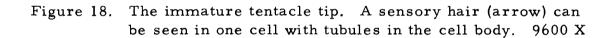


Figure 16. Tentacle bud gastroderm showing the heterogeneous granules (HG) adjacent to the mesoglea. 14,300 X

Figure 17. Stage two hydranth showing the enlarged ridge of cells below the hypostome and the small immature tentacles. 900 X







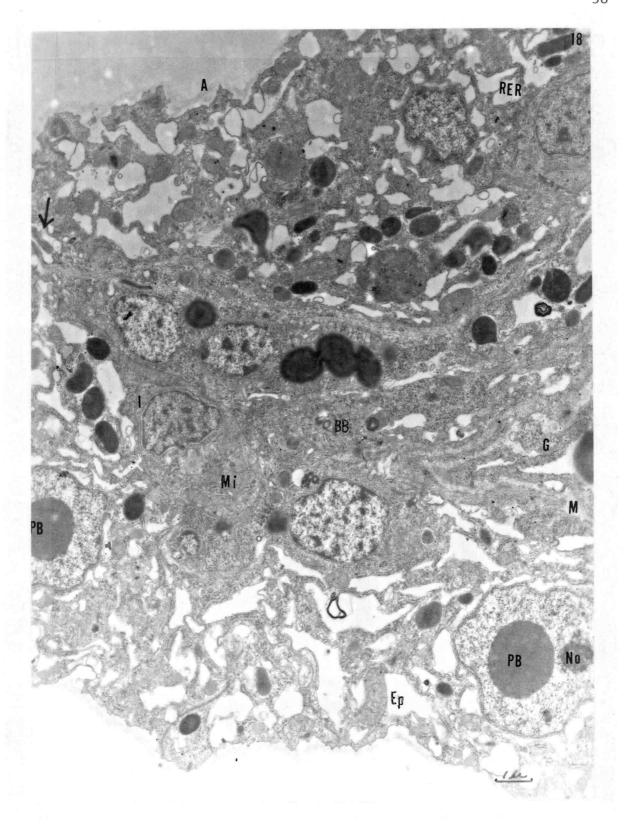
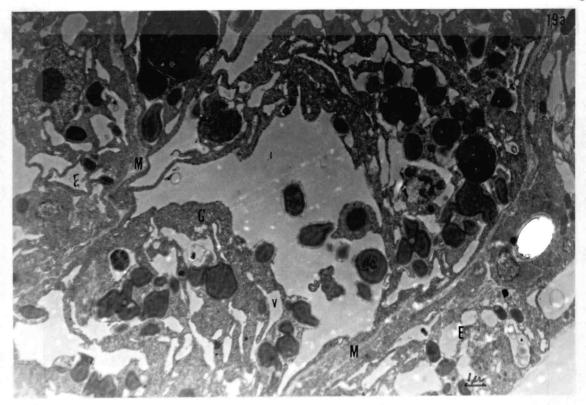
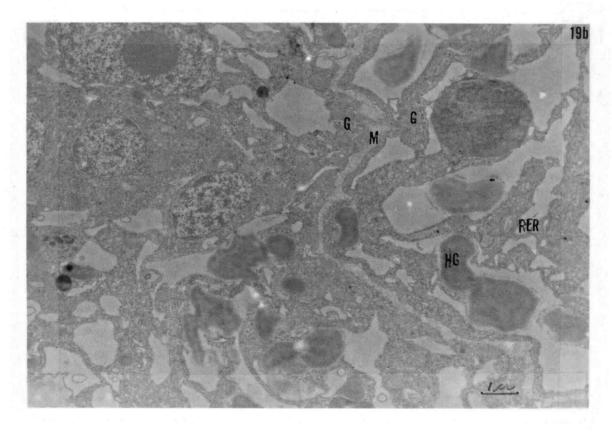
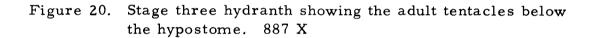


Figure 19a. A median section of the immature tentacle showing the confluent gastroderm between the hydranth and the tentacle. $4900~\mathrm{X}$

Figure 19b. A sagittal section of the immature tentacle showing the mesoglea separating the hydranth and tentacle gastroderm. $9600~\mathrm{X}$







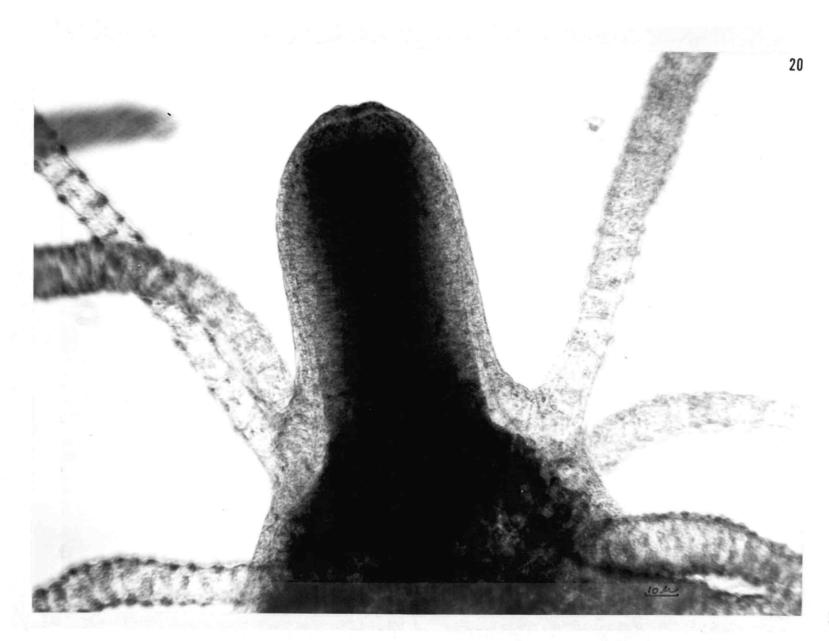
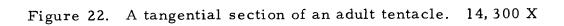
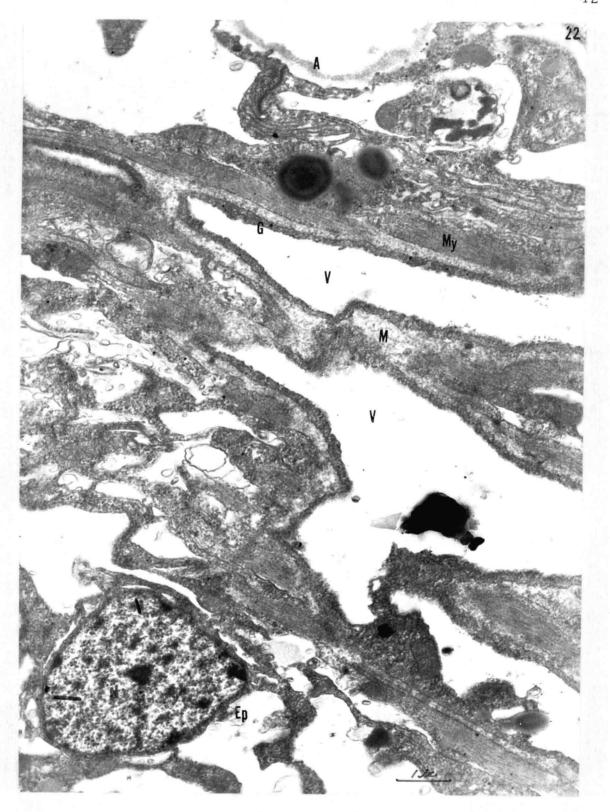


Figure 21. A montage of three electron micrographs of medial longitudinal sections of a tentacle in an advanced state of maturation. The black lines are folds in the sections. 2900 X







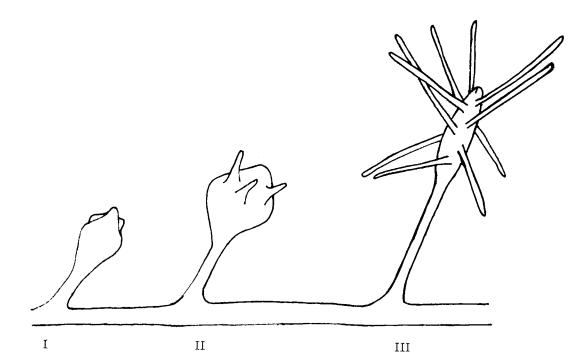


Figure 23. Sketch of the three stages of tentacle development.

- I. The tentacle bud.
- II. The immature tentacle.
- III. The adult tentacle.